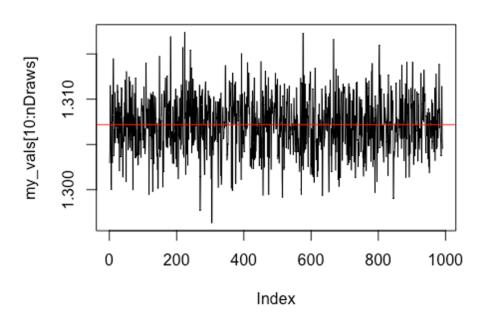
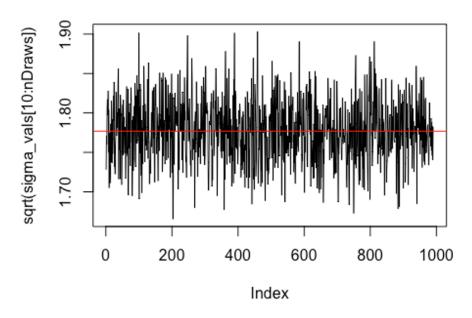
Plot of gibbs sampled distribution for my



Plot of gibbs sampled distribution for sigma



Inefficiency factor my: 0.52 Inefficiency factor sigma: 1.05

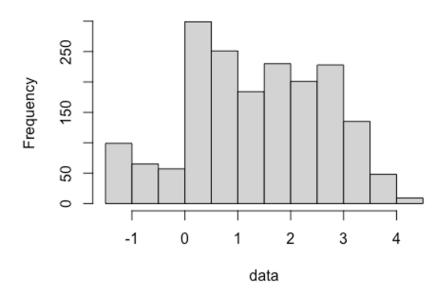
Conclusion: As the plots suggest, we can't distinguish any convergence for the mean and standard deviation as we clearly can see that the values oscillate around the means (red line). Moreover, the inefficiency factors shows how correlated the draws are

```
data=readRDS("Precipitation.rds") #Reading data
data = log(data)
#a)
# Initial setting (prior settings)
mv0 = 1
tao0=1
v0=1
sigma0sg=1
nDraws = 1000
n = nDraws
true mean = mean(data)
vn = v0 + n
#Lists for storing draws
my vals = rep(0, nDraws)
sigma vals = rep(0.nDraws)
sigma vals[1] = 1 #Initial sigmasq value to be used
#Drawing posteriors
for(i in 1:nDraws) {
w = (n/sigma vals[i])/((n/sigma vals[i])+(1/tao0**2)) #Calculating weights
myn = w*true mean+(1-w)*my0 #Calculating myn
taonsq = 1/((n/sigma_vals[i])+(1/tao0)) #Calculating taonsq
my vals[i] = rnorm(1,mean=myn, taonsq) #Calculating posterior my val
<u>if (i <= (nDraws-1)) {</u>
chi val = rchisq(1,vn) #Drawing random value from chi distribution
sigmansq = (v0*sigma0sq+sum((data-my_vals[i])**2))/(n+v0) #Calculating approximation
sigmasg
sigma vals[i+1] = vn*sigmansq/chi val #Calculating posterior sigma val
_}
#Plotting my convergence
plot(my_vals[10:nDraws], _main = "Plot of gibbs sampled distribution for my",type = "I")
abline(h=mean(my vals), col="red") #Expected value
#Plotting sigma convergence
plot(sqrt(sigma_vals[10:nDraws]), ,main = "Plot of gibbs sampled distribution for sigma",type
```

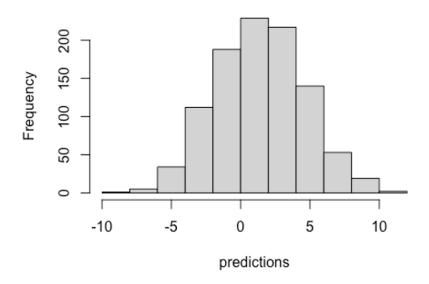
```
=_"|")
abline(h=mean(sqrt(sigma_vals)), col="red") #Expected value
#Calculating inefficienty factor (IF)
#Autocorrelation for my
a my = acf(my vals)
inefficiency_factor_my = 1+2*sum(a_my$acf[-1])
inefficiency factor my
## [1] 0.5193648
#Autocorrelation for my
a_sigma = acf(sqrt(sigma_vals))
inefficiency factor sigma = 1+2*sum(a sigma$acf[-1])
inefficiency factor sigma
## [1] 1.04877
#Conclusion: As the plots suggest, we can't distinguish any convergence for the mean and
standard deviation
#as we clearly can see that the values oscillate around the means (red line)
#Moreover, the inefficiency factors shows how correlated the draws are, as we can see, the
results are
#relatively close to 1 which indicates low correlation
```

b)

Histogram of data



Histogram of predictions



Conclusion: As we can see, the draws from the predictions differ slightly from the true distribution. The main difference is that the normal distribution have a bit longer tails on each end, however the distributions seem to be centred around the same values in both plots

Code:

<u>#b)</u>

#1) Histogram or kernel density estimate of the daily preception y1,...,yn

```
hist(data) #Histogram with true data
```

#2) The resulting posterior predictive density $p(\tilde{y}|y1,...,yn)$ using the simulated posterior draws from (a)

```
predictions = rep(0,nDraws)
for (i in 1:nDraws) {
  predictions[i] = rnorm(1,my_vals[i],sigma_vals[i])
}
```

hist(predictions)

#Conclusion: As we can see, the draws from the predictions differ slightly from the true distribution.

