Class 14: RNA-seq analysis mini-project

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Background:

The data for for hands-on session comes from GEO entry: GSE37704, which is associated with the following publication:

• Trapnell C, Hendrickson DG, Sauvageau M, Goff L et al. "Differential analysis of gene regulation at transcript resolution with RNA-seq". Nat Biotechnol 2013 Jan;31(1):46-53. PMID: 23222703

The authors report on differential analysis of lung fibroblasts in response to loss of the developmental transcription factor HOXA1. Their results and others indicate that HOXA1 is required for lung fibroblast and HeLa cell cycle progression. In particular their analysis show that "loss of HOXA1 results in significant expression level changes in thousands of individual transcripts, along with isoform switching events in key regulators of the cell cycle". For our session we have used their Sailfish gene-level estimated counts and hence are restricted to protein-coding genes only.

Data Import:

Load our files:

```
counts <- read.csv('GSE37704_featurecounts.csv', row.names = 1)
colData <- read.csv('GSE37704_metadata.csv')</pre>
```

Inspect and tidy data:

Q. Complete the code below to remove the troublesome first column from count-Data. Does the counts column match with the colData rows?

head(counts)

	length	SRR493366	SRR493367	SRR493368	SRR493369	SRR493370
ENSG00000186092	918	0	0	0	0	0
ENSG00000279928	718	0	0	0	0	0
ENSG00000279457	1982	23	28	29	29	28
ENSG00000278566	939	0	0	0	0	0
ENSG00000273547	939	0	0	0	0	0
ENSG00000187634	3214	124	123	205	207	212
	SRR4933	371				
ENSG00000186092		0				
ENSG00000279928		0				
ENSG00000279457		46				
ENSG00000278566		0				
ENSG00000273547		0				
ENSG00000187634	2	258				

head(colData)

```
id condition
1 SRR493366 control_sirna
2 SRR493367 control_sirna
3 SRR493368 control_sirna
4 SRR493369 hoxa1_kd
5 SRR493370 hoxa1_kd
6 SRR493371 hoxa1_kd
```

Since it does not match we need to transform the data frame so that they align, since there is an extra length column in the counts data, which needs to be removed.

colnames(counts)

```
[1] "length" "SRR493366" "SRR493367" "SRR493368" "SRR493369" "SRR493370" [7] "SRR493371"
```

```
countData <- counts[,-1]
head(countData)</pre>
```

	SRR493366	SRR493367	SRR493368	SRR493369	SRR493370	SRR493371
ENSG00000186092	0	0	0	0	0	0
ENSG00000279928	0	0	0	0	0	0
ENSG00000279457	23	28	29	29	28	46
ENSG00000278566	0	0	0	0	0	0
ENSG00000273547	0	0	0	0	0	0
ENSG00000187634	124	123	205	207	212	258

Check for matching countData and colData:

```
colnames(countData) == colData$id
```

[1] TRUE TRUE TRUE TRUE TRUE TRUE

Q1. How many genes in total?

19808 genes total in our dataset

nrow(countData)

[1] 19808

Q2. Filter to remove zero count genes (rows where there are zero counts in all columns). How many genes are left?

There are 15975 genes that are not zero count genes, within this data set.

```
to.keep.inds <- rowSums(countData) > 0
```

new.counts <- countData[to.keep.inds,]</pre>

nrow(new.counts)

[1] 15975

Setup for DESeq:

library(DESeq2)

```
Loading required package: S4Vectors
```

Loading required package: stats4

Loading required package: BiocGenerics

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

anyDuplicated, aperm, append, as.data.frame, basename, cbind, colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget, order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind, Reduce, rownames, sapply, saveRDS, setdiff, table, tapply, union, unique, unsplit, which.max, which.min

Attaching package: 'S4Vectors'

The following object is masked from 'package:utils':

findMatches

The following objects are masked from 'package:base':

expand.grid, I, unname

Loading required package: IRanges

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

Loading required package: SummarizedExperiment

Loading required package: MatrixGenerics

Loading required package: matrixStats

Attaching package: 'MatrixGenerics'

