

Breedbase User Manual

Breedbase team

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Introduction

Welcome to the Breedbase manual!

This manual is intended for database users.

If you are a developer looking for software implementation details, please visit the developer wiki instead: <https://github.com/solgenomics/sgn/wiki>

Chapter 1

Basic Website Usage

Breedbase is usually hosted on the cloud and is entirely web-based, with only a browser required to access and use it. The recommended browser is Firefox. To use Breedbase, you or your project will usually need to have your own Breedbase instance. Information how you can obtain your own instance can be obtained from the Breedbase project (<https://breedbase.org/>).

Once an instance is set up, the site needs to be configured and some metadata uploaded before you can design, run and analyze trials:

- define user accounts and their access privileges.
- add a trait ontology with the traits that you require.
- add the locations that you use in your breeding program
- add the foundational germplasm and the respective pedigree data
- “historical” trial data can be uploaded as needed. Usually more recent trials are prioritized over older trials

In this chapter, we will cover how you can manage the user accounts and some basic website features such as lists and how to navigate the menus. The other topics are covered in subsequent chapters. Note that Breedbase instances are highly customizable, and that not every instance of Breedbase will have the same options in the same location. Refer to the site specific documentation, if any, for site specific information.

1.1 Creating a User Account

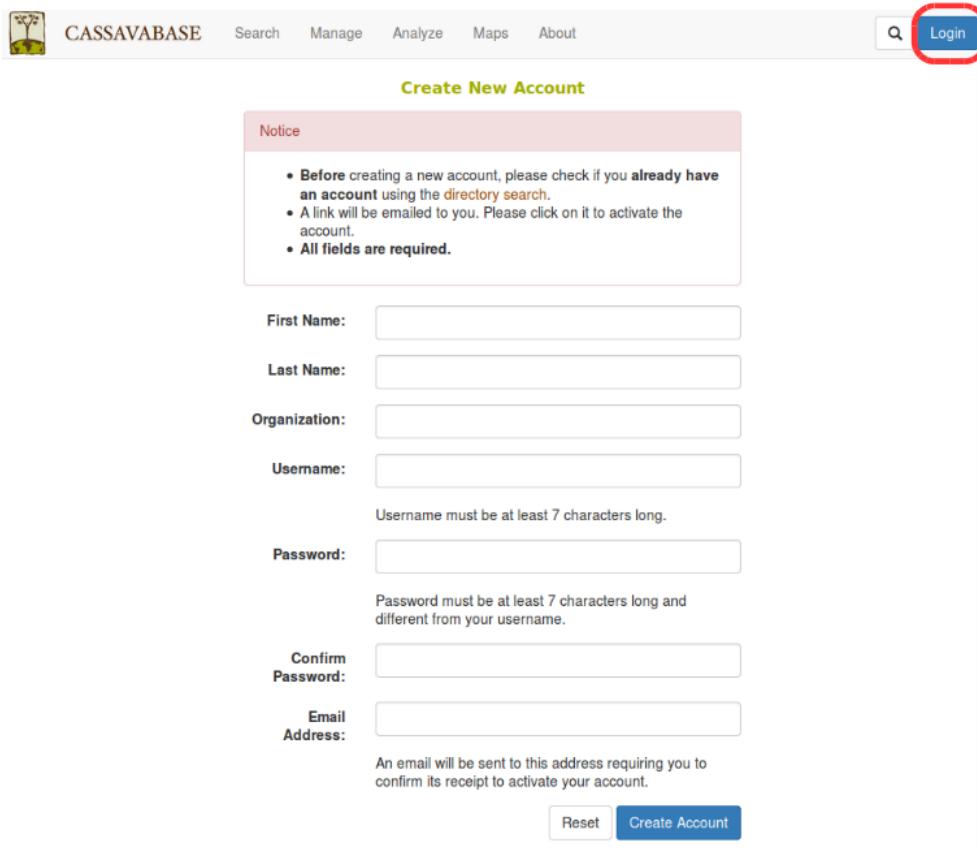
1.1.1 Verifying first that you do not already have an account

Before creating an account, please verify first that you don't already have an account. You can use "Search" menu to check if you already registered as a user.

In the "Search" menu, selecting the "People" tab and search your name. If nothing is found, proceed with the instructions below. Otherwise, continue by clicking the "Login" button. If you have forgotten your password, you can retrieve it by clicking the "Forgot your password?" link on the login page.

1.1.2 Creating a user account

On the right of the toolbar, click on "Login", which will take you to the login dialog. On the login dialog, click on the link "sign up for an account." It will take you to the page below:



The screenshot shows the 'Create New Account' page of the CASSAVABASE website. At the top, there is a navigation bar with links for Search, Manage, Analyze, Maps, and About. To the right of the navigation bar is a search icon and a 'Login' button, which is highlighted with a red circle. Below the navigation bar, the page title 'Create New Account' is displayed in green. A 'Notice' box contains instructions: 'Before creating a new account, please check if you already have an account using the directory search.', 'A link will be emailed to you. Please click on it to activate the account.', and 'All fields are required.' Below the notice, there are input fields for First Name, Last Name, Organization, Username, Password, Confirm Password, and Email Address. Each input field has a corresponding validation message below it. At the bottom of the form are 'Reset' and 'Create Account' buttons.

Filling in all of the information, then click “Create Account.”

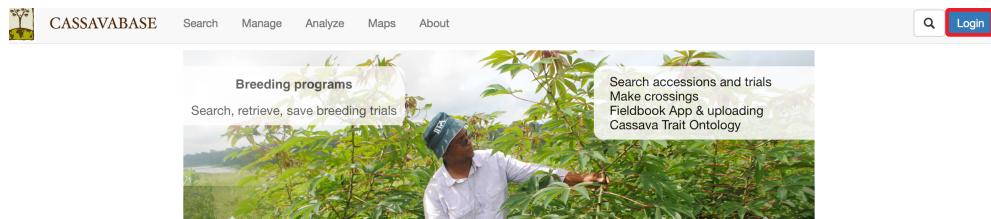
After you submit the information, an email will be sent to the provided email address. Check your email and click on the link to activate your account.

1.2 Managing your Account

1.2.1 Login

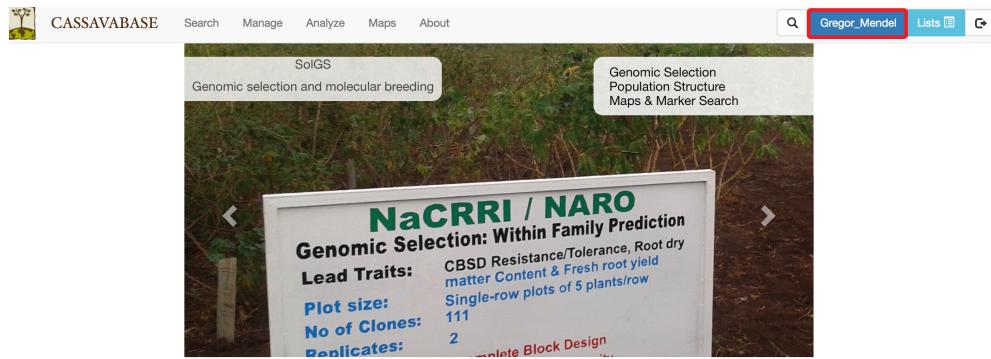
To login, click the “Login” link in the toolbar on any page and enter your username and password.

If you have forgotten your password, you can retrieve it by clicking the “Forgot your password?” link on the login page.



1.2.2 Editing Account Settings

Account settings can be edited by clicking on the “my profile” link displayed as your user name, on the right of the toolbar. You must be logged in order to access and change account settings.



You can add personal information to your account using the “View or update personal information” link.

To change your password, username, or your contact email, click on “Update account information” link. You must provide your old password before you can make any changes.

The screenshot shows a user profile page for 'Gregor Mendel' on the CASSAVABASE platform. At the top, there's a navigation bar with links for Search, Manage, Analyze, Maps, and About. On the right, there are search, user info ('Gregor_Mendel'), lists, and a refresh icon. Below the header, it says 'Welcome Gregor Mendel' and 'Not Gregor Mendel? [log out]'. A red box highlights the 'General Tools' section, which includes links for viewing or updating personal information, updating account info, and posting to the SGN forum. Other sections shown include 'QTL data submission' (upload and analyse QTL data), 'soi6S submitted analysis jobs' (no submitted jobs), 'Loci with Editor Privileges' (none), 'Annotated Loci' (view annotated loci by date), and 'User Status' (current status is 'submitter').

1.2.3 Changing Your Account Status: From “User” to “Submitter”

After you create an account, your account has a “user” status. This account has limited privileges.

Accounts with “user” status are able to:

- Change personal information
- Post comments on pages
- Post to the forum

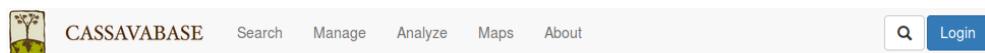
To upgrade your account status to “submitter,” contact the database curators using the “contact” link provided at the footer of each page. Submitter accounts can add data, such as new plots, accessions, phenotypic data and images.

1.2.4 Submitting Feedback on an SGN Database

We appreciate your feedback! Feel free to submit any questions or suggestions by using the “Feedback” link provided at the footer of each page.

1.3 Menu Layout

SGN Database websites have a toolbar on the top of each page with a number of menus for convenient access of major functions. The menus, as pictured below, are “search,” “manage,” “analyze,” and “maps.” The toolbar also provides a quick search, a “log in” button, and a “new user” button. The menus can be customized for each instance, so they may not appear exactly as shown on your instance; for example, most instances do not have a map menu.



1.3.1 Menu Options

Search

In the Search menu, the options are:

Tab	Description
Wizard	Search different accessions and plots by location, year, trial, and trait data. Can also be used to create lists of different types.
Accession and plots	Search accessions and plots using a variety of criteria
Trials	Search trials by name, description, breeding program, year, location, and trial type.
Markers	Search different markers
Images	Search images contained in the SGN database
People	Search database users

Manage

In the Manage menu, the options are:

Tab	Description
Breeding Programs	View, add and delete breeding programs

Tab	Description
Locations	View, add and delete locations
Accessions	Manage and search different accessions
Seedlots	Manage and search different seedlots
Crosses	Create new crosses in the database
Field Trials	Manage field trials. Create trials using different field layouts.
Genotyping Plates	Manage genotyping plates. Create 96 or 384 well plates.
Phenotyping	Upload phenotyping files from the Tablet Field Book application
Field Book App	Manage the field book app data (download files to tablet)
Barcodes	Refers to the old barcode system, mainly historical
Download	Download information in the database based on lists

Analyze

Clicking on the “Analyze” link will give a full menu of all analysis functions

In the Analyze menu, the options are:

Tab	Description
Breeder Tools	
Breeder Home	Access breeding functionalities. Lists important and helpful links.
Barcode Tools	Manage, create, and download barcodes. Also access barcode tools.
Genomic Selection	Can search for traits, start building a GS model, and predict values based on genotypes
Sequence Analysis	
BLAST	Sequence homology search
Other	
Ontology Browser	Browse all recorded ontologies

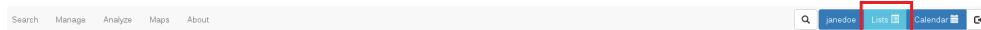
1.4 Working with Lists

Lists are collections of identifiers that are stored in the database. Lists can be composed of accessions, plots, traits, locations, and trials. A given list can only contain one type of items, for example, all items have to be of type location in a list of locations. Lists are attached to the individual user's account, and can only be created and seen by the user while logged in, however, lists can be made public for all other users to see. SGN databases make heavy use of lists in a number of tools on the website. For example, trials are created using lists of accessions.

1.4.1 Creating lists

Lists can be generated in various ways:

One way to create a list is by clicking on the “Lists” link located on the toolbar.



To create a new list, enter the name of your new list and then click on the “New List” button. The name of the list can be anything, but should be unique and should be something to help you easily identify the list.

List Name	Count	Type	View	Delete	Download	Share	Group
acc88	1	accessions					
acc_vk_1	1	accessions					
accessions_for_solsg_tests	374	accessions					
accessions_for_trial2	307	genotyping_trials					
desynonymize_test_list	6	accessions					
geno_trial	1	genotyping_trials					
janedoe_1_private	2	null					
janedoe_1_public	2	null					
m1	1	accessions					
m2	1	accessions					

If the list already exists, it will appear on the “Your Lists” dialog. To add items to your list, click on the “View” icon to open the “List Contents”

section.

ListName	Count	Type	View	Delete	Download	Share	Group
acc88	1	accessions					
acc_wk_1	1	accessions					
accessions_for_sols_tests	374	accessions					
accessions_for_trial2	307	genotyping_trials					
desynonymize_test_list	6	accessions					
geno_trial	1	genotyping_trials					
janedoe_1_private	2	null					
janedoe_1_public	2	null					
m1	1	accessions					
m2	1	accessions					

Showing 1 to 10 of 20 entries

Previous 1 2 Next

[View Public Lists](#) [Close](#)

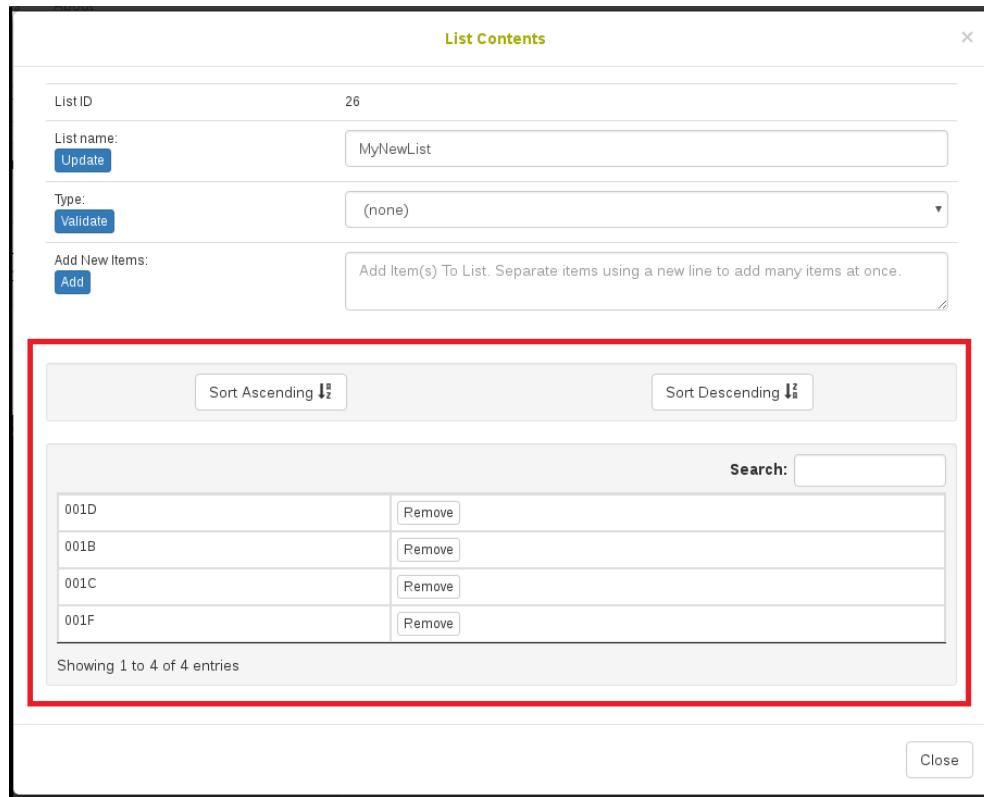
On the “List Contents” page, enter items that you want to add to the list, then click on “Add” button.

List ID	26		
List name:	<input type="text" value="MyNewList"/>		
Type:	(none)		
Add New Items:	<input type="button" value="Add"/>		
<table border="1"> <tr> <td>001D</td> </tr> <tr> <td>001B</td> </tr> </table>		001D	001B
001D			
001B			
<input type="button" value="Sort Ascending"/> <input type="button" value="Sort Descending"/>			
<input type="text" value="Search:"/> <input type="button" value="Search"/>			
No data available in table			
Showing 0 to 0 of 0 entries			

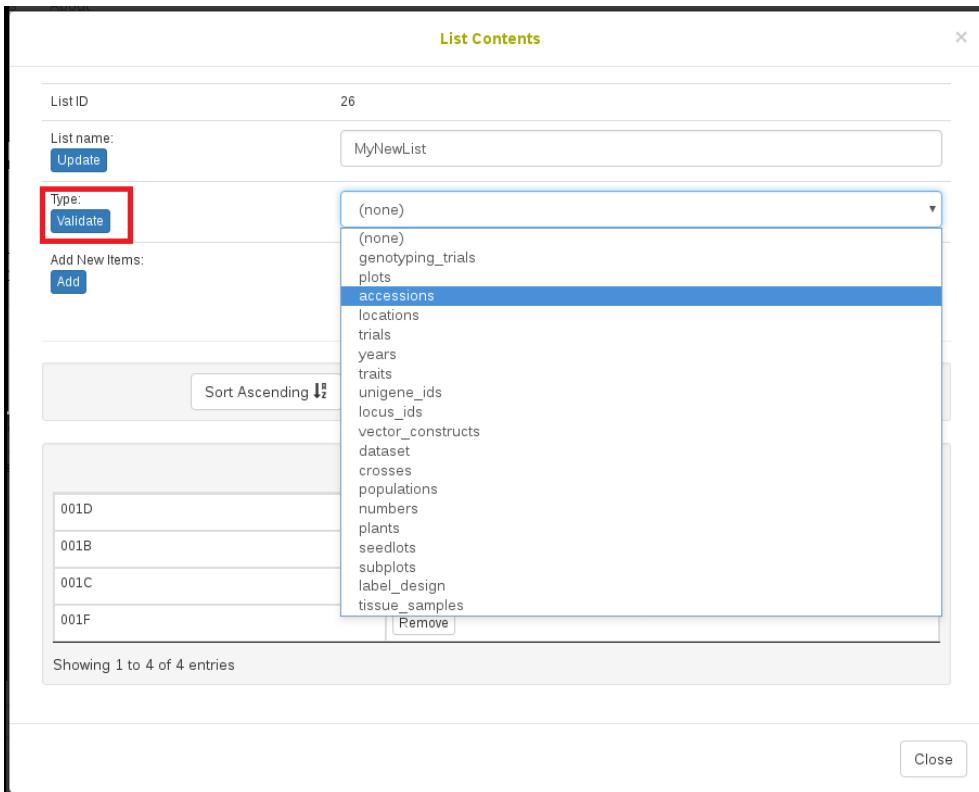
[Close](#)

The page will be updated and will display your items in a table at the bottom

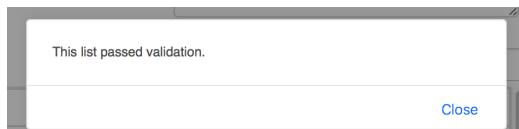
of the dialog It is possible to sort the list if you need.



Select the type of items in your list. To verify that the items that you added to your list are already stored in the database and that you selected a correct type for the items, click on the “Validate” button.



If those items are already in the database, a message will indicate that “This list passed validation”



Note that a list cannot contain duplicate elements. If a duplicate item is entered, the list manager will inform the user that the element is already in the list and will not add it again.

Another easy way to create a list is to use the [2.1](#), which can be accessed from the Search menu.

1.4.2 Viewing and editing lists

Lists can be viewed and edited using the “Lists” link on the toolbar. Clicking on the link will open a dialog that displays all of your lists, as well as an option to create new lists.

List Name	Count	Type	Actions
IITA_WKSHP_D2	20	acquisitions	
IITAwksp16_acquisitions_list	24	acquisitions	
new_acquisition_list	6	acquisitions	

New List

View Public Lists **Close**

This page shows all lists that have been created, including those created by using the Search Wizard. You can view and edit your lists by using “Actions” buttons.

1. Clicking on the “view” icon will open a new window called “List Contents” that allows you to change the list name, the type of the list, add new items, or delete existing items.
2. Clicking on the “delete” icon will delete your list. **Caution: this action cannot be undone.**
3. Clicking on the “download” icon will download the contents of your list to your computer.
4. Clicking on the “make public” icon will make your list available for other users to view and use your list.

1 2 3 4

1.5 User Permissions

Breedbase accounts are assigned one or more of four different roles to determine the level of access they have within the database. The possible roles

are **User**, **Submitter**, **Sequencer**, and **Curator**. Each role grants specific permissions, and careful management of them helps prevent data from being altered or deleted in error.



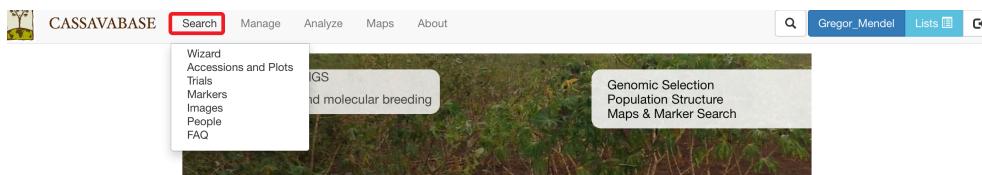
Accounts are also assigned Breeding Program role(s) to grant access to the specific breeding program(s) they work with.

- The **User** role gives an account permission to view and download data throughout the database.
- The **Submitter** role gives an account permission to design field experiments and to upload and edit data using the tools in the “Manage” section. In order to submit and manage breeding data within a given breeding program, a submitter also must have a matching Breeding Program role.
- The **Sequencer** role gives an account permission to design genotyping experiments and submit plates to a genotyping service.
- The **Curator** role gives an account permission to do all of the above, as well as to delete data within the database. The Curator role also enables the addition or deletion of roles for all database accounts in the ‘Manage User Roles’ tool.

Chapter 2

Searching the Database

You can search for information on the database by using the following search options: Wizard, which uses combined criteria specified by users; Accessions and Plots; Trials; Markers; Images; People; FAQ.



2.1 The Search Wizard

The screenshot shows the 'Search Wizard' interface. At the top right are three buttons: 'Don't see your data?', 'Refresh Lists', and 'Update Wizard'. Below this are four identical search panels, each consisting of a 'Select Column Type' dropdown, a 'Search' input field, and a 'Select All' button with '0/0' and 'Clear' options. Below these panels is a section titled 'Load/Create Datasets using' with 'Match' and 'Columns' dropdowns. It includes 'Load Dataset' and 'Create New Dataset' buttons, and a 'Load' button. To the right are three sections: 'Related Genotype Data', 'Related Trial Metadata', and 'Related Trial Phenotypes'.

2.1.1 How the Search Wizard Works

The search wizard presents a number of select boxes, which are initially empty. You start searching by picking a category of data from the dropdown above the left-most select box.

Once a category has been picked, the database will retrieve all the options within this category and display them within the first select box. You then select one or more options from the first select box, which activates the second dropdown.

You can then select a category from the second dropdown, and repeat this same search process through all four dropdowns and select boxes.

The screenshot shows the 'Search Wizard' interface with three main categories: Locations, Years, and Accessions. Each category has a search bar at the top, followed by a list of items with checkboxes. The 'Locations' category shows 'Ibadan' selected. The 'Years' category shows '2011' and '2012' selected. The 'Accessions' category shows several accession numbers selected.

Category	Selected Items
Locations	Ibadan
Years	2011, 2012
Accessions	20_20, 462, 50395, 58308, 79-106

- In the example above, the “locations” category was chosen in the first dropdown. The first select box then displayed all the possible locations in the database. The option Ibadan was selected.
- This activated the second dropdown. The category “years” was chosen in the second dropdown. The second select box then displayed all the years that are linked in the database to the location Ibadan. From that list, the options 2011 and 2012 were selected.
- This activated the third dropdown. A final category, “accessions”, was chosen in the third dropdown. The third select box was then populated with the 3847 accessions in the database that are linked with the location Ibadan in the years 2011 or 2012.

In addition to the basic search operations demonstrated above, users can take advantage of two more features:

Load Selection from List

Load Selection from List:

- Instead of picking a category in the first dropdown, users can instead populate the first selectbox from a list by scrolling down in the first dropdown to the “Load Selection from List” subheading and selecting a list. This is useful for starting queries with a list of plots, as this category is not among the options in the first dropdown.

ANY/MIN/ALL Toggle



- By default, the search wizard combines options within a category using an OR query. In the example above, in the third panel the wizard retrieved accessions associated with the location ‘Ibadan’ in **ANY** of the years “2011 **OR** 2012”
- If the user clicked the toggle below the second select box to change it to **ALL** before choosing accessions in the third dropdown, the wizard would instead retrieve accessions associated with the location ‘Ibadan’ in the years “2011 **AND** 2012”. This will be a smaller set of accessions, because any accessions used only in 2011, or only in 2012 will be excluded.
- A more advanced search could use the **MIN** toggle option. This allows the user to make a query in between an ANY or ALL query, where a minimum number of matches from the selected column will be used as a filter for the next column. The minimum can be provided as either a percentage (%) or an actual count of items (#). In the example above, if the years 2011, 2012, and 2013 were selected in the second column, the user could enter ‘2’ in as the minimum and select ‘#’ as the minimum match type. This would select accessions in the third column that were used in 2 or more of the selected years.

Match	ANY	MIN	ALL
>=	2	% #	

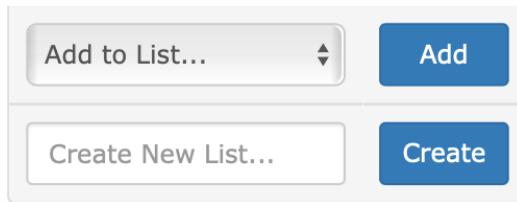
2.1.2 How to use retrieved data

Getting more Info

Any option in the wizard select boxes (except for years) can be clicked to open a page with more details. The new page is opened in a new tab.

Saving to a list

You can store the highlighted items in any selected box to lists. This is done using the inputs and buttons directly below the select box. **Don’t forget, you must be logged in to work with lists!**



- To **add items to an existing list**, first pick an existing list using the “Add to List...” dropdown on the left. Then click the “Add” button. A popup window will confirm the action, and display the number of items added to your existing list.
- To **store items to a new list**, first type a new list name in the “Create New List...” text input on the left. Then click on the “Create” button. A popup window will confirm the action, and display the number of items added to your new list.

Downloading Data

You can download trial metadata, phenotypes and genotypes associated with the highlighted items in the wizard select boxes. This is done using the buttons in the download section at the bottom of the page. **Don't forget, you must be logged in to download data!**



Metadata Trial metadata can be downloaded by selecting a subset of trials from the database or based on your search categories. To download, click on “Related Trial Metadata”, a dialog will appear. Select download format and click the “Metadata” button to complete your download.

The screenshot shows a modal window titled "Related Trial Metadata". It contains a text input field with "3 trials" and a dropdown menu set to "CSV". A blue button labeled "Metadata" is at the bottom.

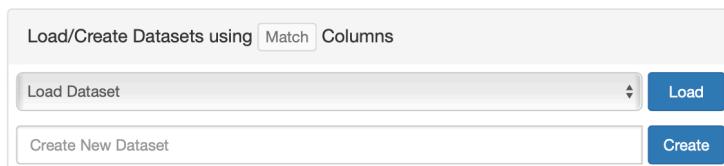
Phenotypes The phenotypes download is quite flexible, and can download a subset of all the trial data in the database based on whichever categories and options you currently have selected. Simply click on the “Related Trial Phenotypes” link, review the options, changing or adding any additional parameters you like, then click ‘Download Phenotypes’.

The screenshot shows a modal window titled "Related Trial Phenotypes". It includes a message "Too few trials", download format options ("Default", "CSV", "All"), and several checkboxes: "Include timestamps", "Include accession entry numbers", "Suppress user defined phenotype outliers", and "Repetitive measurements: Averaged value". It also features date filters for "From" (1960-01-01) and "To" (2024-12-15), a checkbox for "include items without a date", and trait name filtering ("Trait Name Contains: e.g. plant height"). A blue button labeled "Download Phenotypes" is at the bottom.

Genotypes The genotype download is more stringent. It requires a minimum of one accession and one genotyping protocol to be selected in the wizard select boxes. The text box in the download section of the page will help track what has been selected. Once clicked, the “Download Genotypes” button will download a genotype file for the selected accessions.

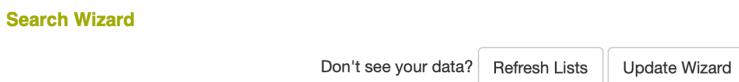
Saving the wizard selections

As discussed above, the selections of the individual select boxes in the wizard can be saved separately to a list. The lists can be used as inputs in other tools on the site. However, sometimes creating a selection is quite time consuming and restoring the selections from four different lists would be cumbersome too. Therefore, the selections can be saved together in a dataset, and named for later retrieval. This is done in the section “Load/Create Datasets” that is below the first two wizard select boxes. To select an existing dataset, one uses the “Load Dataset” dropdown. A particular dataset can be chosen, and the “Load” button can be clicked to retrieve and display the dataset in the wizard. To create a new dataset using items that are selected in the wizard, one can enter the name of the new dataset in the “Create New Dataset” text box. Once the dataset has been given a name, clicking the “Create” button will save the new dataset.



2.1.3 Updating the Wizard

The search wizard uses a copy of the database, or a cache, to return results quickly. If data appears to be missing, it usually means that the cache needs to be updated. Users with submitter privileges or above can do this using the ‘Update Wizard’ button. One can also use the ‘Refresh Lists’ button to update the available lists.



This will take just a few seconds in small databases, but may take a few hours to complete in larger databases.

2.2 Accessions and Plot Search

Accessions and their related materials (cross, plant, plot, population, tissue_sample, training population) can be searched by using “Search Accessions and Plots” page. On this page, “accession” is the default stock type; however, you can change stock type by selecting an option from the dropdown list. From this page you can construct detailed queries for stock types. For example, by using the “Usage” section, the “Properties” section, and the “Phenotypes” section you could search for accessions which were diploids used in a specific year and location and were also phenotyped for height. You can also search for accessions based on genetic properties, such as the location of an introgression on a specific chromosome.

The screenshot shows the 'Search Accessions and Plots' interface. At the top, there is a search bar with a dropdown menu set to 'Uniquename'. Below the search bar are three sections: 'Properties', 'Usage', and 'Phenotypes', each with a radio button. A 'Search' button is located at the bottom of this section. The main area is titled 'Search Results' and contains a table with columns: Stock Name, Stock Type, Organism, Synonyms, Owners, and Organization. The table lists 10 entries, including 'BLANK', various accession numbers, and test entries like 'new_acc_ppp001'. The table includes a 'View Another Property' dropdown and an 'Add' button. At the bottom of the table, it says 'Showing 1 to 10 of 482 entries' and has a navigation bar with pages 1 through 50 and a 'Next' button.

Stock Name	Stock Type	Organism	Synonyms	Owners	Organization
BLANK	accession	Manihot esculenta		John Doe	
IITA-TMS-IBA011412	accession	Manihot esculenta		John Doe	
IITA-TMS-IBA30572	accession	Manihot esculenta		John Doe	
IITA-TMS-IBA880002	accession	Manihot esculenta		John Doe	
IITA-TMS-IBA890581	accession	Manihot esculenta		John Doe	bti
new_acc_ppp001	accession	Manihot esculenta	synp001	Jane Doe	
new_acc_ppp002	accession	Manihot esculenta		Jane Doe	bti
new_acc_ppp003	accession	Manihot esculenta		Jane Doe	
new_test_crossP001	accession	Solanum lycopersicum		John Doe	
new_test_crossP002	accession	Solanum lycopersicum		John Doe	

It is possible to query over any of the available properties, such as “ploidy_level”, “country of origin”, “introgression_chromosome”, etc.

Search Accessions and Plots

Search

Uniquename

Stock Name or Description: Type search here...

Properties

Stock Type: Organism:

Stock Owner: Organization:

Search By Another Property:

accession number:

country of origin:

introgression_start_position_bp:

Usage

Phenotypes

In the search result table it is possible to select any of the available properties to view.

Search Results

View Another Property:

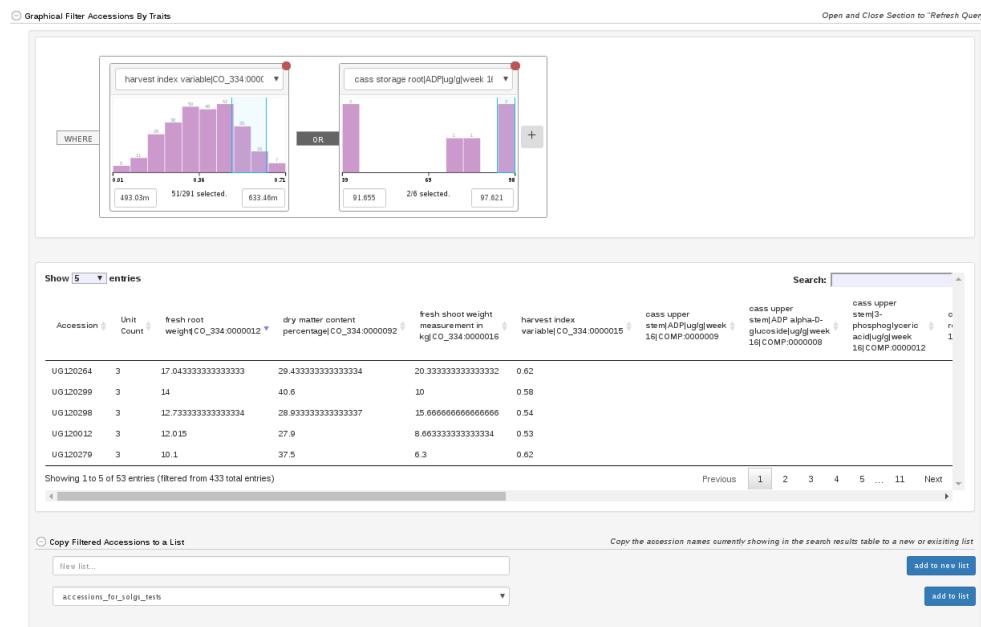
Show 10 entries

Stock Name	Stock Type	Organism	Synonyms	Owners	organization	ploidy_level
BLANK	accession					
IITA-TMS-IBA011412	accession	Manihot esculenta		John Doe		
IITA-TMS-IBA30572	accession	Manihot esculenta		John Doe		
IITA-TMS-IBA980002	accession	Manihot esculenta		John Doe		
IITA-TMS-IBA980581	accession	Manihot esculenta		John Doe	bti	
new_acc_ppp001	accession	Manihot esculenta	symp0001	Jane Doe		2
new_acc_ppp002	accession	Manihot esculenta		Jane Doe	bti	
new_acc_ppp003	accession	Manihot esculenta		Jane Doe		3
new_test_crossP001	accession	Solanum lycopersicum		John Doe		
new_test_crossP002	accession	Solanum lycopersicum		John Doe		

Showing 1 to 10 of 482 entries

Previous ... Next

At the bottom of the accession search there is a phenotype graphical filtering tool. Here you can filter down accessions based on combinations of trait performance. The filtered down accessions are then able to be saved to a list.



For information on adding Accessions please see the Managing Accessions help. For information on how field trial plots, plants, tissue samples, and subplots are added to the database, please see the Managing Field Trials help.

2.3 Trials Search

Trials on the database can be searched based on trial name, description, breeding program, year, location, trial type, design, planting date, and harvest date.

Trial Search						
Show 10 entries		Search: <input type="text"/>				
Trial name	Description	Breeding program	Folder	Year	Location	Trial type
CASS_6Genotypes_Sampling_2015	Copy of trial with postcomposed phenotypes from cassbase.	test		2017	test_location	Preliminary Yield Trial
Kasese solgs trial	This trial was loaded into the fixture to test solgs.	test		2014	test_location	Clonal Evaluation
new_test_cross	new_test_cross	test				
selection_population	selection_population			2015		
test_genotyping_project	test_genotyping_project			2015		
test_population2	test_population2			2015		
test_t	test tets	test		2016	test_location	
test_trial	test trial	test		2014	test_location	
trial2 NaCRRI	another trial for solGS	test		2014	test_location	

Showing 1 to 9 of 9 entries Previous Next

Copy Results to a List Copy the trial names currently showing in the search results table to a new or existing list

New list...

2.4 Trait Search

On the Trait Search page (menu item **Search > Traits**), traits in the database can be searched by ID, name, or description. Optionally, a starting list of traits can be selected to filter down results.

Trait Search		
Subset Traits:		Select A Subset
Show 10 entries		
Trait ID	Trait Name	Definition
<input type="checkbox"/> CO_334.0000008	sprouting proportion	Proportion of stakes germinated scored one month after planting.
<input type="checkbox"/> CO_334.0000009	initial vigor assessment 1-7	Visual assessment of plant vigor during establishment scored one month after planting.
<input type="checkbox"/> CO_334.0000010	plant stands harvested counting	A count of the number of plant stands at harvest.
<input type="checkbox"/> CO_334.0000011	root number counting	A count of the total number of storage roots harvested per plot.
<input type="checkbox"/> CO_334.0000012	f.root.weight	Total fresh weight of storage roots harvested per plot measured in kilogram (kg).
<input type="checkbox"/> CO_334.0000013	fresh root yield	Fresh weight of harvested roots expressed in tons per hectare (tha) per plant.
<input type="checkbox"/> CO_334.0000014	dry yield	Dry weight of harvested roots derived by multiplying fresh storage root yield by dry matter content expressed in tons per hectares (tha).
<input type="checkbox"/> CO_334.0000015	harvest index variable	Proportion of fresh root weight in total biomass.
<input type="checkbox"/> CO_334.0000016	fresh shoot weight measurement in kg	Total fresh weight of harvested foliage and stems in kilograms per plot.
<input type="checkbox"/> CO_334.0000017	top yield	Total fresh weight of harvested foliage and stems expressed in tons per hectare (tha).

Showing 1 to 10 of 245 entries

[Select All](#) [Deselect All](#)

Previous 1 2 3 4 5 ... 25 Next

Selecting traits in the results of the search allows one to add the selected results to a trait list, or create a new trait list from the select results.

Show 10 entries			Search:
Trait ID	Trait Name	Definition	
<input checked="" type="checkbox"/> CO_334:0000008	sprouting proportion	Proportion of stakes germinated scored one month after planting.	
<input type="checkbox"/> CO_334:0000009	initial vigor assessment 1-7	Visual assessment of plant vigor during establishment scored one month after planting.	
<input type="checkbox"/> CO_334:0000010	plant stands harvested counting	A count of the number of plant stands at harvest.	
<input type="checkbox"/> CO_334:0000011	root number counting	A count of the total number of storage roots harvested per plot.	
<input checked="" type="checkbox"/> CO_334:0000012	fresh root weight	Total fresh weight of storage roots harvested per plot measured in kilogram (kg).	
<input checked="" type="checkbox"/> CO_334:0000013	fresh root yield	Fresh weight of harvested roots expressed in tons per hectares (t/ha) per plant.	
<input type="checkbox"/> CO_334:0000014	dry yield	Dry weight of harvested roots derived by multiplying fresh storage root yield by dry matter content expressed in tons per hectares (t/ha).	
<input type="checkbox"/> CO_334:0000015	harvest index variable	Proportion of fresh root weight in total biomass.	
<input checked="" type="checkbox"/> CO_334:0000016	fresh shoot weight measurement in kg	Total fresh weight of harvested foliage and stems in kilograms per plot.	
<input type="checkbox"/> CO_334:0000017	top yield	Total fresh weight of harvested foliage and stems expressed in tons per hectare (t/ha).	

Showing 1 to 10 of 245 entries **4 rows selected**

Previous 1 2 3 4 5 ... 25 Next

[Select All](#) [Deselect All](#)

[Copy Selected Results to a List](#) Copy the trait names currently selected in the search results table to a new or existing list

4 trait(s) selected.

New list... [add to new list](#)

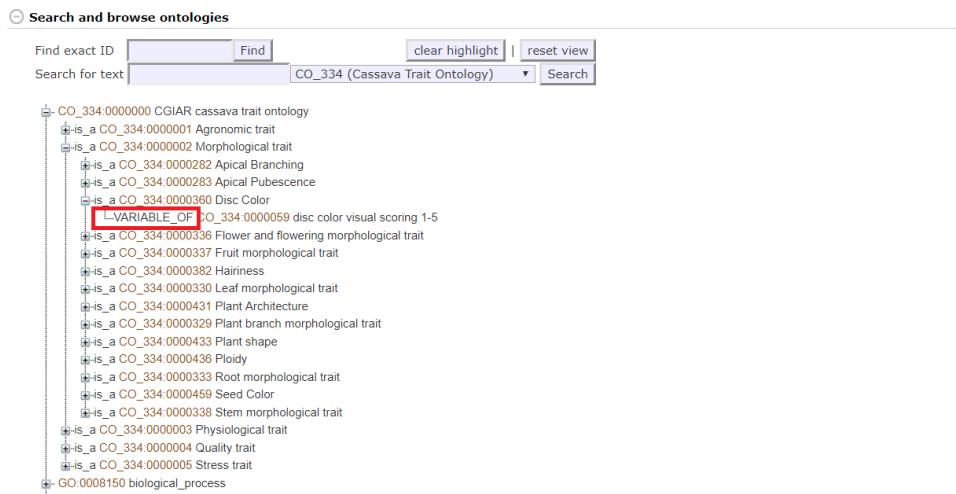
[add to list](#)

2.5 Ontology Browser

A more advanced tool for searching for Traits is the ontology browser, available by clicking on Analyze and Ontology Browser. From here you can search ontologies and see the various classifications of terms in a tree display.



The terms which appear in the Trait Search in 2.4 are only variable terms. The ontology browser shows these variables as different from their grouping terms by indicating VARIABLE_OF like in the following screenshot.



2.6 Search Seedlots

Seedlots are different from Accessions in that they represent the physical seed being evaluated in an experiment. Seedlots have things like physical storage locations and seed quantities, which accessions do not. To search for available seedlots you go to Manage and then click Seed Lots. By clicking Search Seedlots, you can specify query information. The results from your search will be in the table below the search form.

2.6. SEARCH SEEDLOTS

41

Available Seedlots

(1) Search Seedlots

Seedlot Name:	<input type="text"/>
Breeding Program:	<input type="text"/>
Contents (Accession):	<input type="text"/>
Location:	<input type="text"/>
Minimum Count:	<input type="text"/>

Search

(2) About Seedlots

What are seedlots?

- Seedlots represent physical seed in packets.
- This tool can be used to manage seed accessions.
- Seedlots can have a specific location, box, weight(g), and count.
- Seedlots can belong to breeding programs and organizations.

How do I inventory my seed?

- 1) Make sure your seedlots are in the database. Use "Add New Seedlot" to add a single seedlot or "Upload New Seedlots" to add many.
- 2) Make sure your seedlots are barcoded. You can print these barcodes from the database.
- 3) Use the "Seed Inventory" Android Application to scan seedlot barcodes and record weight. Then use "Upload Inventory" to upload this info into database. If you prefer you can create your own CSV file and upload that, if you do not want to use the Seed Inventory Application.
- **For more info about the "Seed Inventory" Android Application go to [Seed Inventory](#).**
- It is also possible to manually enter a transaction by going to the seedlot detail page and clicking "Add New Transaction".

(3) Seedlots

[Add New Seedlot]
[Upload New Seedlots]
[Upload Inventory]

Show 10 ▾ entries	Seedlot Name	Breeding Program	Contents	Seedlot Location	Count	Weight(g)	Owners	Delete
new_test_crossP001_001	test		new_test_crossP001 (accession)	NA	1		X	
new_test_crossP002_001	test		new_test_crossP002 (accession)	NA	1		X	
new_test_crossP003_001	test		new_test_crossP003 (accession)	NA	1		X	
new_test_crossP004_001	test		new_test_crossP004 (accession)	NA	1		X	
new_test_crossP005_001	test		new_test_crossP005 (accession)	NA	1	-7	X	
new_test_crossP006_001	test		new_test_crossP006 (accession)	NA	1		X	
test_accession1_001	test		test_accession1 (accession)	NA	-1	-72	X	
test_accession2_001	test		test_accession2 (accession)	NA	1		X	

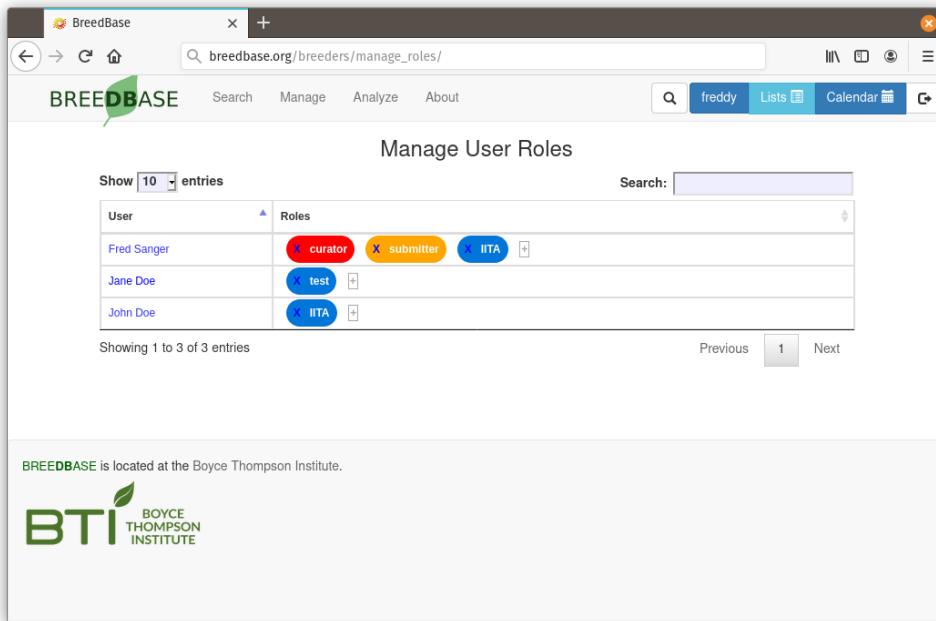
Showing 1 to 10 of 515 entries

Previous 1 2 3 4 5 ... 52 Next

<input style="width: 100%; height: 20px;" type="text"/> <input style="width: 100%; height: 20px;" type="text"/>	[add to new list] [add to list]
--	---

Chapter 3

Managing User Roles



The screenshot shows a web browser window for 'BreedBase' at the URL breedbase.org/breeders/manage_roles/. The page title is 'Manage User Roles'. It features a table with columns 'User' and 'Roles'. The table contains three entries:

User	Roles
Fred Sanger	curator (red), submitter (orange), IITA (blue)
Jane Doe	test (blue)
John Doe	IITA (blue)

Below the table, it says 'Showing 1 to 3 of 3 entries'. At the bottom left is the BTI logo: 'BTI BOYCE THOMPSON INSTITUTE'. At the bottom right are links for 'Previous' (disabled), '1', and 'Next'.

