The Magpie manual

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Contents

1 Introduction

Magpie is a program that simulates an NMR spectrometer environment. It can load sample and pulse sequences, and simulate the outcome of NMR measurements. The goal of the program is to be used in teaching, allowing students a first introduction to practical NMR. The basis for the simulation back-end is the classical description of magnetic resonance based on the Bloch equations including relaxation. This allows already a wide variety of NMR effects to be studied. As no quantum effects are taken into account, coherence transfer experiments and the like are not supported.

2 Running Magpie

2.1   Python and library versions

Magpie should run on python versions starting from 3.8. For the library version, the following are needed:

- `numpy` $>= 1.23.1$

- `matplotlib` $>= 3.5.2$

- `scipy` $>= 1.8.1$

- `PyQt5` $>= 5.15.7$

Older library and python version might also work, but have not been tested.

2.2   Installing

2.2.1   Linux

On Linux, Magpie can be most efficiently run using the python libraries that are in the repositories. For Ubuntu, these can be installed by running:

```
sudo apt install python3 python3-numpy python3-matplotlib python3-scipy
python3-pyqt5
```

Navigating to the `magpie/src` directory in a terminal, Magpie can then be run by executing `python3 magpie.py`.

2.2.2   Windows

On Windows, the relevant python libraries can be installed by installing `anaconda`: https://www.anaconda.com/download/. If you do not have another python version installed already, make sure to add anaconda to the `path` during the install. In this case, Magpie can be run by executing the `WindowsRun.bat` file in the `magpie/src` directory.

   If you already have other version of python installed, adding anaconda to the `path` might create issues. Do not do this in this case. When not added to the path, the `WindowsRun.bat` should be edited in such a way that `pythonw` is replaced with the path to your `pythonw.exe` executable in the anaconda install directory.

2.2.3   OS X

On OS X, the relevant python libraries can be installed by installing `anaconda`: https://www.anaconda.com/download/. Navigating to the `magpie/src` directory in a terminal, Magpie can then be run by `anacondapython magpie.py`, with `anacondapython` replaced by the path to your anaconda python executable.
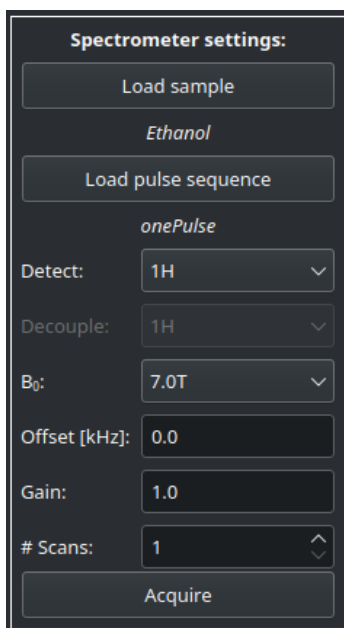
3   Interface

The Magpie interface looks like this:
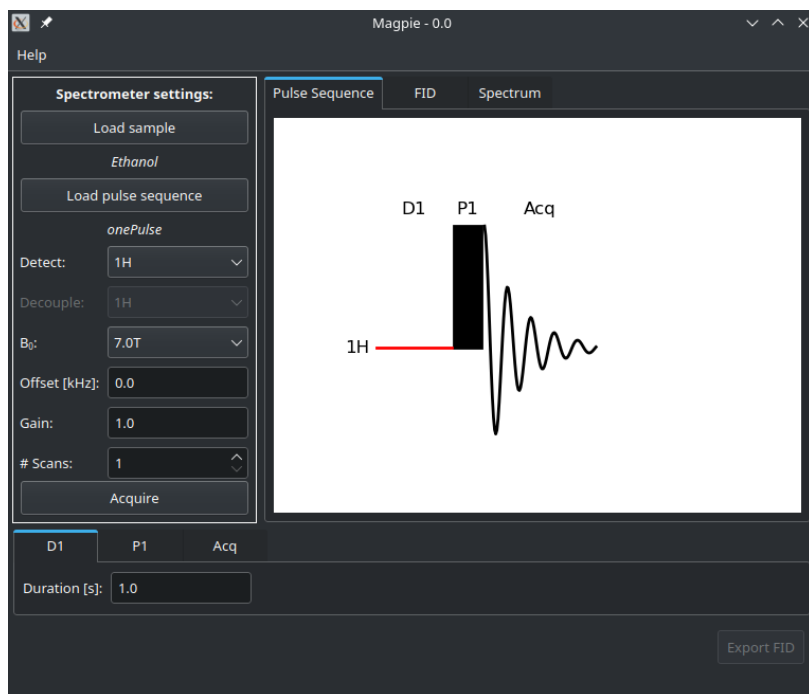
In the sections below, the individual elements are explained.

