# Ldpred2

Bayesian approach to computing polygenic risk scores

Computing Sources: Google Cloud Platform(GCP), Nipa Ubuntu Server(고성능 컴퓨팅 서비스), Texas Advanced Computing Center(TACC)

## Google Cloud Platform

Session logged out too quickly

## Nipa Ubuntu Server

- Extremely slow for finding correlation on one chromosome on R
- Downloaded R studio but could not access it through https://localhost:8787 (Still do not know why)

## **Texas Advanced Computing Center**

A very nice stampede guideline: <a href="https://portal.tacc.utexas.edu/user-guides/stampede2">https://portal.tacc.utexas.edu/user-guides/stampede2</a>

### Accessing TACC

- Create an account on Xsede site and connect it with duo mobile application
- Through terminal, log into xsede site by typing ssh -l your\_xsede\_id login.xsede.org and then your password
- Then, type gsissh stampede2 to go into TACC

#### File location

#### Your directory

ABCD.bed	chr12.OMNI.interpolated_genetic_map
ABCD.bim	chr13.OMNI.interpolated_genetic_map
ABCD.bk	chr14.OMNI.interpolated_genetic_map
ABCD.fam	chr15.OMNI.interpolated_genetic_map
ABCD.rds	chr16.OMNI.interpolated_genetic_map
ABCD.valid.sample	chr17.OMNI.interpolated_genetic_map
ABCD_QCed.bed	chr18.OMNI.interpolated_genetic_map
ABCD_QCed.bim	chr19.OMNI.interpolated_genetic_map
ABCD_QCed.bk	chr2.OMNI.interpolated_genetic_map
ABCD_QCed.fam	chr20.OMNI.interpolated_genetic_map
ABCD_QCed.het	chr21.OMNI.interpolated_genetic_map
ABCD_QCed.log	chr22.OMNI.interpolated_genetic_map
ABCD_QCed.nosex	chr3.OMNI.interpolated_genetic_map

```
ABCD_QCed.prune.in chr4.OMNI.interpolated_genetic_map
ABCD_QCed.prune.out chr5.OMNI.interpolated_genetic_map
ABCD_QCed.rel.id chr6.OMNI.interpolated_genetic_map
ABCD_QCed.snplist chr7.OMNI.interpolated_genetic_map
chr1.OMNI.interpolated_genetic_map chr8.OMNI.interpolated_genetic_map
chr10.OMNI.interpolated_genetic_map chr9.OMNI.interpolated_genetic_map
chr11.OMNI.interpolated_genetic_map pca
```

- GWAS summary stat file: directory private!
- > It should look like this...

```
CNCR AD
                PGC EATING.txt
CNCR ANTISOCIAL.txt PGC OCD
CNCR DEPRESSION.txt PGC SCZ
CNCR_DEPRESSION_SUB.txt PGC_UKB_MDD
CNCR IQ.txt
                SSAGC_ASP.txt
CNCR_NEUROTICISM.txt SSAGC_DRINK.txt
CNCR WORRY SUB.txt SSAGC RISK4PC.txt
ETC INSOMNIA
               SSAGC RISKTOL MA.txt
                SSAGC_SMOKER_MA.txt
ETC PTSD EA
ETC_SNORING.txt UKB_AUDIT.txt
GWAS_CP_all.txt
                 UKB_BMI.txt
GWAS CP all Idpred.txt UKB CANNABIS.txt
GWAS_CP_all_ldpred2.txt UKB_GENERALHAPPINESS.txt
GWAS EA excl23andMe.txt UKB GENERALHAPPINESS HEALTH.txt
PGC_ADHD_EA
                  UKB_GENERALHAPPINESS_MEANINGFUL.txt
PGC_ASD
                UKB HAPPINESS.txt
PGC_BIP_2018
                 adas
PGC CROSS.txt
```