3.1 What are User Roles?

Every user account in Breedbase has one or more associated “roles” that determine the authorizations (what the user is allowed to do) in the database.

There are three fundamental roles, “curator”, “submitter”, and “user”, which determine basic read/write levels. The “curator” status can read and write everything in the database. The “submitter” status can add information and edit or delete previously submitted information. The “user” type can only read data. Additional roles represent the breeding programs, and are sometimes used to fine-tune write and edit capabilities, as it necessary for multiple users in a breeding program to edit each other’s data.

3.2 The Manage User Roles page

In the “Manage” menu, select the item “User Roles”. This will show the current users in the database with their associated roles. If you are logged in as a curator, the table will show system roles as well as breeding program roles; if you are logged in as a submitter or user, it will show breeding program membership.

If logged in as a “curator”, the roles can be added or deleted.

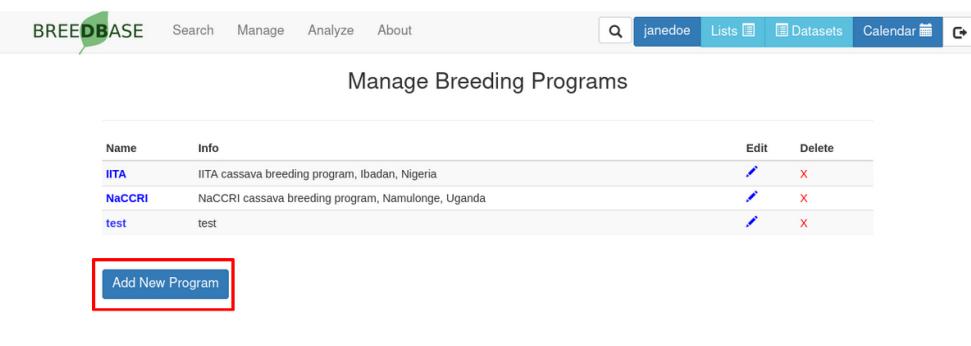
- To delete a role, click on the X in the role name. A confirm dialog will be displayed to prevent accidental deletion.
- To add a role, click on the plus sign next to the roles. A dialog will pop up with a list of roles. Select the desired role and click “Submit”.
- The new role should be displayed next to the user immediately.
- Role deletions and additions will be effective immediately.

It is recommended that few users be given the “curator” privileges to avoid confusion over data ownership and accidental data overwriting and deletion.

Chapter 4

Managing Breeding Programs

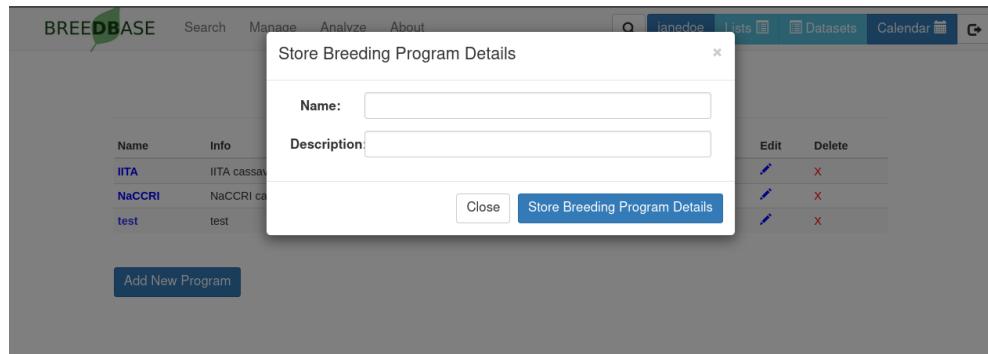
New breeding programs can be added by using “Add New Program” button on the “Manage Breeding Programs” page.



Name	Info	Edit	Delete
IITA	IITA cassava breeding program, Ibadan, Nigeria		
NaCCRI	NaCCRI cassava breeding program, Namulonge, Uganda		
test	test		

Add New Program

Clicking on the “Add New Program” button will generate a blank form for you to fill out the name and description of the breeding program that you want to add. After completing the form, click on “Add Breeding Program” button to finish the process.



Chapter 5

Managing Locations

Field locations can be managed using the “**Manage Locations**” page. On this page, locations in the database are organized based on their breeding programs. Each location has a link to trials conducted in that location. To add a new location, click on the “Upload New Locations” button that links to the “Upload Locations” form.

Location Table											[Upload New Locations]
Filter Locations by List:											<input type="text" value="select"/>
Search:											<input type="text"/>
Column visibility											
Id	Name	Abbreviation	Country	Program	Type	Lat	Long	Elevation(m)	Trials	Nearest NOAA Station ID	
3	Ibadan		NGA Nigeria	IITA & BTI & Cornell & NRCRI	Field	7.3775353	3.9470396		1557 trials		
4	Jos		NG Nigeria	IITA		9.896527	8.858331		26 trials		
5	Ubiaja	UB	NGA Nigeria	NRCRI & IITA	Field	6.6493306	6.3917656		346 trials		
6	Zaria	ZA	NGA Nigeria	IITA	Field	11.085541	7.7199454		203 trials		
7	Malam Maduri		NGA Nigeria	IITA		12.561389	9.881389		65 trials		
8	Mokwa	MK	NGA Nigeria	IITA	Field	9.239425	5.3102503		419 trials		
9	Onne	ON	NGA Nigeria	IITA	Field	4.744334	7.035316		170 trials		
10	Cornell Biotech		US United States	BTI & IITA		42.45345	-76.4735		6 trials		

The “Upload Locations” describes how to build a spreadsheet with location data for upload. Name, abbreviation, country code, country name, program, type, latitude, longitude, and elevation are all required. The NOAA station ID is optional. Link a spreadsheet to the form and click “Upload” to add

those locations to the database.

Upload Locations

Header:
The first row (header) must contain the following:

Name	Abbreviation	Country Code	Country Name	Program	Type	Latitude	Longitude	Elevation	NOAA Station ID
------	--------------	--------------	--------------	---------	------	----------	-----------	-----------	-----------------

Required values:

- Name:** must not conflict with an existing location name
- Abbreviation:** used in the map and in uploads, must not conflict with an existing abbreviation
- Country Code:** must be an [ISO Alpha-3 country code](#)
- Country Name:** must be an uppercase english short name from the [ISO standard](#)
- Program:** must be an existing Breeding Program in the database, or multiple breeding programs separated by '&'
- Type:** must be one of the following: Town, Farm, Field, Greenhouse, Screenhouse, Lab, Storage, Other
- Latitude:** (in degrees) must be a number between 90 and -90.
- Longitude:** (in degrees) must be a number between 180 and -180.
- Elevation:** (in meters) must be a number between -418 (Dead Sea) and 8,848 (Mt. Everest).

Optional values:

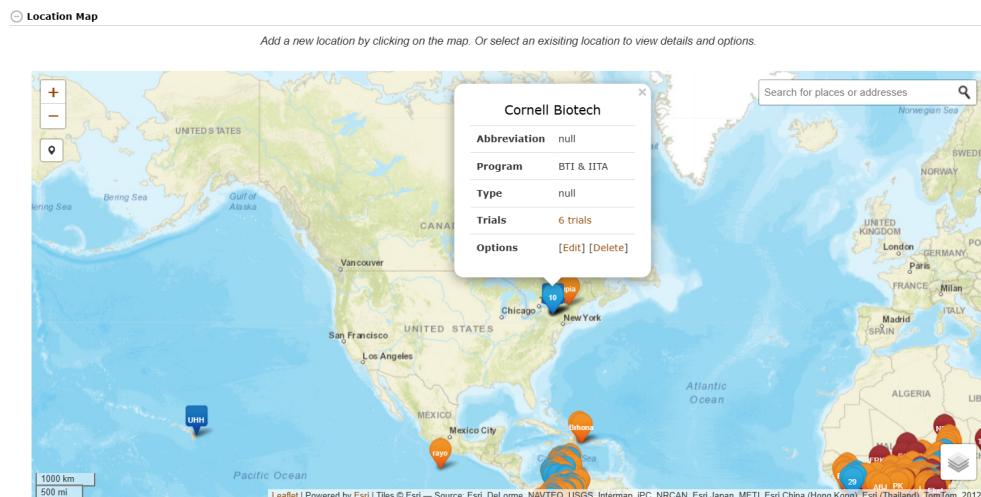
NOAA Station ID: The NOAA Station ID that is nearest and most relevant to the location. [Search Here](#)

Locations may be uploaded using any of the supported file types: MS Excel ([.xls](#) or [.xlsx](#)), comma-separated file ([.csv](#)), tab-delimited file ([.txt](#) or [.tsv](#)), or semicolon-separated file ([.ssv](#)).

Optional columns may be left out, if not used in your data.

[Close](#) [Upload](#)

Alternatively, locations can be viewed and added via the map. Hover over an icon on the map to see the location details and trials linked to that location. Click on the map to open the new location dialog. Fill in the same information that would be used in the spreadsheet upload to add a new location.



Chapter 6

Managing Accessions

The “Manage Accession” page provides links for adding new accessions. New accessions can be added to the database by either using a List or by uploading an Excel file (either XLS or XLSX format). Both options are explained in more detail below. To begin, click on the “Add Accessions or Upload Accession Info” link.

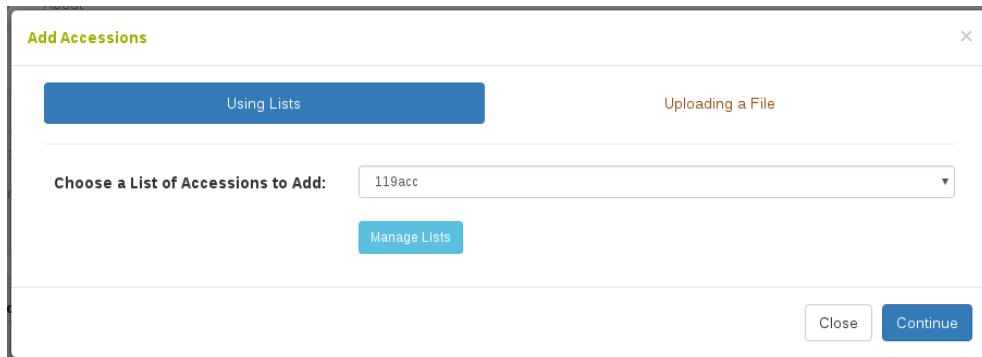
The screenshot shows the "Manage Accessions" page with three main sections:

- Accessions:** Shows a total of 137066 accessions. It includes a "Search Accessions" button and links to "Add Accessions Or Upload Accession Info" and "Upload Pedigree File".
- Find Trials in Common:** Allows selecting an accession list from a dropdown menu and clicking a "Find Trials" button. A note says "Use a list of accessions to search for trials that contain them all".
- Populations:** Includes a "[Create Population]" button.

This will open a dialog allowing you to select either “Using Lists” or “Uploading a File”.

6.1 Add Accessions Using A List

First we will show how to add accessions “Using Lists”.

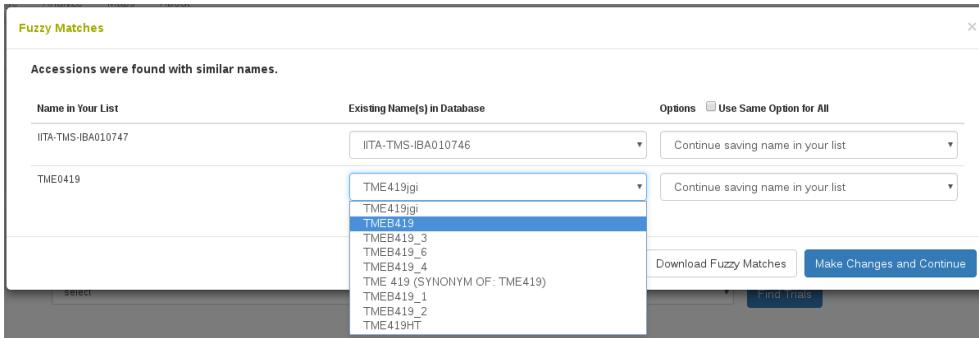


Here you select an accession list which you have previously made (see List Manager chapter). If you need to create or edit your list you can do so now by clicking “Manage Lists”. After selecting your list, click “Continue”.

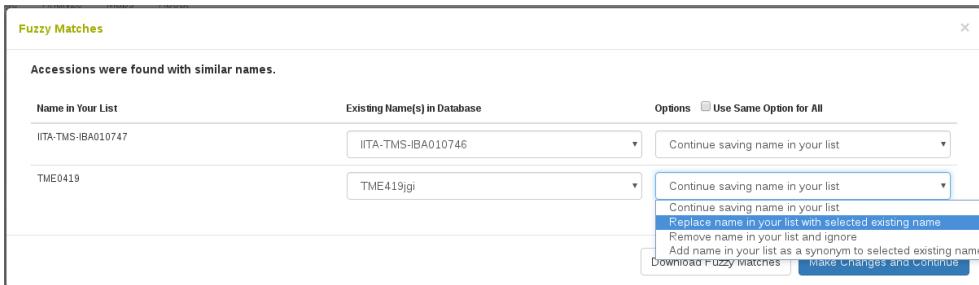
The contents of the list will be checked against the database, and elements that are already present will be flagged. A dialog will appear that will show the accessions which already exist in the database.

Found Accessions	
The following accessions already exist and cannot be added: Total number already in the database(7)	
Show 10 entries	Search:
Search Name	Found in Database
IITA-TMS-IBA010746	IITA-TMS-IBA010746
IITA-TMS-IBA010758	IITA-TMS-IBA010758
IITA-TMS-IBA010760	IITA-TMS-IBA010760
IITA-TMS-IBA010779	IITA-TMS-IBA010779
IITA-TMS-IBA010797	IITA-TMS-IBA010797
IITA-TMS-IBA010816	IITA-TMS-IBA010816
IITA-TMS-IBA010819	IITA-TMS-IBA010819
Showing 1 to 7 of 7 entries	
Previous 1 Next	
Continue	

After clicking on the “Continue” button, the next dialog will appear with accessions that have very similar names as the accession that you are adding. In the example below, there are two accession with very similar names to accessions already in the database. ‘TME0419’ is very similar to ‘TME419’, and probably represent the same line, so it would be a mistake to add this to the database again. Duplicate lines in the database should be avoided, as they cause problems when evaluating lines; data is divided up among several duplicates, making it harder to get the full picture about an accession.



To avoid situations in adding a mistaken duplicate accession, the database gives you options for moving forward with these very similar looking accession names. You can either “continue saving the name in your list”, “replace name in your list with selected existing name”, “remove name in your list and ignore”, or “add name in your list as a synonym to selected existing name”.



Clicking “Download Fuzzy Matches” will return a tabular result of the “fuzzy” accession name results shown. Click “Make changes and continue” to move on.

The final dialog shows the accessions that will be added. Here you need to assign the species of these accessions. You can optionally group the accessions into a population and/or add an organization for the accessions.

Accessions to be Added

Species name for added accessions
Manihot esculenta

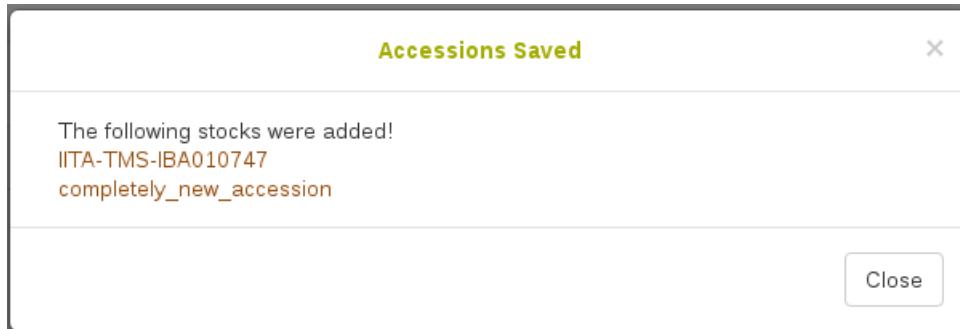
Population name for added accessions (optional)

Organization name for added accessions (optional)

The following accessions are new and will be added to the database:
Total number to be added(2)
ITA-TMS-IBA010747
completely_new_accession

Add Accessions

Once you click “Add Accessions”, the new accessions will be created in the database and you will see the following confirmation dialog, which includes links to the newly created accessions.



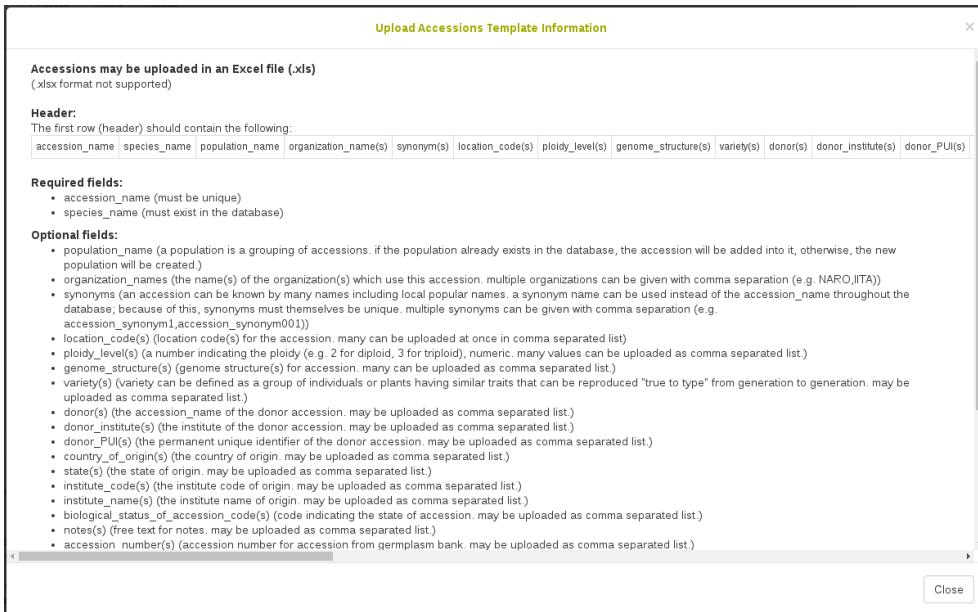
6.2 Uploading Accessions and Accession's Info From A File

Uploading accessions using a file is very similar to using a list, but enables you to add a variety of attributes, such as synonyms or ploidy levels, to the accessions in bulk.

6.2. UPLOADING ACCESSIONS AND ACCESSION'S INFO FROM A FILE53



Clicking on “Spreadsheet format” will show the required structure of the spreadsheet. The file must be XLS or XLSX format and can contain a number of header columns as attributes. It is important that you use exactly the same header column names as listed here. In columns that indicate that many attribute values can be passed at once using (s), such as synonym(s), you can pass a comma separated list of values, such as ‘synonym1,synonym2’.



Once you have selected your XLS or XLSX file for upload, click “Continue”.

The following process is the same way as with lists:

The first dialog which can appear will show accession names which are already in the database.

Click “Continue” and the next dialog that can appear will show “fuzzy” matches for the accession names you are trying to upload. Here you can choose to prevent adding accession names which look very similar to each other as wrongly duplicated accessions.

Click “Continue” and the final dialog that will appear will show the information to be added into the database. Here it is divided into accession names that are new and accession names that already exist in the database; however, for the accession names that already exist it will show additional attributes that originated from your file that will be added to these accessions.

The screenshot shows two stacked dialog boxes. The top dialog is titled "Accessions to be Added" and contains a table of new accession entries. The bottom dialog is titled "The following accessions will be updated:" and contains a table of existing entries with additional attributes.

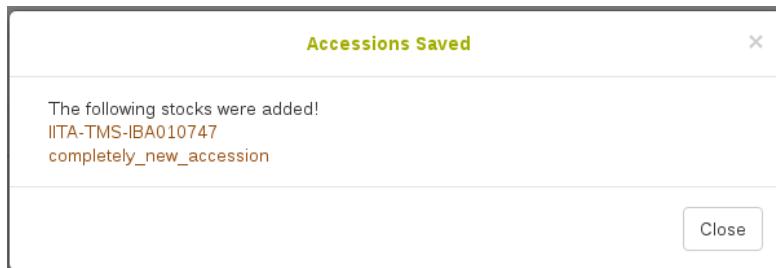
uniquename	properties
new_test_accession01	state:Oyo germplasmName:new_test_accession01 ploidyLevel:2 countyOfOriginCode:Nigeria species:Manihot esculenta defaultDisplayName:new_test_accession01 organizationName:test_organization populationName:test_population locationCode:ITH synonyms:new_test_accession_synonym1,new_test_accession_synonym2,new_test_accession_synonym3
new_test_accession02	organizationName:test_organization populationName:test_population defaultDisplayname:new_test_accession02 synonyms:germplasmName:new_test_accession02 state:Oyo countryOfOriginCode:Nigeria species:Manihot esculenta
new_test_accession03	germplasmName:new_test_accession03 state:Oyo countyOfOriginCode:Nigeria species:Manihot esculenta defaultDisplayname:new_test_accession03 organizationName:test_organization populationName:test_population synonyms:new_test_accession3_synonym1
new_test_accession04	synonyms: populationName:test_population organizationName:test_organization defaultDisplayname:new_test_accession04 species:Manihot esculenta countyOfOriginCode:Nigeria state:Oyo germplasmName:new_test_accession04

Showing 1 to 4 of 4 entries

uniquename	properties
ITATMS-IBA010746	stock_id:4867 synonyms:ITATMS-IBA010746_synonym1,ITATMS-IBA010746_synonym2 species:Manihot esculenta defaultDisplayName:ITATMS-IBA010746 germplasmName:ITATMS-IBA010746 organizationName:null populationName:test_population

Close Add Accessions

Once you click “Add Accessions”, the new accessions and information will be created in the database and you will see the following confirmation dialog, which includes links to the created and updated accessions.



6.3 Email alert for accession upload

When uploading a large number of accessions from a file, uploads can take a while, as the system needs to perform a series of checks on each entry. You have the option to receive an email notification about the status and results of your upload by clicking the “Email Alert” checkbox. By default, the system will use the email address associated with your account, but you have the option of entering a different email address if you prefer. After submitting, the upload process runs in the background, allowing you to continue using the interface without interruptions. Once the process completes, you will receive an email with the upload results, including any warnings or errors that may have occurred during the upload.

A screenshot of a web-based form titled "Add Accessions". The form has two main tabs at the top: "Using Lists" (which is currently selected) and "Uploading a File".

Below the tabs, there is a section for "File format information": "Spreadsheet format" with an info icon.

The "Uploading a File" section contains:

- "Upload File:" with a "Browse..." button and the file path "demo_accessions.xlsx".
- "Email Alert:" with a radio button set to "On" and an input field containing "noreply@breedbase.com".

The "Using Lists" section contains:

- "Use Fuzzy Search:" with a checked checkbox and a note: "Note: Use the fuzzy search to match similar names to prevent uploading of duplicate accessions. Fuzzy searching is much slower than regular search. Only a curator can disable the fuzzy search."
- "Append Synonyms:" with a checked checkbox and a note: "When checked, add synonyms of existing accession entries to the synonyms already stored in the database. When not checked, remove any existing synonyms of existing accession entries and store only the synonyms in the upload file."

A yellow callout box at the bottom provides additional file type information:

Accessions may be uploaded using any of the supported file types: MS Excel (.xls or .xlsx), comma-separated file (.csv), tab-delimited file (.txt or .tsv), or semicolon-separated file (.ssv).
Optional columns may be left out, if not used in your data.

At the bottom right of the form are "Close" and "Continue" buttons.

6.4 Add Parentage (Pedigree) Information to Accessions

Pedigree data can be uploaded from your computer by clicking on “Upload Pedigree File”

The screenshot shows the 'Manage Accessions' interface. At the top right, there is a red box highlighting the 'Upload Pedigree File' button. The page includes sections for 'Accessions' (with a count of 127130), 'Find Trials in Common', and 'Populations'. A search bar and user navigation links are also present.

You can find detailed information on how to prepare pedigree file by clicking on “File format information”. The file format can be tab or comma delimited text file, or Excel files (.xls or .xlsx).

The currently supported format has four columns:

progeny name female parent accession male parent accession type

Type can be biparental, self, backcross, sib, polycross, reselected, or open. In the case of the open type, the male parent accession field can remain blank. For all other types, both columns should be filled, even if they contain the same information as another column (such as self).

The screenshot shows the 'Upload Pedigrees' dialog box. It contains a 'File format information' section with a 'Spreadsheet Format' link, and an 'Upload Pedigree File:' section with a 'Choose File' input field showing 'no file selected'. There are 'Close' and 'Submit File' buttons at the bottom. The background shows the 'Manage Accessions' interface.

Template Information

Pedigrees may be uploaded in tab-delimited text file format
(.xls or .xlsx formats are **NOT** supported)

Header:
The first row (header) should contain the following:

progeny name	female parent accession	male parent accession	type
--------------	-------------------------	-----------------------	------

Required fields:

- progeny name (must exist in the database and can be accession uniquename or accession synonym)
- female parent accession (must exist in the database)
- type (biparental, open, self)

Optional fields

- male parent accession (can be accession uniquename or accession synonym or population name).

Notes

- Always specify the type of the cross (biparental, open, or self).
- If the type is open and no potential parents are known, leave the male parent field empty

Close

6.5 Working with grafts

Grafts are plants that are composed of a rootstock and a scion, which are genetically different and fused together, usually at the stem level.

To work with grafts, the grafts interface needs to be activated by a system administrator. Please contact your Breedbase provider. Briefly, a configuration parameter needs to be added to the sgn_local.conf file, show_grafting_interface. It should be set to 1 in sgn_local.conf, the default is 0 in sgn.conf.

Grafts to be created need to be specified using an Excel file (xlsx format) with two columns. The first column should have the header “scion accession” and should list accession names that will be scions. The second column should have the header “rootstock accession” and should list accession names that will be rootstocks.

In the database, the graft accessions will be created as single accessions. The graft accession will have two relationships, one to the scion accession (`scion_of` relationship) and one to the rootstock (`rootstock_of` relationship). These relationships are displayed on the pedigree viewer. The graft accession name is created from the scion accession name and the rootstock accession name, separated by the graft separator character. By default, the graft separator character is the plus sign ‘+’. The graft separator character can be changed in the `sgn_local.conf` file, using the parameter `graft_separator_string`. The graft separator string should not occur in any other accession names that are not grafts.

When the grafting interface is activated, a new button will be shown on the manage accessions page, called “Upload Grafts”.

Clicking the button brings up the upload grafts dialog.

Select the Excel file containing the grafting information. The system will validate the file, for example, check whether the accessions are in the database, and if the headers are correct.

The validation result will be presented, and if problems are found, they will be listed. In addition, if there are problems, the Upload button will be grayed out and upload will not be possible. Conversely, if there are no problems, the Upload button will be activated and can be clicked to store the data.

If the upload completes, a completion message is displayed with a summary what was uploaded.

Grafted accessions can be used like any other accession, for example, they can be used on field layouts. If you create a list of graft accessions, use the list type ‘accessions’.

Note that you shouldn’t create new grafts based on other grafts. The scion accession and the rootstock accession have to be different, otherwise they will not be created.

6.6 Bulk renaming of accessions

Accessions can be renamed in bulk using the rename accessions feature. To rename accessions, prepare a tab delimited file with two columns: the first

column should have the header “old name” and contain the accession names that need to be changed. The second column should have the header “new name” and contain the names that the accessions in column 1 should be renamed to.

The accession renaming feature is available from the Manage->Accessions page. Click on the “Rename Accessions” button. The first step is the upload of the file with a verification step. The verification step checks whether all the accession names in column 1 exist in the database, and whether all the accession names given in column 2 do NOT exist in the database. Only if both conditions are met, will the “rename” button become active, otherwise an error message is displayed listing the offending accession names.

Optionally, the old name can be automatically added as a synonym to the renamed accession, using the checkbox on the submit form. This option is clicked by default. Unclick the checkbox to NOT save any old names as synonyms.

Note that accession renaming should not be undertaken lightly. This feature is intended for special use cases, such as where accessions are created in a nursery with a name that is different from the accession name in the downstream breeding program.

It can also be used to rename accessions in bulk that have spelling mistakes and other issues. Please note however, that the tool does not make any attempt to change the names of associated elements, such as plots, that may have been constructed using accession names.

Because of the many implications of accession renaming, the feature is limited to accounts with the curator role.

Chapter 7

Managing Seed Lots

Seedlots are different from Accessions in that they represent the physical seed being evaluated in an experiment. Seedlots have things like physical storage locations and seed quantities, which accessions do not. The seed in seedlots can be from crosses or can be named accessions. Seedlots from crosses would represent seed harvested. Click Manage and then Seed Lots to begin.

Available Seedlots

About Seedlots

What are seedlots?

Seedlots represent physical seed in packets.

- This can be from crosses or named accessions.
- Seedlots can have a specific location, box, weight(g), and count.
- Seedlots can belong to breeding programs and organizations.

How do I inventory my seed?

1) Make sure your seedlots are in the database. Use "Add New Seedlot" to add a single seedlot or "Upload New Seedlots" to add many.
2) Make sure your seedlots are barcoded. You can print these barcodes from the database.
3) Use the "Seed Inventory" Android Application to scan seedlot barcodes and record weight. Then use "Upload Inventory" to upload this info into database. If you prefer you can create your own CSV file and upload that, if you do not want to use the Seed Inventory Application.
For more info about the "Seed Inventory" Android Application go to [Seed Inventory](#).
It is also possible to manually enter a transaction by going to the seedlot detail page and clicking "Add New Transaction".

Add New Seedlot | Upload New Seedlots | Upload Inventory

Seedlots

Search Seedlots

Show 10 entries

Seedlot Name	Breeding Program	Contents	Seedlot Location	Count	Weight (g)	Owners	Delete
new_test_crossP001_001	test	new_test_crossP001 (accession)	NA	1		X	
new_test_crossP002_001	test	new_test_crossP002 (accession)	NA	1		X	
new_test_crossP003_001	test	new_test_crossP003 (accession)	NA	1		X	
new_test_crossP004_001	test	new_test_crossP004 (accession)	NA	1		X	
new_test_crossP005_001	test	new_test_crossP005 (accession)	NA	1	-7	X	
new_test_crossP006_001	test	new_test_crossP006 (accession)	NA	1		X	
test_accession1_001	test	test_accession1 (accession)	NA	-1	-72	X	
test_accession5_001	test	test_accession5 (accession)	NA	1		X	

Showing 1 to 10 of 515 entries

Previous 1 2 3 4 5 ... 52 Next

seedlots

7.1 Add New Seedlot(s)

To add a single new seedlot, click on “Add Seedlot”. This will bring up the following dialog where you enter information about where the seedlot exists, what accession or cross is contained in it, and how many seeds there are. A seedlot must contain either an accession or a cross, and not both. A seedlot must have a weight in grams or a seed count or both of these.

Create New Seedlot

Name:	Required
Breeding Program:	test
Location:	Required
Box Name:	Optional
Contents:	
Accession name:	One Content Required
OR	
Cross name:	One Content Required
Amount [number of seeds]:	Amount OR Weight(g) Required
Weight (g):	Amount OR Weight(g) Required
Organization:	Optional
Timestamp:	Wed Mar 14 10:44:34 2018
Description:	Optional

OK

In the case where you have many seedlots to add to the database, you can upload an excel XLS or XLSX file instead. Click “Upload Seedlots” to see the following dialog.

Upload Seedlots

File format information
Spreadsheet format

Breeding Program: NelsonLab

Location: Required

Population Name: Optional

Organization Name: Optional

Upload File (.xls): Choose File No file chosen

Submit

7.2 Seedlot Transactions

Seedlots are capable of tracking where seeds came from, such as from crosses, and to where seeds go, such as to plots in the field. If you navigate to a seedlot detail page you will see the following.

Seedlot test_acquisition2_001

Details

Breeding Program	test
Seedlot Name	test_acquisition2_001
Organization	my org
Location Code	NA
Box Name	box2
Contents	test_acquisition2 (acquisition)
Current count	1
Current weight(g)	34

Transactions

Show 10 entries

Transaction Id	Transaction Date	From	To	Transaction Num Seeds	Transaction Weight (g)	Operator	Description	Options
40008	Mon Sep 18 11:44:00 2017	test_acquisition2 (acquisition)	test_acquisition2_001 (seedlot)	+1	NA	nmares	Auto generated seedlot from accession. DbPatch 00085	[Edit]
41456	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	-34	some user	Seed inventory CSV upload.	[Edit]
41459	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	-34	some user	Seed inventory CSV upload.	[Edit]
41460	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	-34	some user	Seed inventory CSV upload.	[Edit]
41464	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	-34	some user	Seed inventory CSV upload.	[Edit]
41466	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	+170	some user	Seed inventory CSV upload.	[Edit]
41470	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	NA	some user	Seed inventory CSV upload.	[Edit]
41473	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	+0	some user	Seed inventory CSV upload.	[Edit]
41477	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	+0	some user	Seed inventory CSV upload.	[Edit]
41478	2018-04-01-02-04-32	test_acquisition2_001 (seedlot)	test_acquisition2_001 (seedlot)	NA	+0	some user	Seed inventory CSV upload.	[Edit]

Showing 1 to 10 of 10 entries

On this page you see and can edit information regarding a single seedlot,

such as its name and location. You will also see a table indicating all the transactions that a seedlot has been involved in, such as if it was planted in a plot in the field. Transactions to field plots are created when adding or uploading a new trial or from a trial's detail page. Clicking on "Add New Transaction" let you add a transaction from between this seedlot and another seedlot. This kind of transaction is useful for representing if you have distributed seed to different locations.

Add New Seedlot Transaction

Transaction Type: Added to this Seedlot (test_accession2_001)

Taken From Existing Seedlot: test_accession4_001
Only showing seedlots with matching content

Transaction Amount (number seeds): Amount OR Weight Required

Transaction Weight (g): Amount OR Weight Required

Timestamp: Wed Mar 14 10:54:18 2018

Description: Required

OK

7.3 Seed Inventory

To inventory your seed: 1) Make sure your seedlots are in the database. Use "Add New Seedlot" to add a single seedlot or "Upload New Seedlots" to add many. 2) Make sure your seedlots are barcoded. You can print these barcodes from the database. 3) Use the "Inventory" Android Application to scan seedlot barcodes and record weight. Then use "Upload Inventory" to upload this info into database. If you prefer you can create your own CSV file and upload that, if you do not want to use the Inventory Application. For more info about the "Inventory" Android Application go to Inventory.

Clicking the "Upload Inventory" button will bring the following dialog:

Upload Seedlot Inventory

How do I inventory my seed?

- 1) Make sure your seedlots are in the database. Use "Add New Seedlot" to add a single seedlot or "Upload New Seedlots" to add many.
- 2) Make sure your seedlots are barcoded. You can print these barcodes from the database.
- 3) Use the "Seed Inventory" Android Application to scan seedlot barcodes and record weight. Then use "Upload Inventory" to upload this info into database. If you prefer you can create your own CSV file and upload that, if you do not want to use the Seed Inventory Application.
- For more info about the "Seed Inventory" Android Application go to [Seed Inventory](#).**
- It is also possible to manually enter a transaction by going to the seedlot detail page and clicking "Add New Transaction".

i File format information
Spreadsheet format

Upload File (.csv): No file chosen

The CSV file that should contain your inventory should meet these Template requirements. The Seed Inventory Android Application exports this exact file.

Upload Template Information

Seedlots may be uploaded in a CSV file (.csv)
(Excel .xls and .xlsx format not supported)

Header:
The first row (header) should contain the following:

box_id	seed_id	inventory_date	inventory_person	weight_gram
--------	---------	----------------	------------------	-------------

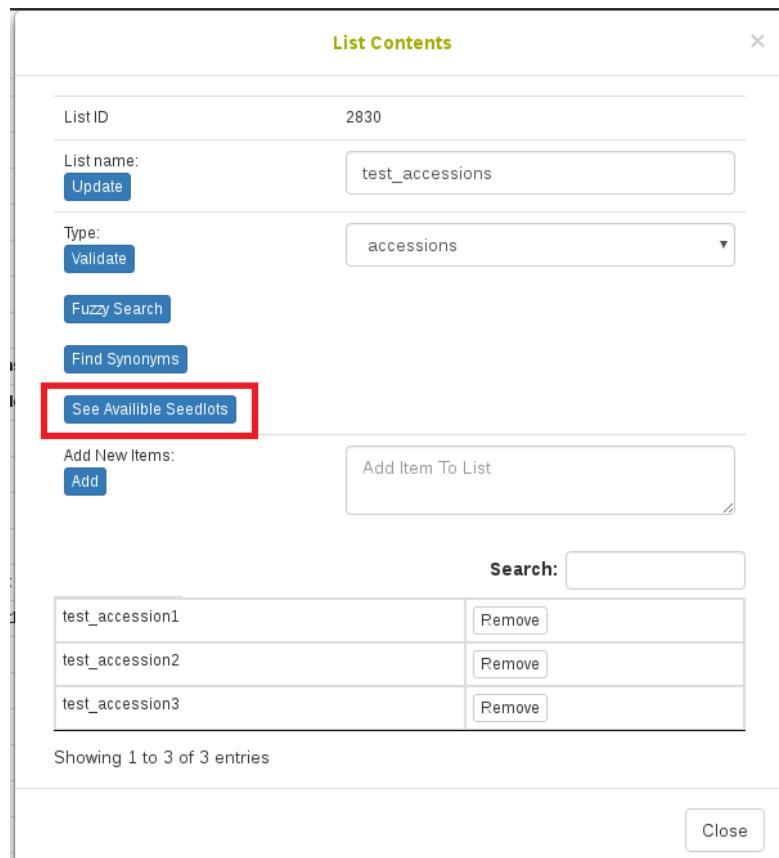
Required fields:

- box_id (the name of the box that the seedlot is in. also called box_name.)
- seed_id (unique identifier for the seedlot. must exist in the database. also called seedlot_name)
- inventory_date (a timestamp for when the seedlot was inventoried)
- inventory_person (the name of the person doing the inventory. can be any name. also called operator_name)
- weight_gram (the weight in grams of the seedlot)

7.4 Find Seedlots For a List of Accessions

A convenient tool for searching available seedlots for a list of accessions is available in the list tool. First open up your list of accessions. For help

opening a list of accessions please see the List section help. There is a button called “See Available Seedlots”.



Once you click this, you will see the following table in a dialog. From here you can create a list of seedlots using the checkboxes and the input at the bottom.

Available Seedlots					
Accessions	Seedlots				
	Breeding Program	SeedlotName	Contents	SeedlotLocation	Count
test_accession3	IITA	test_accession3_001	test_accession3	NA	-71
	IITA	seedtesty004	test_accession3	x1 location	99
	IITA	seedtest2004	test_accession3	x2	48
	IITA	seedtesty004	test_accession3	x2	99
test_accession1	NPCRI	UG120243_0015	test_accession1	NA2	1
test_accession2	IITA	test_accession2_001	test_accession2	NA	-138
	IITA	seedtesty003	test_accession2	x1 location	0
	IITA	seedtest2003	test_accession2	x2	0
	IITA	seedtesty003	test_accession2	x2	0
	IITA	seednx10	test_accession2	x2	0
	IITA	seednx11	test_accession2	x2	26

Create a New List from Selected Seedlots:

7.5 Create a seedlot for an Accession or Cross

Complementary to what we saw above for creating seedlots from the “Manage Seedlots” page, it is possible to create a new seedlot from an accession’s detail page or from the cross detail page. On the accession detail page, this is visible in the “Related Stocks” section as seen below. The cross detail page has an identical section. Notice the link for creating a new seedlot, which streamlines adding the seedlot.

Related stocks
 Related stocks in trials
 Seedlots of this Accession [\[Create New Seedlot\]](#)

Show 10 entries	Search:			
SeedlotName	Breeding Program	Contents	SeedlotLocation	Count
002B_001	IITA	002B (accession)	NA	-17
002B_test1_001	IITA	002B (accession)	Abuja	90

Showing 1 to 2 of 2 entries Previous 1 Next

Copy Seedlots to a List Copy the seedlot names showing in table to a new or existing list

Progenies
 Groups /members
 Related stocks for tissue sample

7.6 Add quality data to a seedlot

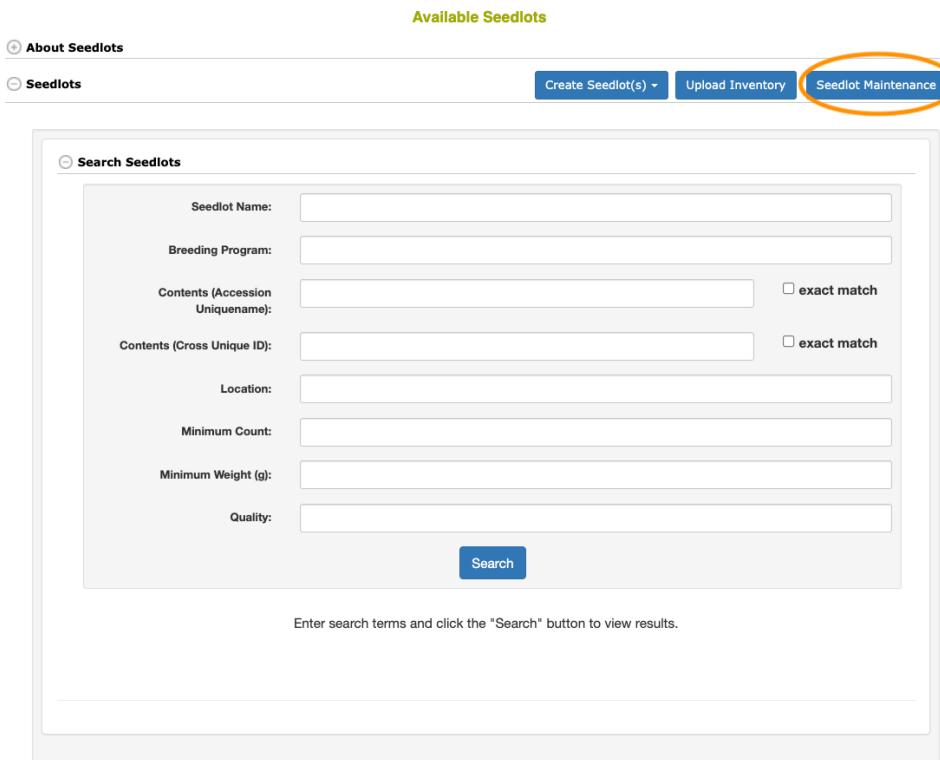
Quality information can be added to a seedlot in the quality field. This is also available as a column in the file upload format. It is recommended to use a controlled vocabulary, defined by the user, for the quality field. For example, good quality seed should be labelled “ok”, whereas other quality descriptors could be “moldy”, “insect damage”, or “low sprouting”, etc.

7.7 Seedlot Maintenance Events

For some crops, such as sugar kelp, a “seedlot” requires routine maintenance for the successful long-term storage of the seedlot. (For example, a Seedlot Maintenance Event for sugar kelp would be the routine change of the water that gametophytes are kept in). Breedbase can now store a record of these Seedlot Maintenance Events associated directly with existing Seedlots. Maintenance Events can be uploaded using a simple Excel template or recorded directly on the website.

7.7.1 Setup

Each Breedbase instance needs to be configured to support the storage of Seedlot Maintenance Events since each crop will have their own distinct set of maintenance events for their seedlots. To check if your Breedbase instance supports this feature, go to the Manage menu and select the Seed Lots page. Make sure you are logged in and look for the **Seedlot Maintenance** button near the top, next to the **Create Seedlot(s)** and **Upload Inventory** buttons. If you don’t see this button, contact the developer(s) supporting your Breedbase instance and ask if they can setup this feature.



The screenshot shows a web interface for managing seed lots. At the top, there are two tabs: 'About Seedlots' and 'Seedlots'. Below these are three buttons: 'Create Seedlot(s)', 'Upload Inventory', and 'Seedlot Maintenance'. The 'Seedlot Maintenance' button is circled in orange. The main area is titled 'Search Seedlots' and contains several search fields: 'Seedlot Name', 'Breeding Program', 'Contents (Accession UniqueName)', 'Contents (Cross Unique ID)', 'Location', 'Minimum Count', 'Minimum Weight (g)', and 'Quality'. Each field has an 'exact match' checkbox next to it. A 'Search' button is located at the bottom of the search form. Below the search form, a note says 'Enter search terms and click the "Search" button to view results.'

The location of the Seedlot Maintenance button on the Manage > Seed Lots page

7.7.2 Adding Events

Seedlot Maintenance Events can be added using two methods: 1) Uploading an Excel template or 2) Recording events directly on the website

Uploading Events with Excel Template

To bulk-upload a file of Seedlot Maintenance Events, first create an Excel (.xls or .xlsx) file with the following headers:

- **seedlot** - the name of the Seedlot to associate the event with (must exactly match an existing Seedlot in the database)
- **type** - the name of the Seedlot Maintenance Event type (these vary between Breedbase instances, a list of supported event types is displayed)

on the upload page)

- **value** - the value of the Seedlot Maintenance Event (these may be different for each event type and vary between Breedbase instances, a list of supported event values is displayed on the upload page)
- **notes** - optional, additional notes/comments about the event
- **operator** - the username of the Breedbase user that recorded the event
- **timestamp** - the date/time the event was recorded, in ‘YYYY-MM-DD HH:MM:SS’ format

Once you have an Excel file with the events filled out, follow these steps to upload the events to the database:

1. Make sure you are logged in to your Breedbase instance
2. Go to the Manage > Seed Lots page
3. Select the **Seedlot Maintenance** button
4. Select the **Upload Maintenance** button
5. Choose your Excel (.xls or .xlsx) file to upload
6. Select the **Upload** button

The dialog box has a header "Upload Seedlot Maintenance Events" and a close button "X". It contains the following sections:

- Select an Excel (.xls) file with the Seedlot Maintenance Events to upload**
- Requirements:**
 - The Maintenance Events are associated with Seedlots, so the name of the Seedlot in the file must match an existing Seedlot in the database. If a Seedlot is not yet in the database, go to the [Manage Seedlots](#) page to create it first.
 - The name of the Maintenance Event must be a valid event type. Valid event types include:

Event Type Name	Event Type Values
Water Change	Successful, Unsuccessful
Blended	Successful, Unsuccessful
Container Scraped	Successful, Unsuccessful
Light Intensity	<10, 20, 30-45, 50-75
Light Color	red, white
Container Size	1 L, 500 mL, 250 mL, 125 mL, vial
Form	Backup vial, Flask, Backup vial sibling
Biomass	high, medium, low
Health	healthy, not healthy
Color	1, 2, 3, 4, 5, 6, 7
Stickiness	yes, no
Clumping	yes, no
Contaminants	green, bacteria, other, all, green and bacteria, green and other, bacteria and other, none
Additional Notes	Any Value

i File format information
Spreadsheet Format

Upload File: No file chosen

The Seedlot Maintenance upload dialog, showing the supported event types and values (for sugar kelp)

Recording Events on Website

To add individual Seedlot Maintenance Events to the database in real time, as they're being recorded, use the **Record Maintenance** page. Follow these steps to record Seedlot Maintenance Events:

1. Make sure you are logged in to your Breedbase instance

2. Go to the Manage > Seed Lots page
3. Select the **Seedlot Maintenance** button
4. Select the **Record Maintenance** button
5. Enter the **Seedlot Name** or scan a barcode that has the Seedlot Name encoded. Once entered, the box at the top of the page will display basic information about the Seedlot as well its recently recorded events.
6. Select or Enter the values of individual events
7. Optionally, notes button next to each event to add additional notes/comments about that specific event
8. Make sure the operator/username and timestamp are correct
9. Select the **Submit** button to add the recorded events to the database.
NOTE: any events that remain selected as “Not Recorded” will not be submitted to the database.