3.1   Spectrometer settings

In the left of the Magpie screen, the spectrometer settings can be put in. These are all settings that are not specific to a pulse sequence. Also, the buttons to load a sample and a pulse sequence

are contained here. Below is a list of the settings and their description.

- **Load sample:** Opens a file dialog to brows for a sample file. More info can be found in section 4. The file should have extension `.txt`.

- **Load pulse sequence:** Opens a file dialog to brows for a pulse sequence file file. More info can be found in section 5. The file should have extension `.csv`.

- **Detect:** The nucleus to detect. In principle, any nucleus can be detected. For the moment, we limit the selection to ^{1}H and ^{13}C though. This is a design choice to not overflow the user with obscure nuclei.

- **Decouple:** The nucleus that is decoupled during a decoupling pulse sequence element. Only active if the pulse sequence has a decouple element.

- **B$_0$:** The main magnetic field strength in Tesla. Magpie supports any magnetic field strength. For now, we decide on using a dropdown box with only common fields. This is the teach the user that in reality the magnetic field cannot easily be changed to whatever we want.

- **Offset [kHz]:** The transmitter offset in kHz. This can be used to fine-tune the center frequency of the experiment. It defines the center of the spectrum, as well as the frequecy on which any pulses are given.

- **Gain:** The gain factor of the receiver. All signals are multiplied by this value before detection. Note that, contrary to most real-life spectrometers, Magpie uses a linear scale, and does not work in dB.