### R script for PRS computation (my case: bipolar disorder)

- Saved in directory private
- The code looks like this.. (Blue comments are for additional info)

```
#install.packages("dplyr")
library(bigsnpr)
options(bigstatsr.check.parallel.blas = FALSE) # For multi-thread

obj.bigSNP <- snp_attach("privaate_______ABCD_QCed.rds")
str(obj.bigSNP, max.level = 2, strict.width = "cut")

G <- obj.bigSNP$genotypes
CHR <- obj.bigSNP$map$chromosome
POS <- obj.bigSNP$map$physical.pos
y <- obj.bigSNP$fam$affection - 1

sumstats <- bigreadr::fread2("/private directory..../ABCD_summarystats/PGC_BIP_2018")
#If you wish to find PRS on other GWAS summarystats change PGC_BIP_2018 part
str(sumstats)</pre>
```

```
set.seed(1)
ind.val <- sample(nrow(G), 400)</pre>
ind.test <- setdiff(rows along(G), ind.val)</pre>
sumstats$beta <- log(sumstats$OR)</pre>
sumstats$n eff <- 4 / (1 / sumstats$Nca + 1 / sumstats$Nco)</pre>
sumstats$Nca <- sumstats$Nco <- NULL</pre>
sumstats$HetPVa <- sumstats$HetDf <- sumstats$Direction <-</pre>
sumstats\$FRQ A 20352 <- sumstats\$FRQ U 31358 <- NULL
names(sumstats) <- c("chr", "rsid", "pos", "a0", "a1", "beta se", "p", "beta", "n eff")
# check the format and contents of sumstats by str(sumstats)
map <- obj.bigSNP$map[-(2:3)]</pre>
names(map) <- c("chr", "pos", "a0", "a1")</pre>
info_snp <- snp_match(sumstats, map)</pre>
library (R.utils)
library(data.table)
library (magrittr)
POS2 <- snp asGeneticPos(CHR, POS, dir = "private ")
# Get maximum amount of cores
NCORES <- nb cores()
# Start doing analysis on each chromosome
fam.order <- as.data.table(obj.bigSNP$fam)</pre>
fam.order[, Inf.est := 0]
# add progress bar
pb = txtProgressBar(min = 0, max = 22, initial = 0)
#inf.model 로 chr 별 결과 각각 print
for (chr in 1:22) {
  setTxtProgressBar(pb, chr)
  # extract current chromosome
  chr.idx <- which(info snp$chr == chr)</pre>
  df_beta <- info_snp[chr.idx,</pre>
                      c("beta", "beta_se", "n_eff")]
  ind.chr <- info snp$` NUM ID `[chr.idx]</pre>
  # calculate LD
  corr0 <- snp cor(</pre>
    G,
   ind.col = ind.chr,
   ncores = NCORES,
    infos.pos = POS2[ind.chr],
    size = 3 / 1000
  corr <- bigsparser::as_SFBM(as(corr0, "dgCMatrix"))</pre>
  # Perform LDSC analysis to get h2 estimate
  ldsc <- snp ldsc2(corr0, df beta)</pre>
  h2 est <- ldsc[["h2"]]
  # Get adjusted beta from infinitesimal model
  beta_inf <- snp_ldpred2_inf(corr, df_beta, h2 = h2_est)</pre>
```

#### Job submission Script

- This script is saved in private directory.... as bipld2script.sh on TACC

```
#!/bin/bash
#_____
# Sample Slurm job script
# for TACC Stampede2 SKX nodes
#-----
#SBATCH -J bipld2  # Job name

#SBATCH -o bipld2.0%j  # Name of stdout output file

#SBATCH -e bipld2.e%j  # Name of stderr error file
#SBATCH -p skx-dev # Queue (partition) name / dev도 있고 skx-normal, long 같은 경우는
kn1
#SBATCH -N 4
                     # Total # of nodes (must be 1 for serial) > glimits 라고
커맨드에 치면 limits 볼 수 있음
#SBATCH -n 32
                     # Total # of mpi tasks (should be 1 for serial)
#SBATCH --mail-user=private_
#SBATCH --mail-type=all  # Send email at begin and end of job
# Other commands must follsw all #SBATCH directives...
private /ld2 #stdout/ err output file goes in! WORKSPACE
module load Rstats
pwd
date
# Launch serial code...
ibrun Rscript bipld2.R # dev 경우 ibrun 했고 아닌경우는 Rscript file명.R
# -----
```

- I wanted to run the R script with skx-large, but I couldn't for whatever reason. So I tried it with long. It uses KNL if queue is long so probably inaccurate partition name. This script is saved in ----- on TACC

```
login3.stampede2(678)$ cat bipld2large.sh
#!/bin/bash
```

```
# Sample Slurm job script
# for TACC Stampede2 SKX nodes
#-----
#SBATCH -J bipld2  # Job name
#SBATCH -o bipld2.o%j  # Name of stdout output file
#SBATCH -e bipld2.e%j  # Name of stderr error file
#SBATCH -p long # Queue (partition) name
                     # Total # of nodes (must be 1 for serial)
#SBATCH -N 1
#SBATCH -n 1
                         # Total # of mpi tasks (should be 1 for serial)
#SBATCH -t 24:00:00
                        # Run time (hh:mm:ss)
#SBATCH --mail-user=blahblah (private)
#SBATCH --mail-type=all # Send email at begin and end of job
# Other commands must follsw all #SBATCH directives...
cd ----- private
module load Rstats
pwd
date
# Launch serial code...
Rscript bipld2.R
```