Record Seedlot Maintenance

Seedlot

Name:	TEST_SEEDLOT_1-LOTA		Barcode
Contents:	SA18-CB-S1-FG1 (accession)		
Location:	WHOI		
Box:	Shelf 1 / Tray 1		
Recent Events:	Event	Value	Notes
	Water Change	Successful	additional notes
	Container Size	1 L	2021-07-22 13:04:24

Maintenance Events

Actions

- Water Change

<input checked="" type="button"/> Not Recorded	<input type="button"/> Successful	<input type="button"/> Unsuccessful	
--	-----------------------------------	-------------------------------------	--
- Blended

<input checked="" type="button"/> Not Recorded	<input type="button"/> Successful	<input type="button"/> Unsuccessful	
--	-----------------------------------	-------------------------------------	--
- Container Scraped

<input checked="" type="button"/> Not Recorded	<input type="button"/> Successful	<input type="button"/> Unsuccessful	
--	-----------------------------------	-------------------------------------	--

Observations

- Light Intensity

<input checked="" type="button"/> Not Recorded	<input type="button"/> <10	<input type="button"/> 20	<input type="button"/> 30-45	<input type="button"/> 50-75	
--	----------------------------	---------------------------	------------------------------	------------------------------	--
- Light Color

<input checked="" type="button"/> Not Recorded	<input type="button"/> red	<input type="button"/> white	
--	----------------------------	------------------------------	--
- Container Size

<input checked="" type="button"/> Not Recorded	<input type="button"/> 1 L	<input type="button"/> 500 mL	<input type="button"/> 250 mL	<input type="button"/> 125 mL	<input type="button"/> vial	
--	----------------------------	-------------------------------	-------------------------------	-------------------------------	-----------------------------	--

Additional Notes
Any additional notes, usually concerning culture termination or partial use for an experiment

Username/Timestamp

Operator:	dwaring87
Timestamp:	2021-08-18 14:29:49

The Seedlot Maintenance record page, as configured for sugar kelp

7.7.3 Displaying Events

Recently recorded Seedlot Maintenance Events are displayed in a table from the main Seedlot Maintenance page, as well as the detail page for individual Seedlots.

Seedlot Maintenance																																																																																																																																																																										
About Seedlot Maintenance		Seedlot Maintenance Tools																																																																																																																																																																								
Seedlot Maintenance Events		Record Maintenance Upload Maintenance																																																																																																																																																																								
Filter Events Filter maintenance events based on date, type, and/or value																																																																																																																																																																										
<table border="1"> <thead> <tr> <th>Excel</th><th>CSV</th><th colspan="7">Search: <input type="text"/></th></tr> <tr> <th>Seedlot</th><th>Event ID</th><th>Event Date</th><th>▼</th><th>Event Type</th><th>Value</th><th>Notes</th><th>Operator</th><th>Options</th></tr> </thead> <tbody> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381860</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Water Change</td><td>Successful</td><td>additional notes</td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381866</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Color</td><td>3</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381865</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Biomass</td><td>medium</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381864</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Container Size</td><td>1 L</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381863</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Light Color</td><td>red</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381862</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Light Intensity</td><td><10</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381861</td><td>Thu Jul 22 13:04:24 2021</td><td></td><td>Blended</td><td>Unsuccessful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381819</td><td>Fri Jul 9 13:22:24 2021</td><td></td><td>Clumping</td><td>yes</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_1-LOTA</td><td>381818</td><td>Fri Jul 9 13:21:46 2021</td><td></td><td>Blended</td><td>Successful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381816</td><td>Fri Jul 9 13:19:08 2021</td><td></td><td>located_in</td><td>Successful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381807</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Light Intensity</td><td>50-75</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381804</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Water Change</td><td>Successful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381805</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Blended</td><td>Successful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381806</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Container Scraped</td><td>Successful</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381808</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Light Color</td><td>red</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> <tr><td>TEST_SEEDLOT_2</td><td>381809</td><td>Fri Jul 9 13:18:05 2021</td><td></td><td>Container Size</td><td>1 L</td><td></td><td>dwaring87</td><td>[Remove]</td></tr> </tbody> </table>									Excel	CSV	Search: <input type="text"/>							Seedlot	Event ID	Event Date	▼	Event Type	Value	Notes	Operator	Options	TEST_SEEDLOT_1-LOTA	381860	Thu Jul 22 13:04:24 2021		Water Change	Successful	additional notes	dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381866	Thu Jul 22 13:04:24 2021		Color	3		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381865	Thu Jul 22 13:04:24 2021		Biomass	medium		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381864	Thu Jul 22 13:04:24 2021		Container Size	1 L		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381863	Thu Jul 22 13:04:24 2021		Light Color	red		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381862	Thu Jul 22 13:04:24 2021		Light Intensity	<10		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381861	Thu Jul 22 13:04:24 2021		Blended	Unsuccessful		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381819	Fri Jul 9 13:22:24 2021		Clumping	yes		dwaring87	[Remove]	TEST_SEEDLOT_1-LOTA	381818	Fri Jul 9 13:21:46 2021		Blended	Successful		dwaring87	[Remove]	TEST_SEEDLOT_2	381816	Fri Jul 9 13:19:08 2021		located_in	Successful		dwaring87	[Remove]	TEST_SEEDLOT_2	381807	Fri Jul 9 13:18:05 2021		Light Intensity	50-75		dwaring87	[Remove]	TEST_SEEDLOT_2	381804	Fri Jul 9 13:18:05 2021		Water Change	Successful		dwaring87	[Remove]	TEST_SEEDLOT_2	381805	Fri Jul 9 13:18:05 2021		Blended	Successful		dwaring87	[Remove]	TEST_SEEDLOT_2	381806	Fri Jul 9 13:18:05 2021		Container Scraped	Successful		dwaring87	[Remove]	TEST_SEEDLOT_2	381808	Fri Jul 9 13:18:05 2021		Light Color	red		dwaring87	[Remove]	TEST_SEEDLOT_2	381809	Fri Jul 9 13:18:05 2021		Container Size	1 L		dwaring87	[Remove]
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TEST_SEEDLOT_1-LOTA	381819	Fri Jul 9 13:22:24 2021		Clumping	yes		dwaring87	[Remove]																																																																																																																																																																		
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TEST_SEEDLOT_2	381809	Fri Jul 9 13:18:05 2021		Container Size	1 L		dwaring87	[Remove]																																																																																																																																																																		

Unfiltered table of recent Seedlot Maintenance events

The events displayed in these tables are sorted by timestamp, with the most recently recorded events displayed first. The displayed events can be filtered using any number of supported filter criteria, such as: - seedlot names (as entered on the page or using an existing seedlot list), - dates (on, on or before, before, on or after, and/or after the entered dates) - event types - event type values - operator/username

Select the properties of the filter(s) you want to apply, then select the **Add** button next to the button to add the filter to the list of applied filters. Once

you're done adding filters, select the **Filter** button to search the database for the filtered events.

Seedlot Maintenance

- [About Seedlot Maintenance](#)
- [Seedlot Maintenance Tools](#)
- [Seedlot Maintenance Events](#)

[Record Maintenance](#) [Upload Maintenance](#)

Filter Events Filter maintenance events based on date, type, and/or value

Add one or more filters to apply to the table of displayed maintenance events. To add a filter, enter the properties for a filter type and click the add button to add the filter to the list. Once you're done adding filters, click the Filter button to display the results.

Seedlot(s): Enter the name(s) of the Seedlot(s) - one per line [Add](#)

TEST_SEEDLOT_2

OR

Select a List of Seedlots [Add](#)

Date: on

Type: healthy not healthy

Operator: [Add](#)

Applied Filters:

Property	Comparison	Value	Remove
name	includes	TEST_SEEDLOT_2	
Water Change	includes	Any Value	
Health	includes	Any Value	

[Filter](#)

Excel CSV Search:

Seedlot	Event ID	Event Date	Event Type	Value	Notes	Operator	Options
TEST_SEEDLOT_2	381804	Fri Jul 9 13:18:05 2021	Water Change	Successful		dwaring87	[Remove]
TEST_SEEDLOT_2	381811	Fri Jul 9 13:18:05 2021	Health	healthy		dwaring87	[Remove]
TEST_SEEDLOT_2	381789	Fri Jul 9 13:15:11 2021	Water Change	Successful		dwaring87	[Remove]

EVENTS: 1 - 3 / 3
PAGE: 1 / 1 ◀ Prev Next ▶

A filtered table of Seedlot Maintenance events

The filtered events can be downloaded directly from the table using the **Excel** or **CSV** buttons at the top of the table. Or Seedlot Maintenance Events can be bulk-downloaded (this includes all events for a Seedlot) using a list of Seedlots from the main downloads page (see below).

7.7.4 Downloading Events

To bulk-download all events for a specific subset of Seedlots:

1. Create a list containing the Seedlots you are interested in.
2. Go to the **Download Using Lists** page (Manage > Download)
3. Find the **Download Seedlot Maintenance Events** section
4. Select your list of Seedlots
5. Select the **Download** button to generate the download file

The downloaded file will follow the same format as the upload template and will contain all recorded Seedlot Maintenance Events for each Seedlot in the list.

7.8 Deleting Seedlots

Seedlots can be deleted on the Manage Seedlots page (/breeders/seedlots) by search the seedlot and then clicking the X to delete one seedlot at a time. To delete a seedlot, the logged in user needs the required delete privileges on the seedlot. The seedlot also should not have any transactions associated with it (except for the initial transaction).

To delete seedlots in bulk, generate a list of type seedlot, for example, using the wizard. Open the section “Delete seedlots using a list” on the Manage Seedlots page and select the list. Seedlot deletion using a list is only available to user with curator status.

Chapter 8

Managing Populations

Populations are modeled as groups of accessions. This grouping can be useful in downstream analyses. To manage these populations go to Manage Accessions and scroll tp the bottom.

The screenshot shows a web-based application for managing accessions. At the top, there's a header titled "Manage Accessions". Below it, there are three main sections:

- Accessions:** Shows a total of 137103 accessions. It includes a "Search Accessions" input field and links to "Add Accessions Or Upload Accession Info" and "Upload Pedigree File".
- Find Trials in Common:** Allows selecting an accession list from a dropdown menu and clicking "Find Trials". A note says "Use a list of accessions to search for trials that contain them all".
- Populations:** This section is highlighted with a red border. It has a "Create Population" link.

To add a new population click “Create Population”. The following dialog will appear where you choose a list of accessions and give a name to the new population. Please note it is also possible to create a population when you are uploading new accessions into the database.

Create A Population

Population Name:	<input type="text"/>
Choose a List of Accessions to Add:	<input type="text" value="119acc"/>
<input type="button" value="Close"/> <input type="button" value="Submit"/>	

Click on the plus (+) button next to Populations to see all the available populations. Click on a population name to see the accessions in the population.

<input checked="" type="radio"/> Populations		[Create Population]
new_test_population	[Go To Population Page]	[Add Accessions To Population] [Delete Population]
NARITA	[Go To Population Page]	[Add Accessions To Population] [Delete Population]

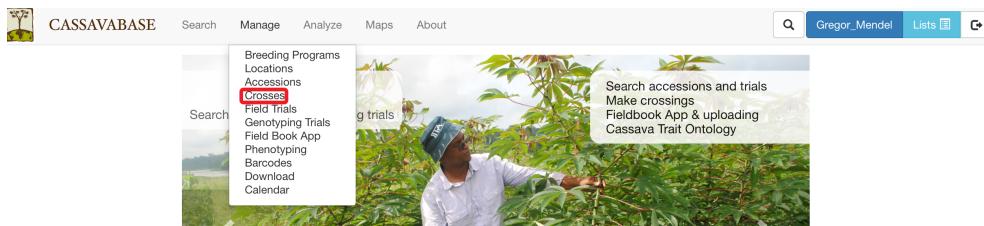
From here you can delete accessions from a population as well as add new accessions to the population.

new_test_population		[Go To Population Page]	[Add Accessions To Population] [Delete Population]
Show 10 ▾ entries			
Accession Name	Description	Synonyms	Remove From Population
037B			X
037D			X
037F			X
038F			X
039B			X
039D			X
039F			X
040B			X
040D			X
041B			X
Showing 1 to 10 of 119 entries			
new_test_population		<input type="button" value="add to new list"/>	
119acc		<input type="button" value="add to list"/>	

Chapter 9

Managing Crosses

Information for crosses can be managed using the “Crosses” option in the Manage menu.



9.1 Crossing Experiment

Different crosses in the same trial/nursery/project are grouped via “**crossing experiment**”. Crossing experiments are organized based on their breeding programs. To find a crossing experiment, you can either type the crossing experiment name in the “Search” box, or look for the crossing experiment directly in its breeding program by clicking on the “+” icon. In each breeding program, crossing experiments can be placed directly in the breeding program, or organized in folders. The “**Folders**” section allows you to place crossing experiments in folders, move a crossing experiment in a folder to another folder, or rearrange your folders within a breeding program.

Manage Crosses

[\[Add Crossing Trial\]](#) [\[Add Cross\]](#) [\[Upload Crosses\]](#) [\[Add Cross Wishlist\]](#)

Information	Breeding Programs – Folders – Crossing Trials
Search <input type="text" value="Search"/> <p><i>Double click crossing trial (⚡) or folder (📁) to view detail page.</i></p> <p>Breeding programs (⚡)</p>	Breeding Programs – Folders – Crossing Trials <input type="button" value="Refresh"/> <ul style="list-style-type: none"> + IITA <ul style="list-style-type: none"> + 2017 <ul style="list-style-type: none"> ⚡ IITA_crossingtrial_2017 ⚡ yam_crossingtrial_2017 + NRCRI + NaCRRI
Folders <input type="button" value="Create new folder"/> <input type="button" value="Move crossing trial to folder"/> <input type="button" value="Move folder"/>	

Manage Crosses

[\[Add Crossing Trial\]](#) [\[Add Cross\]](#) [\[Upload Crosses\]](#) [\[Add Cross Wishlist\]](#)

Information	Breeding Programs – Folders – Crossing Trials
Search <input type="text" value="Search"/> <p><i>Double click crossing trial (⚡) or folder (📁) to view detail page.</i></p> <p>Breeding programs (⚡)</p>	Breeding Programs – Folders – Crossing Trials <input type="button" value="Refresh"/> <ul style="list-style-type: none"> + IITA <ul style="list-style-type: none"> + 2017 <ul style="list-style-type: none"> ⚡ IITA_crossingtrial_2017 ⚡ yam_crossingtrial_2017 + NRCRI + NaCRRI <ul style="list-style-type: none"> ⚡ crossing_nursery_2018

9.1.1 Add New Crossing Experiment

To add a new crossing experiment, click on “Add Crossing Experiment” link.

Manage Crosses

[\[Add Crossing Trial\]](#) [\[Add Cross\]](#) [\[Upload Crosses\]](#) [\[Add Cross Wishlist\]](#)

Information	Breeding Programs – Folders – Crossing Trials
Search <input type="text" value="Search"/>	Breeding Programs – Folders – Crossing Trials <input type="button" value="Refresh"/> <ul style="list-style-type: none"> + IITA + NRCRI + NaCRRI

Required Information:

- **“Crossing Experiment Name”**: enter a name for the crossing experiment. The crossing experiment name must not already exist in the database.

- “**Breeding program**”: select a breeding program that is available in the database. New breeding programs can be added on the “Breeding program” page, accessible from the “Manage” menu. *Breeding Program Page*
- “**Location**”: select a location for the crossing experiment. New locations can be entered on the “**Locations**” page, accessible from the “**Manage**” menu. *Location Page*
- “**Year**”: select a year.
- “**Description**”: enter a description for the crossing experiment.

After filling in the information, click “**Submit**” to generate the crossing experiment.

Add New Crossing Trial ×

Crossing Trial Name:	IITA_crossing_trial_2017
Breeding Program:	IITA
Location:	Ibadan
Year:	2017
Description:	To improve disease resistance

Close Submit

9.2 Cross

9.2.1 Add New Crosses

Add a cross by using the “Add New Cross” dialog

To add a single new cross, click on “Add Cross” link.

The screenshot shows the 'Manage Crosses' interface. At the top, there are buttons for '[Add Crossing Trial]', '[Add Cross]' (which is highlighted with a red box), '[Upload Crosses]', and '[Add Cross Wishlist]'. Below this is a search bar and a 'Double click' instruction. On the right, there's a tree view under 'Breeding Programs – Folders – Crossing Trials' with nodes for IITA (2017, NRCRI), NaCRRI, and a folder for 'crossing_nursery_2018'.

Enter cross information in the popup dialog.

The 'Add New Cross' dialog has the following fields:

- Required:**
 - Crossing Trial: IITA_crossingtrial_2017
 - Location: Ibadan
 - Cross Name: UG120001xUG120002
 - Cross Type: biparental
 - Female Parent: UG120001
 - Male Parent: UG120002 (highlighted with a blue border)
- Optional:**
 - Field Trial: (empty field with a 'Search Plots' button)
 - Female Plot: Choose trial first
 - Male Plot: Choose trial first

Required Information:

- “**Crossing experiment**”: select a crossing experiment available in the database.
- “**Location**”: select a location available in the database.

- “**Cross name**”: enter a name for the cross. The cross name must not already exist in the database.
- “**Cross type**”: the options for cross types are: biparental, self, open pollinated, bulk, bulk selfed, bulk and open pollinated, double haploid, polycross, reciprocal and multicross.

Create New Crosses

Crossing Trial:	Select Crossing Trial
Location:	Cornell Biotech
Cross Name:	
Cross Type:	Select a cross type biparental self open pollinated bulk bulk selfed bulk and open pollinated doubled haploid polycross reciprocal multicross
Optional:	
Field Trial:	
Female Plot:	
Male Plot:	

- The “**Female Parent**” and “**Male Parent**” field are auto-complete fields for accessions that are already in the database. The parents specified will be entered in the pedigree of the new accessions generated by this cross.

Optional Information:

- “**Female Plot and/or Male Plot**”: In addition to the accession names, specific plots used in the cross can also be added to the database. To retrieve plot names associated with each female/male accession, enter your trial name, then click “**Search Plots**”. Plot names of each parental accession in that field trial will be shown in the drop-down list, you can then select the plot used in the cross.

Optional:

Field Trial:	Kasese solgs trial	Search Plots
Female Plot:	KASESE_TP2013_842	
Male Plot:	KASESE_TP2013_1591	

Additional crossing experimental information such as pollination date, number of flowers, number of fruits, number of seeds can be specified during adding new cross. Alternatively, this information can be updated or edited directly on the “**Cross Details**” page.

If you know the number of accessions that are generated from the cross, they can be instantiated immediately in the database by clicking the “**Add accessions for progeny**” checkbox and specifying the number.

Specify Pollination Date:

Specify Number of Flowers:

Specify Number of Fruits:

Specify Number of Seeds:

Add New Accessions for Progeny:

Submit

Click “Submit” to generate the cross.

Upload New Crosses

To upload new crosses from an Excel file (.xls or .xlsx), click on “Upload Crosses” link.



Select a crossing experiment and a location available in the database from drop-down lists and choose a file that you want to upload, then click “**Upload File**”.

Upload Crosses

 File Format Information
Spreadsheet Format

Crossing Trial: IITA_crossingtrial_2017

Location: Ibadan

Upload File: Choose File No file chosen

Close **Upload File**

Please check spreadsheet format carefully. The file must be an Excel file (.xls or .xlsx).

Template Information

Crosses may be uploaded in an Excel file (.xls)
(.xlsx format not supported)

Header:
The first row (header) must contain the following:

cross_name	cross_type	female_parent	male_parent	Tag Number	Pollination Date	Number of Flowers	Number of Fruits	Fruit Harvest Date	Number of Seeds	Seed Harvest Date	Number of Seeds Sown	Number of Seeds Germinated
------------	------------	---------------	-------------	------------	------------------	-------------------	------------------	--------------------	-----------------	-------------------	----------------------	----------------------------

Required columns:

- cross_name (must not conflict with an existing cross name)
- cross_type (must be one of the following: biparental, self, open, bulk, bulk_self, bulk_open, or doubled_haploid)
- female_parent (accession names must exist in the database)
- male_parent (required in the header, but value may be left blank for most cross types. Must be specified for biparental and bulk crosses.
When specified, accession names must exist in the database)

Optional columns (dates must be in the format YYYY/MM/DD):

- Tag Number
- Pollination Date
- Number of Flowers
- Number of Fruits
- Fruit Harvest Date
- Number of Seeds
- Seed Harvest Date
- Number of Seeds Sown
- Number of Seeds Germinated

Close

9.2.2 Update Crosses by Uploading

To upload progenies and/or experimental info of crosses already in the database, go to “**Manage-Upload**” page.

In the “Crosses” section, there are links for uploading progenies and experimental info.

Crosses			
Plan	Add	Manage	Search
Create Cross Wishlist	Upload Many New Crosses Add A Cross Upload Progenies of Existing Crosses Upload Experimental Info of Existing Crosses	Go To Manage Crosses Page	Go To Search Crosses

Please check spreadsheet format in each link carefully. The file must be an Excel file (.xls or .xlsx).

Template Information ×

Progenies of existing crosses may be uploaded in an Excel file (.xls)
(.xlsx format not supported)

Header:
The first row (header) must contain the following:

cross_name	progeny_name
------------	--------------

Required columns:
-cross_name (must exist in the database)
-progeny_name (must not already exist in the database, must have only one progeny for each row, you can add many progenies by adding more rows)

Close

Template Information ×

Experimental Info of existing crosses may be uploaded in an Excel file (.xls)
(.xlsx format not supported)

Header:
The first row (header) must contain the following:

cross_name	At least one column of experimental info listed below
------------	---

Required columns:
-cross_name (must exist in the database, must not have duplicate cross name in the upload file)
-At least one of the following columns: (all of the experimental info of a cross must be in a single row)
 Tag Number
 Pollination Date
 Number of Bags
 Number of Flowers
 Number of Fruits
 Number of Seeds

Note: crossing experimental information is customized based on the need for

each crop. As a result, column headers for experimental info in your database may be different from the information shown in this manual.

9.3 Cross Wishlist

An Android ODK application is being developed to record cross information on a mobile device in the field. To link this mobile application with the database, the Cross Wishlist can be used to create a plan for which crosses to perform.

This tool is available on the Manage Cross page. It is currently only available on certain databases, so when you click this link you may see an alert mentioning that the cross wishlist is not available on your database.

The screenshot shows a web-based application titled 'Manage Crosses'. At the top right, there is a toolbar with three buttons: '[Create Cross Wishlist]', '[Create New Crosses]', and '[Upload Crosses File]'. The main area displays a hierarchical tree structure under the heading 'Breedings Programs -- Folders -- Crosses'. The tree includes nodes for IITA, NRCRI, NaUHR, ARI Tanzania, QAT, CARI, Rayong, KU, CSIR, SCP, BTI, SRI Kibaha, and CNRA_PRT. On the left side of the interface, there are sections for 'Information' (with a 'Search' input field), 'Folders' (with buttons for 'Create new folder', 'Move cross to folder', and 'Move folder'), and 'Breeding programs' (with a link to 'Breedings programs'). A red box highlights the '[Create Cross Wishlist]' button in the toolbar.

9.3.1 Create a Cross Wishlist

Step 1. Select the accessions to be crossed in your trial

There are two interfaces for this step, either “Not Using Lists” or “Using Lists”. Depending on if you already have a list of female and male accessions to use, you can decide on which interface to use. The end result of using either interface is the same.

The screenshot shows the 'Create Cross Wishlist' dialog box. At the top, there are two tabs: 'Using Lists' (orange) and 'Not Using Lists' (blue, selected). Below the tabs, there are two dropdown menus. The first dropdown under 'Trial Name(s)' is labeled 'Please select a trial'. The second dropdown under 'Female Accession Name(s)' is labeled 'First Select A Trial'. At the bottom right of the dialog box are three buttons: 'Available Cross Wishlist(s) for ODK Use', 'Next', and 'Close'.

We will start by showing “Not Using Lists”. First select the trial in which the crosses are to be performed. This will populate a select box with all the accessions used in that trial. From here, one or many accessions can be selected as the female accession.

The screenshot shows the 'Create Cross Wishlist' dialog box with the 'Using Lists' tab selected. The 'Trial Name(s)' dropdown contains '05uyt20interIB'. The 'Female Accession Name(s)' dropdown is expanded, showing a list of accessions: IITA-TMS-IBA000203, IITA-TMS-IBA000210, IITA-TMS-IBA000211, IITA-TMS-IBA000214, IITA-TMS-IBA000222, IITA-TMS-IBA000252, and IITA-TMS-IBA30572. At the bottom right of the dialog box are three buttons: 'Available Cross Wishlist(s) for ODK Use', 'Next', and 'Close'.

Once the female accessions are selected, a table is populated. Each row in this table begins with the female accession that was selected, followed by a select box with all the accessions used in the trial. From here, one or many accessions can be selected as the male to use in the cross.

9.3. CROSS WISHLIST

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The screenshot shows the 'Create Cross Wishlist' dialog box. At the top, there are two tabs: 'Using Lists' (selected) and 'Not Using Lists'. Below this, the 'Trial Name(s)' field contains '05uyt20InterB'. The 'Female Accession Name(s)' section lists three entries: 'ITA-TMS-IBA000210', 'ITA-TMS-IBA000214', and 'ITA-TMS-IBA000222'. To the right of each female accession is a dropdown menu titled 'Select Male Parent' containing a list of male accessions. Red arrows point from each female accession name to its corresponding male selection list. The male lists include:

- For ITA-TMS-IBA000210: ITA-TMS-IBA000203, ITA-TMS-IBA000210, ITA-TMS-IBA000211, ITA-TMS-IBA000214, ITA-TMS-IBA000222, ITA-TMS-IBA000252, ITA-TMS-IBA30572, ITA-TMS-IBA000224
- For ITA-TMS-IBA000214: ITA-TMS-IBA000203, ITA-TMS-IBA000210, ITA-TMS-IBA000211, ITA-TMS-IBA000214, ITA-TMS-IBA000222, ITA-TMS-IBA000252, ITA-TMS-IBA30572, ITA-TMS-IBA000224
- For ITA-TMS-IBA000222: ITA-TMS-IBA000203, ITA-TMS-IBA000210, ITA-TMS-IBA000211, ITA-TMS-IBA000214, ITA-TMS-IBA000222, ITA-TMS-IBA000252, ITA-TMS-IBA30572, ITA-TMS-IBA000224

At the bottom right of the dialog box are buttons for 'Available Cross Wishlist(s) for ODK Use', 'Next', and 'Close'.

Once the male accessions are selected to cross with each female accession, a table indicating priorities appears. Priority is meant to indicate an order in which to attempt the cross; first the highest priority male will be considered, but if this cross is not possible then subsequent males will be considered. An equal priority can be given and this will not indicate a specific order to follow.

Create Cross Wishlist

Using Lists Not Using Lists

Trial Name(s): 05uyt20interIB

Female Accession Name(s):

- ITA-TMS-IBA000203
- ITA-TMS-IBA000210
- ITA-TMS-IBA000211
- ITA-TMS-IBA000214
- ITA-TMS-IBA000222
- ITA-TMS-IBA000252
- ITA-TMS-IBA30572
- ITA-TMS-IBA30574

Female Parent	Select Male Parent	Priority { 1 : highest, 10 : lowest }								
ITA-TMS-IBA000210	<ul style="list-style-type: none"> ITA-TMS-IBA000203 ITA-TMS-IBA000210 ITA-TMS-IBA000211 ITA-TMS-IBA000214 ITA-TMS-IBA000222 ITA-TMS-IBA000252 ITA-TMS-IBA30572 ITA-TMS-IBA30574 	<table border="1"> <thead> <tr> <th>Male Parent</th> <th>Priority</th> </tr> </thead> <tbody> <tr> <td>ITA-TMS-IBA000203</td> <td>1</td> </tr> <tr> <td>ITA-TMS-IBA000211</td> <td>3</td> </tr> <tr> <td>ITA-TMS-IBA000252</td> <td>2</td> </tr> </tbody> </table>	Male Parent	Priority	ITA-TMS-IBA000203	1	ITA-TMS-IBA000211	3	ITA-TMS-IBA000252	2
Male Parent	Priority									
ITA-TMS-IBA000203	1									
ITA-TMS-IBA000211	3									
ITA-TMS-IBA000252	2									
ITA-TMS-IBA000214	<ul style="list-style-type: none"> ITA-TMS-IBA000203 ITA-TMS-IBA000210 ITA-TMS-IBA000211 ITA-TMS-IBA000214 ITA-TMS-IBA000222 ITA-TMS-IBA000252 ITA-TMS-IBA30572 ITA-TMS-IBA30574 	<table border="1"> <thead> <tr> <th>Male Parent</th> <th>Priority</th> </tr> </thead> <tbody> <tr> <td>ITA-TMS-IBA000203</td> <td>1</td> </tr> <tr> <td>ITA-TMS-IBA000210</td> <td>1</td> </tr> </tbody> </table>	Male Parent	Priority	ITA-TMS-IBA000203	1	ITA-TMS-IBA000210	1		
Male Parent	Priority									
ITA-TMS-IBA000203	1									
ITA-TMS-IBA000210	1									

Available Cross Wishlist(s) for ODK Use Next Close

Alternatively, we could have used the “Using List” interface instead. Here we select the trial in which the crosses will be performed and we provide a list of accessions to consider for the females and the males to be crossed.

Create Cross Wishlist

Using Lists Not Using Lists

Trial Name(s):	05uyt20interB
Female Accession List:	acc_test
Male Accession List:	acc_test

Set Cross Priorities: 1 is highest and 10 is lowest
 Female Accessions Are in First Column and Male Accessions Are in Header

Female Accessions	IITA-TMS-IBA010746	IITA-TMS-IBA010758	IITA-TMS-IBA010760	IITA-TMS-IBA010779	IITA-TMS-IBA010797	IITA-TMS-IBA010816	IITA-TMS-IBA010819
IITA-TMS-IBA010746							
IITA-TMS-IBA010758							
IITA-TMS-IBA010760							
IITA-TMS-IBA010779							
IITA-TMS-IBA010797							

Available Cross Wishlist(s) for ODK Use Next Close

Step 2. Select the female plots to be considered in the crosses

After selecting your lists, the table below is populated. The first column has all the female accessions specified and the header row has all the male accessions specified. The males to consider crossing with each female are indicated with priority.

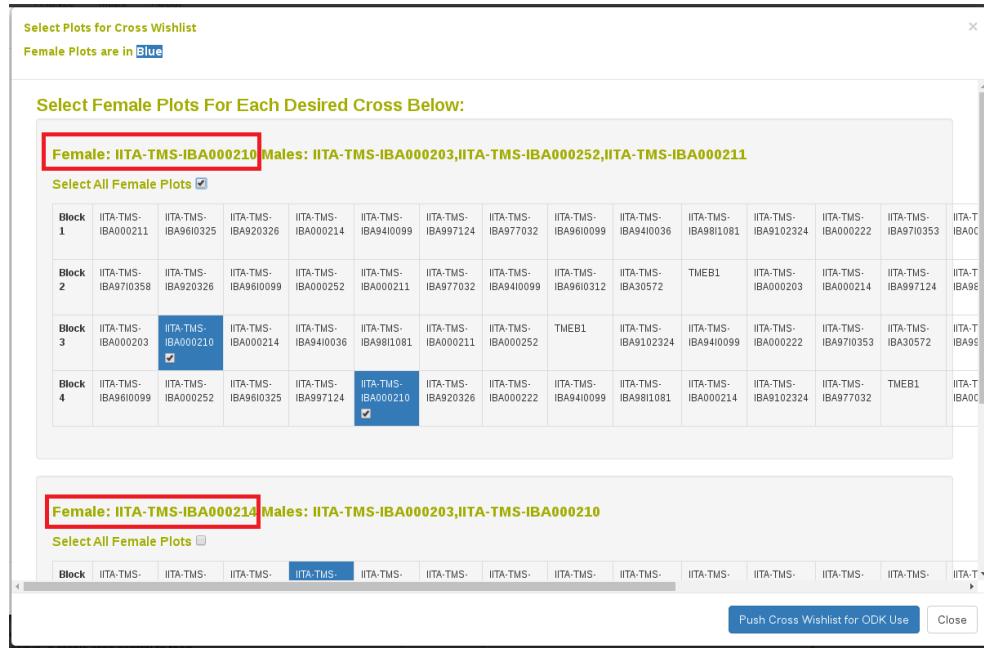
Create Cross Wishlist

Female Accessions	ITA-TMS-IBA010746	ITA-TMS-IBA010758	ITA-TMS-IBA010760	ITA-TMS-IBA010779	ITA-TMS-IBA010797	ITA-TMS-IBA010816	ITA-TMS-IBA010819
ITA-TMS-IBA010746							
ITA-TMS-IBA010758		3		1			2
ITA-TMS-IBA010760							
ITA-TMS-IBA010779				1	1	1	
ITA-TMS-IBA010797							
ITA-TMS-IBA010816							
ITA-TMS-IBA010819							

Set Cross Priorities: 1 is highest and 10 is lowest
Female Accessions Are in First Column and Male Accessions Are in Header

Available Cross Wishlist(s) for ODK Use **Next** **Close**

After female and male accessions are selected to cross, either by the “Not Using List” or “Using List” interface, click Next. The next dialog will allow selection of specific female plots to use for the cross. Sections for each female accession selected will appear with the field layout displayed. Selecting all plots in which the female is present indicates that the cross should be performed on all plots where that female accession is present.



Step 3. Transfer the cross wishlist to your mobile crossing application

Clicking “Push Cross Wishlst for ODK Use” will send the cross wishlist plan to the ONA server for use by the mobile ODK application. Crosses can then be performed and recorded in the field using the mobile application. Afterwards, the crosses are sent back to our database and stored.

9.4 Crossing Experiment Detail Page

Information for crosses in the same crossing experiment is compiled in the crossing experiment detail page.

Details for IITA_crossingtrial_2017

⌚ Crossing Trial details

Crossing Trial Name	IITA_crossingtrial_2017
Breeding Program	IITA
Location	Ibadan
Year	2018
Trial Type	crossing_trial
Planting Date	[No Planting Date]
Harvest Date	[No Harvest Date]
Description	To improve disease resistance

Folder [New Folder] | [Change]

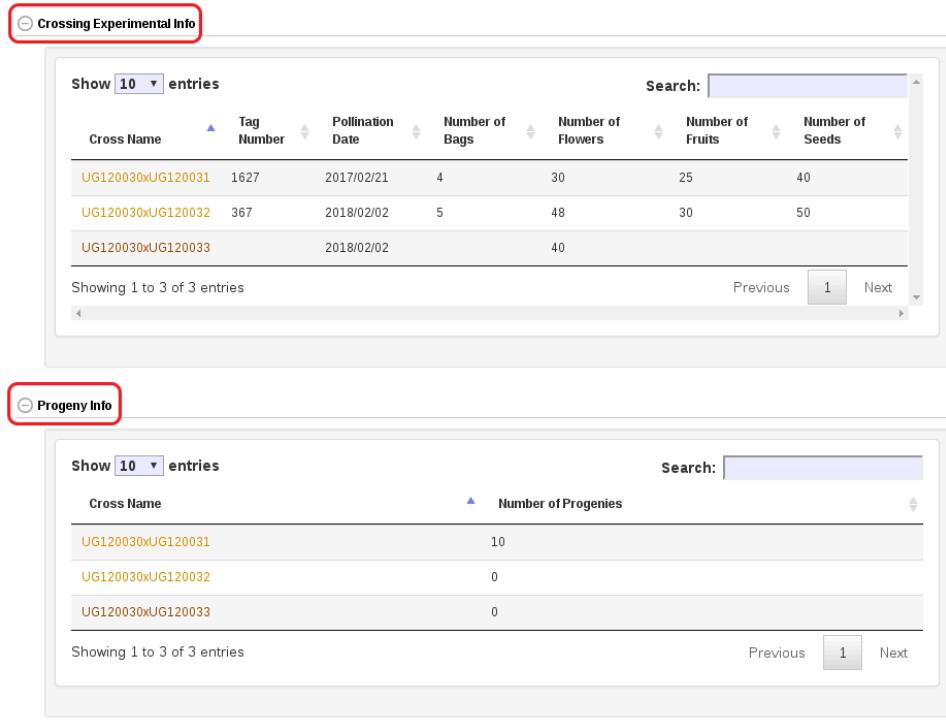
2017

⌚ Crosses in this trial

Show 10 entries Search:

Cross Name	Female Parent	Male Parent	Cross Type	Female Plot	Male Plot
UG120030xUG120031	UG120030	UG120031	biparental	KASESE_TP2013_1627	KASESE_TP2013_909
UG120030xUG120032	UG120030	UG120032	biparental		
UG120030xUG120033	UG120030	UG120033	biparental		

Showing 1 to 3 of 3 entries Previous 1 Next



Crossing Experimental Info

Show 10 entries							Search:
Cross Name	Tag Number	Pollination Date	Number of Bags	Number of Flowers	Number of Fruits	Number of Seeds	
UG120030xUG120031	1627	2017/02/21	4	30	25	40	
UG120030xUG120032	367	2018/02/02	5	48	30	50	
UG120030xUG120033		2018/02/02		40			

Showing 1 to 3 of 3 entries

Previous 1 Next

Progeny Info

Show 10 entries		Search:
Cross Name	Number of Progenies	
UG120030xUG120031	10	
UG120030xUG120032	0	
UG120030xUG120033	0	

Showing 1 to 3 of 3 entries

Previous 1 Next

Each cross name, female parent, male parent, female plot and male plot has a link to its own detail page, which contains information specific to each one. Note: crossing experimental information is customized based on the need for each crop. As a result, the details of the information in your database may be different from the information shown in this manual.

9.5 Cross Detail Page

Information of each cross can also be viewed in its detail page.

Detail for cross 'UG120030xUG120031'

Cross information [Edit]

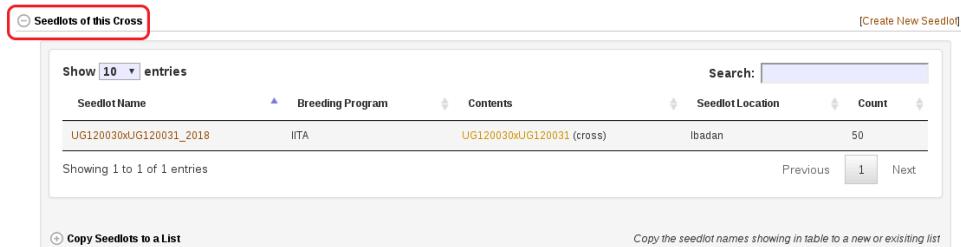
Organism	Solanum lycopersicum
Stock type	cross
Stock name	UG120030xUG120031
Uniquename	UG120030xUG120031
Description	

Parents

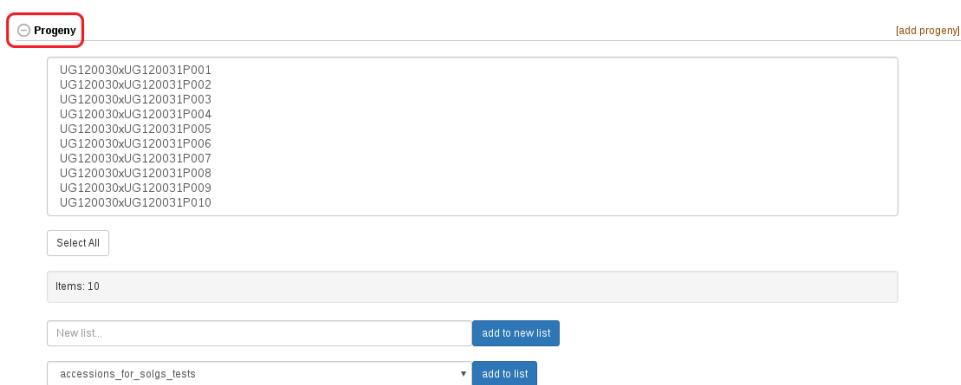
Cross Type	Female Accession	Male Accession	Female Plot	Male Plot
biparental	UG120030	UG120031	KASESE_TP2013_1627	KASESE_TP2013_909

Crossing Experimental Info [Edit]

Tag Number	Pollination Date	Number of Bags	Number of Flowers	Number of Fruits	Number of Seeds
1627	2017/02/21	4	30	25	40

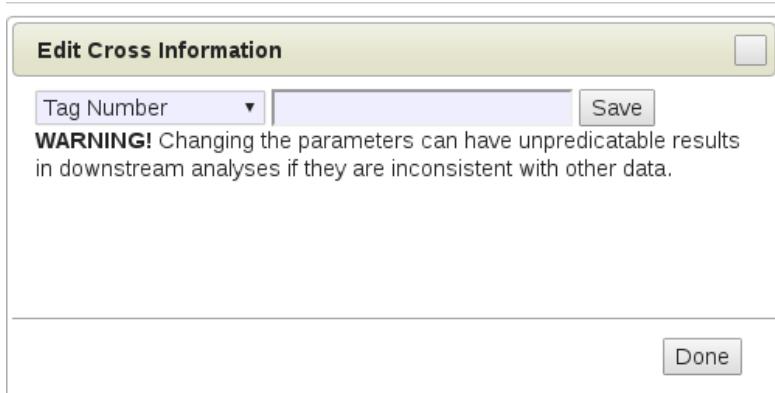


The screenshot shows a table titled "Seedlots of this Cross". The columns are "Seedlot Name", "Breeding Program", "Contents", "Seedlot Location", and "Count". There is one entry: UG120030xUG120031_2018, IITA, UG120030xUG120031 (cross), Ibadan, 50. Below the table, it says "Showing 1 to 1 of 1 entries". At the top right, there is a link "[Create New Seedlot]".

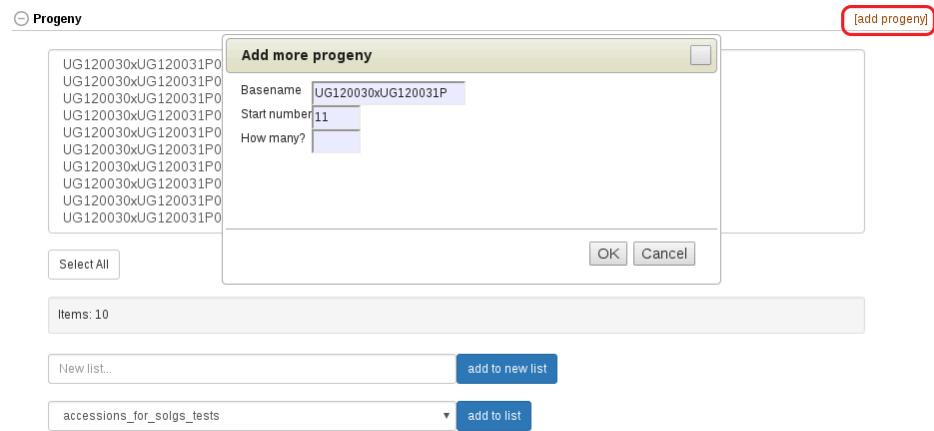


The screenshot shows a list of progeny items under the heading "Progeny". The items are: UG120030xUG120031P001, UG120030xUG120031P002, UG120030xUG120031P003, UG120030xUG120031P004, UG120030xUG120031P005, UG120030xUG120031P006, UG120030xUG120031P007, UG120030xUG120031P008, UG120030xUG120031P009, UG120030xUG120031P010. Below the list are buttons for "Select All", "Items: 10", "New list...", "add to new list", "accessions_for_soilgs_tests", and "add to list".

This page allows you to update or edit crossing experimental information and add progenies related to that cross. Note: crossing experimental information is customized based on the need for each crop. As a result, the details of the information in your database may be different from the information shown in this manual.



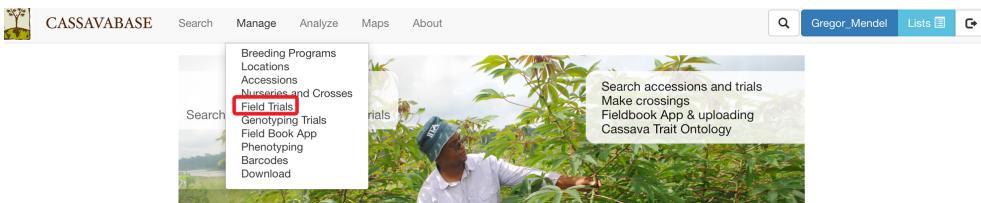
The screenshot shows a dialog box titled "Edit Cross Information". It has a "Tag Number" dropdown menu and a "Save" button. A warning message reads: "WARNING! Changing the parameters can have unpredictable results in downstream analyses if they are inconsistent with other data." At the bottom right is a "Done" button.



Chapter 10

Managing Field Trials

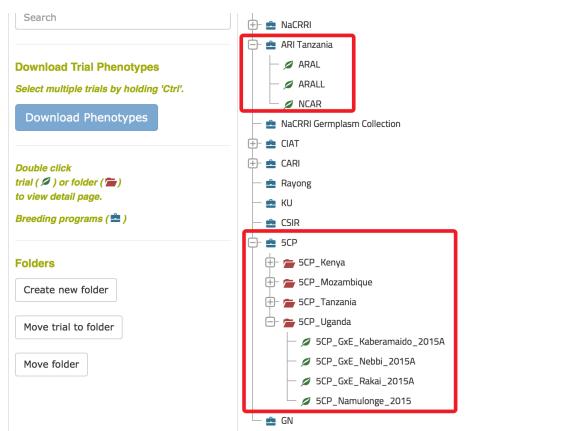
To view trial details on the database, click on the “Field Trials” link under the “manage” menu on the toolbar.



Clicking on the “Field Trials” link will bring you to the “Manage Trials” page. On this page, trials are organized according to their breeding programs. To access trial details, click on the + icon next to your breeding program.