#The main difference is that the normal distribution have a bit longer tails on each end, however

#the distributions seem to be centered around the same values in both plots

Coefficients:

```
Estimate Std. Error z value Pr(>|z|)
(Intercept) 1.07244 0.03077 34.848 < 2e-16 ***
PowerSeller -0.02054 0.03678 -0.558
                                     0.5765
VerifyID -0.39452 0.09243 -4.268 1.97e-05 ***
Sealed
          0.44384 0.05056 8.778 < 2e-16 ***
Minblem
         -0.05220
                     0.06020 -0.867
                                     0.3859
MajBlem
         -0.22087 0.09144 -2.416 0.0157 *
          0.07067
                     0.05633 1.255
LargNeg
                                     0.2096
LogBook
          -0.12068
                     0.02896 -4.166 3.09e-05 ***
MinBidShare -1.89410
                     0.07124 -26.588 < 2e-16 ***
```

the z-value shows that VerifyID, Sealed, MajBlem, LogBook and MinBidShare are significant

```
# (a) Obtain the maximum likelihood estimator of \beta in the Poisson regression model for the
eBay data
# [Hint: glm.R, don't forget that glm() adds its own intercept so don't input the covariate
Const].
# Which covariates are significant?
library(mvtnorm)
ebay data <- (read.table("eBayNumberOfBidderData.dat", header=TRUE))
Xnames <- names(ebay data[,2:ncol(ebay data)]) #Names of columns
n obs <- dim(ebay data)[1] #number of observations
data <- ebay_data[,-2] #training and test data without Const
#y <- as.numeric(data[,1]) #target output
# maximum likelihood estimator of beta
model = glm(formula=nBids~., data=data, family=poisson())
summary(model)
##
## Call:
## gIm(formula = nBids \sim ., family = poisson(), data = data)
##
## Deviance Residuals:
              1Q Median 3Q
       Min
                                    Max
## -3.5800 -0.7222 -0.0441 0.5269 2.4605
##
## Coefficients:
##
              Estimate Std. Error z value Pr(>|z|)
```

```
## (Intercept) 1.07244
                           0.03077 34.848 < 2e-16 ***
## PowerSeller -0.02054
                           0.03678 -0.558 0.5765
             -0.39452
                           0.09243 -4.268 1.97e-05 ***
## VerifyID
## Sealed
             0.44384
                           0.05056 8.778 < 2e-16 ***
                           0.06020 -0.867 0.3859
## Minblem -0.05220
## MajBlem -0.22087
                           0.09144 -2.416 0.0157 *
## LargNeg 0.07067
                           0.05633 1.255 0.2096
## LogBook -0.12068
                           0.02896 -4.166 3.09e-05 ***
## MinBidShare -1.89410
                           0.07124 -26.588 < 2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Dispersion parameter for poisson family taken to be 1)
##
##
      Null deviance: 2151.28 on 999 degrees of freedom
## Residual deviance: 867.47 on 991 degrees of freedom
## AIC: 3610.3
##
## Number of Fisher Scoring iterations: 5
#the z-value shows that VerifyID, Sealed, MajBlem, LogBook and MinBidShare are
```

b)

significant

The posterior mode:

- [1,] 1.06984118 "Const"
- [2,] -0.02051246 "PowerSeller"
- [3,] -0.39300599 "VerifyID"
- [4,] 0.44355549 "Sealed"
- [5,] -0.05246627 "Minblem"
- [6,] -0.22123840 "MajBlem"
- [7,] 0.07069683 "LargNeg"
- [8,] -0.12021767 "LogBook"
- [9,] -1.89198501 "MinBidShare"

Posterior standard deviation:

Const PowerSeller VerifyID Sealed Minblem MajBlem LargNeg 0.03074837 0.03678418 0.09227871 0.05057448 0.06020470 0.09146070 0.05634767 LogBook MinBidShare 0.02895635 0.07109682

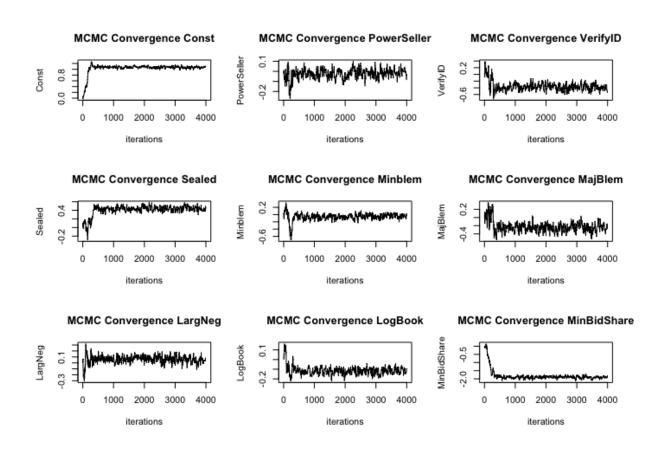
the result seems reasonable since it is similar to a)

```
# (b) Let's do a Bayesian analysis of the Poisson regression. Let the prior be beta ~ N[0,
100*(X_T%*%X)]
# where X is the n \times p covariate matrix. This is a commonly used prior, which is called
Zellner's g-prior.
# Assume first that the posterior density is approximately multivariate normal:
# betaly \sim N(beta\ tilde,\ Jy^{-1}(beta\ tilde))
#where \beta_tilde is the posterior mode and Jy(beta_tilde) is the negative Hessian at the
posterior mode.
#beta tilde and Jy(beta tilde) can be obtained by numerical optimization (optim.R)
# exactly like you already did for the logistic regression in Lab 2
# (but with the log posterior function replaced by the corresponding one for the Poisson
model.
#which you have to code up.).
#function for Poisson logPosterior
logPostPoisson = function(beta,y,X,mu,Sigma) {
 beta <- matrix(beta, nrow=1)
 linPred <- X%*%t(beta)</pre>
 logLik <- sum( - exp(linPred) + linPred*y - log(factorial(y)) ) #loglikelihood
 logPrior <- dmvnorm(beta, mu, Sigma, log=TRUE); #prior
 return(logLik + logPrior) #prior and likelihood are added since we use log
}
#initialize variables
y <- as.numeric(ebay data[,1]) #target output
X <- as.matrix(ebay_data[,-1]) #training data
X \leftarrow apply(X, c(1, 2), as.numeric) #convert from characters to doubles
n_col <- dim(X)[2] #number of columns in training data
initVal <- rep(0,n_col) #initial values for Beta
mu <- as.matrix(rep(0,n col)) # Prior mean vector for Beta
Sigma <- 100*solve(t(X)%*%X) # Prior covariance matrix for Beta
logPost <- logPostPoisson; #function to be minimized</pre>
set.seed(123)
OptimRes <-
optim(initVal,logPost,gr=NULL,y,X,mu,Sigma,method=c("BFGS"),control=list(fnscale=-1),hes
sian=TRUE)
# Printing the results of mean and variance of normally distributed Beta
OptimBeta <- OptimRes$par
names(OptimBeta) <- Xnames # Naming the coefficient by covariates
PostCov <- solve(-OptimRes$hessian) #get covariance matrix by inverting the negative
hessian
approxPostStd <- sqrt(diag(PostCov)) # Computing approximate standard deviations.
names(approxPostStd) <- Xnames # Naming the coefficient by covariates
print('Posterior beta values:')
```

```
## [1] "Posterior beta values:"
print(OptimBeta)
##
      Const PowerSeller
                           VerifyID
                                        Sealed
                                                                    MajBlem
                                                      Minblem
## 1.06984118 -0.02051246 -0.39300599 0.44355549 -0.05246627 -0.22123840
##
                    LogBook MinBidShare
## 0.07069683 -0.12021767 -1.89198501
print('Posterior standard deviation:')
## [1] "Posterior standard deviation:"
print(approxPostStd)
##
      Const PowerSeller
                           VerifyID
                                        Sealed
                                                      Minblem
                                                                    MaiBlem
## 0.03074837 0.03678418 0.09227871 0.05057448 0.06020470 0.09146070
##
                    LogBook MinBidShare
## 0.05634767 0.02895635 0.07109682
```