- **# Scans:** The number of individual scans that need to be recorded. All scans are automatically summed.
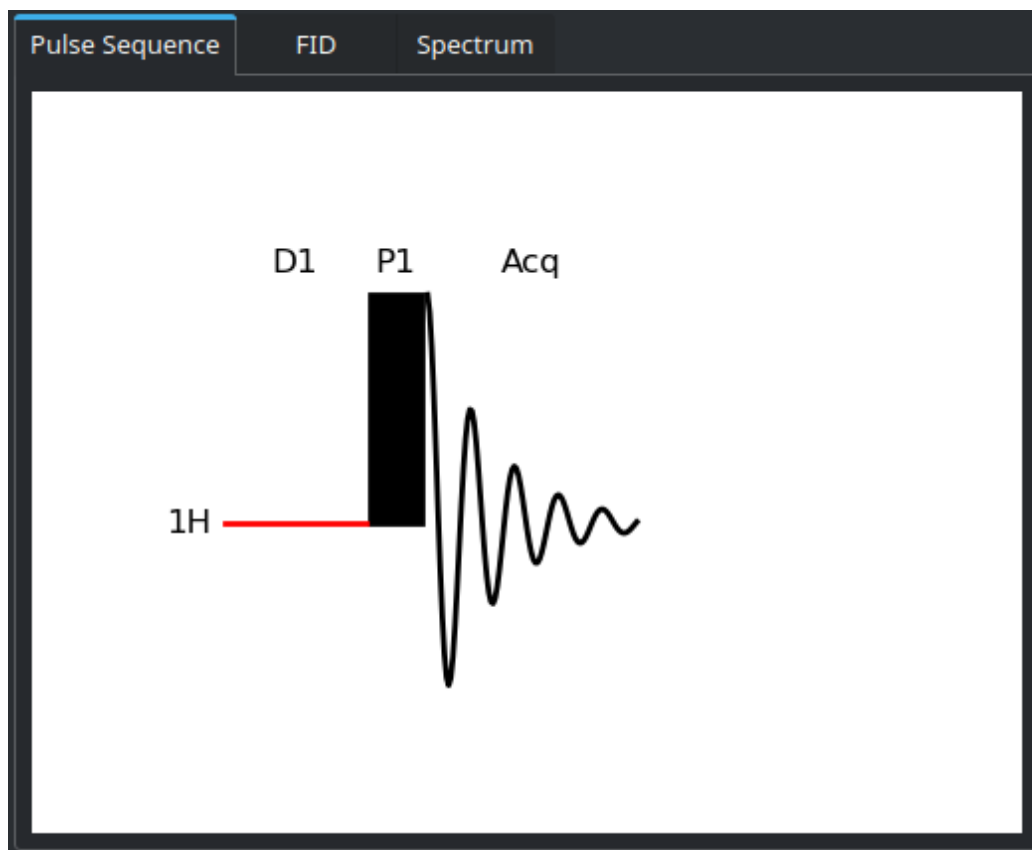
3.2   Pulse sequence settings
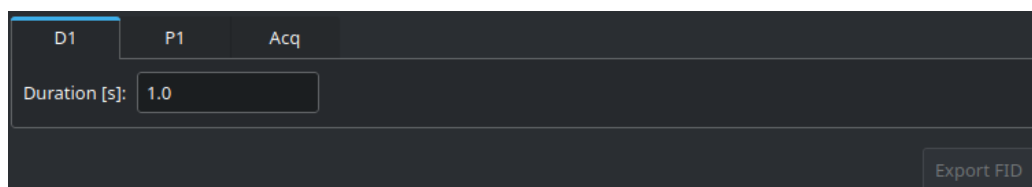
In the bottom of the Magpie interface, the pulse sequence settings are defined. Each tab represent a separate element of the pulse sequence. The names of the tabs are equal to the names of the elements, as defined in the pulse sequence file. The pulse sequence diagram (in the plot window) always highlights the element which settings tab is currently showing. More info on pulse sequences, and the relevant settings per element can be found in section 5.

3.3   Plot windows

The plot window contains three tabs: Pulse sequence, FID, and Spectrum. These are explained below.

3.3.1   Pulse sequence

The pulse sequence tab displays a diagram of the current pulse sequence. The current element that is selected is displayed in red. Left-clicking on an element changes the selection. The sequence settings tab (see subsection 3.2) folows this selection (and vice-versa).

The labels displayed above the sequence elements are those as defined in the pulse sequence file. The tabs in the sequence settings window also have this name.

The elements in the pulse sequence each have a unique pictorial representation. The style of the elements is not updated depending on the pulse sequence settings (e.g. a pulse with amplitude 0 is not shown as a much lower rectangle).