A screenshot of the 'Manage Trials' page. The left side has a search bar and a 'Download Trial Phenotypes' button. The right side shows a tree diagram of breeding programs: IITA, NRCRI, NaCRI, ARI Tanzania, NaCRI Germplasm Collection, CIAT, CARI, and Rayong. Each program node has a plus sign icon to expand it.

Trials can be placed directly in their breeding program. Alternatively, they can be organized by using folders within each breeding program. Clicking on trial name will take you directly to the trial details page.



10.1 Trial Detail Page

The trial detail page displays important information about individual trials including breeding program, location, year, description of the trial, design, and any files associated with that trial.

Trial Details

View and edit basic information about the experiment.

Edit Trial Details

Trial Name	21.Mulungu.EPR
Breeding Program	IITA
Trial Location	Mulungu (Congo, the Democratic Republic of the)
Year	2021
Stock Type Being Evaluated in This Trial	accession
Number of Stocks in This Trial	55
Trial Type	phenotyping_trial
Planting Date	[No Planting Date]
Harvest Date	[No Harvest Date]
Description	21.Mulungu.EPR
Folder	DRC_2021
	New Folder Change Folder
Created by	Dhirkullah Mahmud Kehinde 2021-11-26
Latest Trial Activity	[No Record]

21.Mulungu.EPR SGN9967

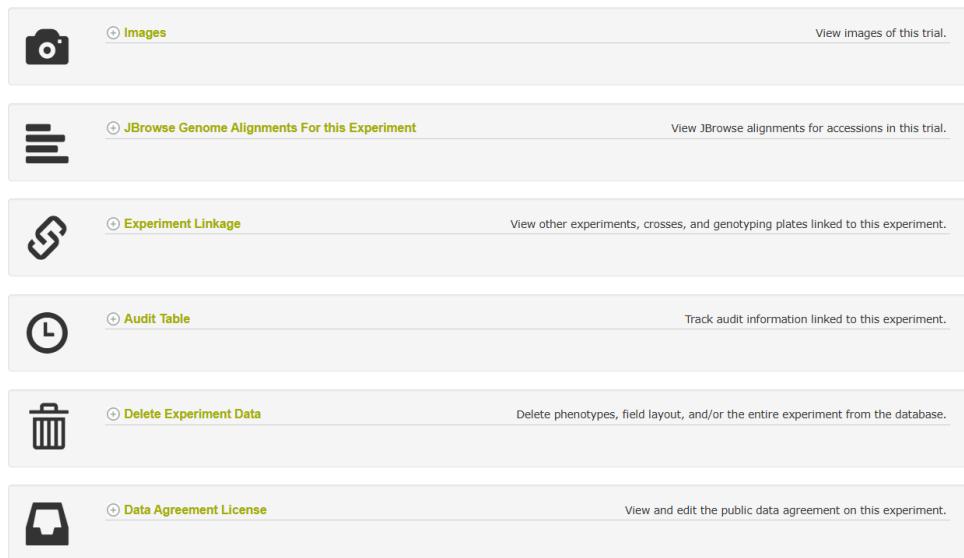
Plot Width (m)	[No Plot Width]
Plot Length (m)	[No Plot Length]
Field Size (ha)	[No Field Size]
Trial Will Be Genotyped	no
Trial Will Be In Crosses	no

Below the trial details you will find various menus for accessing and modifying trial data. There are sections for printing labels for your plots or plants, recording phenotypes, viewing your trial layout or design, viewing

phenotypes for this trial, or conducting analyses.

The screenshot displays a grid of 14 tool cards, each with an icon, a title, and a brief description. The tools are organized into two main sections: top section (6 tools) and bottom section (8 tools).

Section	Icon	Title	Description
Top Section		Trial Status	View progress of this trial.
		Generate barcode labels for plots or plants or accessions in this trial.	Go
		Directly record phenotypes to database for this trial.	Go
		Field Layout Tools and Phenotype Heatmap	View and edit the spatial layout of the experiment. Also view a heatmap for phenotyped traits.
		Experimental Design	View and add experimental design information. Add plant entries and tissue sample entries.
		Phenotype Summary Statistics	View and download uploaded phenotype data.
Bottom Section		Phenotype Raw Data	View phenotyping raw data.
		Compute New Phenotypes	Compute derived traits or compute plot phenotypes from plant phenotypes.
		Upload Data Files	Phenotypic data collection using Excel or Android Fieldbook. Also upload any additional files for this trial.
		Analysis Tools	Run correlation, anova, population structure (PCA), or clustering. Also access HIDAP, if enabled on this site.
		NIRS Data	View and plot NIRS data for this trial.
		Weather Data	View rainfall, temperature, light intensity and other weather data for this location.
		Soil Data	View and upload soil data for this trial.



The “transplanting date” field feature will only be displayed if it has a value. To add a transplanting date after creating a trial, change the show_transplanting_date parameter from 0 to 1 in the SGN config file. As a result, you will be able to add a date under the transplanting date field by clicking the “Edit Trial Details” on the trial detail page.

The screenshot shows the 'Edit Trial Details' dialog box. It contains the following fields:

- Trial Name: 199934HBEP_cara
- Breeding Program: demo
- Location: test_location
- Year: 1999
- Trial Type: Preliminary Yield Trial
- Planting Date: 06/04/1999
- Transplanting Date: Clear
- Harvest Date: July 11, 2024
- Description: EPR
- Field Size (ha): 8
- Plot Width (m): 5
- Plot Length (m): 5
- Trial Will Be Genotyped: No
- Trial Will Be Crossed: No

A small green icon indicating a pending change is visible near the bottom left. At the bottom right are 'Cancel' and 'Save Changes' buttons.

10.2 Adding Trials

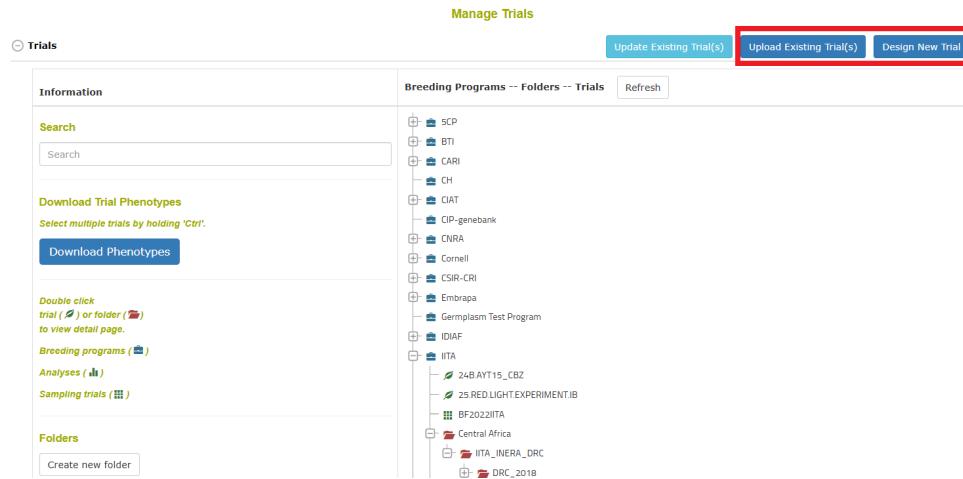
Only users with the account status of “submitter” may create trials. To learn how to change your account status from “user” to “submitter” visit section 1.2.

10.2.1 Prerequisites

- To add a trial, all of your accessions should already exist in the database before you begin to design a trial. If you have accessions that are not in the database, see the instructions in *Managing Accessions*.
- The breeding program and location for your trial should also exist in the database. If you need to add breeding program and/or location to the database, see the instructions in *Managing Breeding Programs* and *Managing Locations* respectively.

On the “Manage Trials” page, there are two methods to create trials: by

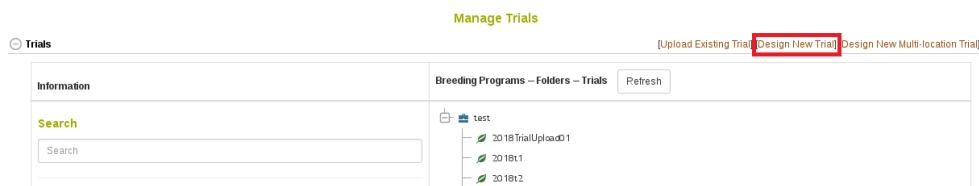
selecting “Upload Existing Trial(s)” to create a trial or trials from a spreadsheet; or by selecting “Design New Trial” and entering the trial data by hand.



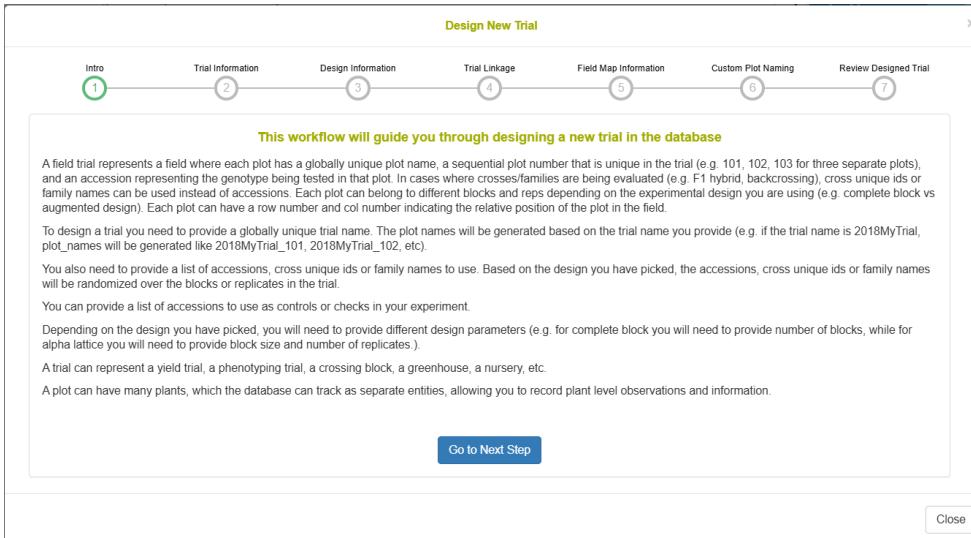
10.2.2 Adding a trial by using “Design New Trial” form

Step 1. Begin the “Design New Trial” workflow

Click on “Design New Trial” to begin.



The first step in this workflow is an introduction that looks like this:



Here it gives information about what is required for a trial, including that to create a new trial, you need to create a list of the accessions that you would like to use in the trial. Lists can be viewed, created, and modified with the “lists” tool at the upper right of the screen. For more information on lists, click [here](#).

Step 2. Enter Trial Information

On this screen you need to enter basic information about the trial, such as breeding program and location(s). You must also select a design type, such as Complete Block Design. The design is important because it influences how your genotypes are distributed and randomized over the trial. You must first click validate before proceeding to the next step.

Design New Trial

Enter basic information about the trial

Breeding Program: BTI

Locations: (One or More)

- Abuja
- Agro-Owo
- Boycott Thompson Institute
- Cornell Biotech
- Ibadan

Locations Selected: 2

Trial Name: 2025_user_manual
Location abbreviation will automatically be added as a prefix if multiple locations are selected.

Trial Type: Preliminary Yield Trial

Year: 2025

Planting Date:

Plot Width (m):

Plot Length (m):

Field Size (ha):

Plants per Plot: 10
Creates plant entries for each plot. Ignore if not adding plant entries.

Interventions Management Factor(s) from Plot:

Description: Demonstrating the trial creation process for the user manual

Stock Type Being Evaluated In Trial: accession

Design Type: Completely Randomized
Generates a Completely Randomized design (CRD) with equal or different repetition, using the methods of random number generation in R. Creates plot entities in the database.

Use same randomization for all locations:

First validate the form **Continue to Next Step**

Close

Step 3. Enter Design Information

On this screen you need to specify a list of accessions to use in the experiment. This list must be a valid list of accessions. You must also specify all required design information, such as number of replicates. In this case, the number of blocks must be given.

Design New Trial

Intro 1 Trial Information 2 Design Information 3 Trial Linkage 4 Field Map Information 5 Custom Plot Naming 6 Review Designed Trial 7

Design your trial layout

Which accessions will be in the field?

List of accessions to include (required):

List of checks to include. Checks list should be separate from accessions list. (optional):

Need to create a list?

Number of replicates (required):

Continue to Next Step

Close

Step 4. Enter Trial Linkage Information (Optional)

This next section allows you to associate this new trial with other field trials, crossing experiments, or genotyping plates already present in the database. This is optional, and can be completed at a later date from the trial detail page.

Design New Trial

Intro 1 Trial Information 2 Design Information 3 Trial Linkage 4 Field Map Information 5 Custom Plot Naming 6 Review Designed Trial 7

Is your trial linked with other field trials, genotyping plates, or crossing experiments in the database? If you are unsure, you can skip this. This information can be added from the trial detail page after the trial is saved.

Is this trial following-up a previous field trial?:

If you go on to collect tissue samples for creating a 96 well plate for genotyping, when adding the genotyping plate (96 well plate layout) to the database you can use plot names or plant names or tissue sample names from this field trial. By doing so, we can create linkage between this field trial and the genotyping plate.

Will this trial be genotyped?:

If you go on to perform crosses on this field trial, each cross can be linked to specific female and male plots. When you upload these crosses we can then automatically link this field trial to the crossing experiment in the database.

Will crosses be done on this trial?:

Continue to Next Step

Close

Step 5. Enter Field Map Information (Optional)

On this screen you can specify how the row and column numbers will be generated for the plots in the trial. The row and column number represent a relative position of the plot in the field. If you are not exactly sure of how you will plant the plots in the field or you have an irregular (non-rectangular) layout, you can skip this step for now. This information can be added on the Trial Detail Page once the trial is saved in the database in order to reflect exactly how the plots were planted in the field.

The screenshot shows the 'Design New Trial' interface with the title 'Design New Trial' at the top. Below it is a horizontal navigation bar with seven steps: 'Intro' (step 1), 'Trial Information' (step 2), 'Design Information' (step 3), 'Trial Linkage' (step 4), 'Field Map Information' (step 5, highlighted in green), 'Custom Plot Naming' (step 6), and 'Review Designed Trial' (step 7). The main content area is titled 'Specify the number of rows and columns for the entire field'. It contains instructions and notes about field map display settings. A section for 'Field map display' has a checked checkbox. Fields for 'Number of rows (required)' and 'Plot layout format' (set to 'Serpentine') are also present. A blue 'Continue to Next Step' button is at the bottom of the form. A 'Close' button is located in the bottom right corner of the window.

Step 6. Custom Plot Naming (Optional)

On this screen it is possible to change the format in which plot names will be generated for your trial. It is recommended to skip this step and just use the format generated by the database by default.

Design New Trial

If you want to change the way in which plot names will be generated by the database

It is recommended to allow the database to create the plot prefixes, so leave the prefix blank unless necessary.

Custom plot naming/numbering:

block based plot numbers (increment leading digit for every block)

consecutive plot numbers throughout the blocks

Plot prefix:

Plot start number:

Plot number increment:

Continue to Next Step

Close

Step 7. Review Designed Trial

On this screen you can review the trial that the database has generated.

You will see a graphical representation of the trial. The numbers on the squares represent the plot_number of each plot and on mouse hover you can see further information about the plot.

Design New Trial

Review the generated trial layout. Make sure to click Submit at the bottom of this page if you approve of the trial!

Check to confirm that your design looks good. If there are any problems you can redo the randomization step.

Legend: Even Block Numbers (e.g. 2,4,...) Odd Block Numbers (e.g. 1,3,...) Checks Odd Rep Numbers (e.g. 1,3,...) Even Rep Numbers (e.g. 2,4,...)

1	1001	1002	1003	1004	1005	1006	1007	1008	1009	1010	1011
2	1022	1021	1020	1019	1018	1017	1016	1015	1014	1013	1012
3	1023	1024	1025	1026	1027	1028	1029	1030	1031	1032	1033
4	1044	1043	1042	1041	1040	1039	1038	1037	1036	1035	1034
5	1045	1046	1047	1048	1049	1050	1051	1052	1053	1054	1055

Close

You will also see a table representation of all the plots and their information.

If you want to redo the randomization, you can click the “Redo Randomization” button.

Design New Trial

Plot Name	Accession Name	Check Name	Plot Number	Row number	Col number	Block Number	Block Row Number	Block Col Number	Rep Number	Seedlot Name	Num Seeds Per Plot
2018MyTrial1_rep1_test_accession1_1001	test_accession1		1001	1	1	1				1	
2018MyTrial1_rep1_test_accession5_1002	test_accession5		1002	1	2	1				1	
2018MyTrial1_rep1_test_accession4_1003	test_accession4		1003	1	3	1				1	
2018MyTrial1_rep1_test_accession3_1004	test_accession3		1004	1	4	1				1	
2018MyTrial1_rep1_test_accession2_1005	test_accession2		1005	1	5	1				1	
2018MyTrial1_rep2_test_accession4_2001	test_accession4		2001	2	5	2				2	
2018MyTrial1_rep2_test_accession2_2002	test_accession2		2002	2	4	2				2	
2018MyTrial1_rep2_test_accession5_2003	test_accession5		2003	2	3	2				2	
2018MyTrial1_rep2_test_accession1_2004	test_accession1		2004	2	2	2				2	
2018MyTrial1_rep2_test_accession3_2005	test_accession3		2005	2	1	2				2	
2018MyTrial1_rep3_test_accession3_3001	test_accession3		3001	3	1	3				3	
2018MyTrial1_rep3_test_accession1_3002	test_accession1		3002	3	2	3				3	
2018MyTrial1_rep3_test_accession2_3003	test_accession2		3003	3	3	3				3	
2018MyTrial1_rep3_test_accession5_3004	test_accession5		3004	3	4	3				3	
2018MyTrial1_rep3_test_accession4_3005	test_accession4		3005	3	5	3				3	

Redo Randomization

Close

At the bottom there is a brief summary of the trial followed by two buttons.

Design New Trial

Trial Is Valid
The following trial will be added

Design type
Randomized Complete Block Design

Number of locations
1

Number of accessions
5

Number of blocks
3

Number of accessions per block
Block 1: 5 accessions
Block 2: 5 accessions
Block 3: 5 accessions

Number of reps
3

Treatments:

Add Field Management Factor(s) to Design **Confirm (Saves Trial In Database)**

Close

Step 8. Add Field Management Factors to your design (Optional)

You can add Field Management Factors by clicking “Add Field Management Factor(s) to Design”. Clicking this opens a dialog to specify your factor. You can name this to account for fertilizer, watering regime, inoculation, or anything else. The types of management factors available is customizable in the SGN configuration file. This is optional and can be added from the trial detail page afterwards.

Add Field Management Factor to Design

Add Field Management Factor Name:	e.g. Fertilizer01_20N40P50K or Weeding01_Manual
Add Field Management Factor Description:	e.g. Fertilizer Brand Sprayed with 20N40P50K
Field Management Factor Type:	Fertilizer
Field Management Factor Year:	2025
Field Management Factor Date:	
Applied To:	Plots

Continue **Close**

Click “Continue” and a dialog will appear where you can specify plots or plants (if you added plants during trial creation) for which the factor was applied. There is a select all button as well.

Add Field Management Factor to Design									
plot_name	accession	plot_number	block_number	rep_number	is_a_control	row_number	col_number	Fertilizer_Nov_9_2017_20% [Select all]	
2017_tutorial_trial_1001	002D	1001	1	1	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1002	0000test2	1002	1	1	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1003	003D	1003	1	1	undefined	undefined	undefined	<input checked="" type="checkbox"/>	
2017_tutorial_trial_1004	004D	1004	1	1	undefined	undefined	undefined	<input checked="" type="checkbox"/>	
2017_tutorial_trial_1005	002B	1005	1	1	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1006	002B	1006	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1007	002D	1007	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1008	001B	1008	1	1	1	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1009	0000test1	1009	1	1	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1010	002B	1010	1	3	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1011	003D	1011	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1012	004D	1012	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1013	0000test1	1013	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1014	001F	1014	1	1	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1015	001F	1015	1	2	undefined	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1016	001D	1016	1	1	1	undefined	undefined	<input type="checkbox"/>	
2017_tutorial_trial_1017	001D	1017	1	2	1	undefined	undefined	<input type="checkbox"/>	

Apply Field Management Factors to Plants (if available): Continue Close

Step 9. Saving new trial in the database

Once you are done reviewing the trial you can click “Confirm” to save the generated trial into the database. Once the trial has saved you will see the final completion screen:

Design New Trial

Complete! Your trial was saved in the database.

The trial was saved successfully

- You may want to proceed to the trial detail page for the trial you just created.
- You can print barcodes for the plots or plants or tissue samples in this trial.
- You can add phenotypes for the plots or plants in this trial now.

The trial was saved to the database with no errors! Congrats Click Here

Close

10.2.3 Adding a trial from an uploaded file

If you already have trial design layout in a spreadsheet, you can add your trial into the database by using the “Upload Existing Trial(s)” button on the

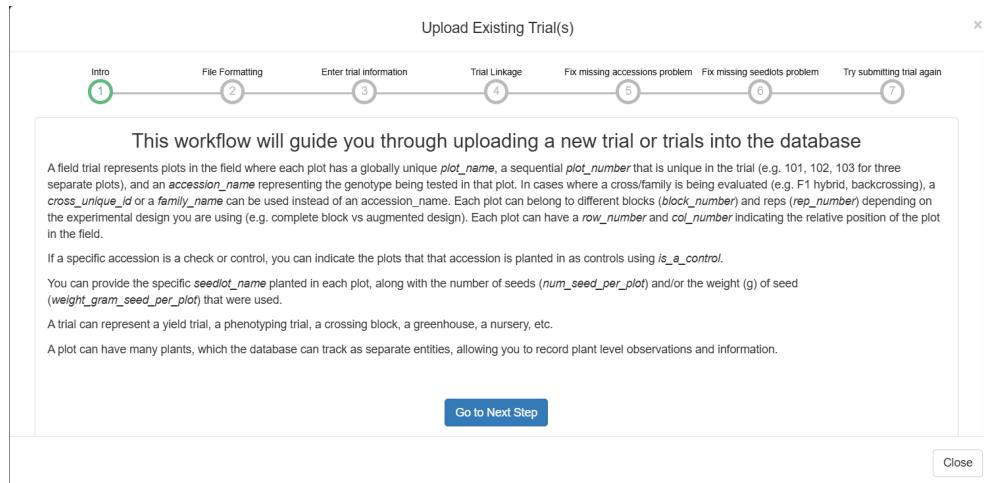
Manage Trials page.

The screenshot shows the 'Manage Trials' interface. At the top, there are three buttons: 'Update Existing Trial(s)', 'Upload Existing Trial(s)' (which is highlighted with a red box), and 'Design New Trial'. Below these buttons is a breadcrumb navigation path: 'Breeding Programs -- Folders -- Trials'. On the left side, there is a sidebar with sections for 'Information', 'Search' (with a search bar), 'Download Trial Phenotypes' (with a 'Download Phenotypes' button), 'Double click trial () or folder () to view detail page.', 'Breeding programs ()', 'Analyses ()', and 'Sampling trials ()'. On the right side, there is a tree view of breeding programs and their associated folders. The tree structure includes: SCP, BTI, CARI, CH, CIAT, CIP-genebank, CNRA, Cornell, CSIR-CRI, Embrapa, Germplasm Test Program, IDIAF, ITA, 24BAY15_C6Z, 25 RED LIGHT EXPERIMENT.IB, BF202IIITA, Central Africa, IITA_INERA_DRC, and DRC_2018.

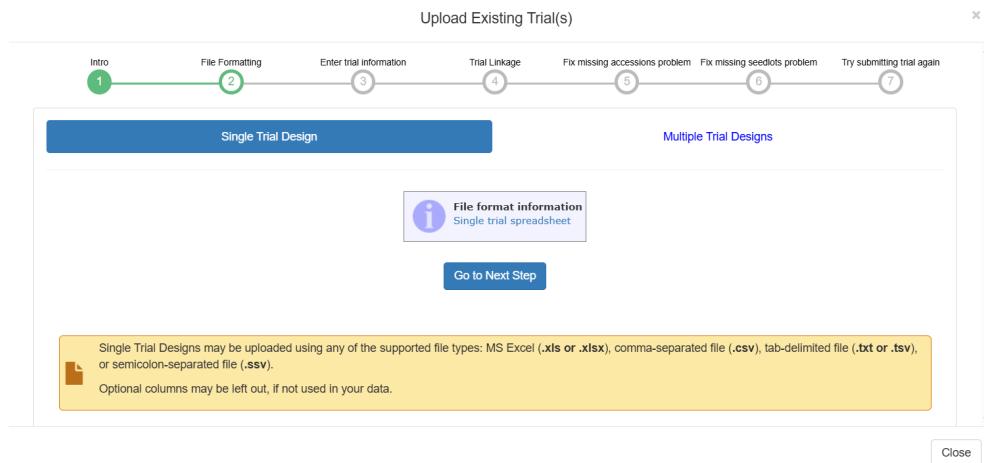
Enter the information in the workflow to upload a trial from a spreadsheet.

Step 1:

The first step is to understand what the format of the trial upload is. It is important to understand that the field layout represents plots in the experiment. Each plot has a globally unique plot_name, a sequential plot_number that is unique in the trial (but not globally unique. e.g. 101, 102, 103 for three separate plots), an accession_name representing what genotype is planted in that plot, and a block_number representing design replication. Each plot can be thought of as having a row_number and a column_number representing the relative position of the plot in a grid (e.g. the top left plot is row 1 column 1 following by row 1 column 2). Each plot can be planted with an amount of seed from a seedlot, where the seedlot_name represents the specific seed packet that was used, and num_seed_per_plot and weight_gram_seed_per_plot represent amount that were transferred from the seedlot_name to the plot_name. Treatments can be applied onto plots using additional column names in your file, where a 1 represents if the treatment was applied to the plot and an empty cell means it was not applied.



The following page will allow you to pick a file for upload, including uploading multiple trials at once. On this page you can also inspect the file requirements for both single trial and multi-trial uploads.



Upload Template Information

Trials may be uploaded in an Excel file (.xls or .xlsx)

Stock type being evaluated in this trial: accession

Stock type being evaluated in this trial: cross unique id

Stock type being evaluated in this trial: family name

Required fields:

- accession_name or cross_unique_id or family_name (must exist in the database. This is the accession or cross unique id or family name being tested in the plot.)
- plot_number (a sequential number for the plot in the field (e.g. 1001, 1002, 2001, 2002). These numbers should be unique for the trial.)
- block_number (a design parameter indicating which block the plot is in)

Optional fields:

- plot_name (must be unique across entire database. If not provided in the file, it will be automatically generated as `{trial_name}-PLOT_{plot_number}`.)
- is_a_control (type 1 in this field if the plot is a control, otherwise leave blank. generally you will have accessions/cross unique ids/family names that are controls, so you should indicate the plots of those accessions/cross unique ids/family names as a control.)
- rep_number (replicate number, numeric)
- range_number (range number, often synonymous with col_number, numeric)
- row_number (row number. If the field is a grid, this represents the y coordinate, numeric, required for field map generation.)
- col_number (column number. If the field is a grid, this represents the x coordinate. Sometimes called range_number, numeric, required for field map generation.)
- seedlot_name (the seedlot from where the planted seed originated. Must exist in the database)
- num_seed_per_plot (number seeds per plot. Seed is transferred from seedlot mentioned in seedlot_name. Numeric)
- weight_gram_seed_per_plot (weight in gram of seeds in plot. Seed is transferred from seedlot mentioned in seedlot name. Numeric)
- entry_number (a trial-level entry number assigned to the stock. Numeric)

Treatments:

- treatment columns (additional column(s) that specify the name of a treatment (e.g. inoculated, drought, etc). The value for each plot should be 1 if the treatment was applied or empty.)

Only the required fields are necessary to include in the upload template. You may add any additional optional fields. The fields can be in any order.

Minimum File requirements

- All accession names in the file must exist in the database. See adding accessions for more information.
 - The uploaded file can be excel (.XLXS or .XLS), comma-separated values (.CSV), tab-separated values (.TSV) or semicolon-separated values (.SSV).
 - The first row (header) must contain the column names: plot_name accession_name plot_number block_number is_a_control rep_number range_number row_number col_number seedlot_name num_seed_per_plot weight_gram_seed_per_plot entry_number
 - Only accession_name, plot_number, and block_number are required.

Minimal Example:

plot_ancessplot_bandsareimportedfromotherheadbutmaxweightperphotiseaddedper_plot
2018plot1 ad01ssidn1 1
2018plot2 ad02ssidn2
2018plot3 ad02ssidn2

```
plot_analysplot_handsomeplot_mgrowingnumberofplotsmaxweightperplotusedper_
2018plot4a2022si0n1 1
```

When uploading multiple trials, the requirements are the same, but with additional headers to differentiate trials in the same file: trial_name breeding_program location year transplanting_date design_type description trial_type trial_stock_type plot_width plot_length field_size planting_date harvest_date. Additionally, when uploading multiple trials, you may choose to get an email when the upload is complete (see section 10.2.4)

File validation

- In case of errors in the uploaded file such as missing or invalid data, a window will appear listing the specific errors in the file that must be corrected before a successful upload.

Uploading a trial with Treatments

- You can upload a trial with treatments by adding additional column(s). The column header will be the treatment e.g. fertilizer, watering regime, inoculation, etc. and the values in these columns will be either 1 or empty, indicating that the treatment was applied to the plot or not.

Step 2:

Once you feel that your experiment field layout is in the right format, click on to the Next Step. You will see the following form which must be filled in completely:

Upload Existing Trial(s)

Enter information about the experiment and upload your trial layout

Trial Name:

Breeding Program:

Location:

Trial Type:

Year:

Planting Date (MM-DD-YYYY):

Transplanting Date(MM-DD-YYYY):

Plot Width (m):

Plot Length (m):

Field Size (ha):

Plants per Plot:
Creates plant entries for each plot. Ignore if not adding plant entries.

Inherits Management Factor(s) From Plots:

Assign row and column data to plants within plots?

Description:

Stock Type Being Evaluated in Trial:

Design Type:

Upload File: No file chosen

[Go To Next Step](#)

[Close](#)

The trial name must be globally unique in the database. Please try to follow standard naming conventions for your group.

Step 3:

Go to the next page where you can link this trial to other projects. Validate the form, and then you can click “Upload Trial”.

X

Upload Existing Trial(s)

Intro 1
 File Formatting 2
 Enter trial information 3
 Trial Linkage 4
 Fix missing accessions problem 5
 Fix missing seedlots problem 6
 Try submitting trial again 7

Is your trial linked with other field trials, genotyping plates, or crossing experiments in the database? If you are unsure, you can skip this. This information can be added from the trial detail page after the trial is saved.

Is this trial following-up a previous field trial?:

If you go on to collect tissue samples for creating a 96 well plate for genotyping, when adding the genotyping plate (96 well plate layout) to the database you can use plot names or plant names or tissue sample names from this field trial. By doing so, we can create linkage between this field trial and the genotyping plate.

Will this trial be genotyped?:

If you go on to perform crosses on this field trial, each cross can be linked to specific female and male plots. When you upload these crosses we can then automatically link this field trial to the crossing experiment in the database.

Will crosses be done on this trial?:

Check this box to ignore any possible warning messages and save the trial to the database.

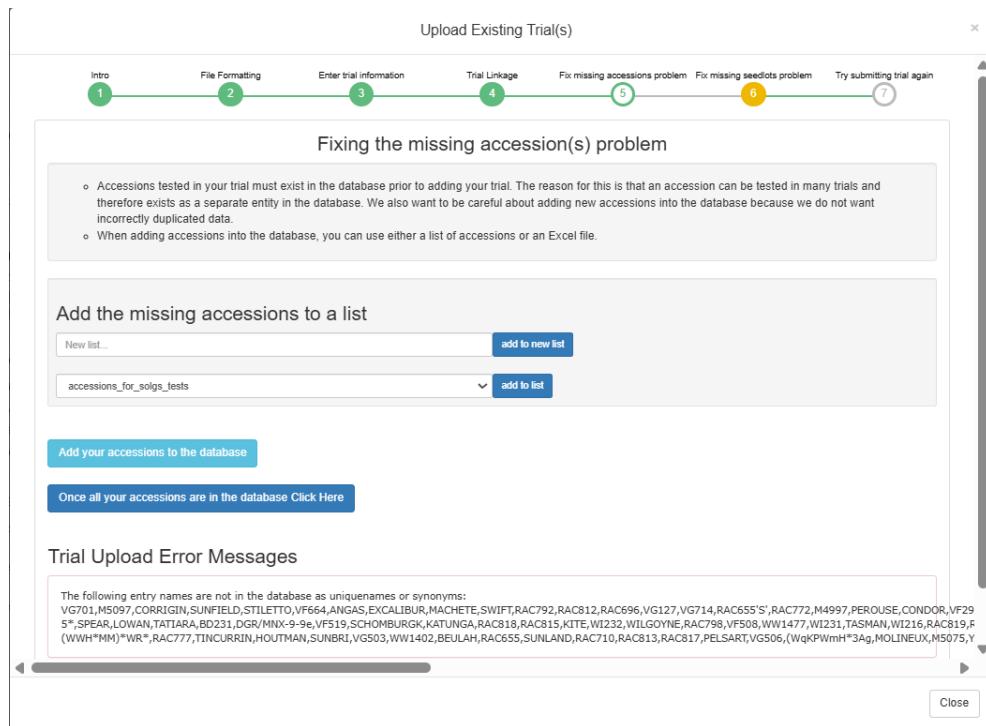
Ignore Warnings?:

First validate the form
 Upload Trial

Close

Step 4:

In the case where you have uploaded an experiment using accession_names that are not already present in the database, you will be taken to this screen. If the accession_names in your file are all already in the database, this step will be skipped. The reason it is necessary for your accessions to be in the database before you can add a trial using them is that a single accession can be used among many trials and therefore must exist as a separate entity in the database; because of this it is also very important to be careful about adding wrongly duplicated accession_names into the database. From this screen it is possible to make a new list with the missing accession_names and then click “Add Accessions to the database” to immediately resolve the issue. Once all your accessions are in the database, click to move to the Next Step.



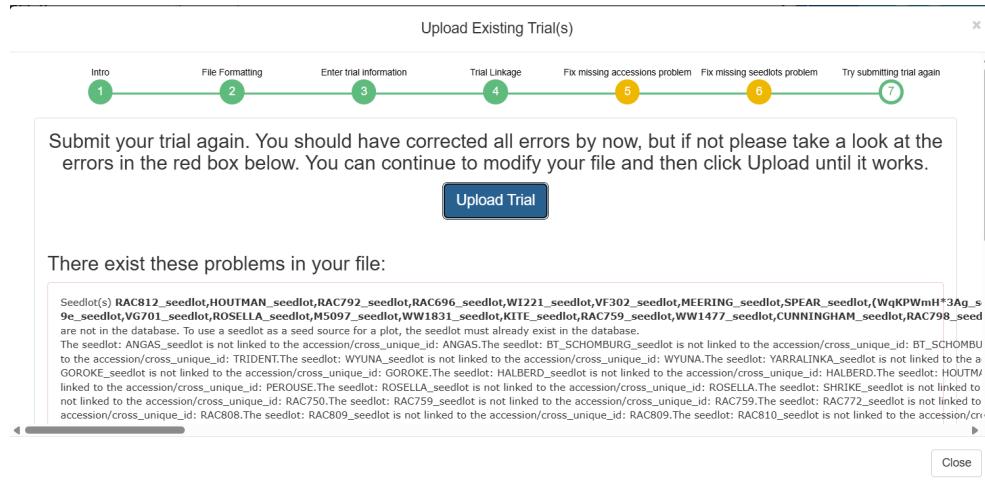
Step 5:

In the case where you have uploaded an experiment using seedlot_names that are not already present in the database, you will be taken to the next screen. If the seedlots in your file are all already in the database, this step will be skipped. The reason it is necessary for your seedlots to be in the database before you can add a trial using them is that a single seedlot can be used among many trials and therefore must exist as a separate entity in the database. Once all your seedlots are in the database, click to move to the next step.

Step 6:

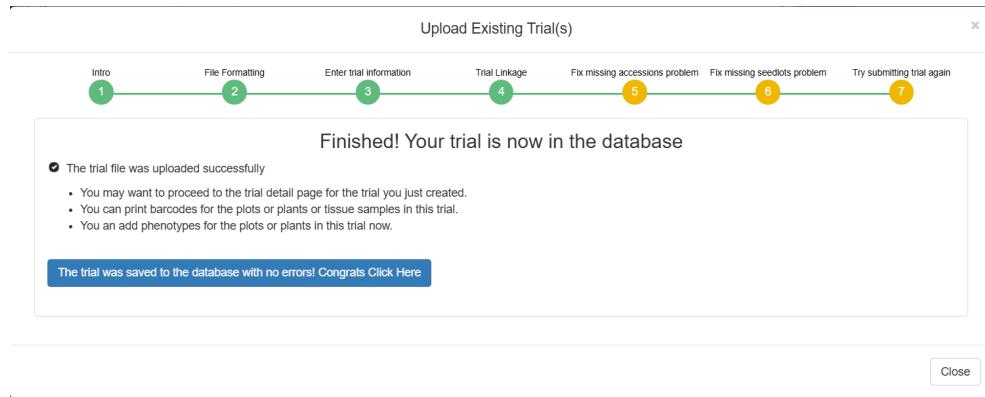
If there are any other errors with your file, such as if the plot_names are not globally unique in the database or your plot_numbers are not unique in your trial or row_number is not an integer or any other error, you will see the errors listed in the red box. It is up to you to correct these errors in your file. Simply open up the file you selected earlier in Excel and correct

the issues and then save the file. Then you can click “Submit Trial” and it will resubmit it for you. You can continue to edit your file here and submit as many times as you need until it is accepted.



Completion screen

Whether you were lucky enough to submit your trial successfully on Step 2 or if you tried many times on Step 5, once your trial has been saved in the database you will see the following screen:



10.2.4 Multi-location trials

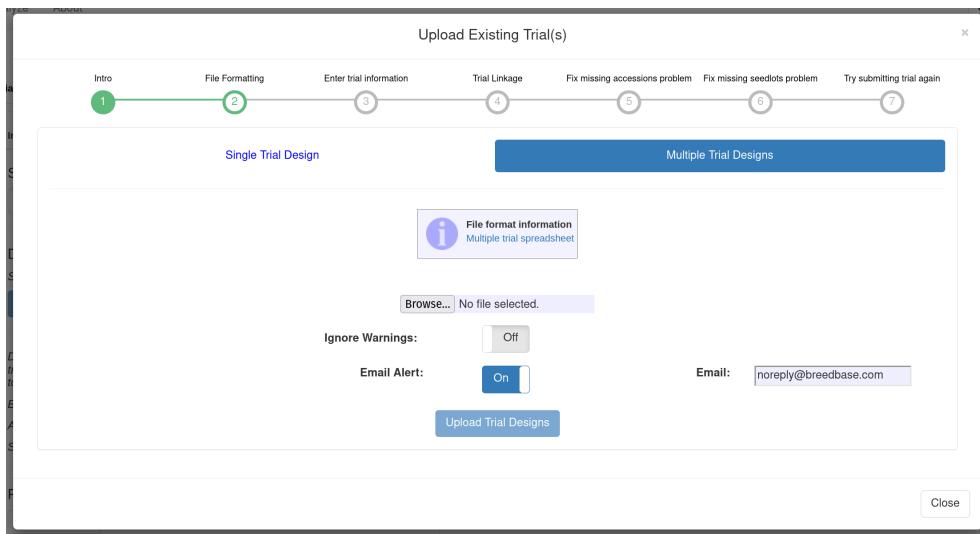
To add multi-location trials, simply select the multiple locations while using the ‘Add Trial’ form.

This will create a separate trial for each selected location, but they will share the same design and will be grouped in a single folder.

By default each trial design will have a fresh randomization, but if desired you may check the “Use same randomization for all locations” option.

Email alert for multiple trial design upload

When uploading multiple trials from a file, you have the option to receive email notifications by clicking the “Email Alert” checkbox. By default, the system will use the email address associated with your account, but you have the option to enter a different email address if you prefer. After submitting, the upload process runs in the background, allowing you to continue using the interface without interruptions. Once the process completes, you will receive an email with the upload results.

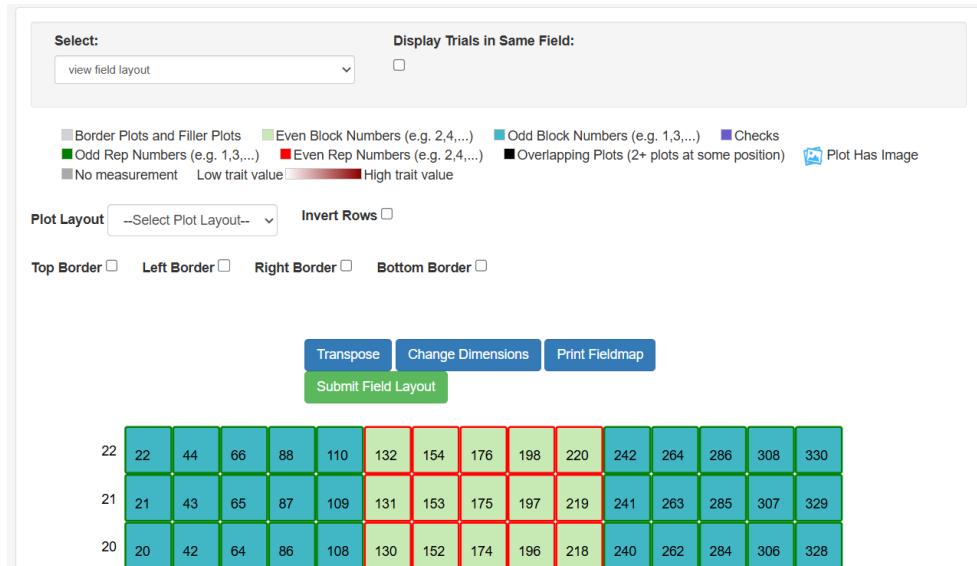


10.2.5 Viewing Plot Layout and Trait HeatMap

10.2.5.1 Viewing plot layout

In the “Field Layout Tools and Phenotype Heatmap” section of a Trial Detail page, the trial physical layout is displayed by default. The relative position of the plots will be displayed based on the row and column positions given to the plots during the trial creation or upload steps. The plots are color-coded

based on the plot's rep and block numbers and whether or not it is used as a check. Hover the mouse over the plot to see details about a specific plot.



10.2.5.2 Viewing plot layout for multiple trials

If there is more than one trial grown in the same physical field, the trial layouts of all of the trials can be shown together if the trials share these properties:

Each trial has the same year

Each trial has the same location

The location type of the trials' location is set to Field

The row and column positions of all of the plots (across the related trials) don't overlap. For example, trial #1 starts at row 1 and trial #2 starts at row 10.

When these conditions are met and you check the "Select Trials in Same Field" checkbox, the plots from all of the related trials will be displayed on the same field layout. The plots will be color-coded by trial. The planting order and harvest order downloads will include the plots from all of the displayed trials in the order in which the plots occur in the field.

Select: view field layout Display Trials in Same Field:

Trials in Same Field:

- SAMPLE_A_2022
- SAMPLE_B_2022

Border Plots and Filler Plots Even Block Numbers (e.g. 2,4,...) Odd Block Numbers (e.g. 1,3,...) Checks Odd Rep Numbers (e.g. 1,3,...)
 Even Rep Numbers (e.g. 2,4,...) Overlapping Plots (2+ plots at some position) Plot Has Image

Download Plot Order Include Borders

Plot Layout Invert Rows

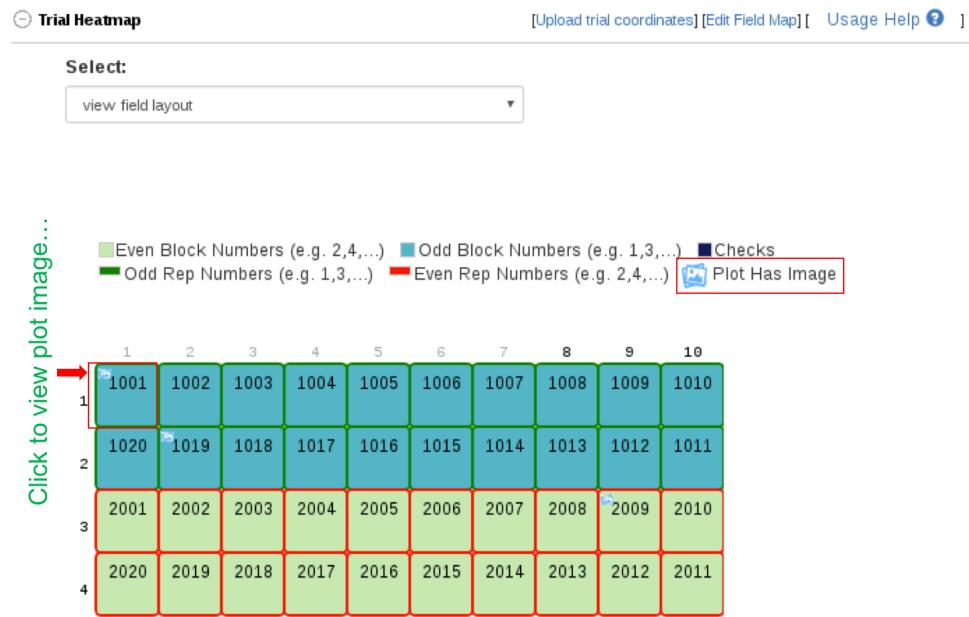
Top Border Left Border Right Border Bottom Border

NOTE: Field Map cannot be modified and borders are not shown when linked trials are displayed.