#the result seems reasonable since they are similar to a)

c)



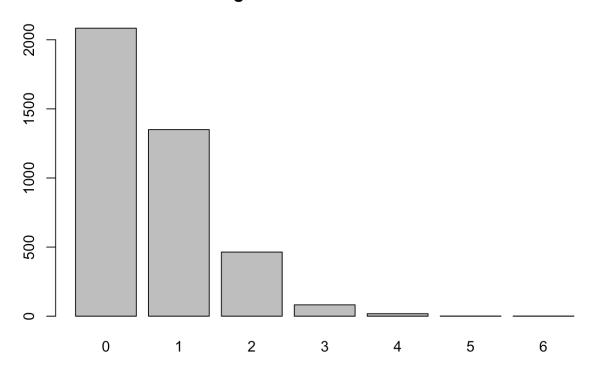
Betavalues are converging as we can see in the plot.

average_alpha = 0.2531, so we have average acceptance probability between 25-30% which means c=0.7 was appropriate.

```
# (c) Let's simulate from the actual posterior of beta using the Metropolis algorithm
# and compare the results with the approximate results in b).
# Program a general function that uses the Metropolis algorithm to generate random draws
from an
# arbitrary posterior density. In order to show that it is a general function for
# any model, we denote the vector of model parameters by \theta. Let the proposal density
# be the multivariate normal density mentioned in Lecture 8 (random walk Metropolis):
# \theta p | \theta(i-1) \sim N(\theta(i-1), c \cdot \Sigma) where \Sigma = Jy^{-1} (beta?tilde) was obtained in b).
# The value c is a tuning parameter and should be an input to your Metropolis function.
# The user of your Metropolis function should be able to supply her own posterior density
function, not
# necessarily for the Poisson regression, and still be able to use your Metropolis
# function. This is not so straightforward, unless you have come across function
# objects in R. The note HowToCodeRWM.pdf in Lisam describes how you can do this in R.
#Now, use your new Metropolis function to sample from the posterior of \beta
# in the Poisson regression for the eBay dataset. Assess MCMC convergence by graphical
methods.
RWMSampler <- function(oldTheta, logPostFunc, c, PostCov, ...) {
 proposedTheta <- rmvnorm(1, mean=oldTheta, sigma=c*PostCov)</pre>
 alpha <- min(1, exp(logPostFunc(proposedTheta, ...) - logPostFunc(oldTheta, ...) ))
 t <- runif(1)
 if (alpha>t) {
       return(list(proposedTheta, alpha))
 } else {
       return(list(oldTheta, alpha))
}
}
nDraws <- 4000
c < -0.7
beta <- matrix(0, nDraws, n col) #matrix to fill with samples from RWM
alpha vector <- rep(0, nDraws) #will be used to calculate mean alpha
logPost <- logPostPoisson
for(i in 1:nDraws) {
 output <- RWMSampler(beta[i,], logPost, c, PostCov, y, X, mu, Sigma)
 alpha_vector[i] <- output[[2]]
 if(i<nDraws) {</pre>
 beta[i+1,] <- output[[1]]
```

d)

Histogram of number of bidders

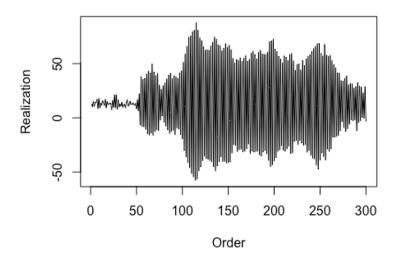


Calculations give 74% chance that we have no bidders with the given characteristics.

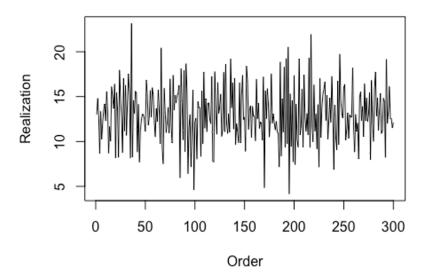
```
# (d) Use the MCMC draws from c) to simulate from the predictive distribution of # the number of bidders in a new auction with the characteristics below. #Plot the predictive distribution. What is the probability of no bidders in this new auction? # PowerSeller = 1
```

```
# VerifyID = 0
# Sealed = 1
# MinBlem = 0
# MajBlem = 1
# LargNeg = 0
\# LogBook = 1.2
# MinBidShare = 0.8
nDraws <- 10000
x \leftarrow c(1,1,0,1,0,1,0,1.2,0.8)
post_beta <- beta[(1000:nrow(beta)),] #select samples after convergence
mean <- exp(post_beta%*%x) #vector of means</pre>
samples <- rpois(nDraws, mean)</pre>
barplot(table(samples), main="Histogram of number of bidders")
prob_no_bidders <- sum(samples==0)/sum(samples) #probability of no bidders
print(prob_no_bidders)
## [1] 0.739524
#the probability of no bidders is 74%
```