3.3.2   FID
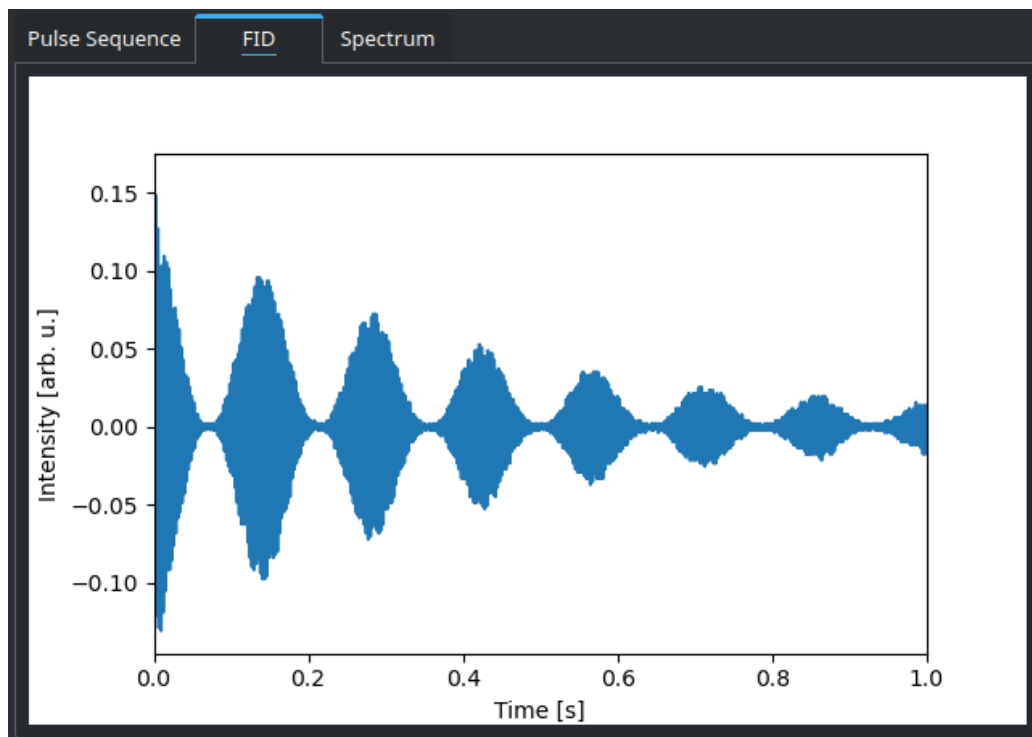
The FID tab shows the FID (time signal) of the simulated data. If there is no data, no plot is shown. The y-axis has the signal intensity. Although it is in arbitrary units, the maximum value is 1. Any signals high than that will be rounded to 1. For negative intensity the limit is -1. These situations relate the 'receiver overflow' in real spectrometers, that happens when signals are to strong to properly digitize.

The plot supports the following mouse-based zoom-controls:

- Dragging a box while holding the left mouse button creates a zoombox, and will zoom to that region.

- Dragging while holding the right mouse button drags (i.e. pans) the spectrum. Doing this while holding the Control button pans only the x-axis, holding Shift pans only the y-axis.

- Double clicking with the right mouse button resets the view to fit the whole plot (both x and y).

- Scrolling with the mouse wheel zooms the y-axis. Doing this while also holding the right mouse button zooms the x-axis. By holding Control the zooming will use a larger step size.

3.3.3 Spectrum
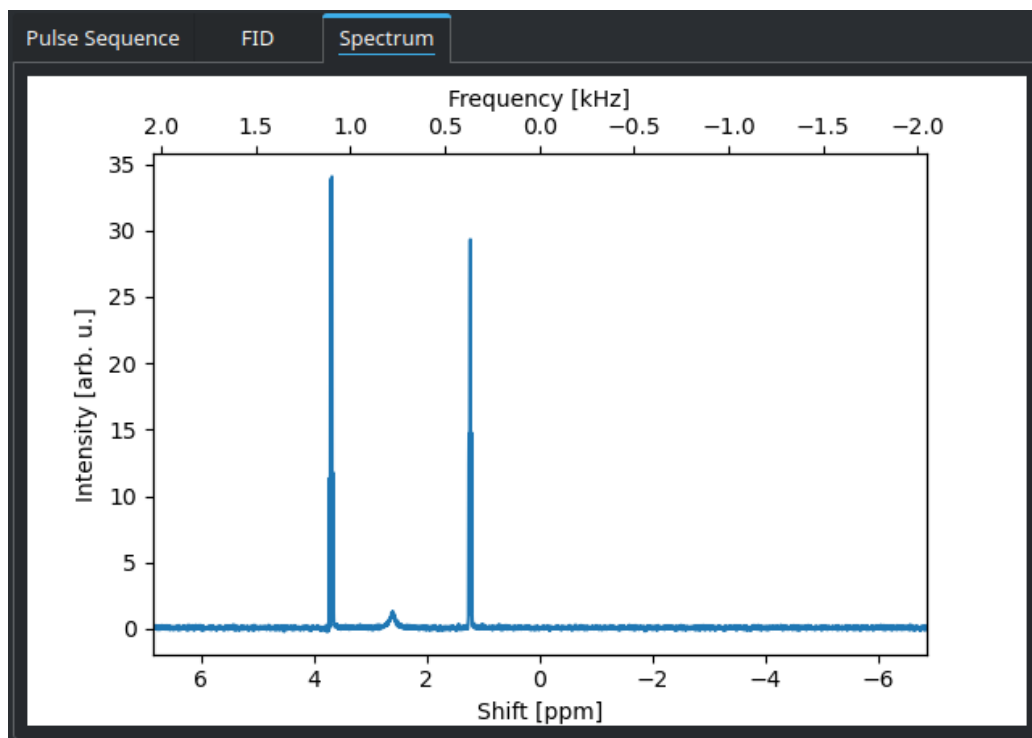
The Spectrum tab shows the signal after Fourier transformation. Two x-axes are plots:

- Bottom x-axis: Has the axis in ppm. Here, 0 ppm relates to the signal of the ppm reference for the measured nucleus. If the spectrometer offset is changes, the 0 ppm position can shift in the spectrum.

- Top x-axis: Has the axis in kHz. For this axis, 0 kHz is always defined in the center of the spectrum. When adjusting the spectrometer offset, this axis can be used to find the desired shift.

The spectrum plots has the same zoom controls as the FID plot (see subsubsection 3.3.2).

4 Sample definition

Sample files for Magpie need to be in a specific definition. They need to be text files with `.txt` extension.

The file should start with a header:

`###SAMPLE###`

Below this, on optional statement regarding the sample amount (concentration) can be made:

`amount 1`

In this case, the scaling is set to 1, so it has no effect.

After this, a molecule can be defined. The sample file can hold multiple molecule definitions. Within this block, the spins are set, as well as the overall relaxation times and the J-couplings.

The statements are:

Statement	Input	Description
`amount`	amount	molecule amount
T1	T1	Overall T1, in seconds (optional)
T2	T2	Overall T2, in seconds (optional)
T2prime	T2prime	Overall T2prime, in seconds (optional)
`spin`	isotope shift multiplicity	For example: `1H 1.2 3`
spin	isotope shift multiplicity T1 T2 T2prime	Spin definition including relation times. These are used in favour of the overall relaxation times, if set. Values can be set empty by an underscore (_)
J	spin1 spin2 strength	Set J-coupling in Hz between two spin indexes. Number order as the spins are defined. For example: `J 1 2 9.6`
`Jmatrix`	matrix	Set J-coupling in Hz between all spins in matrix format. Example for two spins: `[[0,10],[10,0]]`, setting 10 Hz coupling. Size needs to be n_spins x n_spins. When used the, regular J statement cannot be used.
pair	isotope shift1 shift2 k amp1 amp2 T1_1 T1_2 T2_1 T2_2	Sets a spin-pair with exchange between them. amp sets the multiplicity. Individual T1 and T2 times can be set, or set to empty by an underscore (_). No J-couplings can be set to these spins.

Either `Jmatrix` or J statements are allowed. Not both. Spin statements can lack all the relaxation times, or have a _ instead. If a relax time not defined for a spin, it needs to use the global molecule relaxation time. These are also optional, but if they lack, the individual spin lifetimes need to be all there.

An example definition could be the following sample:

```
###SAMPLE###
amount 1
```

```
###MOLECULE###
amount 0.1
T1 5
T2 0.5
T2prime 10
spin 1H 1.2 3
spin 1H 3.6 2
J 1 2 7
```

Here we set a two spin system, with shifts 1.2 and 3.6 ppm, multiplicities 3 and 2. There is a 7 Hz J-coupling between the spins. All nuclei have the same relaxation times, set at 5, 0.5 and 10 s for T1, T2 and T2prime respectively. The overall intensity scaling is set at 1, and the molecule itself has scaling 0.1.