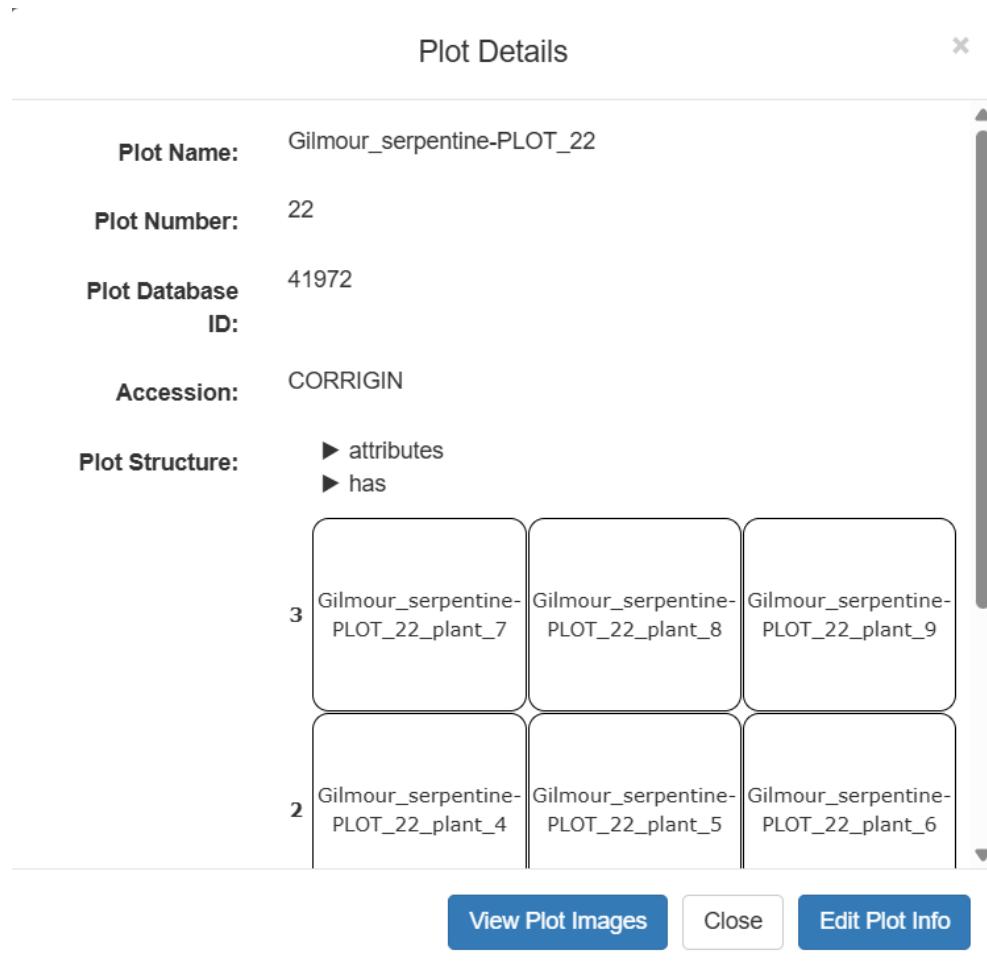
	1	2	3	4	5	6	7	8	9	10
6	301	302	303	304	305	306	307	308	309	310
5	201	202	203	204	205	206	207	208	209	210
4	101	102	103	104	105	106	107	108	109	110
3	301	302	303	304	305	306	307	308	309	310
2	201	202	203	204	205	206	207	208	209	210
1	101	102	103	104	105	106	107	108	109	110

Tracking plot images on fieldMap

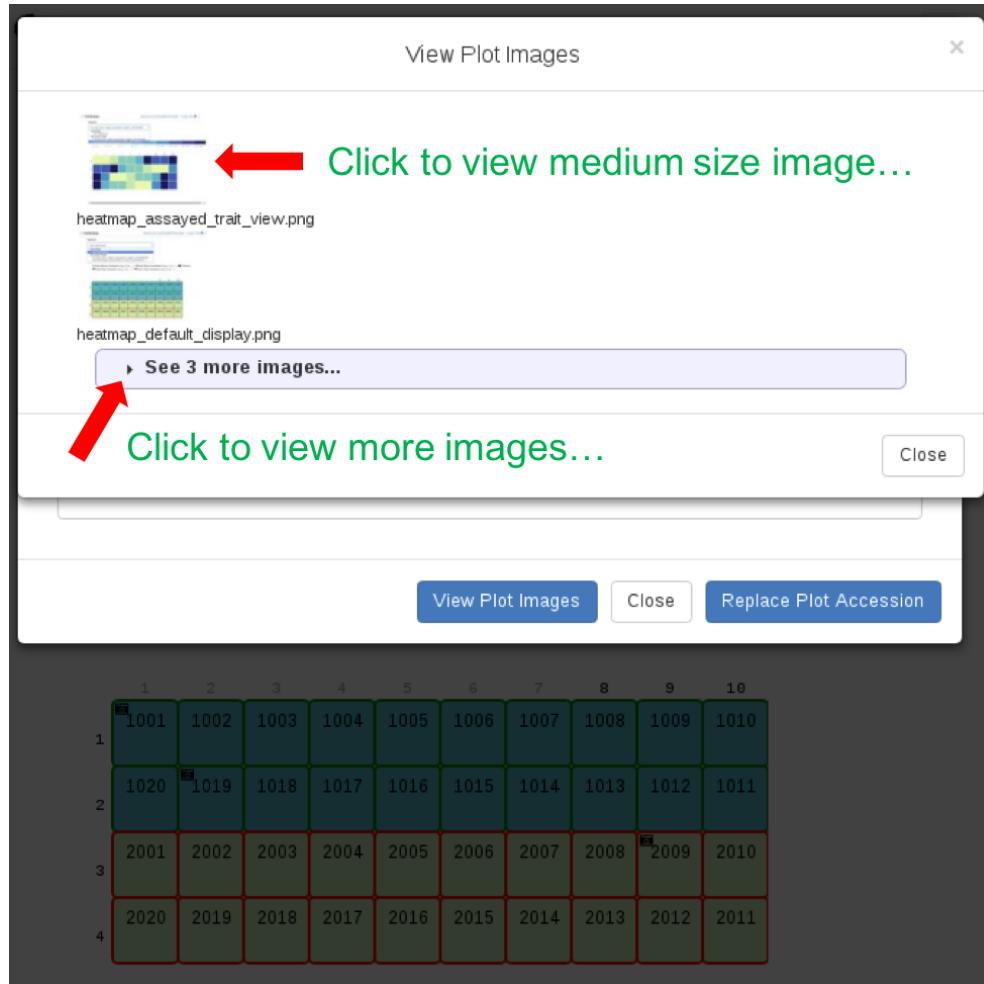
Plot images can be seen on fieldMap if a plot is associated to any image.



To view plot image(s), click on a plot, a dialog will appear. In this dialog you will see a detailed overview of the plot, including what stock(s) are contained within it and their layout in the plot, if applicable. At the bottom of the dialog, you will see options to change the plot name or accession as well as view plot images.



On the resulting dialog, click on “View Plot Images.” To see more images if a plot has more than 2 images, click on See more images... Medium size of an image can be viewed by clicking on an image.



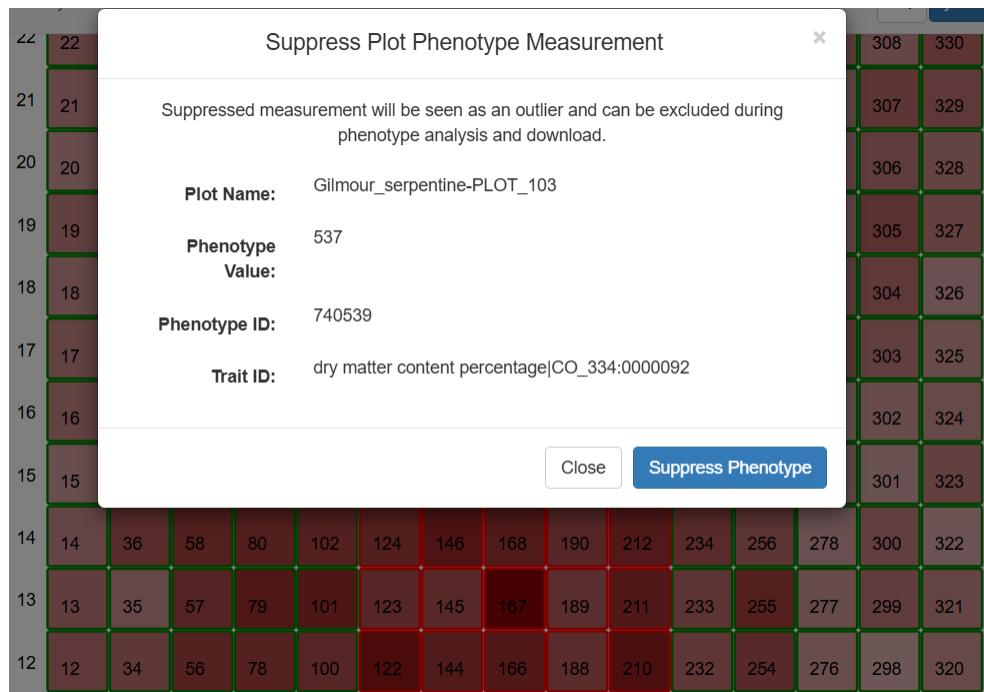
Viewing assayed trait heatmap

The phenotype heatmap can be viewed by selecting a specific assayed trait from the selectbox drop-down. Mousing over the plots highlights the plot in green and also displays the plot's field information including the selected trait's phenotype value.



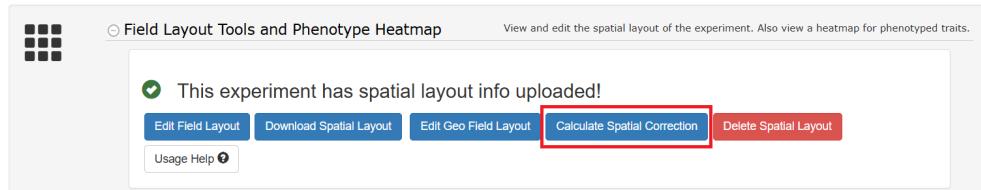
Suppressing Plot Phenotype

Clicking on a plot on the heatmap would display a dialog that has a button for suppressing a plot phenotype value for a given trait. A suppressed plot value can be excluded during trial analysis and phenotype download.



10.2.5.3 Correcting spatial autocorrelation

For trials with spatial layout information and stored phenotypes, you can check and correct for spatial autocorrelation by clicking the “Calculate Spatial Correction” button above the field map. Doing so will open the spatial corrections dialog. For an in-depth review, visit the chapter on *data analysis tools*.

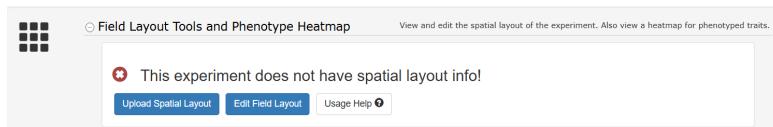


10.2.6 Adding additional information in the *Trial Detail* page

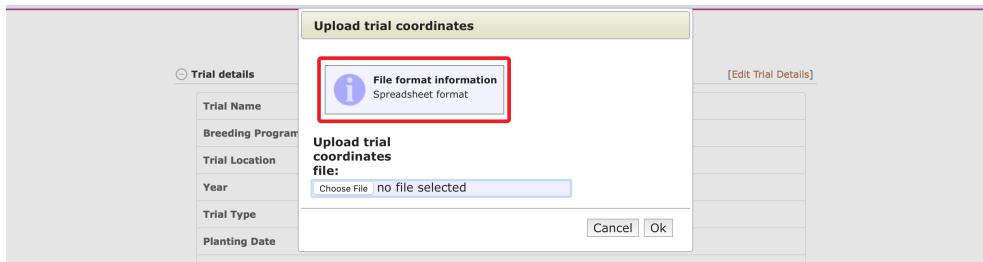
After you added a new trial to the database, you can edit trial details or add more information for that trial through the *Trial Detail* page.

Uploading Physical Trial Layout {-}

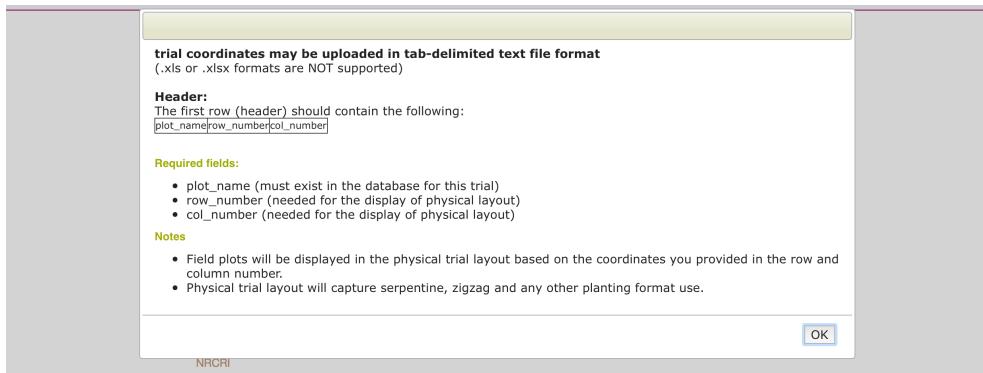
You can upload physical trial layout by clicking on the “Upload Spatial Layout” button on the *Trial Detail* page.



Please check file format carefully. You can find file format information by clicking on the “Spreadsheet format” on the “Upload Spatial Layout” window.



Spreadsheet format:



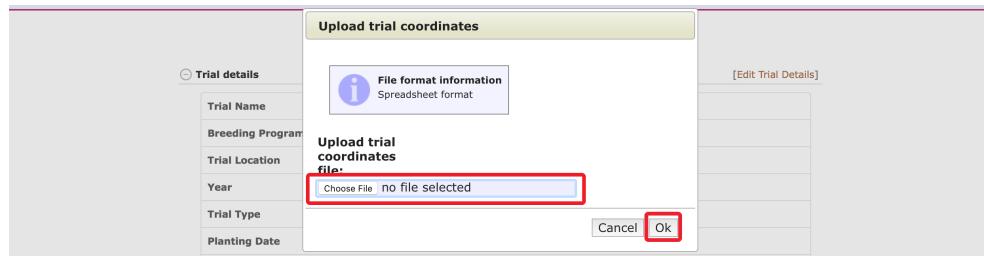
Physical Trial Layout File requirements

- All plot names in the file must exist in the database.
- The uploaded file should be tab delimited (txt).
- The first row (header) must contain the column names

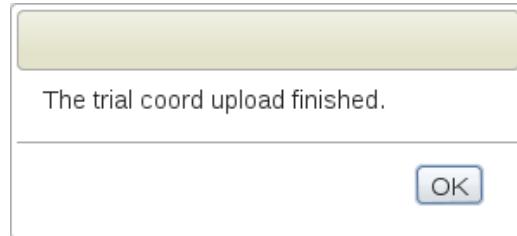
Example:

plot_name	row_number	col_number
plot1	1	1
plot2	1	2
plot3	1	3

Select the trial layout coordinates file that you want to upload for this trial, then click “OK” button to upload the file.

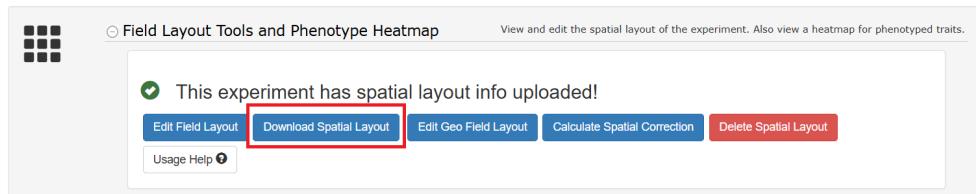


The following message is displayed after the coordinates are uploaded.



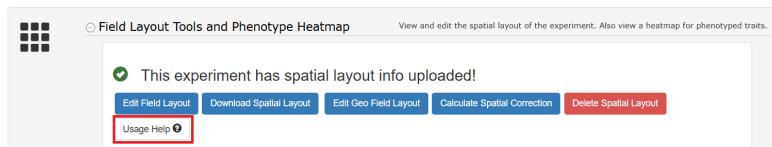
Downloading Field Map Spreadsheet

Field map spreadsheet can be downloaded if the trial has field coordinate (row and column numbers) uploaded for its plots. To download, click on the Download FieldMap Layout link on the Trial Heatmap section.



Editing Physical Trial Layout

The “Usage Help” button contains information on how to edit physical trial layout.



How to Use and Edit Field Map

Background:

Field map is a tool that enable users to view the physical layout of plots in a trial. Maps can be generated on the fly while adding a trial or uploading a trial. Field map coordinates can also be uploaded independently after trials have been added or uploaded. It is a very intuitive, flexible and user friendly tool for manipulation/making changes to field trial layouts before phenotypes are uploaded.

Editing Options:

Replace Plot Accession

A plot accession can be replaced by an accession within or outside of the trial. To do this, **click on the plot and provide the name of the new accession** (must already exist in the database).

Replace Trial Accession

An accession used in a trial can be replaced by a new accession or another accession from the trial. When this replace option is used, it replaces every instances (plots and plants) of that accession in the trial. To do this, **click on the Edit Field Map link** by the top right of the physical trial layout section; **click on Replace Accession button**; **select accession** to replace from the trial and **provide a new accession** (must already exist in the database); **click on Replace Trial Accession button** to complete the operation.

Change dimensions

Click on the change dimensions button to alter the dimensions of the field map, and the respective row and column attributes of the plots in your trial. Clicking on this button will open a dialogue that prompts you to enter an accession name if the change you are making will result in dimensions that are larger than the amount of plots in your trial. The result is that the map will fill in the extra space with filler plots, and the filler plots will have the accession entered in input in the dialogue.

Invert rows

Check invert rows to observe your trial from the top down instead of the bottom up. This change is only present on your visualization, and the trial data will not be changed.

Plot Layout

You can select how you want your map to be ordered. The options include serpentine and zigzag.

Download Harvesting Order

This button allows you to download an excel file which has an incrementally ordered list of the harvesting order for your trial based on the current layout of the field map. You can play around with the four options given to experiment with the different traversal patterns available. The harvesting order file will omit border plots, but it will include filler plots.

Download Planting Order

This button allows you to download an excel file which has an incrementally ordered list of the planting order for your trial. It is very similar to the harvesting order, but since you will need to plant the borders too, it includes the border plots in the traversal of your map.

Add Borders

These checkboxes allow you to add borders around your field map on any sides you would like. These plots are purely visual and have no real data associated with them. They will not be included in the database, but you can see them in your harvesting order if they are present.

Submit Field Layout

This button allows you to update the databases with the changes you have made to your trial. You can continuously change your trial data, just remember to submit the changes when you are satisfied so that it is saved.

HeatMap

If your trial has trait data uploaded, you can view a heatmap of that data by selecting a trait from the dropdown menu. You will see a color weighting on the plots that corresponds with the value of each plot for a given trait.

Transpose Fieldmap

This button transposes the x and y coordinates of each plot in your map. Borders will adjust accordingly.

Features:**Mouse Over**

Displays plot field information.

Double Click

Double clicking on a plot, opens the stock page for that plot.

Download Map

Field Map can be downloaded as image using the download button below the map.

Delete Map

Field Map can be deleted if the user have the right privilege.

Note:

- You have to be a **curator** or a **submitter and associated to the breeding program** of the trial to use the features of this tool.

- **Input boxes** used within the field map tool will automatically (**autocomplete**) give accession name options from the database when you start typing the accession name.

- **Changes can only be made to the physical layout when phenotypes are not yet upload for the trial.**

There are two different options for editing trial layout:

- Replacing plot accession by clicking on the plot in the layout.
- Replacing trial accession by using the “Edit Field Layout” button.

To edit a specific plot, click on that plot. Enter a new accession on the “Edit Plot Info” form, then click the “Replace Plot Accession” button.

Plot Details

Plot Name: Gilmour_serpentine-PLOT_22

Plot Number: 22

Plot Database ID: 41972

Accession: CORRIGIN

Plot Structure:

- ▶ attributes
- ▶ has

3	Gilmour_serpentine-PLOT_22_plant_7	Gilmour_serpentine-PLOT_22_plant_8	Gilmour_serpentine-PLOT_22_plant_9
2	Gilmour_serpentine-PLOT_22_plant_4	Gilmour_serpentine-PLOT_22_plant_5	Gilmour_serpentine-PLOT_22_plant_6

[View Plot Images](#) [Close](#) [Edit Plot Info](#)

Replace Plot Accession ×

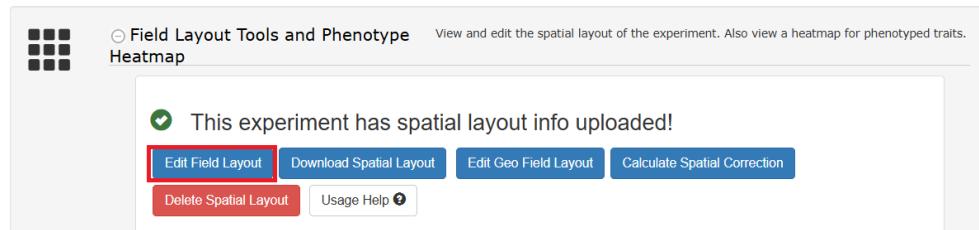
Selected Plot Information:

Enter New Accession:

New Plot Name:
(Optional)

Close Replace Plot Accession

To replace an accession (in every plot/plant of that accession), click on the “Edit Field Layout” button.



On the “Edit Field Layout” window, click the “Replace Accession” button.

Edit Field Map For Gilmour_serpentine ×

Replace Trial Accessions Within A Trial Plots

Close Replace Accession

Select any accession that you want to replace, enter your new accession, then click the “Replace Trial Accession” button.

Replace Trial Accession in Gilmour_serpentine ×

Select Accession:	<input type="text" value="CORRIGIN"/>
Enter New Accession:	<input type="text"/>

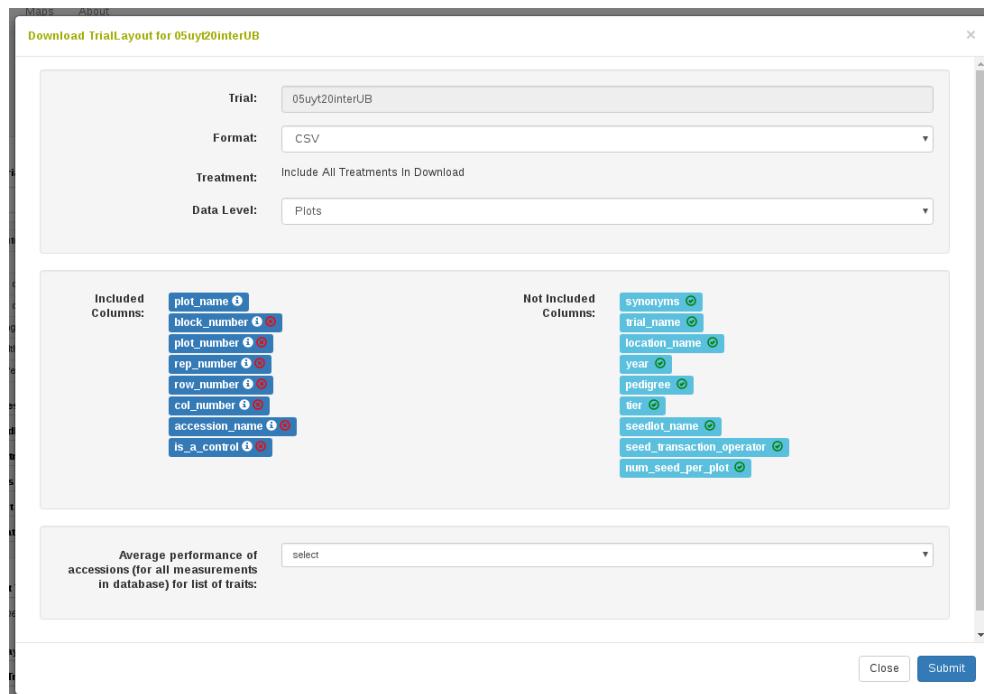
Close
Replace Trial Accession

10.2.7 Downloading the Trial Layout from the *Trial Detail* page

Click on “Download Layout” on the *Trial Detail* page under “Experimental Design”.

Attribute	Value
Design	CRD
Number of Blocks	3
Number of Replicates	3
Plot Length	
Plot Width	
Subplots Per Plot	
Plants Per Plot	9

The trial layout includes all information regarding the observation units in the experiment. The observation units can be plots, plants, or subplots. The trial layout can include trial design information such as the block_number and rep_number. It can also include physical map information such as the row_number and col_number, if that information is available for the trial. The trial layout also includes information regarding treatments that have been applied in the field. Optionally, the layout can give information regarding accession's global performance for a list of traits.



10.2.8 Adding Plant Entries To Your Trial

After you added a new trial to the database you can choose to add plant entries to your trial. Adding plant entries enables plant level phenotyping. It is generally better to enter data at the plant level into the database because it is always possible to calculate plot level phenotypes from the individual plant data.

Plant entries can be added to your trial in four ways: (1) Automatically generated by the database. The only input required is the number of plants per plot. (2) Uploaded in an XLS or XLSX file. This allows you to specifically name your plant entries. (3) Uploaded using plant index number instead of name (the name is automatically generated) (4) Uploaded using the number of plants per plot. This allows different numbers of plant per plot, as opposed to option 1.

Additionally, if a trial has had subplots added, each of these options can be used to add plants to subplots. Note that once subplots are added, plants cannot be added directly to plots, and once plants have been added, subplots cannot be created.

These options are available in the “Plant Entries” section on the *Trial Detail* page under “Experimental Design,” as shown below.

The screenshot shows the 'Experimental Design' section of a web application. At the top, there are several buttons: 'Download Layout', 'Upload Seedlots Planted In Trial', 'Upload GPS Coordinates', 'Add Management Factor', and 'Update Experimental Design Type'. Below these are buttons for 'Change Accessions' and 'Add Management Factor'. The main area contains a table of experimental design attributes with their values:

Attribute	Value
Design	CRD
Number of Blocks	1
Number of Replicates	3
Plot Length	
Plot Width	
Subplots Per Plot	
Plants Per Plot	

Below this table is a sidebar with expandable sections: 'Accessions' (Number of accessions: 307), 'Seedlots' (Number of seedlots: 0), 'Controls' (Number of controls: 0), 'Plots' (Number of plots: 921), 'Subplots' (Number of subplots: 0), and 'Plant Entries'. The 'Plant Entries' section is expanded and highlighted with a red arrow pointing to it. It contains a sub-section 'Add Plants to Plots:' with four buttons: 'Add plant entries', 'Upload plant entries using plant_name', 'Upload plant entries using plant index number', and 'Upload plant entries using number of plants per plot'.

Automatically Generate Plant Entries

Clicking on “Add plant entries” opens the following dialog box. The only input required is the number of plants per plot. This will create plant entries that are named as a concatenation of the plot_name and the plant’s index number e.g. plot_name_plant_1. You may optionally add row and column data to give plants a spatial layout within each plot.

The dialog box has a title bar 'Add plants to test_t' and a close button 'x'. The main area contains the following settings:

- Maximum number of plants per plot:** An input field with a value of 1.
- Inherits Management Factor(s) From Plots:** A checked checkbox.
- Assign row and column data to plants within plots?** An unchecked checkbox.

At the bottom right are 'Close' and 'Save' buttons.

Upload Plant Entries

Alternatively, you can choose to upload an XLS or XLSX file that contains the names of the plant entries, the plant index numbers, or the number of plants per plot. Each option comes with a dialog that specifies the file formats.

10.2.9 Adding Tissue Sample Entries To Your Trial

Some trials require tissue samples to be collected from plants in a field trial. The database will generate these tissue sample identifiers for you and will maintain all relationships with the plant, plot, accession, etc. To begin, go to the Design section of a trial's detail page and open the "tissue sample entries" section. Please note that tissue samples are directly related to plants, therefore your trial requires plants before you can add tissue samples.

The screenshot shows a web-based application interface for managing field trials. At the top left, there is a radio button labeled 'Design'. On the right side, there is a link 'Download Layout'. Below these, there are two columns: 'Attribute' on the left and 'Value' on the right. The 'Attribute' column lists: Design, Number of Blocks, Number of Replicates, Plot Length, Plot Width, and Plants Per Plot. The 'Value' column lists: CRD, 2, 2, and empty rows for the last three. Below this table, there is a sidebar with several sections: Accessions, Seedlots, Controls, Plots, Plant Entries, Tissue Sample Entries, and Field Management Factors. The 'Tissue Sample Entries' section is highlighted with a red box. Inside this box is a button labeled 'Add tissue sample entries'. To the right of the sidebar, there are links: '[Upload Seedlots Planted In Trial]' and '[Upload GPS Coordinates]'. At the bottom right, there is a link '[Add Management Factor]'. The entire interface has a clean, modern design with a light gray background and white text.

When you click on "Add tissue sample entries" you will see a dialog where you specify the number of tissue samples you require per plant. Once you have specified how many tissue samples, you can give specific words to distinguish samples, such as "root" or "stem", as seen below.

Add tissue samples to plotxx

WARNING: This trial does not have plant entries. Tissue samples are added for each plant entry, so you must add plant entries first. You can do so on the "Plant Entries" section of the trial detail page.

Number of tissue samples per plant:	<input type="text" value="3"/>
❶ Tissue Name 1:	<input type="text" value="leaf"/>
❷ Tissue Name 2:	<input type="text" value="root"/>
❸ Tissue Name 3:	<input type="text" value="examples: leaf or root or stem"/>
Inherits Management Factor(s) From Plots:	<input checked="" type="checkbox"/>

[Close](#) [Save](#)

Once you have added tissue sample entries they will appear in the design section of the trial as seen below.

Tissue Sample Entries

spt9903_1003_subplot_1_plant_1_leaf1 spt9903_1003_subplot_1_plant_1_root2 spt9903_1003_subplot_1_plant_2_leaf1 spt9903_1003_subplot_1_plant_2_root2 spt9903_1003_subplot_1_plant_3_leaf1 spt9903_1003_subplot_1_plant_3_root2 spt9903_1003_subplot_2_plant_4_leaf1 spt9903_1003_subplot_2_plant_4_root2 spt9903_1003_subplot_2_plant_5_leaf1 spt9903_1003_subplot_2_plant_5_root2

[Select All](#)

Items: 240 Selected: 1

New list... [add to new list](#)

119acc [add to list](#)

Each tissue sample has a detail page where you can add information about the sample, such as if it is in transit or in storage somewhere.

Tissue_sample: spt9903_1003_subplot_2_plant_4_leaf1

[\[New\]](#) [\[Edit\]](#) [\[Delete\]](#)

Organism	Manihot esculenta
Stock type	tissue_sample
Stock name	spt9903_1003_subplot_2_plant_4_leaf1
Unique name	spt9903_1003_subplot_2_plant_4_leaf1
Description	



SGN stock 787404 (spt9903_1003_subplot_2_plant_4_leaf1)

Stock editors:

Synonyms [\[Add...\]](#)

Additional information [\[Add...\]](#)

notes This tissue is on the way to Erlangen [\[X\]](#)

The related stocks section near the bottom of this detail page displays the relationships between all stocks, including tissue samples.

Related stocks

- Related stocks in trials
- Seedlots of this Accession
- Progenies
- Groups /members
- Related stocks for tissue sample

[\[Create New Seedlot\]](#)

Show 10 entries	Search: <input type="text"/>
Name	Type
i6_1015_plant_1_leaf1	tissue_sample
i6_1015_plant_1_root2	tissue_sample
i6_1015_plant_1_stem3	tissue_sample
i6_1015_plant_2_leaf1	tissue_sample
i6_1015_plant_2_root2	tissue_sample
i6_1015_plant_2_stem3	tissue_sample
i6_1015_plant_3_leaf1	tissue_sample
i6_1015_plant_3_root2	tissue_sample
i6_1015_plant_3_stem3	tissue_sample
i6_1018_plant_1_leaf1	tissue_sample

Showing 1 to 10 of 234 entries

Previous [1](#) [2](#) [3](#) [4](#) [5](#) ... [24](#) Next

Copy Stocks to a List

Copy the stock names showing in table to a new or existing list

10.2.10 Uploading GPS Coordinates For Plots

You can upload GPS coordinates for the plots in your trial. There is a link on the Trial Detail Page as shown below.

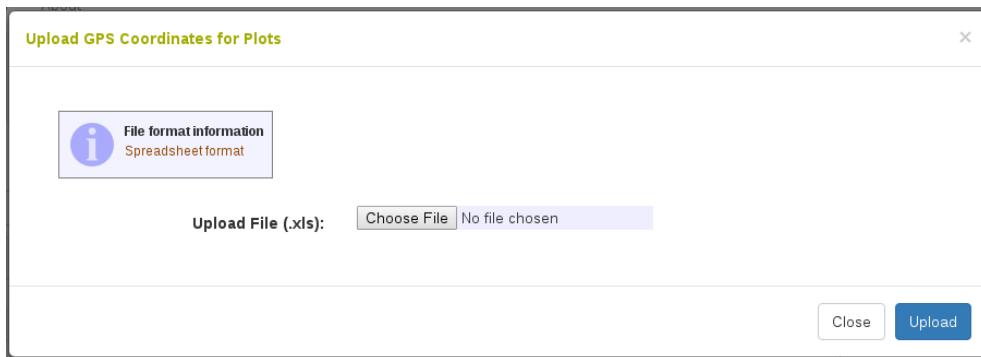
[Download Layout](#)

Attribute	Value
Design	CRD
Number of Blocks	1
Number of Replicates	3
Plot Length	
Plot Width	
Plants Per Plot	3

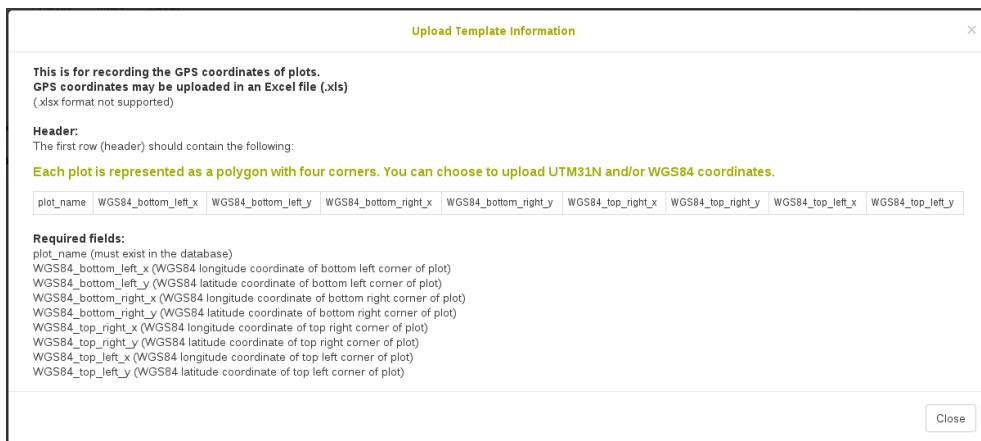
[\[Upload GPS Coordinates\]](#)

[\[Add Management Factor\]](#)

Clicking on this link will bring up the following dialog.



Here you can upload an XLS or XLSX file. To see information on the format of the file that should be uploaded, click on “Spreadsheet format”. This will bring up the following dialog.



This dialog tells you that the file must be XLS or XLSX and must contain: plot_name WGS84_bottom_left_x WGS84_bottom_left_y WGS84_bottom_right_x WGS84_bottom_right_y WGS84_top_right_x WGS84_top_right_y WGS84_top_left_x WGS84_top_left_y The GPS coordinates should be WGS84 format and specify a four-pointed polygon around the plot.

10.2.11 Repetitive Measurements Section

If a trial includes repetitive traits or time-series values, you can effectively view and analyze these values through the Repetitive Measurements Section. Start by selecting the desired trait from the trait drop-down menu. Next, define the date range by either using the date-range picker or an interactive slider, which allows you to dynamically adjust the period you wish to examine. Once the date range is set, determine how to handle the repetitive measurements by choosing from various options such as First Value, Last Value, Averaged Value, Sum Values, or All Values. Choosing the “All Values” option enables an additional feature that visualizes the trend of the values over time, helping you identify patterns and trends within the data.

Observation Unit Name	Accession Name	Repetitive Values	Repetitive line graph
199934HBEPR_cara_rep1_UG120002_1	UG120002	4, 7, 1	
199934HBEPR_cara_rep1_UG120004_2	UG120004	8, 2, 5	
199934HBEPR_cara_rep1_UG120006_3	UG120006	3, 6, 9	

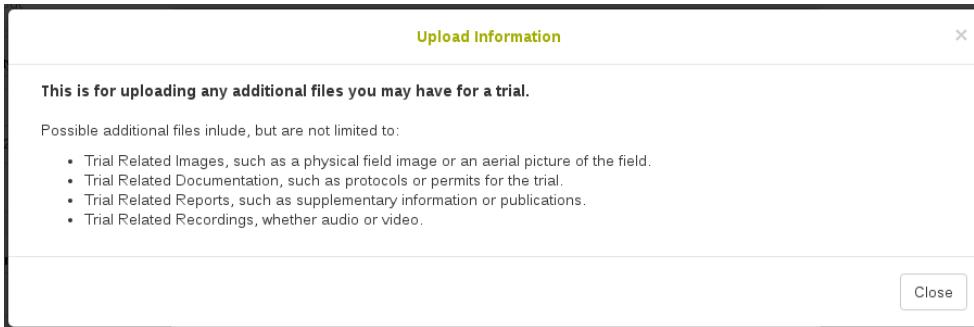
10.2.12 Uploading Additional Files To Trial

It may be of interest to you to upload additional documents, images, or recordings to your trial. To do this, scroll down to the “Uploaded Additional File” section on the trial detail page. From here you can view and download any of these additional files.

Uploaded Additional Files				
Upload Additional Files				
Show 10 entries <input type="text" value="Search:"/>				
Filename	Date Uploaded	Uploaded By	Options	
2018-01-17_15:30:45_2016_mchare_pollination_block	2018-01-17 15:30:49.967178+00	nmorales	Download	
2018-01-17_18:03:42_2016_mchare_pollination_block	2018-01-17 18:03:47.092829+00	nmorales	Download	
2018-01-17_18:12:36_Screenshot from 2017-04-28 12:35:05.png	2018-01-17 18:12:40.924951+00	nmorales	Download	
2018-01-17_18:14:26_Screenshot from 2017-04-28 12:35:05.png	2018-01-17 18:14:30.73281+00	nmorales	Download	
2018-01-17_18:15:38_Screenshot from 2017-04-28 12:35:05.png	2018-01-17 18:15:42.328389+00	nmorales	Download	
2018-01-17_18:17:25_Screenshot from 2017-04-28 12:35:05.png	2018-01-17 18:17:29.467101+00	nmorales	Download	

Showing 1 to 6 of 6 entries Previous [1](#) Next

To upload an additional file, click on the “Upload Additional Files” link. A dialog will appear where you simply select your desired file. For information, you can click “Upload information” to see the following message.



10.3 Updating Trial Data

To updated the trial-level metadata (such as the planting date, design type, description, etc) of one or more existing trials, click the “Update Existing Trial(s)” button from the Manage > Field Trials page. This upload can also be used to rename trials or move trials to a different breeding program. In order to update a trial, you must be a curator or a submitter (that is associated with the breeding program of the trials).



Here you can upload a file that contains the new metadata for the existing trials in the database. The first column is labeled 'trial_name' and includes the name of the existing trial. Additional columns can be included for the metadata you want to update. Any columns not included in the file or values left blank will leave the existing metadata unchanged. The columns that can be included are:

- new_trial_name: A new name for the trial, must not already exist in the database
- breeding_program: The name of breeding program that managed the trial, must exist in the database.
- location: The name or abbreviation of the location where the trial was held, must exist in the database.
- year: The year the trial was held.
- transplanting_date: The transplanting_date of the trial was conducted. Date in YYYY-MM-DD format or 'remove' to remove the date
- planting_date: Date of Planting in YYYY-MM-DD format or 'remove' to remove the date
- harvest_date: Date of Harvest in YYYY-MM-DD format or 'remove' to remove the date
- design_type: The shorthand for the design type, must exist in the database. Possible values include CRD: Completely Randomized Design, RCBD: Randomized Complete Block Design, RRC: Resolvable Row-Column, DRRC: Doubly-Resolvable Row-Column, ARC: Augmented Row-Column, Alpha: Alpha Lattice Design, Lattice: Lattice Design, Augmented: Augmented Design, MAD: Modified Augmented Design, greenhouse: undesigned Nursery/Greenhouse, splitplot: Split Plot, p-rep: Partially Replicated, Westcott: Westcott Design
- description: Additional text with any other relevant information about the trial.
- trial_type: The name of the trial type, must exist in the database. Possible values include Seedling Nursery, phenotyping_trial, Advanced Yield Trial, Preliminary Yield Trial, Uniform Yield Trial, Variety Re-

lease Trial, Clonal Evaluation, genetic_gain_trial, storage_trial, heterosis_trial, health_status_trial, grafting_trial, Screen House, Seed Multiplication, crossing_block_trial, Specialty Trial

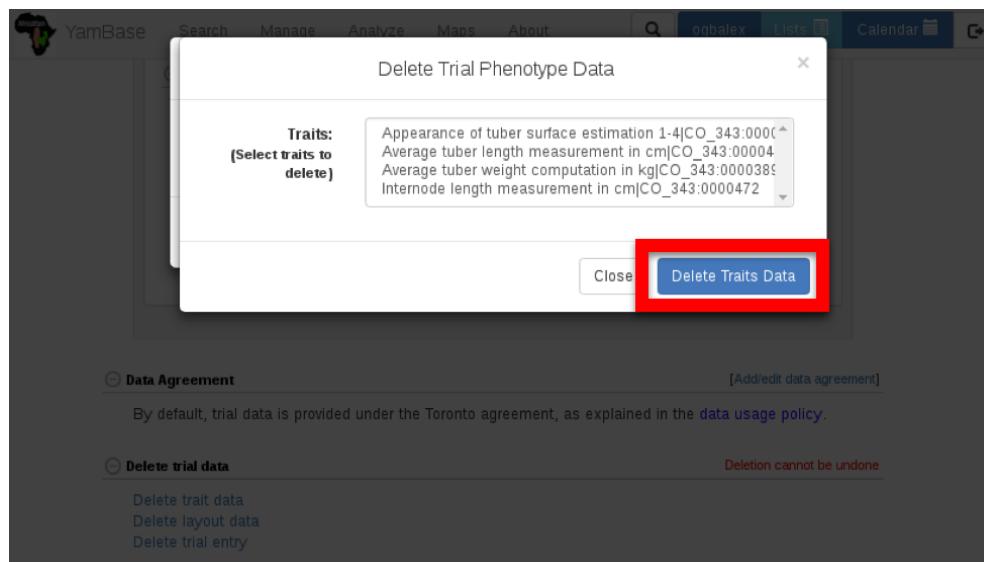
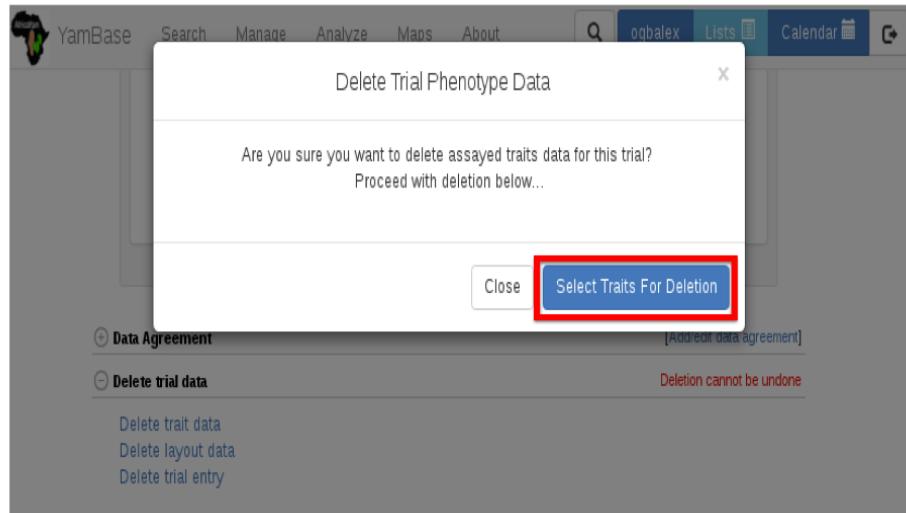
- plot_width: plot width in meters
- plot_length: plot length in meters
- field_size: field size in hectares

10.4 Deleting Trial Data

To delete a trial data, click on the “Delete trial data” section. There are links to delete traits, layout and trial entry data.

The screenshot shows the YamBase web application interface. At the top, there is a navigation bar with links for YamBase, Search, Manage, Analyze, Maps, About, a search icon, a user icon labeled 'ogbalex', Lists, Calendar, and a gear icon. Below the navigation bar, there is a table with a single row containing the text 'No data available in table'. Underneath the table, it says 'Showing 0 to 0 of 0 entries' and has 'Previous' and 'Next' buttons. At the bottom of the page, there is a section titled 'Delete trial data' which is highlighted with a red box. This section includes links for 'Delete trait data', 'Delete layout data', and 'Delete trial entry'. To the right of this section, there is a note in red text: 'Deletion cannot be undone'. Above the 'Delete trial data' section, there is a link '[Add/edit data agreement]'.

To delete assayed trait data, click on “Delete trait data” link. On the appeared dialog, confirm deletion by clicking on the “Select Traits For Deletion” button, then select one or more traits to delete from the trial.



To delete trial layout data, click on the “Delete layout data” link. Confirm deletion on the appeared dialog.

To Delete trial entry, click on “Delete trial entry” link. Confirm deletion on the appeared dialog.

Chapter 11

Managing Genotyping Plates

Genotyping Plates represent the content of a genotyping plate sent to a genotyping facility (e.g. samples in specific wells). To streamline this process, it is possible to upload this information or let the database create a plate for you. Once the genotyping plate is saved in the database it is then possible to export the information directly to genotyping facilities that are BrAPI compliant. The genotyping facility can then provide status information to us via BrAPI.

To begin go to Manage->Genotyping Plates.

Manage Genotyping Trials

[About Genotyping Trials](#)

What are genotyping trials?

- Genotyping trials represent 96 or 384 well plates.
- Each well in the plate has a unique source sample ID.
- The "contents" of each well can be either a tissue sample, plant name, plot name, or accession name. This "source" name must be in the database (e.g. as tissue samples or plants or plots from a trial, or just as accession names). Ideally you will have the barcodes from the field with you.
- Use the "Coordinator" Android Application to scan your "source" barcodes and record the position of the tissue sample in the 96 or 384 well plate. If you prefer you can create your own XLS file and upload that, if you do not want to use the Coordinator Application. Alternatively you can let the database generate the genotyping trial for you, and then produce the plate in that layout.
- For more information on the "Coordinator" Android Application go to [Coordinator](#).
- Click "Add Genotyping Trial" and fill in the form completely.
- To ease shipping materials to the genotyping facility, we can generate the required templates for you after the data is in the database.

Genotyping Trials

[Add Genotyping Trial]

Information	Breeding Programs – Folders – Genotyping Trials
Search <input type="text" value="Search"/> <p>Double click genotyping trial () or folder () to view detail page.</p> <p>Breeding programs ()</p> Folders Create new folder Move genotyping trial(s) to folder Move folder	Refresh <ul style="list-style-type: none"> test <ul style="list-style-type: none"> 18DIA0001 testing_folder_g test_coord2

Here the genotyping plates are divided by Breeding Program. These sections can be expanded by clicking on one.