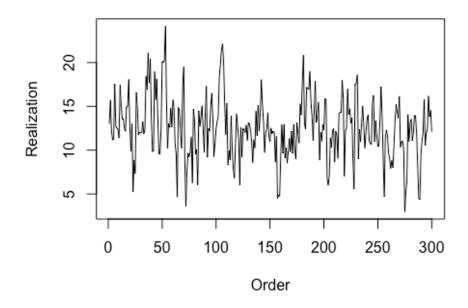
Realization for phi = -1



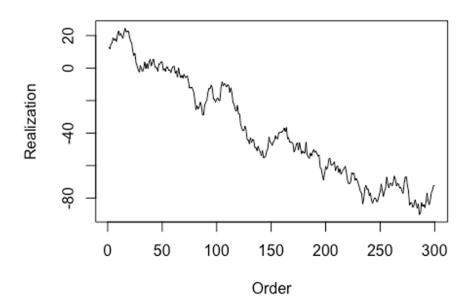
Realization for phi = -0.5



Realization for phi = 0.5



Realization for phi = 1



Conclusion: Increased phi values reduces oscillations. We can see clear oscillations for phi=-1 and that the output becomes more and more correlated with increased phi values

Code:

<u>#a)</u>

library(rstan)

Loading required package: StanHeaders

```
## Loading required package: ggplot2
## rstan (Version 2.21.8, GitRev: 2e1f913d3ca3)
## For execution on a local, multicore CPU with excess RAM we recommend calling
## options(mc.cores = parallel::detectCores()).
## To avoid recompilation of unchanged Stan programs, we recommend calling
## rstan options(auto write = TRUE)
# Initial values
my=13
sigmasg=3
T=300
x1 = my
phi = seq(-1,1,0.25)
#Autoregression function
ar function = function(my,sigmasq,T,x1,phi){
result = rep(0,T)
result[1] = x1
<u>for(i in 2:T) {</u>
epsilon = rnorm(1,0,sigmasq)
<u>result[i] = my + phi*(result[i-1]-my) + epsilon</u>
}
return(result)
}
#Calculating output
result = matrix(0,T,length(phi))
for(i in 1:length(phi)) {
result[,i] = ar function(my,sigmasq,T,x1,phi[i])
}
#Plotting result for phi=-1 and phi=1
plot(seq(1,300,1),result[,1], xlab="Order", ylab="Realization", main="Realization for phi =
-1",type="l")
plot(seg(1,300,1),result[,3], xlab="Order", ylab="Realization", main="Realization for phi =
-0.5",type="l")
plot(seq(1,300,1),result[,7], xlab="Order", ylab="Realization", main="Realization for phi =
0.5",type="l")
plot(seq(1,300,1),result[,9], xlab="Order", ylab="Realization", main="Realization for phi =
1",type="l")
#Conclusion: Increased phi values reduces oscilations. We can see clear oscilations for
#and that the output becomes more and more correlated with increased phi values
```

Table 1: Values for x model

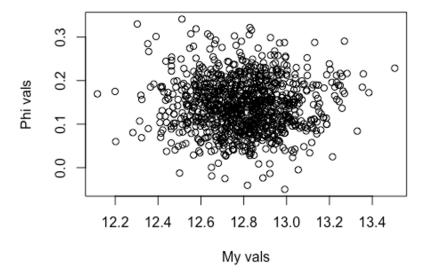
	mean	se_mean	sd	2.5%	25%	50%	75%	97.5%	n_eff	Rhat
mu	12.84	0.00	0.20	12.43	12.71	12.84	12.97	13.24	3440	1
phi	0.12	0.00	0.06	0.01	0.08	0.12	0.16	0.24	3620	1
sigma2	9.02	0.01	0.76	7.66	8.49	8.99	9.51	10.61	3611	1
lp	-474.33	0.03	1.30	-477.78	-474.90	-473.98	-473.39	-472.89	1970	1

Table 2: Values for y model

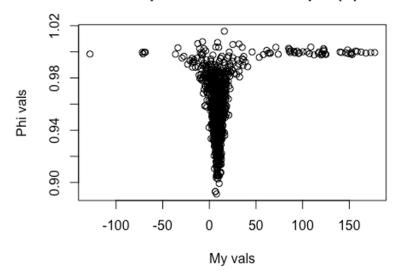
mu phi	mean 14.76 0.98	1.33	24.52	2.5% -40.56 0.94		14.82	19.58	97.5% 76.75 1.00		1.01
sigma	2 8.64	0.02	0.72	7.37	8.14	8.58	9.11	10.20	1273 1	L.00
lp	-467.64	0.05	1.38	-470.83	-468.53	-467.35	-466.51	-465.89	764 1	1.01

Conclusion: As the values in the credible interval are more narrow for the one where phi=0.2 were used, we get a better understanding of the true values for this one compared to the one where phi=0.95 were used. For this one, the values are really wide in the credible intervals

Joint posterior of mu and phi (X)



Joint posterior of mu and phi (Y)