The following objects are masked from 'package:matrixStats':

colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse, colCounts, colCummaxs, colCummins, colCumprods, colCumsums, colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs, colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats, colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds, colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads, colWeightedMeans, colWeightedMedians, colWeightedSds, colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet, rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods, rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps, rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins, rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks, rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars, rowWeightedMads, rowWeightedMeans, rowWeightedMedians, rowWeighted

Loading required package: Biobase

```
Welcome to Bioconductor
```

```
Vignettes contain introductory material; view with 'browseVignettes()'. To cite Bioconductor, see 'citation("Biobase")', and for packages 'citation("pkgname")'.

Attaching package: 'Biobase'

The following object is masked from 'package:MatrixGenerics': rowMedians

The following objects are masked from 'package:matrixStats': anyMissing, rowMedians
```

Setup input objects for DESeq:

Warning in DESeqDataSet(se, design = design, ignoreRank): some variables in design formula are characters, converting to factors

Run DESeq:

```
dds <- DESeq(dds)

estimating size factors

estimating dispersions

gene-wise dispersion estimates

mean-dispersion relationship</pre>
```

final dispersion estimates

fitting model and testing

```
res <- results(dds)</pre>
```

head(res)

log2 fold change (MLE): condition hoxa1 kd vs control sirna Wald test p-value: condition hoxa1 kd vs control sirna DataFrame with 6 rows and 6 columns

```
baseMean log2FoldChange
                                             lfcSE
                                                          stat
                                                                    pvalue
                <numeric>
                               <numeric> <numeric>
                                                     <numeric>
                                                                 <numeric>
ENSG00000279457
                  29.9136
                               0.1792571 0.3248216
                                                     0.551863 5.81042e-01
ENSG00000187634 183.2296
                               0.4264571 0.1402658
                                                     3.040350 2.36304e-03
ENSG00000188976 1651.1881
                              -0.6927205 0.0548465 -12.630158 1.43989e-36
                 209.6379
                               0.7297556 0.1318599
                                                     5.534326 3.12428e-08
ENSG00000187961
ENSG00000187583
                  47.2551
                               0.0405765 0.2718928
                                                     0.149237 8.81366e-01
                               0.5428105 0.5215599
                                                     1.040744 2.97994e-01
ENSG00000187642
                  11.9798
                       padj
                  <numeric>
ENSG00000279457 6.86555e-01
ENSG00000187634 5.15718e-03
ENSG00000188976 1.76549e-35
ENSG00000187961 1.13413e-07
ENSG00000187583 9.19031e-01
ENSG00000187642 4.03379e-01
```

Q3. Call the summary() function on your results to get a sense of how many genes are up or down-regulated at the default 0.1 p-value cutoff.

summary(res)

```
out of 15975 with nonzero total read count
adjusted p-value < 0.1
LFC > 0 (up) : 4349, 27%
LFC < 0 (down) : 4396, 28%
outliers [1] : 0, 0%
low counts [2] : 1237, 7.7%
(mean count < 0)
[1] see 'cooksCutoff' argument of ?results
[2] see 'independentFiltering' argument of ?results</pre>
```

Volcano plot of results:

```
library(ggplot2)

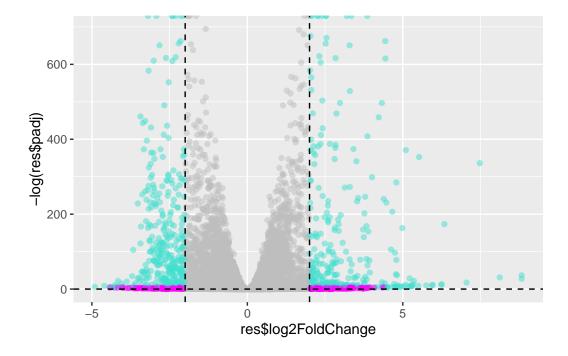
mycols <- rep('gray', nrow(res))
mycols[ abs(res$log2FoldChange) > 2 ] <- "magenta"

inds <- (res$padj < 0.01) & (abs(res$log2FoldChange) > 2 )
mycols[ inds ] <- "turquoise"</pre>

ggplot(res) +
```

```
ggplot(res) +
  aes(res$log2FoldChange, -log(res$padj)) +
  geom_point(col = mycols, alpha = 0.5) +
  geom_vline(xintercept = c(-2,2), linetype = 'dashed', color = 'black') +
  geom_hline(yintercept = 0.01, linetype = 'dashed', color = 'black')
```