▸ SCP	
▸ ARI Tanzania	
▸ BTI	
▸ CARI	
▸ CIAT	
▸ CNRA_PRT	
▸ CSIR	
▸ IITA	
▸ KU	
▼ NRCRRI	
NRCPOLYCR02 NRCPOLYCR010 NRCPOLYGS4 NRCPOLYGM5 NRCPOLYGS3 NRCPOLYGS7 NRCPOLYGS5 NRCPOLYCR01 NRCPOLYGM1 NRCPOLYGM4 NRCPOLYPP01 NRCPOLYGS6 NRCPOLYGM4 NRCPOLYCR03 NRCPOLYCR007 NRCPOLYCR009 NRCPOLYCR08 NRCPOLYCR05 NRCPOLYGS9 NRCPOLYGS8 NRCPOLYCR006 NRCPOLYGS2 NRCPOLYGM6 NRCPOLYCR04 NRCPOLYGS1	
▸ NaCRRI	
▸ Other	
▸ Rayong	
▸ SRI Kibaha	

11.1 Adding a New Genotyping Plate

To begin, click on “Add Genotyping Plate”. Notice that this form is split into three sections: “Plate Information”, “Well Information”, and “Confirm”. The first section is for defining information about the genotyping plate, such as a Plate identifier, plate format (96 well), etc. The second section is for defining the samples in the wells, such as sample names, sample concentrations, well position, etc. The final section is for Submitting the info.

All fields in the Plate Information section are required.

The screenshot shows a modal dialog box titled "Add Genotyping Trial". At the top, there are three tabs: "Plate Information" (selected), "Well Information", and "Confirm". The "Plate Information" tab contains the following fields:

- Genotyping Project Name:** Should match Vendor Project (text input: e.g. NextGenCassava)
- Genotyping Plate ID:** (text input: e.g. 18DNA00001)
- Plate Format:** (dropdown menu: 96 Well)
- Sample Type:** (dropdown menu: DNA)
- Breeding Program:** (dropdown menu: IITA)
- Location:** (dropdown menu)
- Year:** (dropdown menu: 2017)
- Description:** (text area)
- Genotyping Facility:** (dropdown menu: None)

At the bottom right of the dialog box is a "Close" button.

In the Well Information section you can choose to either 1) Upload an XLS or XLSX spreadsheet with your sample layout or 2) let the database create the sample layout.

Add Genotyping Trial

Plate Information Well Information Confirm

1. Do you already have a plate layout created?

 File format information
Spreadsheet format

Select Plate Layout XLS File: No file chosen

2. Or do you want us to generate a plate layout for you?

- Select a list for the source material going into each well. Your list should be a one to one pairing to each well e.g. if you want to fill 95 wells you should supply a list of 95 elements.
- Note: From the most desirable to least desirable list type you can choose: tissue samples, plants, plots, or accessions

Source Observation Unit List:

Blank Well: (Cornell IGD requires a specific well to be blank.)
e.g. A01

Well Concentration (ng/u): (If you used the same conc for all wells)

Well Volume (uL): (If you used the same vol for all wells)

Tissue: (If used the same tissue for all wells)

Extraction: (If used the same extraction for all wells)

Person: (If same person prepared all wells.)

If you choose to upload an XLS or XLSX spreadsheet, the Spreadsheet Template info requires the following:

Upload Template Information

This is for uploading a pre-existing genotyping plate layout.
File must be Excel file (.xls)
(.xlsx format not supported)

Header:
The first row (header) must contain the following:

date	sample_id	well_A01	row	column	source_observation_unit_name	dna_person	notes	tissue_type	extraction	concentration	volume	is_blank
------	-----------	----------	-----	--------	------------------------------	------------	-------	-------------	------------	---------------	--------	----------

Required fields:

- date (should be YYYY/MM/DD)
- sample_id (the unique identifier for the sample in the well)
- well_A01 (the position of the sample in the plate)
- row (the row position of the sample in the plate e.g. A)
- column (the column position of the sample in the plate e.g. 10)
- source_observation_unit_name (must exist in the database, the identifier of the origin material, in order of most desirable identifier to least desirable identifier that can be used here: tissue sample name, plant name, plot name, accession name)

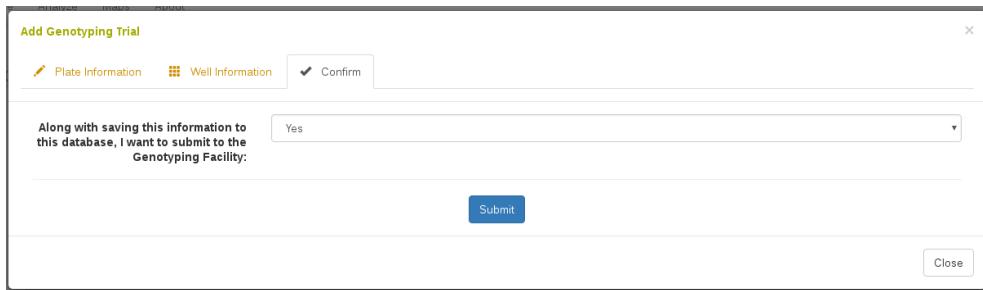
Optional fields:

- dna_person (the name of the person who prepared the well)
- notes (any additional notes on the well)
- tissue_type (free-text for what type of tissue is present in the well)
- extraction (free-text for the extraction method e.g. CTAB)
- concentration (concentration in ng/u)
- volume (volume in uL)
- is_blank (indicates if well is blank, write 1 if blank, otherwise leave empty.)

In either case, the sample identifier is generally a concatenation of Plate name and well position, e.g. MyGenotypingTrial1_A01. In either case, you need to provide a “source_observation_unit_name” for each sample. This

can be a tissue sample name, a plant name, a plot name, or an accession name; however, in any case, the identifier must already exist in the database. This allows us to link the sample in the well to specific field trial plots, or, plants, or tissue_samples. If you only know which accession is in the well, you can use the accession name.

In the final Confirm section you can decide whether to submit this information to the genotyping facility you selected. This requires that the genotyping facility is BrAPI compliant to work.



11.2 Genotyping Plate Detail Page

If you open a specific genotyping plate, it will take you to the detail page. Here you can see the Accessions used in the plate (if you created the trial and the source_observation_unit_names you used were plots, this will still work because we know the accession of the plot or plant or tissue sample).

Genotyping trial genou31

Breeding Program	IITA (IITA cassava breeding program, Ibadan, Nigeria)
Trial Type	Genotyping Trial
Plate Format	96
Plate Sample Type	DNA
Genotyping Facility	lgd
Submitted to Genotyping Facility	yes
Genotyping Facility Status	

Live Status From Genotyping Facility	
Download PDF	



SGN trial 3391 (genou31)

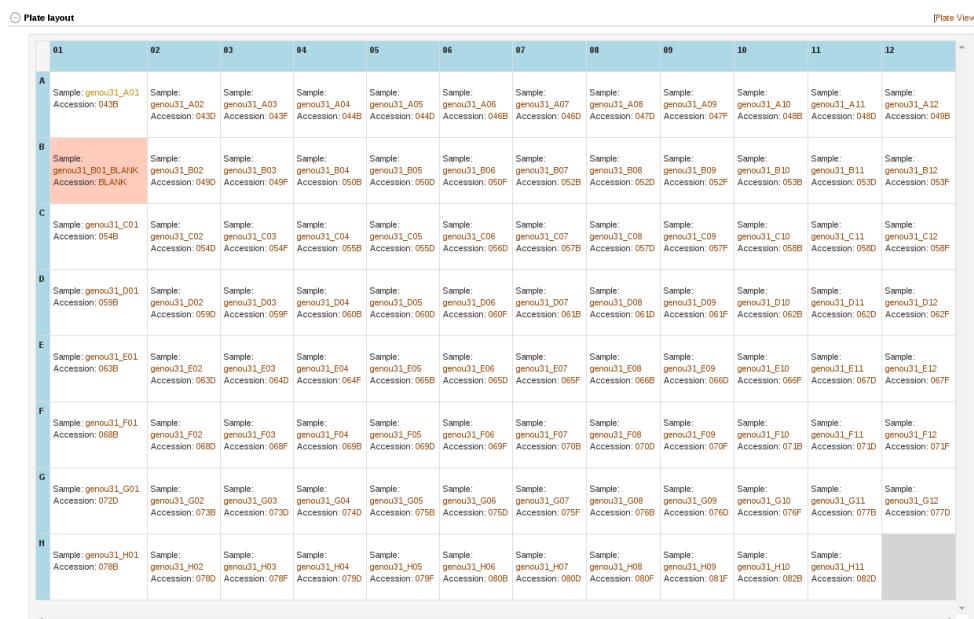
Design [Download layout \[xls\] \[cs\]](#)

Accessions

Tissue Sources

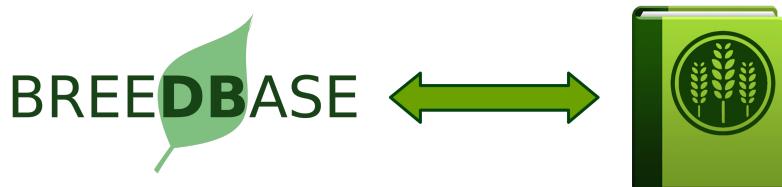
Tissue Samples

Further down you can see a graphical representation of your plate with well positions. This can be 96 well or 384 well depending on your plate format.



Chapter 12

Using Field Book and BreedBase for Data Collection and Storage



12.1 Introduction

Field Book is an open-source Android application designed to streamline phenotypic data collection, replacing traditional paper field books and reducing transcription errors.

BreedBase supports integration with Field Book by enabling users to generate properly formatted field layout and trait files, import collected data, and optionally exchange data via **BrAPI**.

This section provides step-by-step guidance on preparing Field Book for data collection using trial data stored in BreedBase, including file generation, manual file import, and data export. It assumes that trial fields and trait defi-

nitions already exist in BreedBase and that users have access to an Android device.

[Field Book User Manual](#)

12.2 Installing Field Book

Field Book can be installed on an Android device via the Google Play Store or manually from GitHub. To ensure access to the latest features and fixes, we recommend checking for updates regularly.

12.2.1 Option 1: Downloading from the Google Play Store

Download the Field Book app directly from the [Google Play Store](#) or scan the QR code below.



[Walkthrough: Installation and Setup](#)

12.2.2 Option 2: Manual Installation from Github

[Walkthrough: Installing Field Book from GitHub](#)

12.2.3 Keeping Field Book Updated

Regular updates are recommended to access new features and bug fixes.

[Walkthrough: Updating the Field Book App.](#)

12.3 Preparing Field Layout Files in Breed-Base

Field Book requires a **field layout file** containing a unique identifier for each plot or plant to be observed. Optionally, columns for row, column, pedigree, or previous phenotype data may be included for context.

BreedBase provides tools to generate layout files directly from trial metadata. These files can be exported and then uploaded to Field Book for data collection.

[Walkthrough: Creating a Field Layout File in BreedBase](#)

12.4 Importing Field Layout Files into Field Book

Field layout files can be imported into Field Book via cloud storage or directly from the device.

12.4.1 Option 1: Import from Cloud Storage

Requirements:

1. A generated field layout file.
2. Linked cloud service (e.g., Google Drive) on the device.
3. File uploaded to the cloud account.

[Walkthrough: Importing from Cloud Storage](#)

12.4.2 Option 2: Import from Local Storage Device

Requirements:

1. A generated field layout file.
2. File transferred to the Android device's local storage.

[Walkthrough: Importing from Local Device](#)

12.5 Adding Traits to Field Book Using BreedBase

For exported data to be compatible with BreedBase, trait names in Field Book must match the exact names stored in the database.

BreedBase supports generation of **trait files** containing properly formatted trait names, types, units, and metadata.

12.5.1 Creating Trait Files in BreedBase

[Walkthrough: Creating Trait Files in BreedBase](#)

12.5.2 Importing Trait Files to Field Book

[Walkthrough: Importing Trait Files](#)

12.6 Collecting and Exporting Phenotype Data

This section describes the process for recording and exporting phenotype data using the Field Book application. It assumes that the required field layout and trait files have already been imported into the app.

Users will be guided through entering phenotypic observations in the field and exporting collected data in a format compatible with downstream processing and upload to **BreedBase**. This standardized workflow helps ensure data integrity, traceability, and seamless integration with breeding databases.

12.6.1 Collecting Phenotype Data in Field Book

Once field layout and trait files are loaded, data collection can begin. Field Book allows users to view each plot or plant entry, record trait values in

structured forms, and automatically timestamp and geotag observations (if device settings allow).

Walkthrough: Collecting Data in Field Book

12.6.2 Exporting Phenotype Data from Field Book and Importing to BreedBase

After data collection is complete, Field Book allows users to export data as a .csv file. This file can then be uploaded to BreedBase for storage and analysis.

Walkthrough: Exporting Phenotype Data from Field Book

Once exported, the .csv file can be imported into BreedBase through the trial's data upload interface. Proceed with uploading phenotype observations as described in the **Managing Phenotypic Data** section.

It is important that:

- Trait names match those defined in the database.
 - Plot or plant identifiers align with the original layout.
-

12.7 Using BrAPI for Data Transfer Between Field Book and BreedBase

Field Book supports integration with **BreedBase** via the **BrAPI (Breeding API)** specification. BrAPI enables direct exchange of trial metadata and phenotype observations between Field Book and BreedBase without the need for manual file uploads.

This walkthrough covers:

- Connecting Field Book to a BrAPI-enabled BreedBase server
- Importing trial layout and traits from BreedBase to Field Book
- Exporting collected phenotype data from Field Book to BreedBase

Prerequisites:

- You must have a BreedBase user account with permissions to access trial and trait data.
- Your BreedBase instance must be configured with working BrAPI endpoints.
- The Field Book app must be installed on an Android device with internet access.

Advantages of Using BrAPI:

- Reduces manual errors from file handling.
- Supports real-time syncing of data.
- Enables remote team coordination and efficient workflows.

12.7.1 Connecting Field Book to a BrAPI Server

[Walkthrough: Connecting Field Book to a BreedBase BrAPI Server](#)

12.7.2 Importing Trials and Traits from BreedBase

[Walkthrough: Importing Trials from BreedBase via BrAPI](#)

[Walkthrough: Importing Traits from BreedBase via BrAPI](#)

12.7.3 Collecting Data

Proceed with entering phenotype observations as described in the **Collecting Phenotype Data in Field Book** section.

Ensure that:

- Trait values match expected formats (e.g., numeric, categorical).
- Observations are saved regularly during field use.

12.7.4 Exporting Phenotype Data to BreedBase via BrAPI

[Walkthrough: Sending Phenotype Data with BrAPI](#)

12.7.5 Verifying Data in BreedBase

After exporting data to BreedBase via BrAPI, it is important to verify that the export was successful directly in BreedBase.

1. Log in to your BreedBase instance.
2. Navigate to the Manage Trials or Trial Detail page for the uploaded study.
3. Confirm that:
 - Trait data appears in the Phenotype Data tab.
 - Data points align with expected plot/plant identifiers.
 - Trait units and scales match those defined in the database.

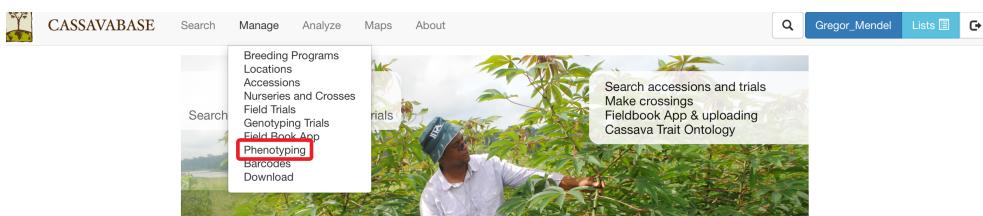
12.7.6 Troubleshooting

- Connection fails: Ensure you have the correct BrAPI server URL and valid credentials.
- Traits do not load: Confirm that the trait exists in BreedBase.
- Upload fails: Check for internet connectivity, correct user permissions, and trait formatting errors.

Chapter 13

Managing Phenotypic Data

To facilitate uploading process for phenotypic data, “Manage Phenotypic Data” page provides two options for uploading: Field Book Phenotype file in database format and phenotype file in Excel (.xls or .xlsx) file format. To access “Manage Phenotypic Data” page, clicking on “Phenotyping” in the “Manage” menu.



13.1 Uploading Fieldbook Phenotypes

13.1.1 Export Field Book Database File

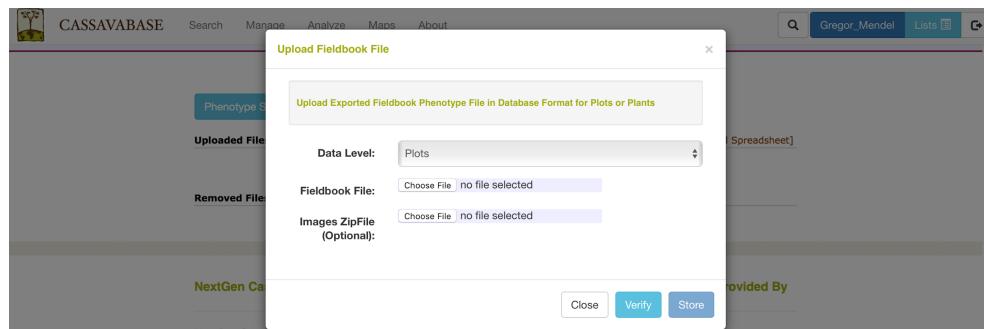
The database upload of Field Book phenotype data relies on the “Database” format from the Field Book. Please make sure to export the “Database” format from the Field Book if you intend to upload the data using the Field Book Upload we describe below. If you prefer to use the “Table” format that the Field Book exports, you can modify this format to work with the Speadsheet Upload we describe below.

13.1.2 Upload Field Book Database File

To upload a Field Book Phenotype file in a database format, click the “Upload Fieldbook” link

The screenshot shows the CASSAVABASE interface with the title "CASSAVABASE" at the top. Below it are navigation links: Search, Manage, Analyze, Maps, and About. A search bar and user profile "Gregor_Mendel" are also present. The main area is titled "Manage Phenotypic Data". It has sections for "Phenotype Search", "Uploaded Files" (with a red box around the "[Upload Fieldbook]" button), and "Removed Files".

The “Upload Fieldbook” link on this page and “Upload” link on the “Field Book Tools” page open the same dialogue. Please follow instructions for uploading phenotypic files on the ?? page.

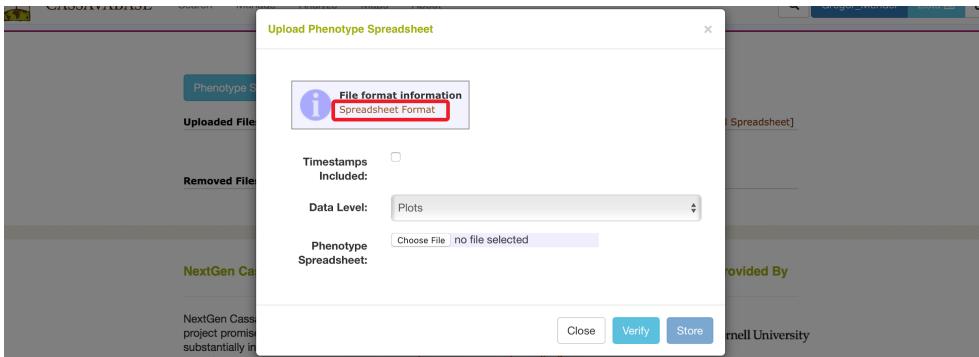


13.2 Uploading Spreadsheet Phenotypes

To upload a phenotype file in an Excel (.xls or .xlsx) file format, click the “Upload Spreadsheet” link.

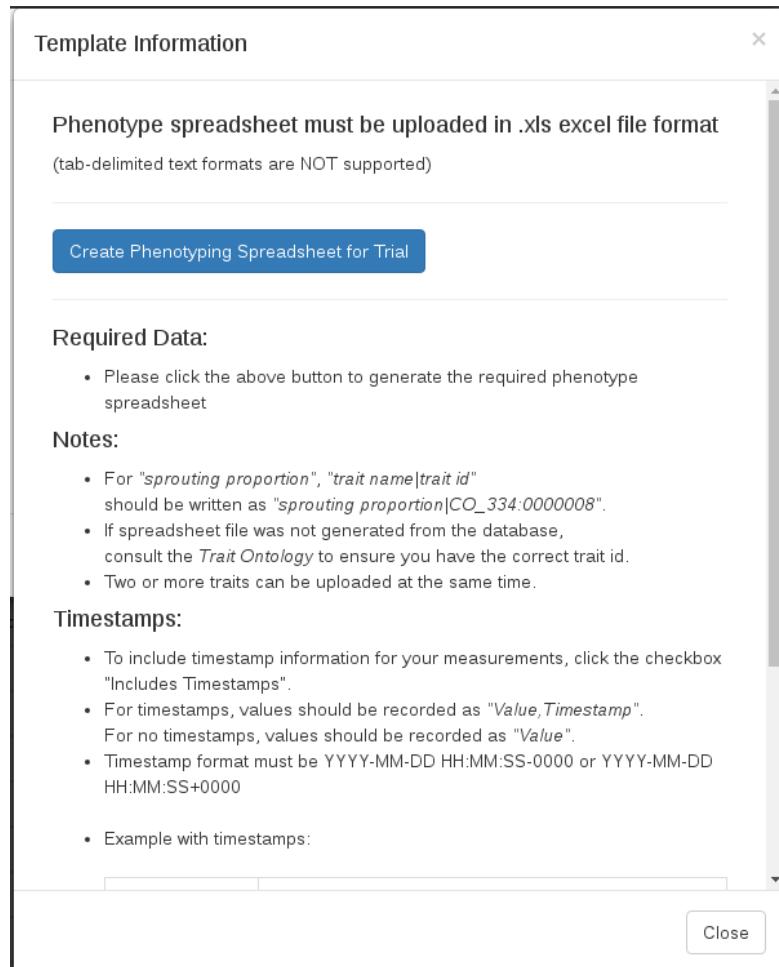
The screenshot shows the CASSAVABASE interface with the title "CASSAVABASE" at the top. Below it are navigation links: Search, Manage, Analyze, Maps, and About. A search bar and user profile "Gregor_Mendel" are also present. The main area is titled "Manage Phenotypic Data". It has sections for "Phenotype Search", "Uploaded Files" (with a red box around the "[Upload Spreadsheet]" button), and "Removed Files".

Please specify “Data Level” (Plots or Plants) and select the Excel file that you want to upload.

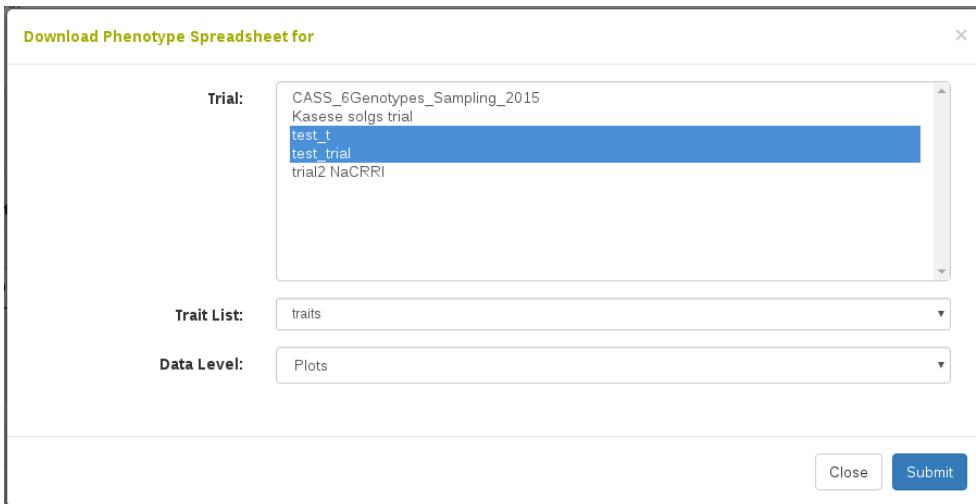


13.2.1 Generating Spreadsheet File

You can find more file format information by clicking on “Spreadsheet Format” link. Clicking on “Spreadsheet Format” will open the following dialog.



Clicking on “Create Phenotyping Spreadsheet” will bring up a dialog where you can indicate the trial(s) you are interested in and the trait list you are interested in. Clicking “Submit” will download the xlsx file onto your computer, where you can then fill in the phenotypes.



13.2.2 Uploading Spreadsheet File

To ensure that the file has a correct format for uploading, click on the “Verify” button. This will check the contents of the file and also perform quality checks on the values in the file. These checks include checking the trait definition for categorical values, minimum and maximum values, and data type checking. It will also check if there are already values uploaded for the given observation units and traits. If there are, there is an option to overwrite the existing values with the new values in your file. If the file is valid, only then can you click “Store” to store the information in the database.

Upload Phenotype Spreadsheet

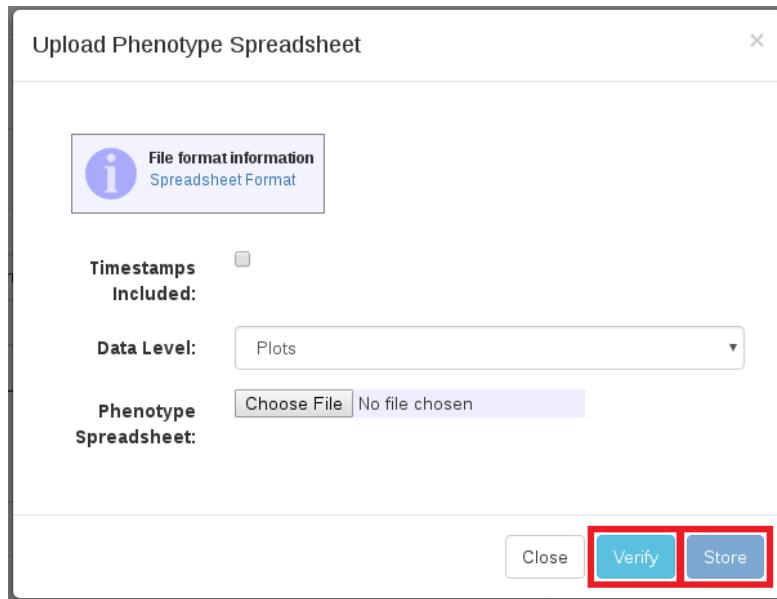
i File format information
Spreadsheet Format

Timestamps Included:

Data Level: Plots

Phenotype Spreadsheet: Choose File No file chosen

Close **Verify** **Store**



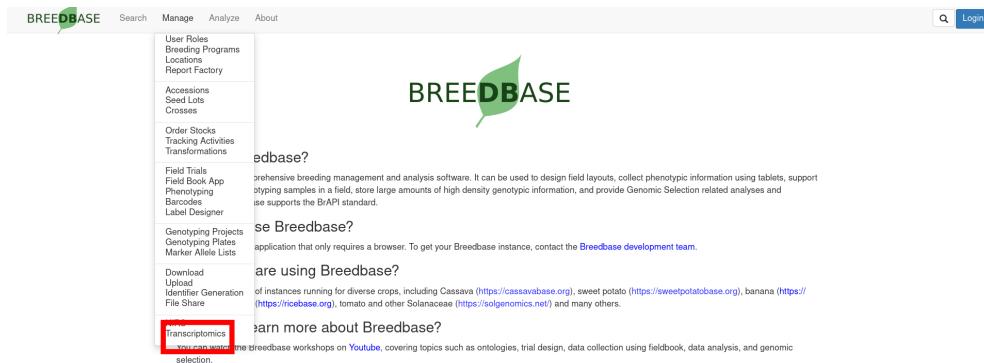
Chapter 14

Managing High Dimensional Phenotyping Data

14.1 Managing Transcriptomic Data

14.1.1 Uploading Transcriptomic Data

To upload transcriptomic data, go to the transcriptomics page by clicking “Transcriptomics” under the Manage tab.



Click the “Upload Transcriptomics Data” button to open the upload workflow dialog. On the second step “Samples”, you will be prompted to make sure that your samples are already in the database. You can refer to the “Managing Tissue Samples” chapter for instructions on how to create a sam-

pling trial. After creating a sampling trial or confirming that your samples exist, move on to the “Protocol Info” step. Here you can select an existing protocol, or create a new protocol. To create a protocol, click the “Protocol Not Shown. Create a New Protocol” button. From here you can fill in information about your new protocol and when finished, click “Go to Next Step”.

Upload High-Dimensional Phenotype Data

X

Protocol Not Shown. Create a New Protocol

Protocol Name:	<input type="text"/>
Protocol Description:	<input type="text"/>
Transcriptomics Unit:	<input type="text"/> RPKM
Genome Version:	<input type="text"/> e.g. v6.1, v7.0, v8.0
Genome Annotation Version:	<input type="text"/> e.g. v6.0, v6.1, v7.0
Instrument Model:	<input type="text"/>
Layout:	<input type="text"/>
Library Method:	<input type="text"/>
Library Comments:	<input type="text"/> e.g. Used the 3'RNA-seq method
Mapping Software:	<input type="text"/>
Sequencing Center:	<input type="text"/>
Sequencing Platform:	<input type="text"/>
Read Length:	<input type="text"/> e.g. 5x depth with 150 bp reads
Nucleic Acid Extraction	<input type="text"/>

Close

You will need to have two csv files when uploading transcriptomics data. The first is the data matrix file, which should have these headers:

Template Information ×

Upload Transcriptomics in a tabular spreadsheet (.csv) format with the following columns:

- **sample_name:** This should be a sample name that exists in the database. The sample can come from a sampling trial or a field trial or a genotyping plate.
- **device_id:** Optional identifier to differentiate device (e.g. serial number). Can be numeric or letter.
- **comments:** Optional comments for any information to include. Can include specific sample preparation for measurement.
- All other columns are **transcript names**. You can add as many columns as you want to upload, there isn't a limit.

sample_name	device_id	comments	transcript name columns
-------------	-----------	----------	-------------------------

The names in the sample_name column should match the sample names you created in your sampling trial file. The header marked “transcript_name_columns” should be replaced with the transcript names you want to upload, each in its own column in the header and the respective expression value below. Example:

sample_name	device_id	comments	ZteamMp004	ZteamMp005	ZteamMp024	ZteamMp032	10.177049366007
transcriptomics_test1			0.847898286111116	0	0	0	

The second file is the transcript details file, which should have these headers:

Upload details about the transcripts in a tabular spreadsheet (.csv) format with the following columns:

- **transcript_name** (required): This should be the same name as present in a column in your transcriptomics data matrix file.
- **chromosome** (required): The chromosome where the transcript exists.
- **start_position** (required): The start position in base pairs of where the transcript begins.
- **end_position** (required): The end position in base pairs of where the transcript ends.
- **gene_description** (optional): Descriptors for gene annotations.
- **notes** (optional): Any additional descriptions.

transcript_name	chromosome	start_position	end_position	gene_description	notes

Each row under the transcript_name column should contain each transcript name from your data matrix file. For example:

transcript_name	chromosome	start_position	end_position	gene_descrip	notes
ZeamMp004	Mt	8752	9915	orf387	
ZeamMp005	Mt	11111	13315	orf734	
ZeamMp024	Mt	161140	164661	rps3	
ZeamMp032	Mt	76132	84976	nad4	

Once these two files are uploaded, click verify to check that they are formatted correctly and that the samples exist. If there are no errors, you can then click store to store your transcriptomics data.

14.1.2 Downloading Transcriptomics Data

To download your data, click the “Uploaded Transcriptomics Data” dropdown menu on the Transcriptomics page. Then click the “Your Uploaded Transcriptomic Data” dropdown. From here you will be able to see the transcriptomics data that you have uploaded. Click the “Download” button next to the data that you wish to download.

The screenshot shows the Transcriptomics interface. At the top, there are buttons for "Upload Transcriptomic Data" and "Download Transcriptomic Data". Below this, a section titled "Your Uploaded Transcriptomic Data" displays a table of uploaded files. The table has columns for "Filename", "Date Uploaded", and "Options". The "Options" column contains "View" and "Download" links. The "Download" link for the first file is highlighted with a red box. The table shows two entries:

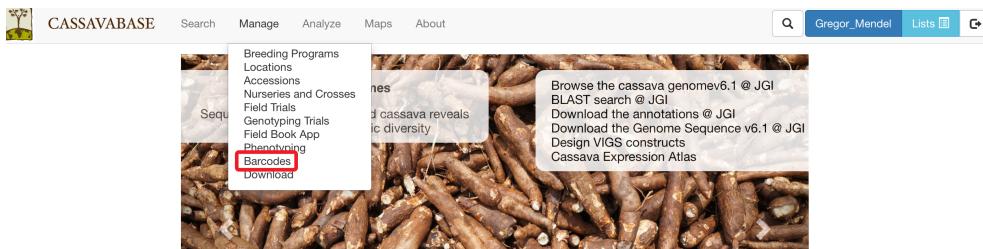
Filename	Date Uploaded	Options
2025-10-20_21:30:34_transcript_upload_data.csv	2025-10-20 21:30:34.720471+00	View Download
2025-10-21_19:27:26_transcript_upload_data.csv	2025-10-21 19:27:26.55596+00	View Download

Below the table, it says "Showing 1 to 2 of 2 entries". There are "Previous" and "Next" buttons. At the bottom, there are sections for "Your Obsoleted Transcriptomic Data" and "All Uploaded Transcriptomic Data".

Chapter 15

Managing Barcodes

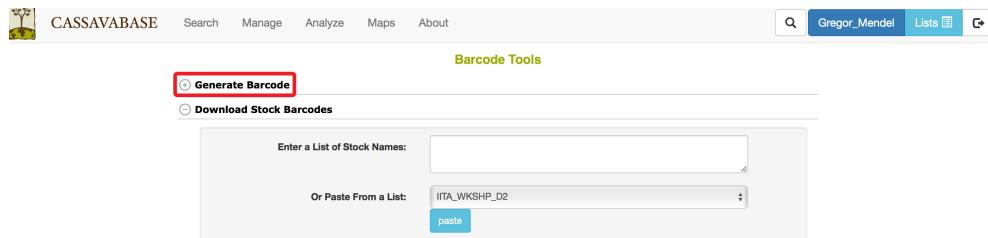
SGN databases provide tools for generating barcodes for stock identification. To access “Barcode Tools” page, clicking on “Barcodes” in the “Manage” menu.



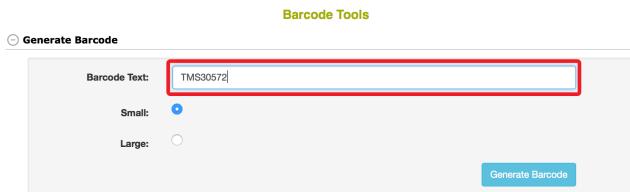
“Barcode Tools” page provides four options for generating barcodes:

- Single barcode
- Multiple barcodes
- Plot phenotyping barcodes
- Trial barcodes

To generate single barcode, clicking on “Generate Barcode” link on the “Barcode Tools” page.



In the “Generate Barcode” section, specify the name of the barcode, size of the barcode, then clicking on “Generate Barcode”



The database will generate a barcode for your stock. The barcode can be printed for your stock identification. It also appears on its corresponding stock page.



If you have a list of stocks that you want to generate barcodes, you can use “Download Stock Barcodes” section. You have three options for entering stock names:

1. Typing in stock names, or copy and paste from other file into the box (1)
2. Choosing a list of stocks from your “Lists” (2), and transferring the list into the box (1) by clicking on “paste” button.
3. Uploading a “Tab-delimited Text File” with stock names.
4. Select an optional printing format from the available formats.

You can select printer settings that you prefer in the “Printer Settings” sec-

tion. After you enter stock names and specify printer settings, clicking on “Download Barcodes” button at the bottom of the page.

The screenshot shows the 'Barcode Tools' section of the CASSAVABASE website. At the top, there are navigation links: Search, Manage, Analyze, Maps, About, and a user profile section with 'Gregor_Mendel' and 'Lists'. Below the navigation is a search bar and a 'Barcode Tools' button.

Generate Barcode

Download Stock Barcodes

1 Enter a List of Stock Names:

2 Or Paste From a List:

3 Or Upload Tab-delimited Text File With Stock Names:
no file selected

Print Duplicate Labels Per Row:

Print Field Information For Plots: Useful for Printing Field Information of Trials.

Print Parents For Nurseries: Useful for Printing Pedigree Information for Nurseries.

Printer Settings

Number of Label Rows:	10	Number of Label Columns Per Page:	3
Page Format:	Letter	Add text to label, e.g. location:	<input type="text"/>
Top Margin (mm):	12	Left Margin (mm):	70
Bottom Margin (mm):	12	Right Margin (mm):	20

If you have a list of plots that you want to generate phenotyping barcodes, you can use “Download Plot Phenotyping Barcodes” section. You have three options for entering plot names:

1. Typing in plot names, or copy and paste from other file into the box (1)
2. Choosing a list of plots from your “Lists” (2), and transferring the list into the box (1) by clicking on “paste” button.
3. Uploading a “Tab-delimited Text File” with plot names.

[Download Plot Phenotyping Barcodes](#)

Enter a List of Stock Names:

Or Paste From a List:

Or Upload Tab-delimited Text File With Stock Names: Choose File No file chosen

Add Text to Label, e.g. location:

If you have a list of trials that you want to generate barcodes, you can use “Download Trial Barcodes” section. You have three options for entering trial names:

1. Typing in trial names, or copy and paste from other file into the box (1)
2. Choosing a list of trial from your “Lists” (2), and transferring the list into the box (1) by clicking on “paste” button.
3. Uploading a “Tab-delimited Text File” with trial names.

[Download Trial Barcodes](#)

Enter a List of Trial Names:

Or Paste From a List:

Or Upload Tab-delimited Text File With Trial Names: Choose File No file chosen

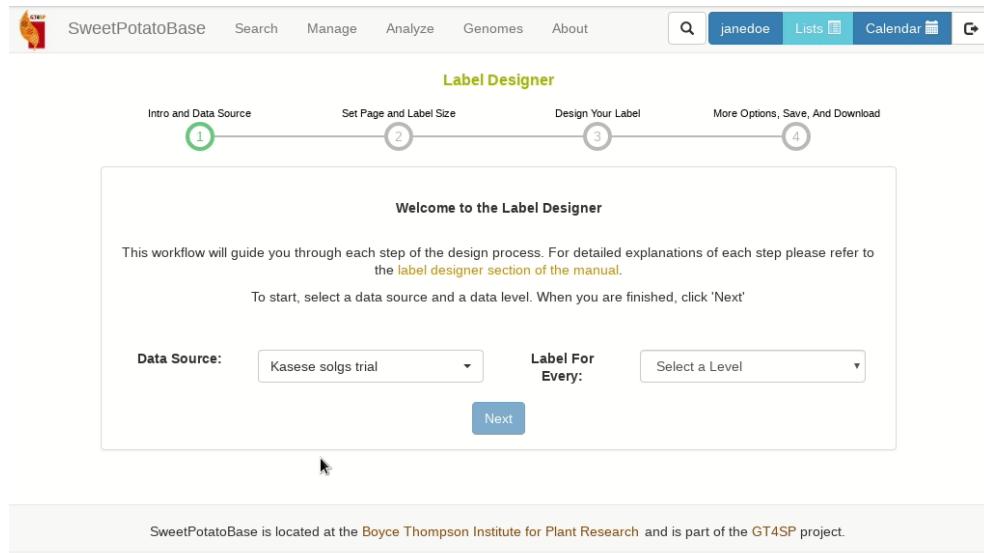
Chapter 16

Using the Label Designer

Breedbase provides an interactive design tool for creating custom labels. To access the label design tool, click on “Label Designer” in the “Manage” menu. The following sections explain your many options as you advance through each step of the design workflow.

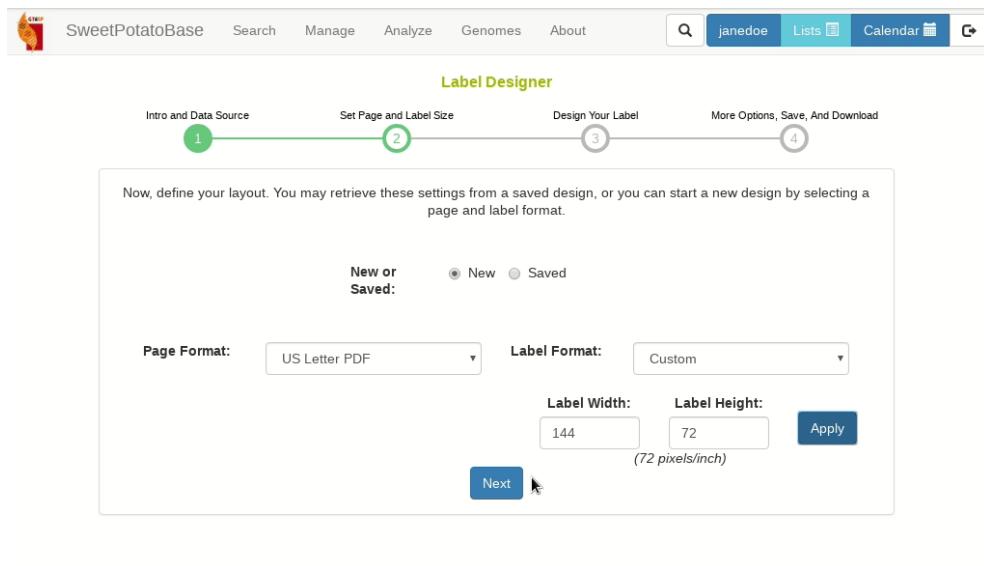
16.1 Select a Data Source

The first step is to select a data source. To do so, you may directly select a specific data source from the “Data Source” dropdown menu (either a field trial, genotyping trial, or crossing trial) to populate your labels with the trial design information. Alternately, from the same dropdown menu, you may select a list to populate your labels with the list contents. Prior to making a selection from the “Data Source” dropdown menu, you may also filter the data sources by using the “Datatype” dropdown menu. You must then choose a level (plot, plant, etc.) before proceeding, using the “Label for Every” dropdown menu. To generate plot-level labels for more than one trial at once, select a list of trials as the source and plot as the level. Once you have made your selections, click the ‘Next’ button to move to the next step in the workflow.



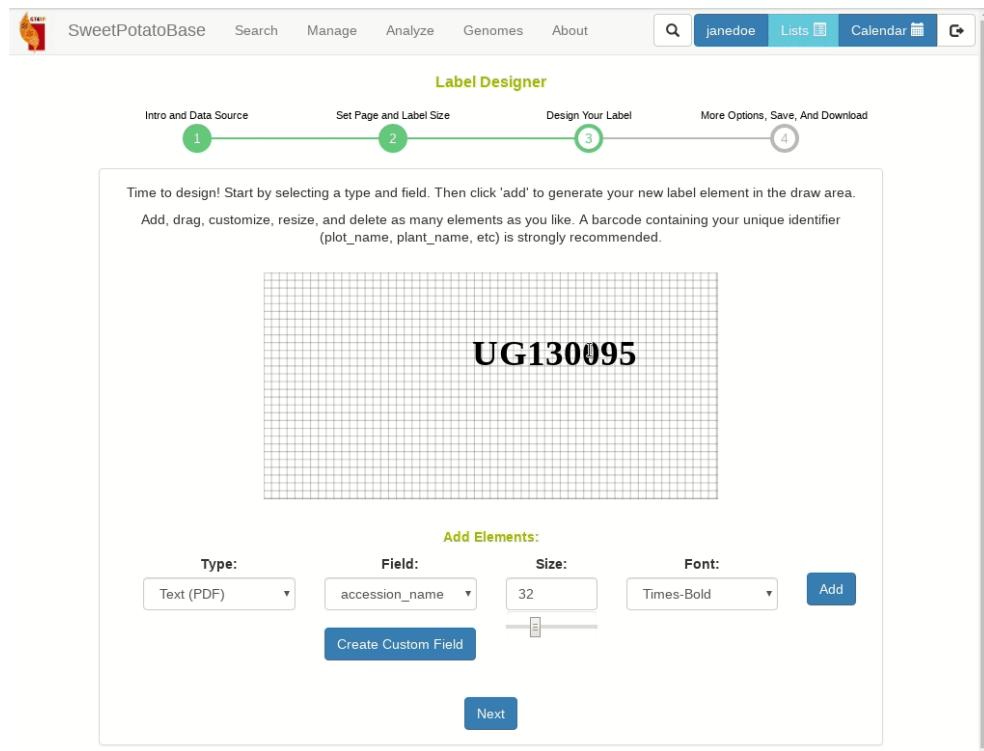
16.2 Set Page and Label Size

After selecting the data source, you must choose whether to create a new design or load a saved design. If you choose the “New” selector, you will be prompted to select a page size and label size. If you do not see your page size or label size as an option, then select “Custom” from the relevant dropdown menu and enter your desired dimensions in pixels (each pixel is 1/72nd of an inch). If you choose the “Saved” selector, you will be prompted to select a saved design. After selecting a saved design, you will be taken directly to the design step with the saved design elements pre-loaded.



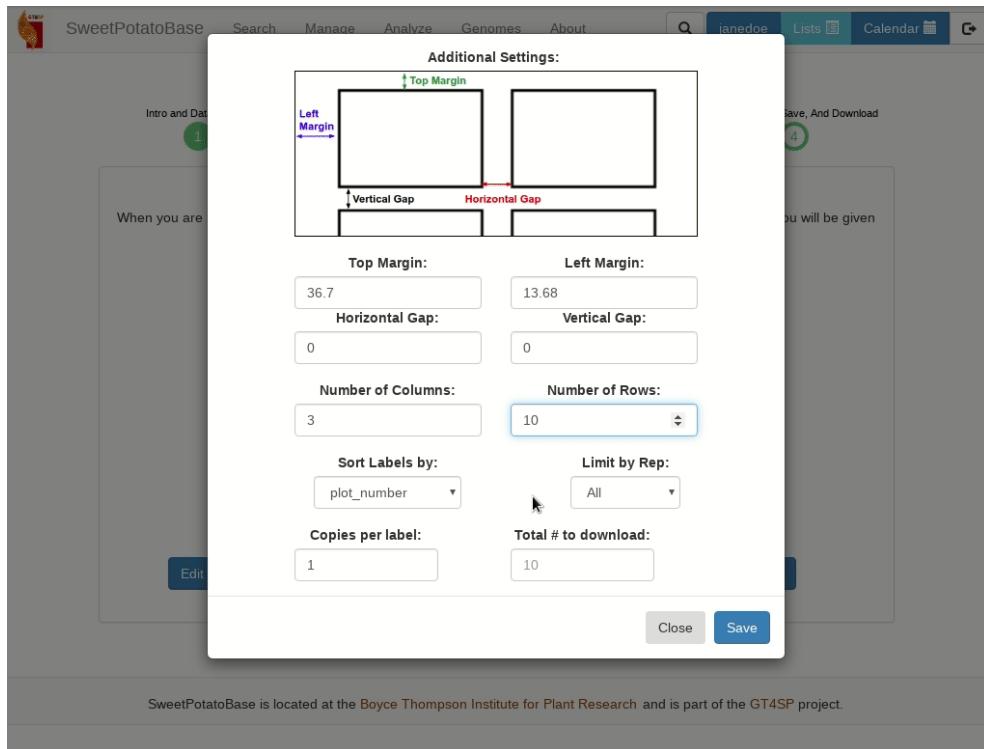
16.3 Design Your Label

Having set the page and label formats, you will be presented with a label canvas where you can begin adding elements to your label. Select a type, field, size, and font, then click “Add”. You can add text to an existing field or create a completely custom field by clicking “Create Custom Field”. Once added, you can drag and drop elements, or you can delete them by clicking on the red box in their upper left corners. Barcodes can also be resized by dragging the green box in their lower right corners. If you are creating labels for a trial, it is highly recommended that you include a barcode encoding your plot, plant, or tissue sample names. These are the unique identifiers that will need to be included with any phenotypic or genotypic measurements loaded into the database. When you are satisfied with your label design, click “Next”.