A more complicated sample could be ethanol:

```
###SAMPLE###
amount 1
###MOLECULE###
amount 0.98
T1 5
T2 0.5
T2prime 10
spin 1H 1.226 3
spin 1H 3.692 2
spin 1H 2.605 1 _ 0.01 0.1
J 1 2 7
###MOLECULE###
amount 0.01
T1 5
T2 1
T2prime 10
spin 1H 1.224 3
spin 1H 3.692 2
spin 1H 2.605 1 _ 0.01 0.1
spin 13C 18.1 1 _ _ 7
J 1 2 7
J 1 4 125
###MOLECULE###
amount 0.01
T1 7
T2 1
```

```
T2prime 10
spin 1H 1.226 3
spin 1H 3.692 2
spin 1H 2.605 1 _ 0.01 0.1
spin 13C 57.8 1 _ _ 7
J 1 2 7
J 2 4 125
```

Here we set three molecules: with no 13C nucleus, and both options with a single 13C nucleus (the 13C-13C variant is too low in intensity to care about). The amount of the molecules is set to follow the natural abundance. For the OH peak (at 2.6 ppm), a separate set of relaxation times is set, as to have a shorter T2 and T2prime, to include the fact that this signal is usually exchanging causing line broadening. J-couplings are ste between the 1H nuclei of the CH_3 and CH_2 groups, as well as between the 13C nuclei and their directly bonded 1H nuclei.

5   Pulse sequence definition

Magpie reads pulse sequence in comma separated format (`.csv`). Each sequence element is on its own row. Default settings for its parameters are given in columns with the correct column name. For each sequence element, magpie only looks for the columns needed for that element. Other columns can be left empty, if a specific sequence element does not need those.

Below, the currently supported sequence elements are listed, along with the required columns.

Element type	Required columns	Description
delay	time	Set a delay during which nothing is done.
pulse	time, amp, phase	Defines a pulse with amplitude in kHz and a phase in degrees. Phase can also be a list (e.g. [0,90]) to define a phase cycle.
FID	time, amp	Measure the FID, with `time` the acquisition time in seconds, and `amp` the amount of points. The receiver phase is always 0, and cannot be phase cycled.
sat		Defines a saturation element. Sets all magnetization to 0. Has no further settings.
shapedPulse	time, amp, phase, shapeFile	Defines a pulse with an amplitude shape. Has the same definitons as the `pulse` element. Additional is the `shapeFile` column, which should point to a shape file. Can be just a name if the shape file is one of the default shapes. See more in subsection 5.1.

The supplied `.csv` file should contain several elements as described above. The element type should be supplied, as well as a name of the element. The name is used in the Magpie interface to identify the element in the pulse sequence visualization, as well as in the settings tabs.

An example for a one pulse sequence is given below:

```
name,type,time,amp,phase
D1,delay,1
P1,pulse,2.5e-6,100e3,-90
Acq,FID,1,4096
```

The first column supplies the name of each element, the second the type. Other columns are used to define the starting values of required inputs. The elements defined in this sequence are:

- A delay with name 'D1' and default time of 1 s.

- A pulse with name 'P1' with time 2.5 µs, amplitude of 100 kHz and pulse phase $-90°$.

- An FID detection block called 'Acq' with time of 1 s, and 4096 points.

The resulting sequence is the one displayed in subsubsection 3.3.1.

An echo sequence can be defined as:

```
name,type,time,amp,phase
D1,delay,1,,
P1,pulse,2.5E-06,100000,"[90,270,90,270]"
t1,delay,0.1,,
P2,pulse,5E-06,100000,"[90,0,270,180]"
t2,delay,0.1,,
Acq,FID,1,4096,
```

Here, we have two pulses and two extra delays. The pulses have a 4 step phase cycle. Note that phase cycles always need to be the same length for all pulse element in the sequence. The phases are defined with respect to the receiver phase, which is always 0, and cannot be phase cycled.

5.1 Shape files

Shape files are defined as a text file with a list of float values. Every entry is on a separate row. Each value indicates the relative pulse amplitude at that point. The phase cannot be changed at the moment.