Warning: Removed 1237 rows containing missing values or values outside the scale range (`geom_point()`) .



Gene annotation:

```
library(AnnotationDbi)
library(org.Hs.eg.db)
```

```
columns(org.Hs.eg.db)
```

```
[1] "ACCNUM"
                    "ALIAS"
                                    "ENSEMBL"
                                                   "ENSEMBLPROT"
                                                                  "ENSEMBLTRANS"
 [6] "ENTREZID"
                    "ENZYME"
                                   "EVIDENCE"
                                                   "EVIDENCEALL"
                                                                  "GENENAME"
                                    "GOALL"
                                                   "IPI"
                                                                  "MAP"
[11] "GENETYPE"
                    "GO"
[16] "OMIM"
                    "ONTOLOGY"
                                   "ONTOLOGYALL"
                                                   "PATH"
                                                                  "PFAM"
[21] "PMID"
                    "PROSITE"
                                   "REFSEQ"
                                                                  "UCSCKG"
                                                   "SYMBOL"
[26] "UNIPROT"
```

Q4. Use the mapIDs() function multiple times to add SYMBOL, ENTREZID and GENENAME annotation to our results by completing the code below.

Add gene SYMBOL, GENENAME, and ENTREZID:

```
res$symbol <- mapIds(org.Hs.eg.db,
    keys = rownames(res),
    keytype = 'ENSEMBL',
    column = 'SYMBOL')</pre>
```

'select()' returned 1:many mapping between keys and columns

```
res$genename <- mapIds(org.Hs.eg.db,
    keys = rownames(res),
    keytype = 'ENSEMBL',
    column = 'GENENAME')</pre>
```

'select()' returned 1:many mapping between keys and columns

```
res$entrez <- mapIds(org.Hs.eg.db,
    keys = rownames(res),
    keytype = 'ENSEMBL',
    column = 'ENTREZID')</pre>
```

head(res)

log2 fold change (MLE): condition hoxa1 kd vs control sirna Wald test p-value: condition hoxa1 kd vs control sirna DataFrame with 6 rows and 9 columns

```
baseMean log2FoldChange
                                             lfcSE
                                                          stat
                                                                    pvalue
                <numeric>
                               <numeric> <numeric> <numeric>
                                                                 <numeric>
ENSG00000279457
                  29.9136
                               0.1792571 0.3248216
                                                      0.551863 5.81042e-01
ENSG00000187634 183.2296
                               0.4264571 0.1402658
                                                      3.040350 2.36304e-03
ENSG00000188976 1651.1881
                              -0.6927205 0.0548465 -12.630158 1.43989e-36
ENSG00000187961
                 209.6379
                               0.7297556 0.1318599
                                                      5.534326 3.12428e-08
ENSG00000187583
                  47.2551
                               0.0405765 0.2718928
                                                      0.149237 8.81366e-01
ENSG00000187642
                  11.9798
                               0.5428105 0.5215599
                                                      1.040744 2.97994e-01
                       padj
                                 symbol
                                                       genename
                                                                     entrez
                  <numeric> <character>
                                                    <character> <character>
ENSG00000279457 6.86555e-01
                                                             NΑ
ENSG00000187634 5.15718e-03
                                 SAMD11 sterile alpha motif ..
                                                                     148398
                                  NOC2L NOC2 like nucleolar ..
ENSG00000188976 1.76549e-35
                                                                      26155
ENSG00000187961 1.13413e-07
                                 KLHL17 kelch like family me..
                                                                     339451
ENSG00000187583 9.19031e-01
                                PLEKHN1 pleckstrin homology ...
                                                                      84069
ENSG00000187642 4.03379e-01
                                  PERM1 PPARGC1 and ESRR ind..
                                                                      84808
```