16.4 Adjust Formatting, Save, and Download

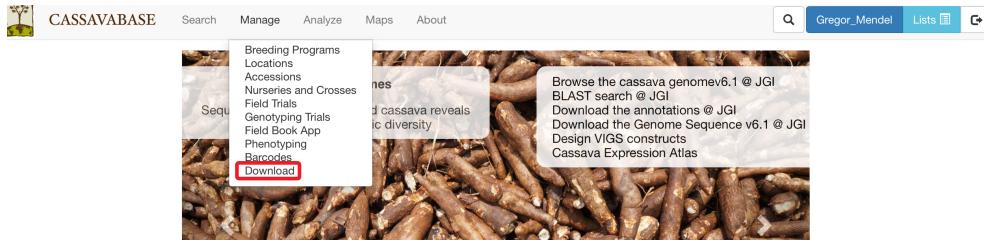
In the last step of the workflow, you can tweak your formatting and page layout, save your design, or download your labels. The “Additional Settings” dialog will allow you to adjust the page margins and margins between labels. The units are pixels (each pixel is 1/72nd of an inch). It is not recommended that you change these settings until you have already printed a test page. You can also set the number of copies per label, filter by replicate, or download only the first page for test purposes. To save your design, type a unique name in the text field and click “Save”. This will save your design to your list manager, where you can make it public in order to share it with others. Finally, if you are ready, click download to generate and download your labels!



Chapter 17

Managing Downloads

You can download phenotype, trial meta-data, pedigree, GBS genotype and GBS genotype QC files from the database to your computer by using “Lists”. To download, clicking on “Download” in the “Manage” menu.



For each category, you can select a list of accessions from your “Lists” to download their phenotypes, pedigree, GBS genotype, GBS genotype QC. In the case of downloading trial meta-data, you would provide a list of trials, while for downloading phenotype and GBS genotype QC, you can also use a list of trials or traits.

A screenshot of a 'Download Metadata' form. The form has a title 'Download Metadata' and a subtitle 'Select Parameters:'. It contains a table with three columns: 'Trials', 'Options', and 'Action'. The 'Trials' column has a dropdown menu with 'select' as the current value. The 'Options' column has a 'Format:' dropdown menu with 'XLS' as the current value. The 'Action' column contains a blue 'Download' button. The entire form is contained within a light gray box.

CASSAVABASE Search Manage Analyze Maps About Gregor_Mendel Lists  

Download Using Lists

Choose a list for each parameter and click "Download".

Download Phenotype

Select parameter:

Accessions	Trials	Traits	Format	Timestamps	Data Level	Action
select	select	select	<input type="checkbox"/> .xls (default)	No	All	Download
			<input type="checkbox"/> .csv			
			<input type="checkbox"/> html			

Download Pedigree

Select parameter:

Accessions	Action
select	Download

Download GBS Genotype

Select parameter:

Accessions	Genotyping Protocol	Action
select	<input checked="" type="checkbox"/> GBS ApeKI Cassava genome v5 GBS ApeKI Cassava genome v6 protocol GBS ApeKI Cassava genome v6_Oct2015	Download

GBS Genotype QC

Select parameter:

Trials	Accessions	Action
select	select	GBS ApeKI Cassava genome v5 Quality Control

Chapter 18

Managing ODK Data Collection

To access this page go to Manage and then ODK Data Collection. ODK is used for remotely collecting data on Android and IOS devices. We currently are working to support two ODK service providers, namely ONA and SMAP. We are using ONA to collect crossing information, including all lab activities following seed production. We are using SMAP for phenotypic data collection.

18.1 ONA Crossing Information

18.1.1 Managing ONA Crossing Information

Manage ODK Data Collection

What is ODK?

- ODK is an application that allows mobile data collection using user defined forms on Android or iOS devices. Data collected on the device can be instantaneously synched to the ODK server. To find out more go to the [ODK site](#). Many services have developed web interfaces to better streamline the ODK experience. These services assist in creating forms, deploying forms to your mobile application, and visualizing data uploaded back from the mobile device. Currently we are working with [SMAP](#) and [ONA](#) as two ODK services.

What do I do from this page?

- ONA is currently being used for collecting crossing information. This requires exporting a crossing plan from here to the OHA server. The crossing plan guides collection of cross information and this data is synched with ONA using ODK. From here, we run a script twice a day, which pulls data on OHA into our database.
- SMAP is currently being used for collecting phenotype information. The user collects phenotypes using a form they previously created. The questions in the form map directly to terms in the ontology. As they collect data on the mobile device, the data is synched to SMAP. From here, we run a script twice a day, which pulls data on SMAP into our database.

Crossing Data: ONA ODK Application

Select A Cross Wishlist: cross_wishlist_Abuja

Select An ODK Form on ONA: BTracT_NM2018_01_BTracT - Nelson Mandela

Management

Export Cross Wishlist (Crossing Plan) to Selected Form on ONA

Import Crossing Data from Selected Form on ONA

Schedule Import For Selected Form: Once per day at midnight

Scheduled Time: Once per day at midnight

Confirm

To begin collecting data using the ONA ODK form you must first have a crossing plan in the form of a Cross Wishlist. To do this from this page, click the “Export Cross Wishlist to ONA” button. Please refer to the “Create Cross Wishlist” help section for more information. It is possible to view the current available cross wishlists by clicking the “Export Cross Wishlist to ONA” button and then clicking “Available Cross Wishlists”.

Once your cross wishlist is available, you can use your mobile ODK application to record crosses being done realtime. You can also record all laboratory activities following seed extraction up to greenhouse plantlet hardening.

As you collect data using your mobile ODK application, your responses will be synchronized with our database. The “Schedule Import for Selected Form” section gives you options to perform the import daily or more frequently. It is also possible to initiate a data import from ONA at anytime by clicking “Import Crossing Data from Selected Form on ONA”.

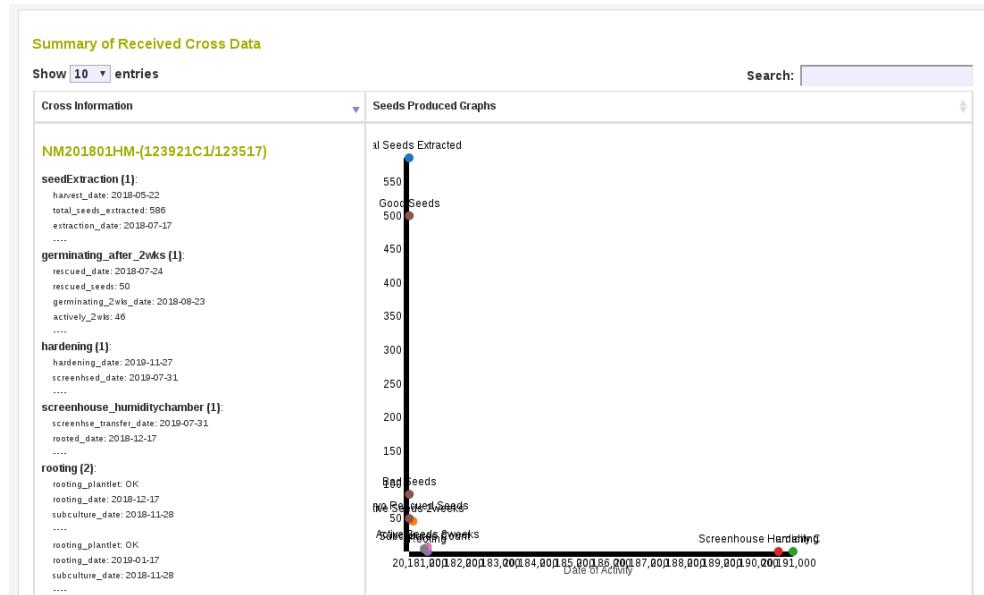
18.1.2 Reviewing Plant Status

The mobile ODK application has options to collect information about the status of plants in the field, such as if they are flowering. Images for each plant can also be recorded. The database will report this information here in a summary table that looks like the following. Notice that images are also transferred to the database.

Summary of Received Plant Status			
Show 10 entries			Search:
PlotName	Date	Status	Search:
	null	Status: Accession Name: null Trial Name: null User: null Status Location: Status: fallen Note: null Image: undefined	
16-Huti-white_r8c12_plot157	2018-02-12	Status: Accession Name: Huti-white Trial Name: 2016 mchare pollination block User: HM Status Location: in_field Status: destroyed Note: destroyed by elephants 	
16-ITC1460-Ijihulnkundu_r17c8_plot344	2018-01-18	Flowering: Accession Name: ITC1460-Ijihu Inkundu Plant Sex: female	
16-ITC0712-AAcvRose_r1c1_plot1	2018-01-17	Flowering: Accession Name: ITC0712-AAcv Rose Plant Sex: male	
16-ITC1468-Kahuti_r1c2_plot2	2018-01-17	Flowering: Accession Name: ITC1468-Kahuti Plant Sex: female	

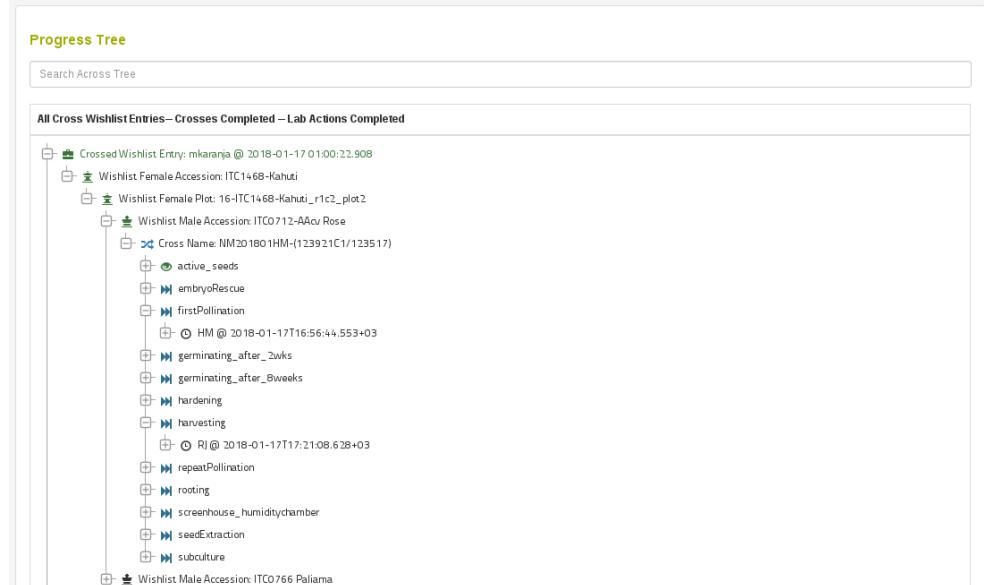
18.1.3 Graphical Summary For Performed Crosses

There is a section to summarize activities done for each cross. In this table each row represents a single cross performed. All the activities that have been performed will be shown here, such as “first pollination” and “embryo rescue”. The scatter plot shown tracks seed numbers generated on the Y axis and date of activity on the X axis.



18.1.4 Summary Information For Performed Crosses

There is a secondary section to summarize what has been done across the entire Cross Wishlist. This tree structure shows all activities performed for a cross and shows how these crosses relate to the Cross Wishlist.



Chapter 19

Managing Tissue Samples

To access this page go to Manage and then Tissue Samples.

19.1 Tissue samples from field trials

A field trial contains plots planted with a specific accession. Each plot can contain many plants, which in turn can contain many tissue samples. On the manage tissue sample page we can see the field trials that contain tissue samples already. We can choose to download the tissue sample layout as seen in the below picture.

Manage Tissue Samples

Field Trial Tissue Samples

Create tissue samples for field trial

View and create tissue samples for field trials. Tissue samples come from a plant in a plot.

Field Trials With Tissue Samples

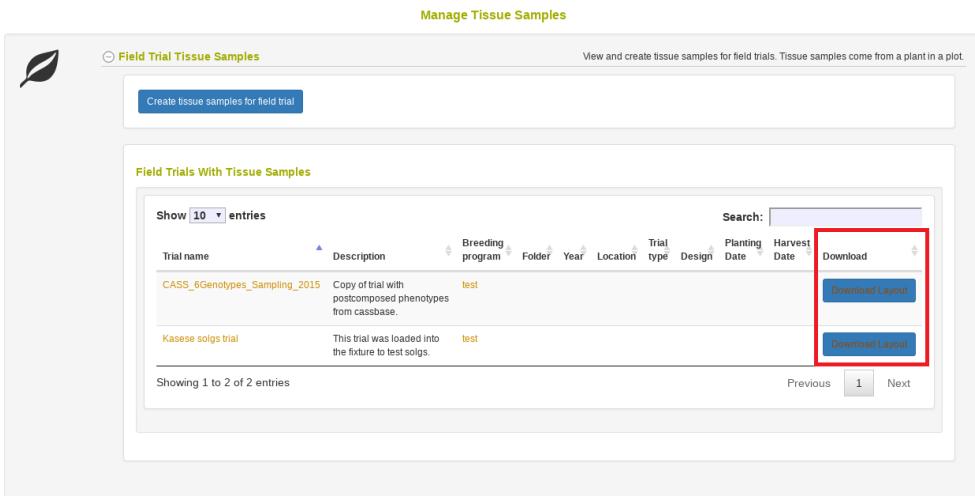
Show 10 entries

Trial name	Description	Breeding program	Folder	Year	Location	Trial type	Design	Planting Date	Harvest Date	Download
CASS_6Genotypes_Sampling_2015	Copy of trial with postcomposed phenotypes from cassbase.	test								Download Layout
Kasese solgs trial	This trial was loaded into the fixture to test solgs.	test								Download Layout

Search:

Showing 1 to 2 of 2 entries

Previous 1 Next



If the field trial you want to collect tissue samples from is not in the above table, you can click the button highlighted below.

The screenshot shows a web-based application titled "Manage Tissue Samples". At the top left is a leaf icon. Below it is a button labeled "Create tissue samples for field trial". The main area is titled "Field Trials With Tissue Samples". It includes a search bar and a table with the following columns: Trial name, Description, Breeding program, Folder, Year, Location, Trial type, Design, Planting Date, Harvest Date, and Download. Two entries are listed:

Trial name	Description	Breeding program	Folder	Year	Location	Trial type	Design	Planting Date	Harvest Date	Download
CASS_6Genotypes_Sampling_2015	Copy of trial with postcomposed phenotypes from cassbase.	test								<button>Download Layout</button>
Kasese solgs trial	This trial was loaded into the fixture to test solgs.	test								<button>Download Layout</button>

At the bottom of the table, it says "Showing 1 to 2 of 2 entries". There are "Previous" and "Next" buttons. A "Search:" input field is also present at the top right.

Once you have clicked this button, you will enter a workflow that begins with the following introduction.

The screenshot shows a modal window titled "Create Tissue Samples for a Field Trial". It has four numbered steps: "Intro" (1), "Select a field trial" (2), "Plant Entries" (3), and "Create Tissue Sample Entries" (4). Step 1 is highlighted with a green circle. The main content area contains the following text:

This workflow will guide you through creating tissue samples for your field trial

Tissue samples are linked to a single plant, which is in turn linked to a single plot.
Many tissue samples can be created for each plant.
Each tissue sample needs a globally unique name.
Tissue samples can then be transferred into genotyping trials (96 or 384 well plates).

A "Go to Next Step" button is located at the bottom of the content area, and a "Close" button is in the bottom right corner.

Once you click next, you will need to select your trial.

Create Tissue Samples for a Field Trial

Intro 1 Select a field trial 2 Plant Entries 3 Create Tissue Sample Entries 4

Select a field trial

Select	Trial name	Description	Breeding program	Folder	Year	Location	Trial type	Design	Planting Date	Harvest Date	Download
<input checked="" type="checkbox"/>	CASS_6Genotypes_Sampling_2015	Copy of trial with postcomposed phenotypes from cassbase.	test		2017	test_location	Preliminary Yield Trial	RCBD			<button>Download Plot List</button>
<input type="checkbox"/>	Kasese soils trial	This trial was loaded into the fixture to test soils.	test		2014	test_location	Clonal Evaluation	Alpha			<button>Download Plot List</button>
<input type="checkbox"/>	PVA20	asd	test		2018	Cornell Biotech	Seedling Nursery	RCBD			<button>Download Plot List</button>
<input type="checkbox"/>	new_test_cross	new_test_cross	test								<button>Download Plot List</button>
<input type="checkbox"/>	selection_population	selection_population			2015						<button>Download Plot List</button>
<input type="checkbox"/>	test_genotyping_project	test_genotyping_project			2015						<button>Download Plot List</button>
<input type="checkbox"/>	test_population2	test_population2			2015						<button>Download Plot List</button>

Show 10 entries Search:

Close

Next, if your trial currently only has plot entries saved, you will be asked to enter how many plants are in each plot.

Create Tissue Samples for a Field Trial

Intro 1 Select a field trial 2 Plant Entries 3 Create Tissue Sample Entries 4

Plant entries in your field trial

Please create plant entries for this trial.

Number of plants per plot:

Inherits Management Factor(s) From Plots:

Submit

Close

Finally you will be asked how many tissue samples you want for each plant. You can specify a string to include in the tissue sample name, such as leaf or root.

Create Tissue Samples for a Field Trial

Intro 1 Select a field trial 2 Plant Entries 3 Create Tissue Sample Entries 4

Create tissue sample entries for this trial

① Number of tissue samples per plant: 3

② Tissue Name 1: leaf

③ Tissue Name 2: leaf

④ Tissue Name 3: stem

Inherits Management Factor(s) From Plots:

Submit

Close

Afterwards you should see the following success message, indicating that the tissue samples are saved.

Create Tissue Samples for a Field Trial

Intro 1 Select a field trial 2 Plant Entries 3 Create Tissue Sample Entries 4

Complete! Your field trial's tissue samples were saved.

② Tissue samples saved successfully

- You may want to go to the trial detail page for the trial now that it has plants.
- You can print barcodes for the new tissue samples.
- You can use these tissue samples as source material for a genotyping trial (96 or 384 well plate)

Close

19.2 Genotyping Plate Tissue Samples (96 or 384 well plates)

A genotyping plate represents a 96 or 384 well plate. You can use the Coordinate Android application to create your plate layout, or you can upload your own Excel plate layout, or you can use the database to generate a plate layout. Ideally, you will use tissue sample names originating from a field trial as the “source” for each well tissue sample, but you can also use plant names, plot names, or accession names.

From the manage tissue samples page, you can see the genotyping plates

19.2. GENOTYPING PLATE TISSUE SAMPLES (96 OR 384 WELL PLATES)191

saved in the database. You can also download the layouts as shown below.

The screenshot shows a web-based application for managing genotyping trials. At the top, there's a header with a grid icon, the title "Genotyping Trial Tissue Samples", and a sub-instruction "View and create tissue samples for genotyping trials. Genotyping trials represent tissue samples in a 96 or 384 well plate layout." Below the header are two buttons: "Create or upload a genotyping trial" and "Export to Genotyping Vendor". The main area is titled "Genotyping Trials" and contains a table with two entries:

Show 10 entries	Search:				
Trial name	Description	Breeding program	Folder	Year	Location
18DNA101	A 96 well DNA sequencing plate	test		2017	Cornell Biotech
18Ngeno1	asd	test		2017	Cornell Biotech

At the bottom of the table, it says "Showing 1 to 2 of 2 entries". To the right of the table, there are "Previous", "1", and "Next" buttons. On the far right of the table, there's a "Download" section containing two buttons: "Download Layout" and "Download Layout". The "Download Layout" button for the second entry is highlighted with a red box.

If you need to create a new genotyping plate, you can click the button shown below. This will guide you through a workflow for uploading or creating the new plate layout.

This screenshot is similar to the one above, showing the "Genotyping Trial Tissue Samples" interface. The "Create or upload a genotyping trial" button at the top left is highlighted with a red box. The rest of the interface, including the table of genotyping trials and the "Download Layout" buttons, is identical to the previous screenshot.

Genotyping vendors require you to send a plate layout during submission. You can download the plate layout as shown above, or you can go to a genotyping plate detail page to download the Intertek formatted file.

In the future you will be able to directly export your genotyping plate plate layout to vendors.

Chapter 20

Managing Observation Variables

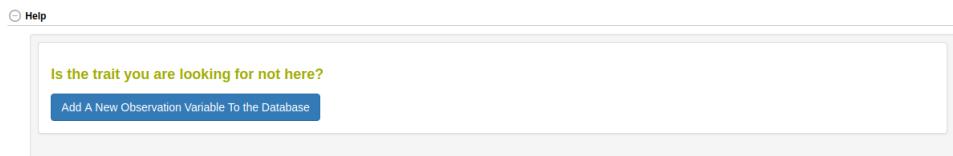
20.1 Managing Observation Variables with Traits, Methods, and Scales

Observation variables are the identifiers used when collecting phenotypic data. An observation variable is composed of a trait, a method, and a scale. The trait describes the attribute being measured e.g. ‘Plant Height’. The method defines the protocol in which the trait was observed e.g. ‘Using a one meter long measuring stick’. The scale defines the units or dimensions for which the measurement was taken e.g. ‘Meters’.

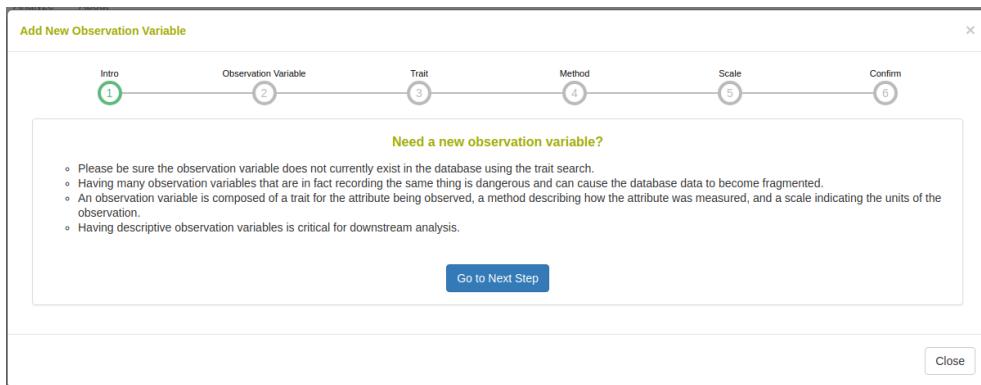
Generally, observation variables are defined in ontologies that are predefined. We often use ontologies from cropontology.org. In this case, you will not be able to define your own observation variables directly; instead, you will need to contact us and we will add the observation variable for you.

For databases where the user has greater control, we have an interface to allow addition of observation variables, along with traits, methods, and scales. To begin, go to the Search->Traits page.

If the database you are on allows you to directly add observation variables, you will see the following button at the bottom of the page.



When you click the button, the following workflow will appear. You should be logged in or else it will not allow addition of the observation variable. The workflow begins with an introduction.



On the next workflow step, you select the ontology that you want to insert the new observation variable into. You must also give a name and a definition for the new observation variable.

A screenshot of the 'Add New Observation Variable' workflow. The title bar says 'Add New Observation Variable'. Below it is a horizontal timeline with six numbered circles: 1 (Intro), 2 (Observation Variable), 3 (Trait), 4 (Method), 5 (Scale), and 6 (Confirm). Step 2 is highlighted with a green circle. A large text area contains the message 'Define your observation variable' followed by a note: 'An observation variable is composed of a trait for the attribute being observed, a method describing how the attribute was measured, and a scale indicating the units of the observation.' Below this are two input fields:

Observation Variable Ontology Name:

- COMP (Composed traits)
- ISOL (ISOL)

New Observation Variable Name: Plant height in meters extracted from RGB image using ImageJ

New Observation Variable Definition: Plant height in meters extracted from an red green blue (RGB) image using ImageJ

At the bottom of the text area is a blue 'Go to Next Step' button, and at the bottom right is a 'Close' button.

On the next workflow step, you select the trait ontology to use. Once you

20.1. MANAGING OBSERVATION VARIABLES WITH TRAITS, METHODS, AND SCALES 195

select a trait ontology, a select containing all the terms in the selected ontology will appear. You can either select a trait or if it does not exist in the select, you can create a new one by giving a name and a definition for the new trait.

The screenshot shows a workflow titled "Add New Observation Variable" with six steps: Intro, Observation Variable, Trait, Method, Scale, and Confirm. Step 3, "Trait", is currently active. The interface includes a "Define your trait" section with instructions and two radio button options: "COMP (Composed traits)" (unchecked) and "ISOL (ISOL)" (checked). Below this is a dropdown menu labeled "Existing Traits in Selected Ontology" showing "None". A note states: "If the trait does not exist in the ontology you selected above, you can add a new trait into the ontology here." At the bottom, there are fields for "New Trait Name" (containing "Plant height from base of plant to highest point on branch") and "New Trait Definition" (containing "The height of a plant from the base of the plant to the highest possible point on a branch"). A "Go to Next Step" button is at the bottom right, and a "Close" button is at the bottom center.

On the next workflow step, you select the method ontology to use. Once you select a method ontology, a select containing all the terms in the selected ontology will appear. You can either select a method or if it does not exist in the select, you can create a new one by giving a name and a definition for the new method.

Add New Observation Variable

A method defines the how it was measured. It is one component of an observation variable; the others are a trait and a scale.

Method Ontology Name: CASSTISS (cass_tissues)

Existing Methods in Selected Ontology: None

If the method does not exist in the ontology you selected above, you can add a new method into the ontology here.

New Method Name: ImageJ for plant height extraction from RGB image

New Method Definition: A script in [ImageJ](#) for extracting plant height from a red, green, blue (RGB) image

Go to Next Step

Close

On the next workflow step, you select the scale ontology to use. Once you select a scale ontology, a select containing all the terms in the selected ontology will appear. You can either select a scale or if it does not exist in the select, you can create a new one by giving a name and a definition for the new scale. You can also define a format, minimum, maximum, categories, and default value for the new scale.

20.1. MANAGING OBSERVATION VARIABLES WITH TRAITS, METHODS, AND SCALES 197

Add New Observation Variable

Intro 1 Observation Variable 2 Trait 3 Method 4 Scale 5 Confirm 6

Define your scale

Scale Ontology Name: UO (Units)

Existing Scales in Selected Ontology: mjUO:03000001

If the scale does not exist in the ontology you selected above, you can add a new scale into the ontology here.

New Scale Name: e.g. mass in kilograms

New Scale Definition: e.g. the mass in kilograms

New Scale Format: Select One

New Scale Minimum: e.g. 1

New Scale Maximum: e.g. 10000

New Scale Categories ('/' separated): e.g. 1/3/5/7

New Scale Default Value: e.g. 5

Go to Next Step

Close

On the last page of the workflow, you confirm the submission.

Add New Observation Variable

Intro 1 Observation Variable 2 Trait 3 Method 4 Scale 5 Confirm 6

Confirm you are creating this observation variable in the database

Submit

Close

Afterwards, you can use the newly created observation variable ontology term in your phenotyping.

Chapter 21

Managing Drone Imagery

21.1 Image-Phenotyping Dashboard

1. Upload raw image-captures in a compressed file (.zip) for orthophoto-mosaic assembly or upload previously stitched orthophotomosaic raster (.PNG, .JPG) imagery.
2. Dashboard shows all field trials and uploaded imaging events in collapsible sections.
3. Follow standard processes to manually create templates for assignment of plot-polygon images to the field experiment design.
4. All imagery is shown with the spectral category within collapsible sections. Figure shows NIR imagery.
5. Apply Fourier transform filtering, thresholding, and vegetation index masking. Plot-polygon images for all image processes are shown.
6. Extract and export phenotypic values from plot-polygon images for analyses and model training.

21.2 Image Input

Clicking “Upload Imagery” will open the following dialog.

This workflow will guide you through uploading aerial images to the database

Your field trial must already be in the database before you can upload images for it. Please go to [Manage->Field Trials](#) if it is not.

A field trial represents plots in the field where each plot has a globally unique *plot_name*, a sequential *plot_number* that is unique in the trial (e.g. 101, 102, 103 for three separate plots), and an *accession_name* representing the genotype being tested in that plot. Each plot can belong to different blocks (*block_number*) and reps (*rep_number*) depending on the experimental design you are using (e.g. complete block vs augmented design). Each plot can have a *row_number* and *col_number* indicating the relative position of the plot in the field. A field trial can represent a yield trial, a phenotyping trial, a crossing block, a greenhouse, a nursery, etc.

If you have raw aerial images that have not been stitched into an orthophotomosaic image of the whole field, your raw images should be uploaded using a zipfile (.zip). You can have several drone runs for a single field trial. For an individual drone run, once you have uploaded all photos, you can stitch an orthophotomosaic together. Afterwards, you will have options to cut the ortho image into plot polygons and extract phenotypes for those plots into the database. The maximum zipfile size is 2GB.

If you already have an orthophotomosaic image of your entire field, you can upload that image under a field trial and a drone run. Afterwards, you will have options to cut the ortho-image into plot polygons and extract phenotypes for those plots into the database. The maximum size for each image is 200MB. The preferred upload format is PNG.

Example Data: Micasense 5 Band Raw Images (Unstitched image-captures) (Upload zipfile for ImageBreed to stitch.)
Example Data: Micasense 5 Band Panel Images (Micasense calibration panel images.) (Upload zipfile for ImageBreed to calibrate Micasense raw-captures during stitching.)
Example Data: Micasense 5 Band Previously Stitched Orthophotomosaic Images (PNG Files in provided zipfile. Can upload each band separately into ImageBreed.)

[Go to Next Step](#)

Raw-captures can be uploaded in a compressed (.zip) file so that they can be assembled into an orthophotomosaic. If orthophotomosaic assembly is not required, raster images (.PNG, .JPG) can be uploaded. Example data is given for raw Micasense RedEdge 5-band multispectral captures and for stitched orthophotomosaics.

Select your field trial

Field Trial: 2015_NYH2

[Go to Next Step](#)

To begin uploading images, a field trial must be selected. The field trial must already be saved in the database. For information about adding a field trial, please read the Field Trial documentation.

Upload Drone Imagery

Select or create new drone run

Show 10 entries Search:

Select	Imaging Event Name	Imaging Event Type	Imaging Event Description	Imaging Event Date	Camera	Field Trial Name	Field Trial Description
<input checked="" type="checkbox"/>	2015_NYH2_07212015	Aerial Medium to High Res	Orthos from Nick Kaczmar from Pix4d	2015-July-21	micasense_5	2015_NYH2	G2F NYH2 2015

Showing 1 to 1 of 1 entries Previous Next

[Create new drone run if not present in table](#)

Imaging Event Name:

Imaging Event Type:

Camera Type:

Imaging Event Description:

Imaging Event Date:

[Go to Next Step](#)

[Close](#)

The image data is added to an imaging (drone run) event. Here you can select a previously saved imaging event or you can create a new one by defining a name, description, and date.

Upload Drone Imagery

Stitched vs Unstitched and Number of Bands (Image Sets) To Upload

- Raw images (unstitched) coming from your drone can be uploaded in a zip file. We can then stitch them together into an orthophotomosaic of the entire drone run.
- Or you can choose to upload a single image and skip any stitching

- It is possible to upload regular RGB or Black and White photos.
- For multi-spectral cameras, it is possible to upload individual spectra orthomosaic photos.
- When uploading many separate bands of unstitched images, you will upload a single zipfile (.zip) which contains all images. In the zipfile each image is named following the template IMG_0001_1.tif, IMG_0001_2.tif, ..., IMG_0001_5.tif, ..., IMG_9999_5.tif. The final number represents the 5 bands coming from the camera, while the middle number is an index for the image capture. The middle number can be as many digits long as needed. The images should be in order in the zipfile. You will also need to upload a zipfile (.zip) containing the Micasense radiometric calibration panel images, so that ImageBreed can produce the best orthomosaic possible.

Do you require stitching an ortho image of the drone run:

Select One
 Yes, I am uploading a zipfile of images to stitch
 No

Number of Spectral Bands (Image Sets) To Upload:

[Go to Next Step](#)

[Close](#)

The uploaded data can be raw image-captures or complete raster images. Here you can select whether orthophotomosaic stitching is required.

Stitched vs Unstitched and Number of Bands (Image Sets) To Upload

- Raw images (unstitched) coming from your drone can be uploaded in a zip file. We can then stitch them together into an orthophotomosaic of the entire drone run.
- Or you can choose to upload a single image and skip any stitching.
- It is possible to upload regular RGB or Black and White photos.
- For multi-spectral cameras, it is possible to upload individual spectra orthomosaicphotos.
- When uploading many separate bands of unstitched images, you will upload a single zipfile (.zip) which contains all images. In the zipfile each image is named following the template IMG_0001_1.tif, IMG_0001_2.tif, ..., IMG_0001_5.tif, ..., IMG_9999_5.tif. The final number represents the 5 bands coming from the camera, while the middle number is an index for the image capture. The middle number can be as many digits long as needed. The images should be in order in the zipfile. You will also need to upload a zipfile (.zip) containing the Micasense radiometric calibration panel images, so that ImageBreed can produce the best orthomosaic possible.

Do you require stitching an ortho image of the drone run:

Yes, I am uploading a zipfile of images to stitch

Go to Next Step

In the case that orthophotomosaic stitching is required, select ‘yes’. On the next step you will see the following: Upload a zipfile with the raw-captures. When uploading Micasense RedEdge raw-captures, provide images of the Micasense calibration panels in a zipfile as well.

Select Image(s) to Upload

Drone Images ZipFile (.zip) (2GB Maximum): No file chosen

Micasense Radiometric Calibration Images ZipFile (.zip): No file chosen

Working Image Scale (Megapixels): 0.6

Submit

In the case that orthophotomosaic assembly is not required, simple upload the raster images. Select the number of image bands that will be uploaded

e.g. for a five band multispectral camera, select 5.

In the cases that orthophotomosaic stitching is not required, select ‘no’. On the next step you will see the following:

Upload an image at each band with a unique name, description, and spectral type.

21.3 Standard Process

Once imagery is uploaded, it will appear on the dashboard under the field trial. Clicking the “Run Standard Process” button will begin extracting plot-polygon phenotypes from the imagery.

The screenshot shows the 'Manage Drone Imagery' dashboard. At the top, there are buttons for 'Upload Imagery', 'Download Image-Phenotypes', and 'Calculate Statistics'. Below this is a search bar and a table titled 'Field Trials -> Imaging Events'. The table lists two imaging events:

- 2015_NYH2_07212015** (2 Imaging Events)
 - 2015_NYH2_07212015** 2015-July-21
 - 2015_NYH2_08072015** 2015-August-07

For the second imaging event (2015_NYH2_08072015), there is a detailed description table and a button labeled 'Run Standard Process For 2015_NYH2_08072015' which is highlighted with a red box. Below this, there is a section for 'No Plot Images Saved' and a table of image bands with their names, descriptions, and types, each with a 'View Images' button.

Image Band(s)	Images/Actions
Name: 2015_NYH2_08072015_Blue Description: Ortho from Nick Kaczmar from Pix4d Type: Blue (450-520nm)	View Images
Name: 2015_NYH2_08072015_Green Description: Ortho from Nick Kaczmar from Pix4d Type: Green (515-600nm)	View Images
Name: 2015_NYH2_08072015_Red Description: Ortho from Nick Kaczmar from Pix4d Type: Red (600-690nm)	View Images
Name: 2015_NYH2_08072015_NIR Description: Ortho from Nick Kaczmar from Pix4d Type: NIR (780-3000nm)	View Images
Name: 2015_NYH2_08072015_RedEdge Description: Ortho from Nick Kaczmar from Pix4d Type: Red Edge (690-750nm)	View Images

Clicking the button will open the following dialog.

The dialog is titled 'Manage Drone Imagery: Run A Standard Process'. It features a horizontal navigation bar with numbered steps: 1 (Intro), 2 (Drone Run Band), 3 (Rotate), 4 (Cropping), 5 (Thresholding), 6 (Plot Polygons), 7 (Apply), 8 (Indices), and 9 (Phenotypes). Step 2 is highlighted with a green circle. Below the navigation bar is a text box containing the following text:

This workflow will guide you through applying a standard process to your aerial imaging bands

Here you can take one of the drone run bands you uploaded all the way through the process to plot image saving. This will require manual steps such as image rotation, cropping, and plot polygon templating. After you have completed this process for one drone run band, you can apply it to all other drone run bands and other calculated vegetative indices.

At the bottom right of the dialog is a blue button labeled 'Go to Next Step'.

Select a drone run band to use in this process. In the case of the Micasense 5 band multispectral camera there will be 5 bands shown here; select the NIR channel in this case because it has the highest contrast. In the case of standard color images, there will only be the RGB Color Image option here.

Manage Drone Imagery: Run A Standard Process

Select a drone run band

Please select one drone run band to take through the process. It is recommended to select a band that has high contrast, such as a NIR band.

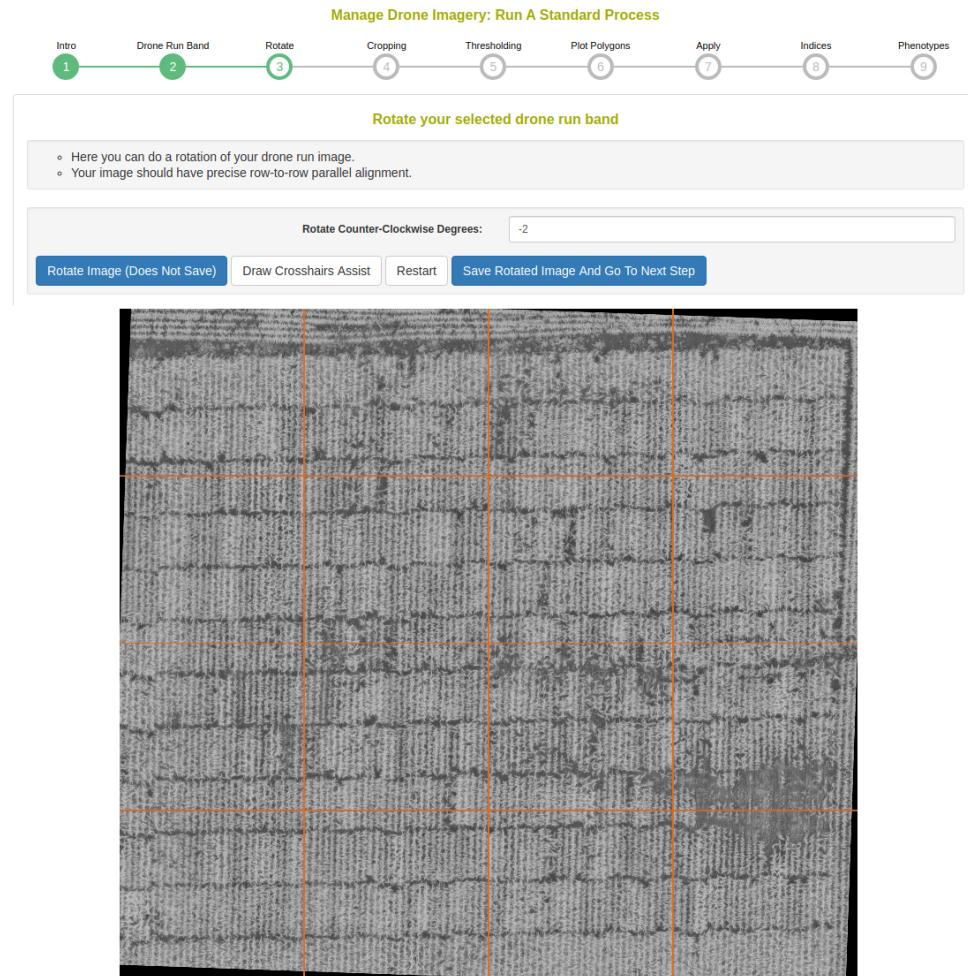
Show 10 entries									Search: <input type="text"/>	
Select	Drone Run Band Name	Drone Run Band Description	Drone Run Band Type	Drone Run Name	Drone Run Description	Drone Run Date	Field Trial Name	Field Trial Description		
<input type="checkbox"/>	2015_NYH2_08072015_Blue	Ortho from Nick Kaczmar from Pix4d	Blue (450-520nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015		
<input type="checkbox"/>	2015_NYH2_08072015_Green	Ortho from Nick Kaczmar from Pix4d	Green (515-600nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015		
<input type="checkbox"/>	2015_NYH2_08072015_Red	Ortho from Nick Kaczmar from Pix4d	Red (600-690nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015		
<input type="checkbox"/>	2015_NYH2_08072015_NIR	Ortho from Nick Kaczmar from Pix4d	NIR (780-3000nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015		
<input type="checkbox"/>	2015_NYH2_08072015_RedEdge	Ortho from Nick Kaczmar from Pix4d	Red Edge (690-750nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015		

Showing 1 to 5 of 5 entries

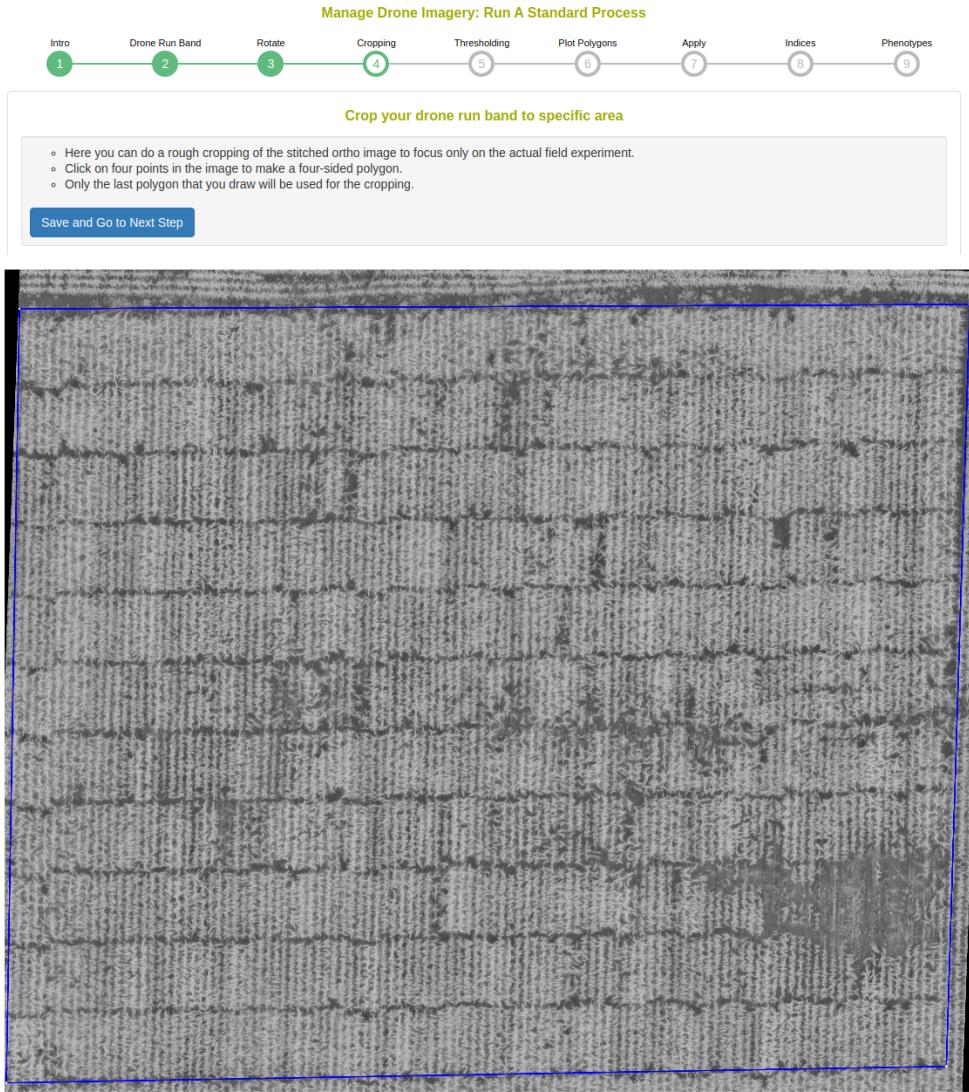
Previous 1 Next

[Go to Next Step](#)

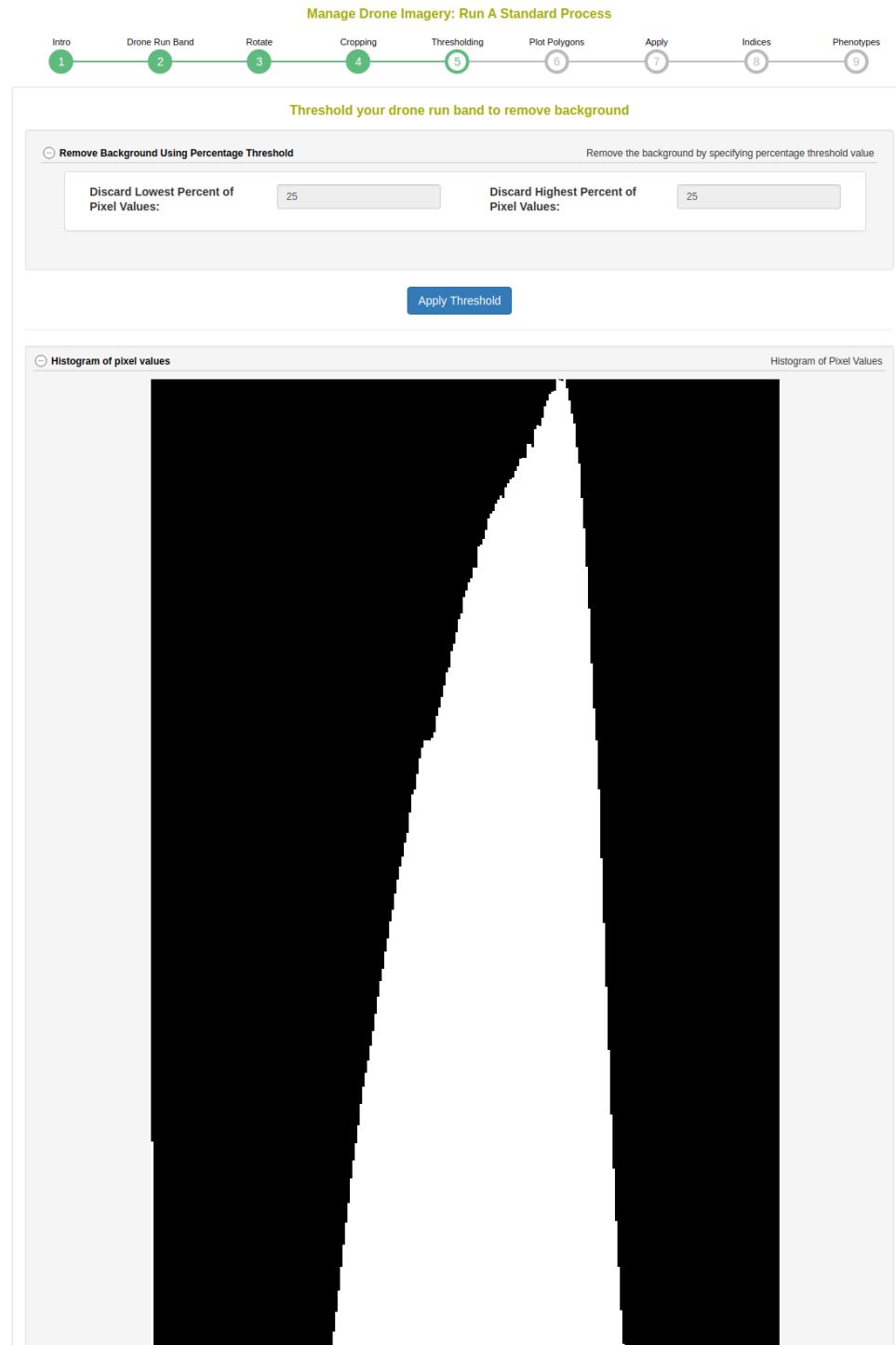
Rotate the image so that there the plots are oriented in a grid fashion. There can be a skew in the field layout, as seen in the following example.