#### How to run r script on TACC by job shell file

1. Send necessary files for job submission- my case: shell file(.sh) and your rscript

```
(base) Desktop % scp shell_file_name
tacc_id@stampede2.tacc.utexas.edu:any_directory_you_wish_to_sen
d file
```

 Then you will need to enter your tacc account password and tacc return token code(6 digits) from duo mobile

```
My case) (base) heewon@Heewonui-MacBookPro Desktop % scp
bipld2script.sh
    _____private_____@stampede2.tacc.utexas.edu:------priv
ate
```

2. Login xsede through terminal

```
ssh -l xsede_id login.xsede.org
My case: ssh -l private_ login.xsede.org
```

- And then, enter your password

3. Access to tcaa, stampede2

```
gissh stampede2
```

- 4. Go to directory or folder that you sent files to and check if they are correctly sent
- 5. Submit job using shell file

```
sbatch shell_file_name
My case: sbatch bipld2script.sh
```

6. Monitor your job schedule

```
login1$ squeue -u your_tacc_id

My id: private_____ so squeue -u private____
```

7. If you chose email option in .sh file, you will get an email at the beginning and the end of the job as below. Also, in the same directory/folder in stempede2, output and error files will be created as project\_name.ejob\_number, and project\_name.ojob\_number

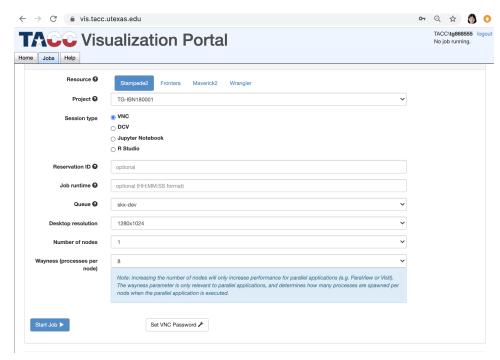


8. After the job is finished, open output file and error files to see how it went

```
cat bipld2.06233457 # Output file for job number 6233457
cat bipld2.e6228979 # Error file for job number 6228979
```

### Using R studio on TACC Visualization Portal

- I wasn't a hundred percent sure if my Rscipt is working in adding pred\_info for all chromosomes at the end in getting the final PRS score
- So I used interactive web based Rstudio on TACC Visualization portal, which was really helpful in checking what values are in data sets, objects, and data frames
- I tried to find snp correlation for chromosome 22
  - 1) Go to this site <a href="https://vis.tacc.utexas.edu/">https://vis.tacc.utexas.edu/</a>
  - 2) Sign in to your TACC account( not the same with xsede account be connected) at the top right corner
  - 3) Go to job section and start interactive R studio which looks like this



- the screen freezes often / only 2 hours

### Errors and why we failed to get PRS using Ldpred2

 Whether the job was queued in skx-dev or skx-normal, there were error messages saying...

에러: 크기가 18.2 Gb인 벡터를 할당할 수 없습니다

실행이 정지되었습니다

경고메시지(들):

시스템 호출에 실패했습니다: 메모리를 할당할 수 없습니다

slurmstepd: error: \*\*\* JOB 6229759 ON c458-064 CANCELLED AT 2020-08-13T09:37:21 DUE TO TIME LIMIT \*\*\*

#### Error in validityMethod(as(object, superClass)):

아직까지는 지원되지 않는 긴 벡터들입니다: ../../src/include/Rinlinedfuns.h:519

Calls: snp\_cor ... validObject -> anyStrings -> isTRUE -> validityMethod

실행이 정지되었습니다

경고메시지(들):

시스템 호출에 실패했습니다: 메모리를 할당할 수 없습니다

In Rstudio TACC visualization portal, it got stuck in the line where the code finds snp\_correlation no matter what chromosome number was. (Probably the smallest 22)

This line: corr0 <-snp\_cor(G,ind.col=ind.chr2,ncores=NCORES,infos.pos=POS2[ind.chr2],size=3/1000)