Q5. Finally for this section let's reorder these results by adjusted p-value and save them to a CSV file in your current project directory.

```
res = res[order(res$pvalue),]
write.csv(res, file = "deseq_results.csv")
```

Pathway analysis:

```
library(gage)
```

```
library(gageData)
library(pathview)
```

Pathview is an open source software package distributed under GNU General Public License version 3 (GPLv3). Details of GPLv3 is available at http://www.gnu.org/licenses/gpl-3.0.html. Particullary, users are required to formally cite the original Pathview paper (not just mention it) in publications or products. For details, do citation("pathview") within R.

The pathview downloads and uses KEGG data. Non-academic uses may require a KEGG license agreement (details at http://www.kegg.jp/kegg/legal.html).

Load up the kegg gene sets:

```
data(kegg.sets.hs)
head(kegg.sets.hs, 2)
$`hsa00232 Caffeine metabolism`
[1] "10"
          "1544" "1548" "1549" "1553" "7498" "9"
$`hsa00983 Drug metabolism - other enzymes`
 [1] "10"
           "1066" "10720" "10941" "151531" "1548"
                                                        "1549"
                                                                "1551"
             "1576"
 [9] "1553"
                                      "1807"
                     "1577"
                              "1806"
                                               "1890"
                                                        "221223" "2990"
[17] "3251"
             "3614" "3615"
                              "3704"
                                      "51733" "54490"
                                                       "54575"
                                                                "54576"
[25] "54577" "54578" "54579" "54600" "54657" "54658" "54659"
                                                                "54963"
[33] "574537" "64816" "7083"
                              "7084"
                                      "7172"
                                               "7363"
                                                        "7364"
                                                                "7365"
[41] "7366"
             "7367"
                     "7371"
                              "7372"
                                      "7378"
                                               "7498"
                                                       "79799"
                                                                "83549"
[49] "8824"
             "8833"
                     "9"
                              "978"
```

```
foldchanges <- res$log2FoldChange
names(foldchanges) <- res$entrez</pre>
```

```
keggres <- gage(foldchanges, gsets = kegg.sets.hs)</pre>
```

```
head(keggres$less)
```

```
p.geomean stat.mean
hsa04110 Cell cycle 8.995727e-06 -4.378644
hsa03030 DNA replication 9.424076e-05 -3.951803
hsa05130 Pathogenic Escherichia coli infection 1.405864e-04 -3.765330
```

hsa03013	RNA transport		1.375901e-03	-3.028500
hsa03440	Homologous recombination		3.066756e-03	-2.852899
hsa04114	Oocyte meiosis		3.784520e-03	-2.698128
			p.val	q.val
hsa04110	Cell cycle		8.995727e-06	0.001889103
hsa03030	DNA replication		9.424076e-05	0.009841047
hsa05130	Pathogenic Escherichia coli i	nfection	1.405864e-04	0.009841047
hsa03013	RNA transport		1.375901e-03	0.072234819
hsa03440	Homologous recombination		3.066756e-03	0.128803765
hsa04114	Oocyte meiosis		3.784520e-03	0.132458191
			set.size	exp1
hsa04110	Cell cycle		121 8.99	95727e-06
hsa03030	DNA replication		36 9.42	24076e-05
hsa05130	Pathogenic Escherichia coli i	nfection	53 1.40	05864e-04
hsa03013	RNA transport		144 1.3	75901e-03
hsa03440	Homologous recombination		28 3.00	66756e-03
hsa04114	Oocyte meiosis		102 3.78	34520e-03

Cell cycle figure:

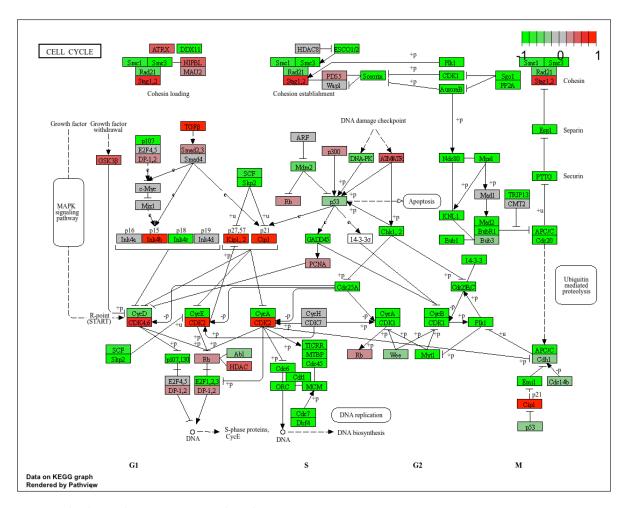
```
pathview(foldchanges, pathway.id = 'hsa04110')
```

'select()' returned 1:1 mapping between keys and columns

Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

Info: Writing image file hsa04110.pathview.png

Insert this figure in my report



We can look at the top 5 upregulated genes:

```
keggrespathways <- rownames(keggres$greater)[1:5]
keggresids = substr(keggrespathways, start=1, stop=8)
keggresids</pre>
```

[1] "hsa04060" "hsa05323" "hsa05146" "hsa05332" "hsa04640"

```
pathview(gene.data = foldchanges, pathway.id = keggresids, species = "hsa")
```

^{&#}x27;select()' returned 1:1 mapping between keys and columns

Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

Info: Writing image file hsa04060.pathview.png

'select()' returned 1:1 mapping between keys and columns

Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

Info: Writing image file hsa05323.pathview.png

'select()' returned 1:1 mapping between keys and columns

Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

Info: Writing image file hsa05146.pathview.png

'select()' returned 1:1 mapping between keys and columns

Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

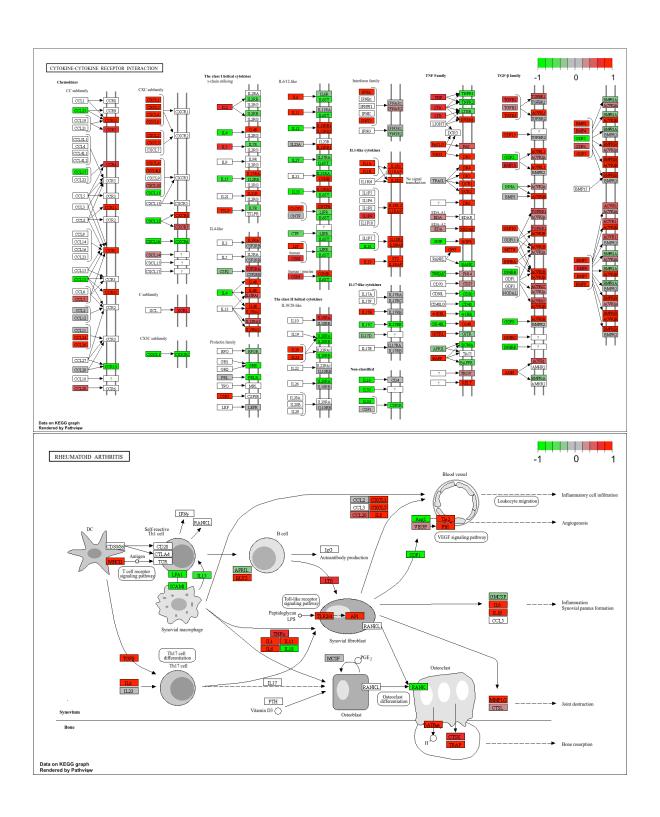
Info: Writing image file hsa05332.pathview.png