Conclusion: We can distinguish a convergence for the first sampler. However it is more difficult to distinguish a convergence for the second sampler as shown in the figure. This follows the previous reasoning, that the second sampler have a wider span for its values compared to the first sampler

```
#b)
#Drawing x1:T values using phi=0.2
x=rep(0,T)
x=ar function(my,sigmasq,T,x1,0.2)
#Drawing y1:T values using phi=0.95
y=rep(0,T)
y=ar function(my,sigmasq,T,x1,0.95)
#Implementing Stan model
StanModel = '
data {
int<lower=0> N;
vector[N] z;
}
parameters {
real mu;
real phi;
real<lower=0> sigma2;
model {
```

```
for(i in 2:N){
z[i] \sim normal(mu+phi*(z[i-1]-mu),sqrt(sigma2));
_}
<u>}'</u>
#For X
data <- list(N=T, z=x)
fit x <- stan(model code=StanModel,data=data)</pre>
#For Y
data <- list(N=T, z=y)
<u>fit_y <- stan(model_code=StanModel,data=data)</u>
#Extract posterior draws
post x = extract(fit x)
post y = extract(fit y)
#i)
#AR-process for X
#Calculating mean and credible interval for all parameters
#Can be achived with print(fit x) too
mean mu = mean(post x mu)
cred int mu = quantile(post x$mu,probs=c(0.025,0.975))
print(mean_mu)
## [1] 12.79963
print(cred int mu)
## 2.5% 97.5%
## 12.43112 13.17352
mean phi= mean(post x$phi)
cred int phi = quantile(post xphi,probs=c(0.025,0.975))
print(mean phi)
## [1] 0.1432679
print(cred_int_phi)
## 2.5% 97.5%
## 0.03507495 0.25712395
mean sigma2 = mean(post x$sigma2)
cred int sigma2 = quantile(post xsigma2,probs=c(0.025,0.975))
print(mean sigma2)
## [1] 7.918592
print(cred_int_sigma2)
```

```
## 2.5% 97.5%
## 6.726024 9.330297
#AR-process for Y
#Calculating mean and credible interval for all parameters
mean_mu = mean(post_y$mu)
cred int mu = quantile(post y$mu,probs=c(0.025,0.975))
print(mean mu)
## [1] 10.04247
print(cred int mu)
## 2.5% 97.5%
## -16.08935 50.19349
mean phi= mean(post v$phi)
cred int phi = quantile(post y$phi,probs=c(0.025,0.975))
print(mean phi)
## [1] 0.957765
print(cred int phi)
## 2.5% 97.5%
## 0.9116279 1.0007403
mean sigma2 = mean(post y$sigma2)
cred int sigma2 = guantile(post v$sigma2,probs=c(0.025,0.975))
print(mean sigma2)
## [1] 9.138192
print(cred int sigma2)
## 2.5% 97.5%
## 7.744661 10.745055
#Plotting using print
print(fit x)
## Inference for Stan model: d85eaf939f1fe064d5bdde49c5af5b23.
## 4 chains, each with iter=2000; warmup=1000; thin=1;
## post-warmup draws per chain=1000, total post-warmup draws=4000.
##
##
      mean se mean sd 2.5% 25% 50% 75% 97.5% n eff Rhat
## mu 12.80 0.00 0.19 12.43 12.68 12.80 12.92 13.17 3586 1
## phi 0.14 0.00 0.06 0.04 0.10 0.14 0.18 0.26 3594 1
## sigma2 7.92 0.01 0.66 6.73 7.46 7.88 8.33 9.33 4058 1
## lp
       <u>-455.35</u> <u>0.03 1.22 -458.63 -455.90 -455.02 -454.47 -453.96 2033 1</u>
##
## Samples were drawn using NUTS(diag_e) at Tue May_9 10:30:51 2023.
```

For each parameter, n_eff is a crude measure of effective sample size, ## and Rhat is the potential scale reduction factor on split chains (at ## convergence, Rhat=1).

print(fit y)

Inference for Stan model: d85eaf939f1fe064d5bdde49c5af5b23.

4 chains, each with iter=2000; warmup=1000; thin=1;

post-warmup draws per chain=1000, total post-warmup draws=4000.

##

mean se_mean sd 2.5% 25% 50% 75% 97.5% n_eff Rhat

mu 10.04 1.21 17.77 -16.09 5.62 8.79 11.80 50.19 215 1.02

phi 0.96 0.00 0.02 0.91 0.94 0.96 0.97 1.00 584 1.01

sigma2 9.14 0.02 0.76 7.74 8.61 9.07 9.64 10.75 1608 1.00

Samples were drawn using NUTS(diag_e) at Tue May 9 10:30:56 2023.

For each parameter, n eff is a crude measure of effective sample size.

and Rhat is the potential scale reduction factor on split chains (at ## convergence, Rhat=1).

#Conclusion: As the values in the credible interval are more narrow for the one where phi=0.2

#were used, we get a better understanding of the true values for this one compared to the one where

#phi=0.95 were used. For this one, the values are really wide in the credible intervals

#ii)

#Joint posterior for X

plot(post_x\$mu[1000:2000],post_x\$phi[1000:2000],xlab="My vals",ylab="Phi vals", main="Joint posterior of mu and phi (X)")

#Joint posterior for X

plot(post_y\$mu[1000:2000].post_y\$phi[1000:2000],xlab="My vals",ylab="Phi vals", main="Joint posterior of mu and phi (Y)")

#Conclusion: We can distinguish a convergence for the first sampler. However it is more difficult to distinguish a

#convergence for the second sampler as shown in the figure. This follows the previous reasoning, that the

#second sampler have a wider span for its values compared to the first sampler

Appendix

Exercise 1

```
data=readRDS("Precipitation.rds") #Reading data
data = log(data)
#a)
# Initial setting (prior settings)
mv0 = 1
tao0=1
v0 = 1
sigma0sq=1
nDraws = 1000
n = nDraws
true mean = mean(data)
vn = v0 + n
#Lists for storing draws
my vals = rep(0, nDraws)
sigma vals = rep(0,nDraws)
sigma vals[1] = 1 #Initial sigmasq value to be used
#Drawing posteriors
for(i in 1:nDraws) {
w = (n/sigma vals[i])/((n/sigma vals[i])+(1/tao0**2)) #Calculating weights
myn = w*true mean+(1-w)*my0 #Calculating myn
taonsq = 1/((n/sigma vals[i])+(1/tao0)) #Calculating taonsq
my_vals[i] = rnorm(1,mean=myn, taonsq) #Calculating posterior my_val
if (i <= (nDraws-1)) {</pre>
chi val = rchisq(1,vn) #Drawing random value from chi distribution
sigmansq = (v0*sigma0sq+sum((data-my vals[i])**2))/(n+v0) #Calculating approximation
sigmasg
sigma vals[i+1] = vn*sigmansq/chi val #Calculating posterior sigma val
_}
}
#Plotting my convergence
plot(my vals[10:nDraws], _main = "Plot of gibbs sampled distribution for my",type = "I")
abline(h=mean(my vals), col="red") #Expected value
#Plotting sigma convergence
plot(sgrt(sigma_vals[10:nDraws]), ,main = "Plot of gibbs sampled distribution for sigma",type
= "|")
<u>abline(h=mean(sqrt(sigma_vals)), col="red")</u> #Expected value
```

```
#Calculating inefficienty factor (IF)
#Autocorrelation for my
a my = acf(my vals)
inefficiency factor my = 1+2*sum(a my$acf[-1])
inefficiency factor my
## [1] 0.5193648
#Autocorrelation for my
a sigma = acf(sqrt(sigma vals))
inefficiency factor sigma = 1+2*sum(a sigma$acf[-1])
inefficiency factor sigma
## [1] 1.04877
#Conclusion: As the plots suggest, we can't distinguish any convergence for the mean and
standard deviation
#as we clearly can see that the values oscillate around the means (red line)
#Moreover, the inefficiency factors shows how correlated the draws are, as we can see, the
results are
#relatively close to 1 which indicates low correlation
#b)
#1) Histogram or kernel density estimate of the daily preception y1,...,yn
hist(data) #Histogram with true data
#2) The resulting posterior predictive density p(\tilde{y}|y1,...,yn) using the simulated posterior
draws from (a)
predictions = rep(0,nDraws)
for (i in 1:nDraws) {
predictions[i] = rnorm(1,my vals[i],sigma vals[i])
}
hist(predictions)
#Conclusion: As we can see, the draws from the predictions differ slightly from the true
distribution.
#The main difference is that the normal distribution have a bit longer tails on each end.
however
#the distributions seem to be centered around the same values in both plots
```