Note that the overall maximum amplitude of the pulse is equal to $\text{amp}_{\text{pulse}} \cdot \text{amp}_{\text{shape}}$. It is recommended therefore to set the maximum amplitude in the shape file to 1, to avoid confusion.

When using the shape, Magpie treats it as a series of pulses. The length of each sub-pulse is set by the total pulse length divided by the number of steps defined in the shape file. Higher resolution pulses can therefore be obtained by supplying a higher resolution shape file. Note that the simulation time will increases by doing this.

6   Data output

Magpie is designed to interface with the `ssNake` program.[1] The default output is a `.mat` file in the same style as `ssNake` saves. The button 'Export FID' can be used to save the current dataset.

7   Simulation background

Magpie uses a classical, time-domain simulator (i.e., Bloch based), so no quantum effects are included. J-couplings are therefore included only as their weak coupling limit, and no coherence between the J-states exists.

Firstly, the sample file is processed, and divided in individual spins. J-coupled states are treated as separate spins, with a different Larmor frequency. For example, a spin which couples to a single ^{1}H nucleus with a coupling constant of 10 Hz is subdivided in two spins with +5 and -5 Hz Larmor frequency and half the magnitude.

After this is done, we can define each 'spin' with 6 properties:

$$
S = \begin{pmatrix} M_x \\ M_y \\ M_z \\ M_0 \\ T_1 \\ T_2 \end{pmatrix} \quad . \tag{1}
$$

At the start of the pulse sequence, the spins start with $M_x == M_y == 0$. The total magnetization in thermal equilibrium, due to the amount of spins of a particular site in a molecule, and the amount of the molecule in the sample, is available in the M_0 value. This thermal equilibrium magnetization is used to calculate the T_1 decay.

With the information per spin, the system changes over time can be calculated using the Bloch equations. This is done by using a ODE solver, for each spin individually per block in the pulse sequence. For blocks where no RF is applied (i.e., during delays or the FID), each spin in propagated in a reference frame rotating at its Larmor frequency, to improve the speed of the ODE solver. In that case, the M_x and M_y can be determined from the rotating frame values by a time-dependent phase shift.

The final z magnetization (M_Z) at the end of a scan will be used as the initial state for the next scan. In this way, system history is taken into account, and a quantitative experiment needs a recovery time of 3-5 times T_1 between scans, equalling reality.

For each element in the pulse sequence, the initial state of the spin is taken, and the final a the end of the sequence element is calculated. Only for the FID element, more intermediate values are calculated, in line with the sampling frequency that is specified in the experimental settings.

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[1] https://www.ru.nl/science/magneticresonance/software/ssnake/

7.1   T2prime

T2prime effects are a collective property relate to the measurement setup of an NMR experiment and the sample being measured. While T_2 represent the unrecoverable loss of coherence of a state, T2prim effects describe the loss of coherence due to macroscopic effects. This relates for example to a sample tube in an inhomogeneous magnetic field. Signals will decay faster than the true T_2, as the XY-magnetization of spins in different locations in the tube will precess at a different rate. Using a spin echo, for example, the full alignment can be recovered, making it a different process than T_2.

Magpie can include T2prime effects in its calculations. It does this by creating a series of duplicate spins, at slightly different Larmor frequency. The sum of all these lines in the spectrum follows a Lorentzian shape, and can be seen as the magnetic field profile over the sample. The width of the envelope is calculated based on the supplied T2prime for each spin. The amount of sub-sampled spins is defined by the T2prime value, and the T_2 value of the original spin.

7.2   Exchange

Magpie also supports simulation of two-site exchange. Exchanging spin pairs can be defined in the sample file as described in section 4. To include exchange in the Block equations, both spins are simulated simultaneously with an additional exchange rate parameter k. One thing to note is that for exchanging spin pairs there is only one rotating frame, which means the speedup is limited. So for rapidly exchanging spins which are separated in frequency, the simulation times can start to increase noticeably.

8   Contact

To contact the `magpie` team write to `ssnake@science.ru.nl`.

Bibliography