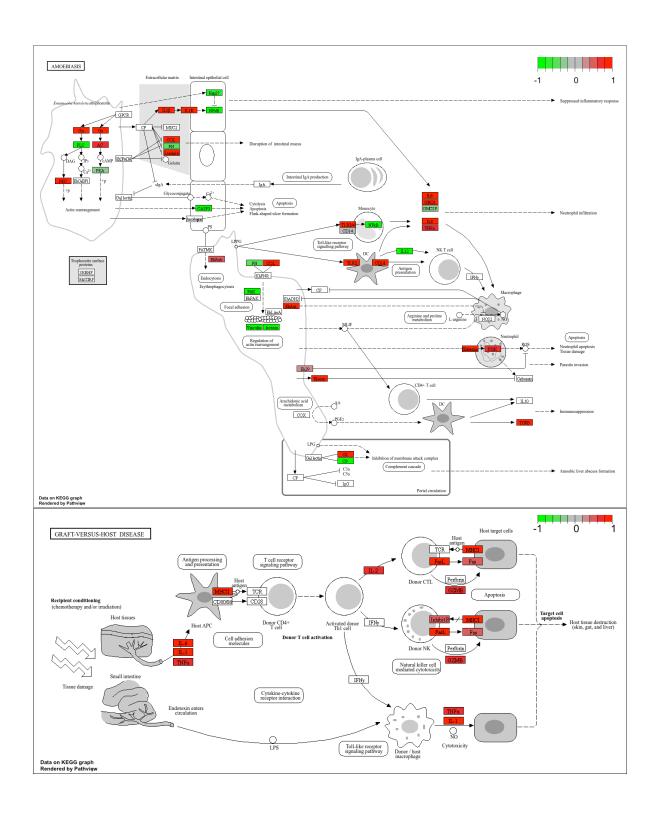
'select()' returned 1:1 mapping between keys and columns

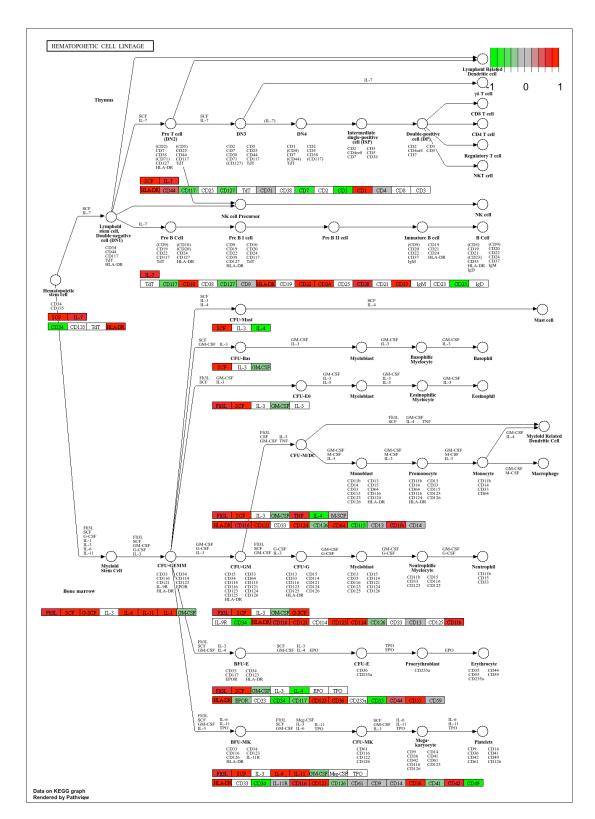
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

Info: Writing image file hsa04640.pathview.png

Below are illustrations of the top 5 upregulated genes:







Can you do the same procedure as above to plot the pathview figures for the top 5 down-reguled pathways?