Exercise 2

```
# Metropolis Random Walk for Poisson regression.
#Consider the following Poisson regression model y i|beta(iid) ~
Poisson[exp(x i transpose%*%β)], i=1,...n
# where v i is the count for the ith observation in the sample
# and xi is the p-dimensional vector with covariate observations for the ith observation.
# Use the data set eBayNumberOfBidderData.dat. This dataset contains observations from
1000 eBay auctions of coins.
# The response variable is nBids and records the number of bids in each auction.
# The remaining variables are features/covariates (x):
# Const (for the intercept)
# PowerSeller (equal to 1 if the seller is selling large volumes on eBay)
# VerifyID (equal to 1 if the seller is a verified seller by eBay)
# Sealed (equal to 1 if the coin was sold in an unopened envelope)
# MinBlem (equal to 1 if the coin has a minor defect)
# MajBlem (equal to 1 if the coin has a major defect)
# LargNeg (egual to 1 if the seller received a lot of negative feedback from customers)
# LogBook (logarithm of the book value of the auctioned coin according to expert sellers.
Standardized)
# MinBidShare (ratio of the minimum selling price (starting price) to the book value.
Standardized).
# (a) Obtain the maximum likelihood estimator of \( \beta \) in the Poisson regression model for the
eBay data
# [Hint: glm.R, don't forget that glm() adds its own intercept so don't input the covariate
Constl.
# Which covariates are significant?
<u>library(mvtnorm)</u>
ebay data <- (read.table("eBayNumberOfBidderData.dat", header=TRUE))</pre>
Xnames <- names(ebay data[,2:ncol(ebay data)]) #Names of columns
n obs <- dim(ebay data)[1] #number of observations
data <- ebay data[,-2] #training and test data without Const
#v <- as.numeric(data[,1]) #target output
# maximum likelihood estimator of beta
model = glm(formula=nBids~., data=data, family=poisson())
summary(model)
##
## Call:
## glm(formula = nBids \sim ., family = poisson(), data = data)
## Deviance Residuals:
       Min 1Q Median 3Q
                                     Max
## -3.5800 -0.7222 -0.0441 0.5269 2.4605
```

```
##
## Coefficients:
              Estimate Std. Error z value Pr(>|z|)
## (Intercept) 1.07244
                            0.03077 34.848 < 2e-16 ***
## PowerSeller -0.02054
                            0.03678 -0.558 0.5765
                            0.09243 -4.268 1.97e-05 ***
## VerifvID -0.39452
## Sealed
              0.44384
                             0.05056 8.778 < 2e-16 ***
## Minblem -0.05220
                            0.06020 -0.867 0.3859
## MaiBlem -0.22087
                            0.09144 -2.416 0.0157 *
                            0.05633 1.255 0.2096
## LargNeg 0.07067
## LogBook -0.12068
                            0.02896 -4.166 3.09e-05 ***
                            0.07124 -26.588 < 2e-16 ***
## MinBidShare -1.89410
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Dispersion parameter for poisson family taken to be 1)
##
##
       Null deviance: 2151.28 on 999 degrees of freedom
## Residual deviance: 867.47 on 991 degrees of freedom
## AIC: 3610.3
##
## Number of Fisher Scoring iterations: 5
#the z-value shows that VerifyID, Sealed, MajBlem, LogBook and MinBidShare are
significant
# (b) Let's do a Bayesian analysis of the Poisson regression. Let the prior be beta ~ N[0,
100*(X T%*%X)1
# where X is the n \times p covariate matrix. This is a commonly used prior, which is called
Zellner's q-prior.
# Assume first that the posterior density is approximately multivariate normal:
# betaly ~ N(beta tilde, Jy^{-1}(beta tilde))
#where \beta tilde is the posterior mode and Jy(beta tilde) is the negative Hessian at the
posterior mode.
#beta_tilde and Jy(beta_tilde) can be obtained by numerical optimization (optim.R)
# exactly like you already did for the logistic regression in Lab 2
# (but with the log posterior function replaced by the corresponding one for the Poisson
model.
#which you have to code up.).
#function for Poisson logPosterior
logPostPoisson = function(beta,y,X,mu,Sigma) {
beta <- matrix(beta, nrow=1)</pre>
linPred <- X%*%t(beta)
 logLik <- sum( - exp(linPred) + linPred*y - log(factorial(y)) ) #loglikelihood</pre>
logPrior <- dmvnorm(beta, mu, Sigma, log=TRUE); #prior
 return(logLik + logPrior) #prior and likelihood are added since we use log
ł
```

```
v <- as.numeric(ebay data[,1]) #target output</pre>
X <- as.matrix(ebay data[,-1]) #training data
X \leftarrow apply(X, c(1, 2), as.numeric) #convert from characters to doubles
n col <- dim(X)[2] #number of columns in training data
initVal <- rep(0,n col) #initial values for Beta
mu <- as.matrix(rep(0,n col)) # Prior mean vector for Beta
Sigma <- 100*solve(t(X)%*%X) # Prior covariance matrix for Beta
logPost <- logPostPoisson; #function to be minimized</pre>
set.seed(123)
OptimRes <-
optim(initVal,logPost,gr=NULL,y,X,mu,Sigma,method=c("BFGS"),control=list(fnscale=-1),hes
sian=TRUE)
# Printing the results of mean and variance of normally distributed Beta
OptimBeta <- OptimRes$par
names(OptimBeta) <- Xnames # Naming the coefficient by covariates</pre>
PostCov <- solve(-OptimRes$hessian) #get covariance matrix by inverting the negative
<u>hessian</u>
approxPostStd <- sqrt(diag(PostCov)) # Computing approximate standard deviations.
names(approxPostStd) <- Xnames # Naming the coefficient by covariates
print('Posterior beta values:')
## [1] "Posterior beta values:"
print(OptimBeta)
      Const PowerSeller VerifyID Sealed Minblem
                                                                      MaiBlem
## 1.06984118 -0.02051246 -0.39300599 0.44355549 -0.05246627 -0.22123840
     LargNeg LogBook MinBidShare
## 0.07069683 -0.12021767 -1.89198501
print('Posterior standard deviation:')
## [1] "Posterior standard deviation:"
print(approxPostStd)
      Const PowerSeller VerifyID Sealed
                                                        Minblem
                                                                      MaiBlem
## 0.03074837 0.03678418 0.09227871 0.05057448 0.06020470 0.09146070
     LargNeg LogBook MinBidShare
## 0.05634767 0.02895635 0.07109682
#the result seems reasonable since they are similar to a)
```

(c) Let's simulate from the actual posterior of beta using the Metropolis algorithm

and compare the results with the approximate results in b).

#initialize variables

```
# Program a general function that uses the Metropolis algorithm to generate random draws
from an
# arbitrary posterior density. In order to show that it is a general function for
# any model, we denote the vector of model parameters by \theta. Let the proposal density
# be the multivariate normal density mentioned in Lecture 8 (random walk Metropolis):
# \theta p | \theta(i-1) \sim N(\theta(i-1), c \cdot \Sigma) where \Sigma = Jy^{-1} (beta?tilde) was obtained in b).
# The value c is a tuning parameter and should be an input to your Metropolis function.
# The user of your Metropolis function should be able to supply her own posterior density
function, not
# necessarily for the Poisson regression, and still be able to use your Metropolis
# function. This is not so straightforward, unless you have come across function
# objects in R. The note HowToCodeRWM.pdf in Lisam describes how you can do this in R.
#Now, use your new Metropolis function to sample from the posterior of β
# in the Poisson regression for the eBay dataset. Assess MCMC convergence by graphical
methods.
RWMSampler <- function(oldTheta, logPostFunc, c, PostCov, ...) {
proposedTheta <- rmvnorm(1, mean=oldTheta, sigma=c*PostCov)</pre>
alpha <- min(1, exp(logPostFunc(proposedTheta, ...) - logPostFunc(oldTheta, ...) ))
t <- runif(1)
<u>if (alpha>t) {</u>
       return(list(proposedTheta, alpha))
} else {
       return(list(oldTheta, alpha))
_}
nDraws <- 4000
c < -0.7
beta <- matrix(0, nDraws, n_col) #matrix to fill with samples from RWM
alpha vector <- rep(0, nDraws) #will be used to calculate mean alpha
logPost <- logPostPoisson</pre>
for(i in 1:nDraws) {
output <- RWMSampler(beta[i,], logPost, c, PostCov, y, X, mu, Sigma)
alpha vector[i] <- output[[2]]
if(i<nDraws) {</pre>
beta[i+1,] <- output[[1]]
_}
}
iterations=seq(1,nDraws,1)
par(mfrow=c(3,3))
for (i in 1:n col) {
plot(iterations, beta[,i], type="l", main=paste("MCMC Convergence", Xnames[i]),
```

```
vlab=Xnames[i])
par(mfrow=c(1,1), new=FALSE)
# Calculate average alpha
average alpha <- mean(alpha vector)</pre>
average alpha #0.2531, has average acceptance probability between 25-30%
## [1] 0.2621183
# (d) Use the MCMC draws from c) to simulate from the predictive distribution of
# the number of bidders in a new auction with the characteristics below.
#Plot the predictive distribution. What is the probability of no bidders in this new auction?
# PowerSeller = 1
# VerifyID = 0
# Sealed = 1
# MinBlem = 0
# MajBlem = 1
\# LargNeg = 0
\# LogBook = 1.2
# MinBidShare = 0.8
nDraws <- 10000
x \le c(1,1,0,1,0,1,0,1,2,0.8)
post beta <- beta[(1000:nrow(beta)),] #select samples after convergence</pre>
mean <- exp(post_beta%*%x) #vector of means
samples <- rpois(nDraws, mean)</pre>
barplot(table(samples), main="Histogram of number of bidders")
prob_no_bidders <- sum(samples==0)/sum(samples) #probability of no bidders</pre>
print(prob no bidders)
## [1] 0.739524
#the probability of no bidders is 74%
```

```
Exercise 3
#a)
<u>library(rstan)</u>
## Loading required package: StanHeaders
## Loading required package: ggplot2
## rstan (Version 2.21.8, GitRev: 2e1f913d3ca3)
## For execution on a local, multicore CPU with excess RAM we recommend calling
## options(mc.cores = parallel::detectCores()).
## To avoid recompilation of unchanged Stan programs, we recommend calling
## rstan options(auto write = TRUE)
# Initial values
my=13
sigmasq=3
T=300
x1 = my
phi = seq(-1,1,0.25)
#Autoregression function
ar function = function(my,sigmasq,T,x1,phi){
result = rep(0,T)
result[1] = x1
<u>for(i in 2:T) {</u>
epsilon = rnorm(1,0,sigmasq)
 result[i] = my + phi*(result[i-1]-my) + epsilon
_}
return(result)
#Calculating output
result = matrix(0,T,length(phi))
for(i in 1:length(phi)) {
result[,i] = ar_function(my,sigmasq,T,x1,phi[i])
ł
#Plotting result for phi=-1 and phi=1
plot(seq(1,300,1),result[,1], xlab="Order", ylab="Realization", main="Realization for phi =
-1",type="l")
plot(seq(1,300,1),result[,3], xlab="Order", ylab="Realization", main="Realization for phi =
-0.5",type="l")
plot(seg(1,300,1),result[,7], xlab="Order", ylab="Realization", main="Realization for phi =
0.5",type="l")
```

```
plot(seg(1,300,1),result[,9], xlab="Order", ylab="Realization", main="Realization for phi =
1",type="l")