Let us also observe the top 5 down-regulated genes:

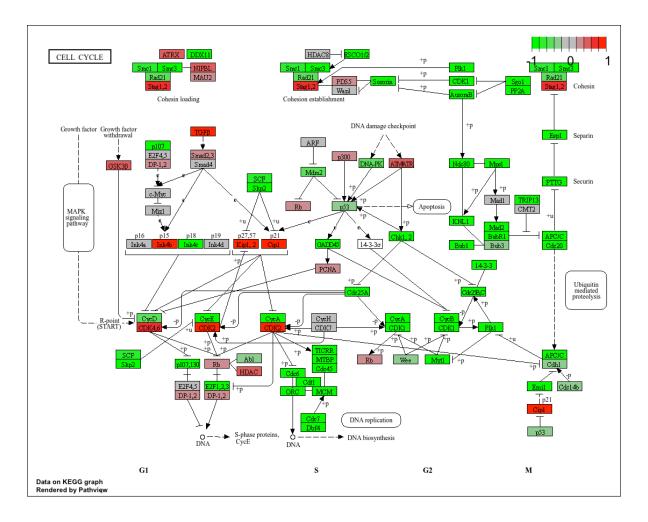
```
keggrespathways_down <- rownames(keggres$less)[1:5]</pre>
keggresids_down = substr(keggrespathways_down, start=1, stop=8)
keggresids_down
[1] "hsa04110" "hsa03030" "hsa05130" "hsa03013" "hsa03440"
pathview(gene.data = foldchanges, pathway.id = keggresids_down, species="hsa")
'select()' returned 1:1 mapping between keys and columns
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14
Info: Writing image file hsa04110.pathview.png
'select()' returned 1:1 mapping between keys and columns
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14
Info: Writing image file hsa03030.pathview.png
'select()' returned 1:1 mapping between keys and columns
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14
Info: Writing image file hsa05130.pathview.png
'select()' returned 1:1 mapping between keys and columns
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14
```

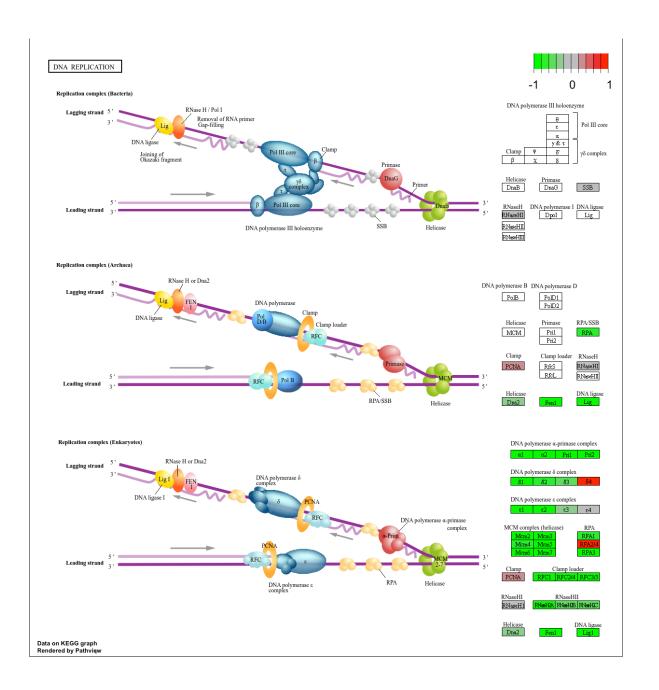
Info: Writing image file hsa03013.pathview.png