#Conclusion: Increased phi values reduces oscilations. We can see clear oscilations for
#and that the output becomes more and more correlated with increased phi values
#b)
#Drawing x1:T values using phi=0.2
x=rep(0,T)
x=ar function(my, sigmasq, T, x1, 0.2)
#Drawing y1:T values using phi=0.95
y=rep(0,T)
y=ar_function(my,sigmasq,T,x1,0.95)
#Implementing Stan model
StanModel = '
data {
int<lower=0> N;
vector[N] z;
parameters {
real mu;
real phi;
real<lower=0> sigma2;
model {
for(i in 2:N){
z[i] ~ normal(mu+phi*(z[i-1]-mu),sqrt(sigma2));
_}
<u>}'</u>
#For X
data <- list(N=T, z=x)
fit x <- stan(model code=StanModel,data=data)
#For Y
data <- list(N=T, z=y)
fit y <- stan(model code=StanModel,data=data)</pre>
#Extract posterior draws
post_x = extract(fit_x)
post y = extract(fit y)
#i)
#AR-process for X
#Calculating mean and credible interval for all parameters
```

```
#Can be achived with print(fit x) too
mean mu = mean(post x mu)
cred int mu = quantile(post x$mu,probs=c(0.025,0.975))
print(mean mu)
## [1] 12.79963
print(cred int mu)
## 2.5% 97.5%
## 12.43112 13.17352
mean_phi= mean(post_x$phi)
cred int phi = quantile(post xphi,probs=c(0.025,0.975))
print(mean_phi)
## [1] 0.1432679
print(cred int phi)
## 2.5% <u>9</u>7.5%
## 0.03507495 0.25712395
mean_sigma2 = mean(post_x$sigma2)
<u>cred int sigma2 = quantile(post x$sigma2,probs=c(0.025,0.975))</u></u>
print(mean_sigma2)
## [1] 7.918592
print(cred int sigma2)
<u>## 2.5% 97.5</u>%
## 6.726024 9.330297
#AR-process for Y
#Calculating mean and credible interval for all parameters
mean mu = mean(post v\$mu)
cred_int_mu = quantile(post_y$mu,probs=c(0.025,0.975))
print(mean mu)
## [1] 10.04247
print(cred int mu)
## 2.5% 97.5%
## -16.08935 50.19349
mean phi= mean(post y$phi)
cred int phi = quantile(post y$phi,probs=c(0.025,0.975))
print(mean phi)
## [1] 0.957765
```

```
print(cred int phi)
## 2.5% 97.5%
## 0.9116279 1.0007403
mean sigma2 = mean(post y$sigma2)
cred int sigma2 = quantile(post y$sigma2,probs=c(0.025,0.975))
print(mean sigma2)
## [1] 9.138192
print(cred int sigma2)
## 2.5% 97.5%
## 7.744661 10.745055
#Plotting using print
print(fit x)
## Inference for Stan model: d85eaf939f1fe064d5bdde49c5af5b23.
## 4 chains, each with iter=2000; warmup=1000; thin=1;
## post-warmup draws per chain=1000, total post-warmup draws=4000.
##
        mean se mean sd 2.5% 25% 50% 75% 97.5% n eff Rhat
##
        <u>12.80 0.00 0.19 12.43 12.68 12.80 12.92</u> 13.17 3586 1
## phi 0.14 0.00 0.06 0.04 0.10 0.14 0.18 0.26 3594 1
## sigma2 7.92 0.01 0.66 6.73 7.46 7.88 8.33 9.33 4058 1
        <u>-455.35</u> <u>0.03 1.22 -458.63 -455.90 -455.02 -454.47 -453.96 2033 1</u>
## lp
##
## Samples were drawn using NUTS(diag_e) at Tue May_9 10:30:51 2023.
## For each parameter, n eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).
print(fit y)
## Inference for Stan model: d85eaf939f1fe064d5bdde49c5af5b23.
## 4 chains, each with iter=2000; warmup=1000; thin=1;
## post-warmup draws per chain=1000, total post-warmup draws=4000.
##
##
        mean se mean sd 2.5% 25% 50% 75% 97.5% n eff Rhat
        <u>10.04</u> <u>1.21 17.77 -16.09</u> <u>5.62</u> <u>8.79</u> <u>11.80</u> <u>50.19</u> <u>215 1.02</u>
## mu
        0.96 0.00 0.02 0.91 0.94 0.96 0.97 1.00 584 1.01
## sigma2 9.14 0.02 0.76 7.74 8.61 9.07 9.64 10.75 1608 1.00
## lp -476.53 0.06 1.48 -480.00 -477.38 -476.14 -475.37 -474.75 614 1.01
## Samples were drawn using NUTS(diag e) at Tue May 9 10:30:56 2023.
## For each parameter, n eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).
```

#Conclusion: As the values in the credible interval are more narrow for the one where phi=0.2

#were used, we get a better understanding of the true values for this one compared to the one where

#phi=0.95 were used. For this one, the values are really wide in the credible intervals

<u>#ii)</u>

#Joint posterior for X

plot(post_x\$mu[1000:2000].post_x\$phi[1000:2000].xlab="My vals",ylab="Phi vals", main="Joint posterior of mu and phi (X)")

#Joint posterior for X

plot(post_y\$mu[1000:2000],post_y\$phi[1000:2000],xlab="My vals",ylab="Phi vals", main="Joint posterior of mu and phi (Y)")

#Conclusion: We can distinguish a convergence for the first sampler. However it is more difficult to distinguish a

#convergence for the second sampler as shown in the figure. This follows the previous reasoning, that the

#second sampler have a wider span for its values compared to the first sampler