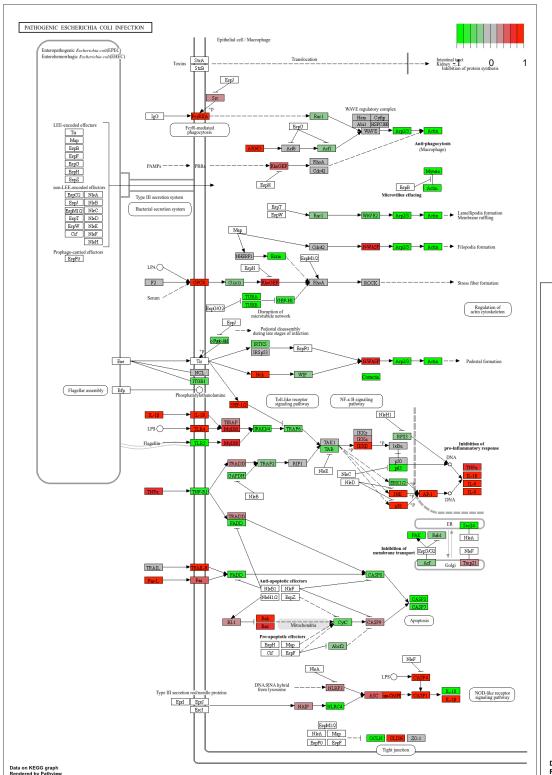
'select()' returned 1:1 mapping between keys and columns

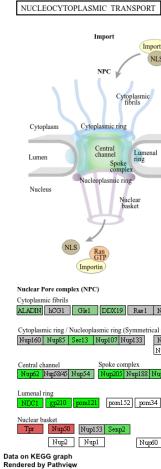
Info: Working in directory /Users/snehita/Desktop/bimm 143/class14

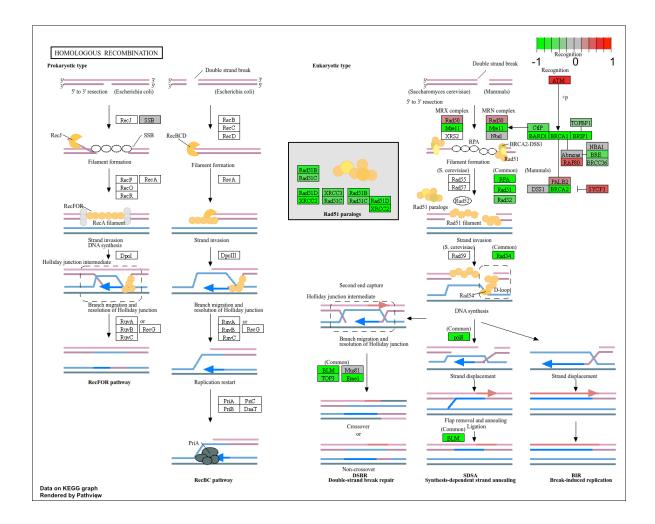
Info: Writing image file hsa03440.pathview.png











Run gene ontology

```
data(go.sets.hs)
data(go.subs.hs)

# Let us focus on just the biological processes of GO
gobpsets = go.sets.hs[go.subs.hs$BP]

gobpres = gage(foldchanges, gsets=gobpsets, same.dir=TRUE)
head(gobpres$less)
```

p.geomean stat.mean

p.val

```
GO:0048285 organelle fission
                                        1.536227e-15 -8.063910 1.536227e-15
GO:0000280 nuclear division
                                        4.286961e-15 -7.939217 4.286961e-15
GD:0007067 mitosis
                                        4.286961e-15 -7.939217 4.286961e-15
GO:0000087 M phase of mitotic cell cycle 1.169934e-14 -7.797496 1.169934e-14
GO:0007059 chromosome segregation
                                        2.028624e-11 -6.878340 2.028624e-11
GO:0000236 mitotic prometaphase
                                        1.729553e-10 -6.695966 1.729553e-10
                                               q.val set.size
GO:0048285 organelle fission
                                        5.841698e-12
                                                         376 1.536227e-15
GO:0000280 nuclear division
                                        5.841698e-12
                                                         352 4.286961e-15
GO:0007067 mitosis
                                        5.841698e-12
                                                         352 4.286961e-15
GO:0000087 M phase of mitotic cell cycle 1.195672e-11
                                                         362 1.169934e-14
GO:0007059 chromosome segregation
                                        1.658603e-08
                                                         142 2.028624e-11
GO:0000236 mitotic prometaphase
                                        1.178402e-07
                                                          84 1.729553e-10
```