Perform a rough cropping of the image by clicking on the four corners of the field. Cropping is important to remove any extraneous parts of the image.



This step shows a histogram of the cropped image. The standard process will magnitude threshold the top and low ends of the distribution.



In this step, the template for the plot polygons in the experimental field design are associated to the image. First, defined the number of rows and columns in the field experiment. Then click the four corners of the image, in respect to the top right, top left, bottom left, and bottom right positions. Next click on “Draw Plot Polygon Template”. Review the template and clear/repeat the process until the template matches well. It is possible to “copy/paste” templates in the case where there are large breaks in the field design. Next, scroll down to the “assign Plot Polygons to Field Trial Entities” section. Select the location of Plot Number 1 as either “top left” or “top right” and whether the field design is serpentine or zigzag. Click on “Generate Assignments” and review that the names of the plots appear correctly in the overlay on the image. Finally, click “Finish and Save Polygons to Plots” when you have confirmed the assignments.

Manage Drone Imagery: Run A Standard Process

Intro 1 Drone Run Band 2 Rotate 3 Cropping 4 Thresholding 5 Plot Polygons Apply Indices Phenotypes

Define plot polygons relative to the field layout

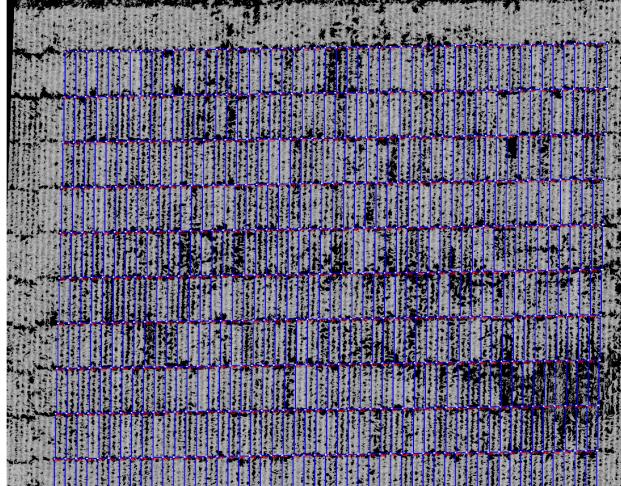
1 Generate Polygon Template Tool Overlay a uniform grid over the image.

Number of Rows: 10 Number of Columns: 50

2 Draw Plot Polygon Template (Does not save. Apply multiple templates if needed.)

3 Previously Used Plot Polygon Templates

Total Image Width: 3316px. Total Image Height: 2680px.



Assign Plot Polygons to Field Trial Entities

Location of First Plot (e.g. plot number): 1³ Second Plot Follows First Plot Going: Right

4 Plot Number Orientation: Serpentine
 5

plot_name	accession_name	plot_number	block_number	is_a_control	rep_number	row_number	col_number	plot_geo_json	Polygon Assigned
2015_NYH2_plot_1	PHV63LH195	1	1	null	1	1	45		
2015_NYH2_plot_10	C0109PHZ51	10	1	null	1	1	54		
2015_NYH2_plot_100	LH145UH82	100	4	null	1	2	45		
2015_NYH2_plot_101	PHN11_LH145_0002PHB47	101	5	null	1	3	45		
2015_NYH2_plot_102	NYH-081PHB47	102	5	null	1	3	46		
2015_NYH2_plot_103	W10005_0029PHB47	103	5	null	1	3	47		
2015_NYH2_plot_104	B109PHZ51	104	5	null	1	3	48		
2015_NYH2_plot_105	PH207_PHG47-13PHB47	105	5	null	1	3	49		
2015_NYH2_plot_106	PH207_PHG47-10PHB47	106	5	null	1	3	50		
2015_NYH2_plot_107	C045LUH82	107	5	null	1	3	51		

Show 10 entries Search:
 ...

Next, the dialog shows you that the standard process will be repeated for all uploaded image bands.

Apply these same steps to other drone run bands in the current drone run

- Here you can apply the same actions you did for the previous steps 1 to 6, to additional drone run bands in this drone run.
- Thresholding will be done dynamically, by removing the top and bottom 20% of pixel values.

Select	Drone Run Band Name	Drone Run Band Description	Drone Run Band Type	Drone Run Name	Drone Run Description	Drone Run Date	Field Trial Name	Field Trial Description
<input checked="" type="checkbox"/>	2015_NYH2_08072015_Blue	Ortho from Nick Kaczmar from Pix4d	Blue (450-520nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015
<input checked="" type="checkbox"/>	2015_NYH2_08072015_Green	Ortho from Nick Kaczmar from Pix4d	Green (515-600nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015
<input checked="" type="checkbox"/>	2015_NYH2_08072015_Red	Ortho from Nick Kaczmar from Pix4d	Red (600-690nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015
<input checked="" type="checkbox"/>	2015_NYH2_08072015_NIR	Ortho from Nick Kaczmar from Pix4d	NIR (780-3000nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015
<input checked="" type="checkbox"/>	2015_NYH2_08072015_RedEdge	Ortho from Nick Kaczmar from Pix4d	Red Edge (690-750nm)	2015_NYH2_08072015	Orthos from Nick Kaczmar from Pix4d	2015-August-07	2015_NYH2	G2F NYH2 2015

Showing 1 to 5 of 5 entries

Previous
1
Next

[Go to Next Step](#)

Next, choose which vegetation indices to apply.

Manage Drone Imagery: Run A Standard Process

Intro
1
Drone Run Band
2
Rotate
3
Cropping
4
Thresholding
5
Plot Polygons
6
Apply
7
Indices
8
Phenotypes
9

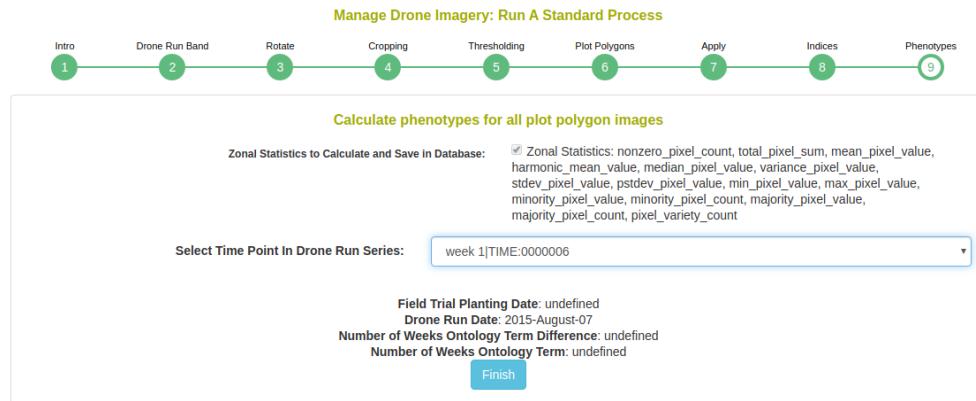
Create and apply these same steps to vegetative indices

Vegetative Indices To Apply:

Triangular Greenness Index (TGI)
 Visible Atmospheric Resistant Index (VARI)
 Normalized Difference Vegetative Index (NDVI)
 Normalized Difference Red Edge Vegetative Index (NDRE)

[Go to Next Step](#)

Next, choose the phenotypic values to extract. You must define the time point for which the phenotype is; if the field trial has a planting date, the time point will automatically be populated as image date minus the planting date.



After completing the standard process, the job will continue in the background until it completes. You can check the status of the job from the dashboard.

21.4 Ground Control Points

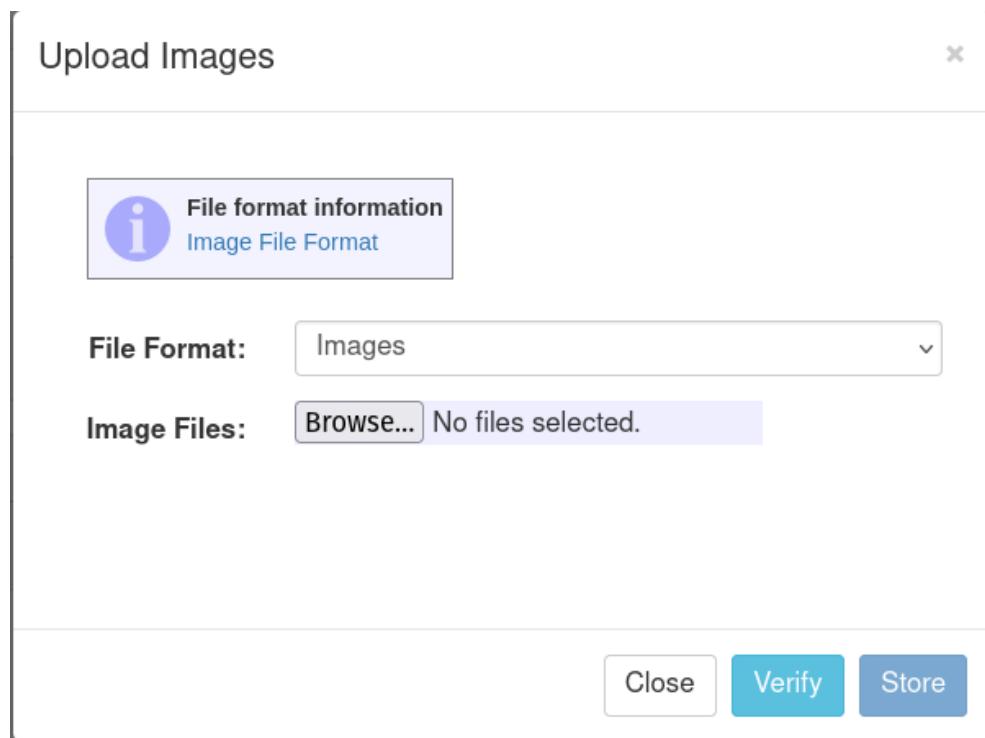
Ground control points can be saved after an imaging event has undergone the standard process on orhomosaics. Ground control points can then be used across imaging events on the same field experiment in order to automate the entire standard process.

Chapter 22

Managing Images

22.1 Uploading Image files

Clicking “Phenotyping” under Manage, and then “Upload Images” will open this dialog:



If uploading images directly, select the file format “Images”. A single image or multiple selected images can be uploaded using this file format. Each of these images must have been downloaded from the Fieldbook app, or have a filename that follows this structure: observationUnitName, traitname, number, timestamp joined by underscores. The filename can not include any underscores other than the ones separating each value. For example: 38873_branching_5_2025-07-28.jpg

If the images have been taken in the Fieldbook app, they will have EXIF data that includes their associated stocks and traits. This EXIF data will be automatically parsed and the associations will be made when the images are uploaded.

After selecting your images for upload, clicking verify will check if your filenames or image metadata is correct and corresponds to existing stocks in the database.

Upload Images ×

File format information
Image File Format

File Format: Images ▼

Image Files: Browse... 38873_branching_5_2025-07-28.jpg

 38873_branching_5_2025-07-28.jpg

and 0 additional image files selected and
ready for verification.

Verification complete. All image files match an existing observationUnit. Ready to store images.

Close Verify Store

If the verification is successful, click store to save these images in the database.

If uploading images with associated phenotypic data, select the file format “Images with Associated Phenotypes”. A spreadsheet with the phenotypic values needs to be uploaded alongside a zipfile of images. The spreadsheet

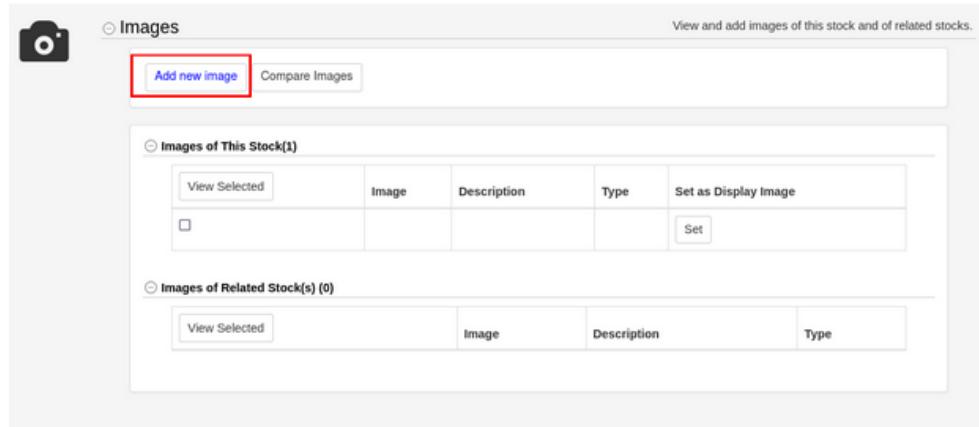
can be in .xls or .xlsx format, and the column headers need to be:

observationUnitName | observationVariableName | value | timestamp | image_name | person

The image_name column should match the file names of the associated images in the zipfile.

22.2 Uploading Images Directly to Stocks

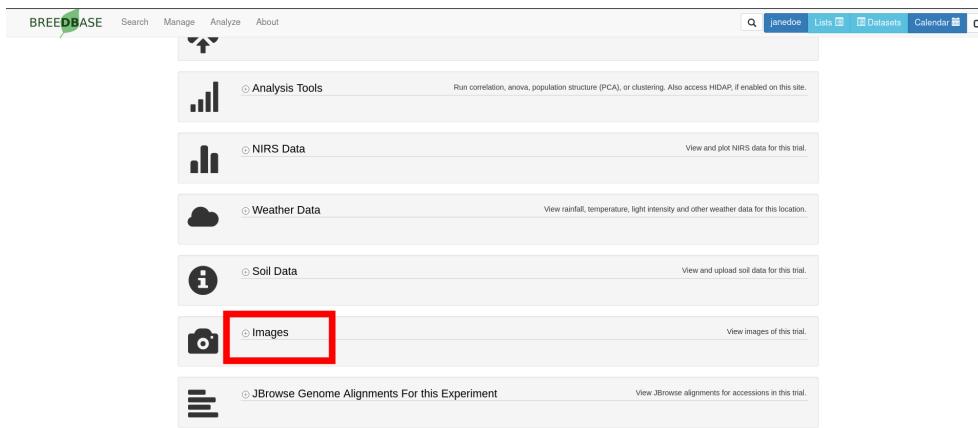
In the Images section on a stock details page you can add a new image associated with that stock by clicking the “Add Image” button. This will associate any images uploaded through this dialog with the corresponding stock.



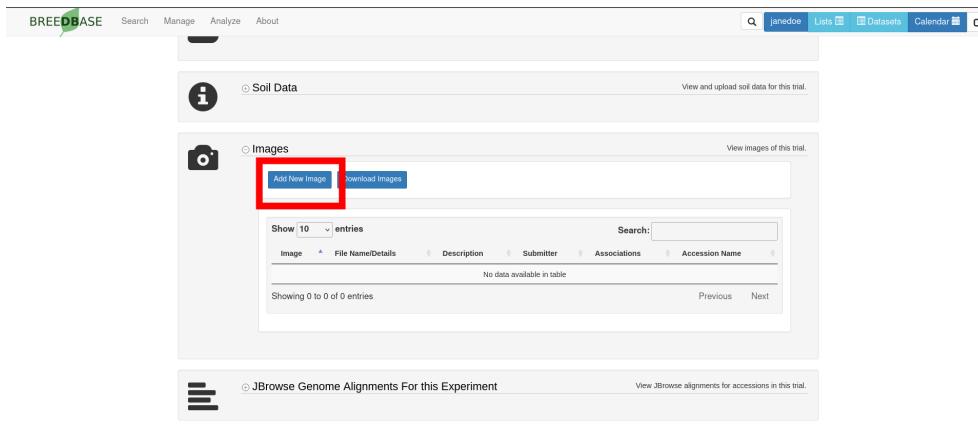
22.3 Uploading Images from the Field Trial and Image Analysis Pages

Another way to upload images is through the field trial details page. Navigate to a field trial details page and open the “Images” section.

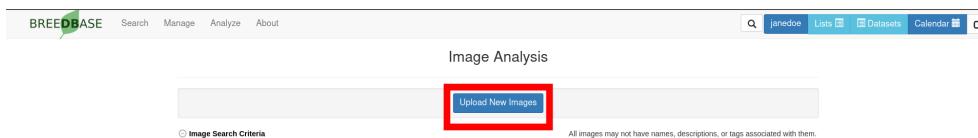
22.3. UPLOADING IMAGES FROM THE FIELD TRIAL AND IMAGE ANALYSIS PAGES 217



Then click the “Add New Image” button to open the image upload dialog, where images can be uploaded in the same formats as described previously.



To upload images from the Image Analysis page, click on “Image Analysis” under the “Analyze” menu. Then click the “Upload New Images” button on this page.



This will open the same image upload dialog.

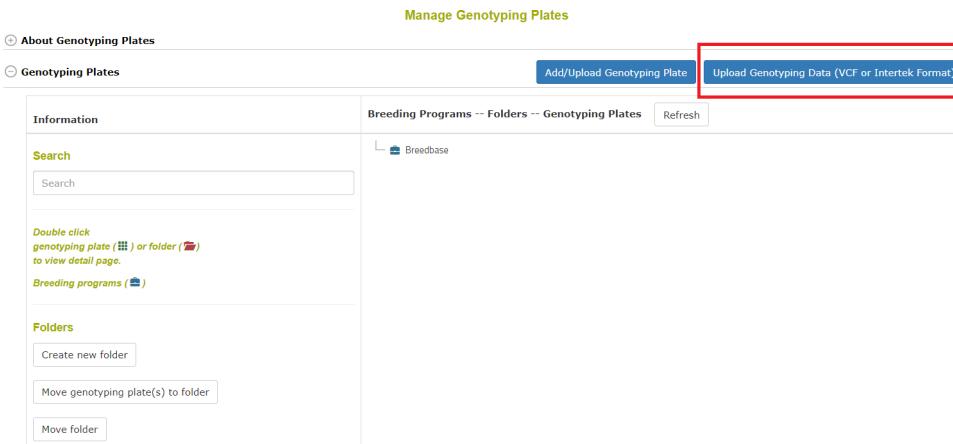
Chapter 23

Managing VCF Data

23.1 Uploading VCF Data

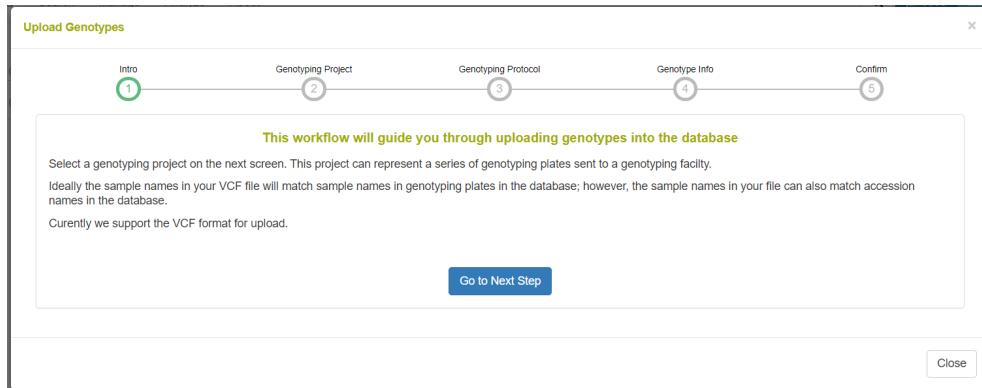
Genotyping data in VCF can be loaded from the web-interface. Breedbase can store any genotypic variants from a VCF, allowing for polyploids, structural variants, etc. without problems.

To begin go to Manage->Genotyping Plates and click the button seen below: Note that you do not need to have genotyping plates uploaded to upload VCF data; you may upload genotyping data to accessions or you can upload genotyping data for tissue samples in genotyping plates.



The screenshot shows the 'Manage Genotyping Plates' page. At the top, there are two tabs: 'About Genotyping Plates' and 'Genotyping Plates'. Below the tabs, there are two main sections: 'Information' and 'Folders'. The 'Information' section contains a search bar, a note about double-clicking, and links for breeding programs and folders. The 'Folders' section contains buttons for creating new folders, moving plates to a folder, and moving a folder. At the top right of the page, there are buttons for 'Add/Upload Genotyping Plate' and 'Upload Genotyping Data (VCF or Intertek Format)'. A red box highlights the 'Upload Genotyping Data (VCF or Intertek Format)' button. The background shows a tree view of the 'Breedbase' folder structure.

The workflow begins with an intro:



On the following step in the workflow, a genotyping project is defined or selected. A genotyping project is a high-level entity for grouping several genotyping events. It is defined with a name, description, name, breeding program, and genotyping facility (IGD, Intertek, etc.).

Select	Genotyping Project Name	Description	Breeding program	Year	Location	Genotyping Facility
<input type="checkbox"/>	GenoTestCassava	asd	Breedbase	2020	igd	
<input type="checkbox"/>	GenoTestMaize	asd	Breedbase	2020	igd	
<input type="checkbox"/>	GenoTestMusa	asd	Breedbase	2020	igd	

Show 10 entries

Search:

Showing 1 to 3 of 3 entries

Previous 1 Next

My project is not here. Create a new one.

Genotyping Project Name: e.g. NextGenCassava

Genotyping Facility: None

Breeding Program: Breedbase

Year: 2020

Description:

Go to Next Step

Close

The following step is to define or select a genotyping protocol. A genotyping protocol represents the set of markers being called against a specific reference genome. A genotyping protocol is defined with a name, description, reference genome name, species name, and a location of data generation. Note in the picture that you can select whether the samples in your file are accessions or tissue samples in the database; tissue samples are for when a genotyping plate is stored in the database. There is an option to parse the sample names for appended sequencing numbers from IGD, where the sample names are like “accession:igdnumber”.

The screenshot shows a web-based form titled "Upload Genotypes". At the top, it says "Showing 1 to 3 of 3 entries" and has navigation buttons for "Previous", "1", and "Next". Below this is a button: "My protocol is not here. Create a new one." The main form area contains the following fields:

- Genotyping Protocol Name: (Input field: e.g. GBS ApeKI Cassava genome v6 Jan2015)
- Genotyping Protocol Reference Genome: (Input field: Mesculenta_511_v7.0)
- Species: (Input field: e.g. Manihot esculenta)
- Description: (Input field)
- Choose Sample Unit:
 - Exported Tissue Sample Name: The sample names in your VCF are tissue_sample_names that already exist in genotyping plates (e.g. 96 well plates) in the database. The sample names in your VCF file can be the tissue_sample_name triple pipe joined to the accession_name (e.g. tissue_sample_name||accession_name) or just simply the tissue_sample_name corresponding to the genotyping plate well.
 - Accession: The sample names are of accession names
- Location of Data Generation: (Input field: Cornell Biotech)
- Exported Tissue Sample Names Include Numbers Generated by Genotyping Facility (e.g. sample_name:IGD1001:09):
The generated number is separated from the tissue sample name in the database by a ':' separating character.

At the bottom of the form is a "Go to Next Step" button and a "Close" button.

The final step is to select the VCF from your computer and upload it. The web interface can be used to upload files arbitrarily large; it is a NGINX configuration to set this size.

The screenshot shows a software interface titled 'Upload Genotypes'. At the top, there is a horizontal navigation bar with five steps: 'Intro' (step 1), 'Genotyping Project' (step 2), 'Genotyping Protocol' (step 3), 'Genotype Info' (step 4, highlighted in green), and 'Confirm' (step 5). Below the navigation bar, the main area is titled 'Provide genotype information'. It contains a dropdown menu labeled 'Select type of genotyping data:' with 'VCF' selected. To the left of this menu is a button labeled 'File format information' with 'VCF format' underneath it. To the right is a file selection input field labeled 'Select VCF File:' with 'Choose File | No file chosen' displayed. At the bottom of the main area is a blue 'Go to Next Step' button. In the bottom right corner of the window is a 'Close' button.

23.2 Searching and Downloading VCF Data

The Search Wizard is the primary means of querying data in the database. Go to Search->Wizard to begin.

Once genotyping protocols are stored, select Genotyping Protocols from the first dropdown menu. Then if you select one or more and select Accessions from the second dropdown menu, you will see the accessions for which genotypes were stored. As seen in the following picture, there is a section for filtering genotypes by chromosome, start position, and end position. Genotypes can be downloaded in VCF or DosageMatrix formats.

Search Wizard

Don't see your data? Refresh Lists | Update Wizard

Genotyping Protocols

Accessions

Select Column Type

Select Column Type

Search

Search

Search

Search

Select All 1/3 Clear

Select All 3/315 Clear

Select All 0/0 Clear

Select All 0/0 Clear

Match ANY ALL

Match ANY ALL

Match ANY ALL

Match ANY ALL

Add to List... Add

Add to List... Add

Create New List... Create

Create New List... Create

Load/Create Datasets using Columns

Related Genotype Data

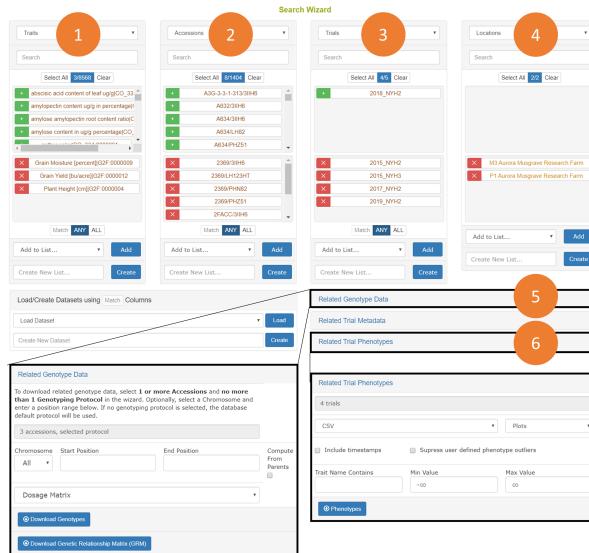
To download related genotype data, select **1 or more Accessions** and **no more than 1 Genotyping Protocol** in the wizard. Optionally, select a Chromosome and enter a position range below. If no genotyping protocol is selected, the database default protocol will be used.

3 accessions, selected protocol

Chromosome	Start Position	End Position
All		

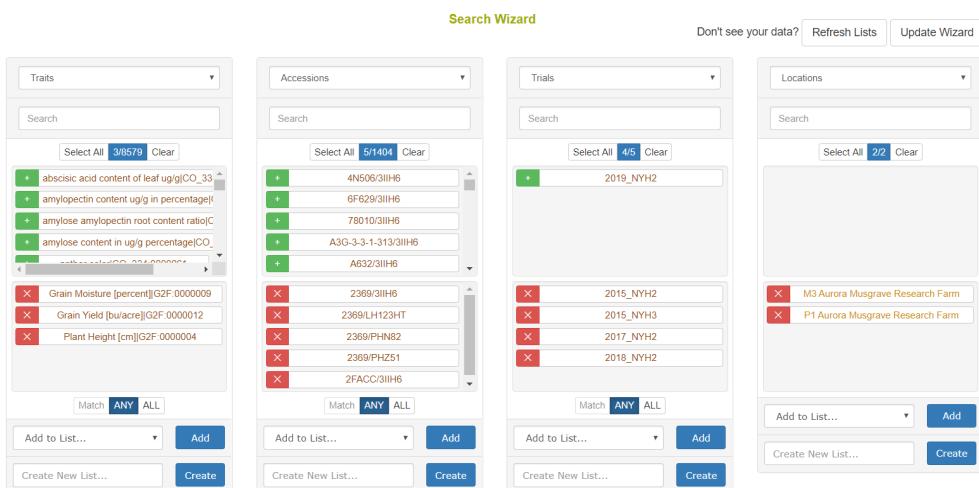
VCF

Using the “Default genotyping protocol” which is configured in a system, you can query over field phenotypic evaluations before downloading genotypes and phenotypes.



23.3 Searching Protocols

Genotyping protocols can be search by going to Search->Genotyping Protocols. To download genotypes accessions must be selected, though any combination of search criteria can be used to filter and select those accessions. If a genotyping protocol is not selected, then the default genotyping protocol set in the configuration will be used. Genotyping protocols can also be selected in the wizard.



The genotyping download menu on the Search Wizard presents options for filtering by chromosome, start position, and end position. Genotypes can be downloaded in VCF or Dosage Matrix formats. The genomic relationship matrix (GRM) can be downloaded for the selected accessions in a tab-delimited matrix format or in a three-column format that is useful in Asreml. Genotypes can be computed from the parents in the pedigree if those parents are genotyped by clicking on the “compute from parents” checkbox. Additionally, the GRM can be computed using genotypes of parents in the pedigree if the “compute from parents” checkbox is selected.

Related Genotype Data

To download related genotype data, select **1 or more Accessions** and **no more than 1 Genotyping Protocol** in the wizard. Optionally, select a Chromosome and enter a position range below. If no genotyping protocol is selected, the database default protocol will be used.

5 accessions, default protocol

Chromosome	Start Position	End Position	Compute From Parents
All			<input checked="" type="checkbox"/>

Dosage Matrix (tsv)

[Download Genotypes](#)

3-Column Format (tsv)

[Download Genetic Relationship Matrix \(GRM\)](#)

As is described elsewhere, the Search Wizard presents a way to filter phenotypic values by minimum and maximum values, and allow for download in CSV and Excel formats.

Related Trial Phenotypes

4 trials

CSV Plots

Include timestamps Suppress user defined phenotype outliers

Trait Name Contains	Min Value	Max Value
	-∞	∞

Phenotypes

23.4 Detail Pages and Deletion

The genotyping protocol detail page will show all information about the protocol such as the reference genome used, the header information lines in the uploaded VCF file, the markers involved, and the samples genotyped.

The markers section will show all markers used and their annotations, such as position, chromosome, alternate allele, reference allele, marker format, etc.

Markers View information about the markers used in this protocol.

Marker Name(s): Search

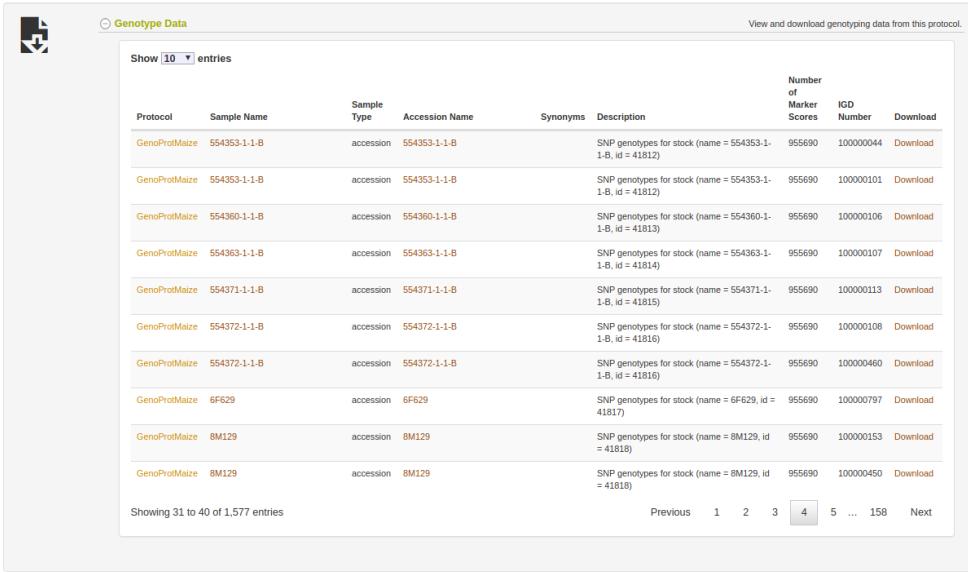
Show 10 entries

Marker Name	Chromosome	Position	Alternate	Reference	Quality	Filter	Info	Format
S0_1000880	0	1000880	T	C	.	PASS	.	GT
S0_1000890	0	1000890	.	G	.	PASS	.	GT
S0_1000912	0	1000912	.	C	.	PASS	.	GT
S0_1000916	0	1000916	.	C	.	PASS	.	GT
S0_1000922	0	1000922	.	C	.	PASS	.	GT
S0_1000924	0	1000924	G	A	.	PASS	.	GT
S0_101126	0	101126	.	G	.	PASS	.	GT
S0_1027188	0	1027188	.	T	.	PASS	.	GT
S0_1152731	0	1152731	.	C	.	PASS	.	GT

Showing 1 to 10 of 955,690 entries Previous 1 2 3 4 5 ... 95569 Next

The samples section will show all samples genotyped. Notice the Download links in the table which can be used to easily get the VCF file results for each

genotyped samples with all markers in the genotyping protocol. For getting multiple samples at once, use the Search Wizard as discussed above.



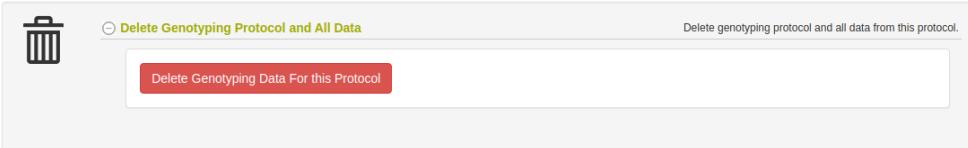
The screenshot shows a table titled "Genotype Data" with the sub-header "View and download genotyping data from this protocol." The table has columns: Protocol, Sample Name, Sample Type, Accession Name, Synonyms, Description, Number of Marker Scores, IGD Number, and Download. There are 10 entries shown, with a total of 1,577 entries. The entries include various accessions like 554353-1-1-B, 554360-1-1-B, 554363-1-1-B, 554371-1-1-B, 554372-1-1-B, 6F629, 8M129, etc., with their respective details and download links.

Protocol	Sample Name	Sample Type	Accession Name	Synonyms	Description	Number of Marker Scores	IGD Number	Download
GenoProtMaize	554353-1-1-B	accession	554353-1-1-B		SNP genotypes for stock (name = 554353-1-1-B, id = 41812)	955690	100000044	Download
GenoProtMaize	554353-1-1-B	accession	554353-1-1-B		SNP genotypes for stock (name = 554353-1-1-B, id = 41812)	955690	100000101	Download
GenoProtMaize	554360-1-1-B	accession	554360-1-1-B		SNP genotypes for stock (name = 554360-1-1-B, id = 41813)	955690	100000106	Download
GenoProtMaize	554363-1-1-B	accession	554363-1-1-B		SNP genotypes for stock (name = 554363-1-1-B, id = 41814)	955690	100000107	Download
GenoProtMaize	554371-1-1-B	accession	554371-1-1-B		SNP genotypes for stock (name = 554371-1-1-B, id = 41815)	955690	100000113	Download
GenoProtMaize	554372-1-1-B	accession	554372-1-1-B		SNP genotypes for stock (name = 554372-1-1-B, id = 41816)	955690	100000108	Download
GenoProtMaize	554372-1-1-B	accession	554372-1-1-B		SNP genotypes for stock (name = 554372-1-1-B, id = 41816)	955690	100000460	Download
GenoProtMaize	6F629	accession	6F629		SNP genotypes for stock (name = 6F629, id = 41817)	955690	100000797	Download
GenoProtMaize	8M129	accession	8M129		SNP genotypes for stock (name = 8M129, id = 41818)	955690	100000153	Download
GenoProtMaize	8M129	accession	8M129		SNP genotypes for stock (name = 8M129, id = 41818)	955690	100000450	Download

Showing 31 to 40 of 1,577 entries

Previous 1 2 3 4 5 ... 158 Next

The genotyping protocol and all associated genotyping data can be deleted from the genotyping protocol page.

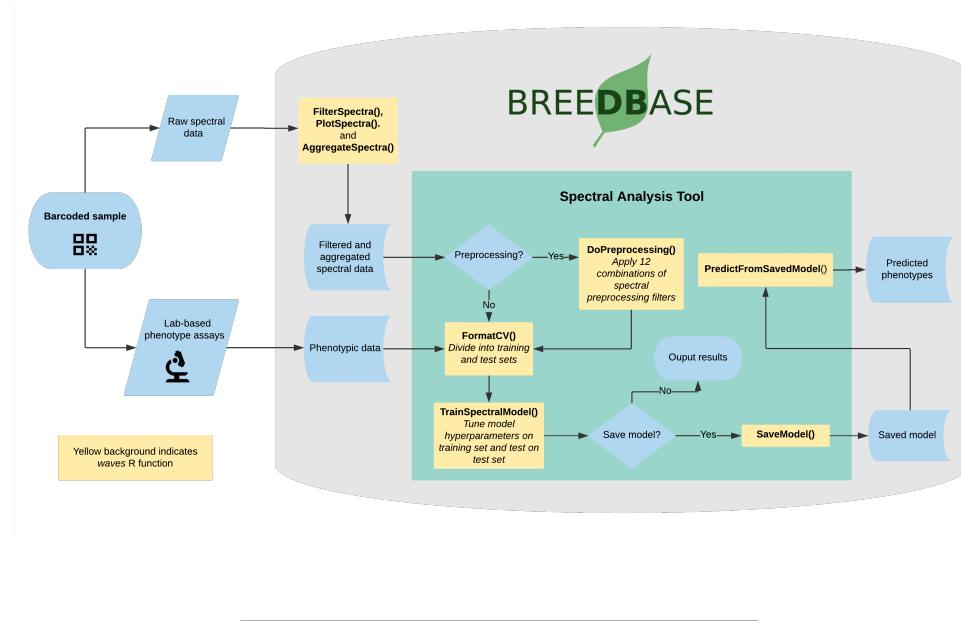


The screenshot shows a confirmation dialog with a trash can icon and the title "Delete Genotyping Protocol and All Data". The sub-header says "Delete genotyping protocol and all data from this protocol." A large red button in the center says "Delete Genotyping Data For this Protocol".

Chapter 24

Managing Spectral Data

Breedbase has implemented a flexible spectral data storage protocol that handles spectral data irrespective of the source spectrometer. Spectral data storage and analysis in Breedbase makes use of the R package *waves* for outlier identification, plotting, sample aggregation, and prediction model training.



24.1 Upload Spectral Data

Spectral data can be added as a CSV file that includes metadata in the leftmost columns followed by one column per spectral measurement to the right. Rows represent a single scan or sample, each with a unique ID that must match to a Breedbase observationUnitName. Future data transfer using BrAPI will allow for interoperability with data collection software.

To upload a spectral dataset, navigate to the ‘Manage NIRS Data’ page by selecting ‘NIRS’ in the ‘Manage’ menu and click the blue ‘Upload NIRS’ button. This will open an upload workflow. A link to the required file format and an example .csv file can be found by clicking in the light blue info box in this workflow. Another example of the file format is shown below.

- **id**: Optional identifier for each NIRS read. The id must be an integer.
- **sampling_id**: Optional identifier for each sample. Strings are allowed.
- **sampling_date**: Optional field. The format allowed is: YYYY-MM-DD.
- **observationunit_name**: Required field that matches existing data in the database. It can be the plot name, subplots, plant name, or tissue sample, depending how your trial is designed.
- **device_id**: Optional field to identify your device. Strings are allowed.
- **device_type**: Required field. It is possible upload data for a single device type. They can be: SCiO, QST, Foss6500, BunchiN500, or LinkSquare.
- **comments**: Optional field for general comments. All other columns are required wavelengths. You can add how many columns you want upload – there is no limit.

Manage NIRS Data

NIRS

Upload and perform analyses using NIRS data

Uploaded NIRS Data

NIRS Analyses

Trained NIRS Models

	A	B	C	D	E	F	G	H	I	J	K	L	M	N
1	Id	sample_id	sampling_date	observationunit_name	device_id	device_type	comments	740	741	742	743	744	745	746
2	1	7a6ac477-d291-4d07-af	2020-6-24	myTriad20_rept_acc_001	503E4BFC4E923999	SCIO		0.885707958	0.88572938	0.885590265	0.885493457	0.885493162	0.885572662	0.885628732
3	2	7a6ac477-d291-4d07-af	2020-6-24	myTriad20_rept_acc_002	503E4BFC4E923999	SCIO		0.89132994	0.8908724223	0.890824451	0.89079170	0.890797281	0.907620115	0.907593838
4	3	7a6ac477-d291-4d07-af	2020-6-24	myTriad20_rept_acc_003	503E4BFC4E923999	SCIO		0.889220207	0.889013119	0.888681812	0.888401257	0.888310362	0.888297321	0.888204052
5	4	73c648ca-5fb-4231-a1f6	2020-6-24	myTriad20_rept_acc_004	503E4BFC4E923999	SCIO		0.8900087	0.888604969	0.888191073	0.888958654	0.888933709	0.888072741	0.88824772
6	5	73c648ca-5fb-4231-a1f6	2020-6-24	myTriad20_rept_acc_005	503E4BFC4E923999	SCIO		0.89101707	0.93686202	0.93620132	0.937873867	0.937742819	0.937775129	0.9374594
7	6	73c648ca-5fb-4231-a1f6	2020-6-24	myTriad20_rept_acc_006	503E4BFC4E923999	SCIO		0.876289461	0.875981159	0.875570263	0.875289867	0.875162225	0.875171404	0.87500442
8	7	d5b55c93-4cb8-4ef6-8c1	2020-6-24	myTriad20_rept_acc_007	503E4BFC4E923999	SCIO		0.87921785	0.878925781	0.878588441	0.8783872	0.87834647	0.87842929	0.87845739
9	8	7a6ac477-d291-4d07-af	2020-6-24	myTriad20_rept_acc_008	503E4BFC4E923999	SCIO		0.890746588	0.890515672	0.89016542	0.88990562	0.889783304	0.889782826	0.889792154
10	9	7a6ac477-d291-4d07-af	2020-6-24	myTriad20_rept_acc_009	503E4BFC4E923999	SCIO		0.850444238	0.85032422	0.850094039	0.8499495	0.849942163	0.850175036	

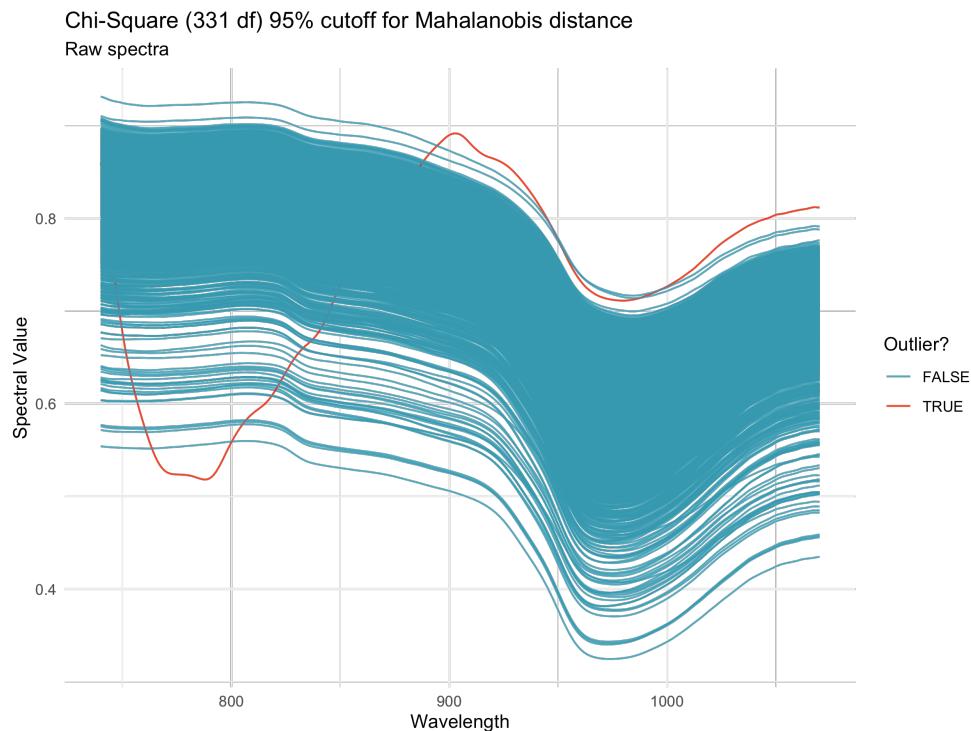
24.2 Evaluate and Remove Outliers

Spectral calibration models can be heavily affected by the presence of outliers, whether they come from spectrometer spectral artifacts or user errors. Mahalanobis distance (Mahalanobis, 1936) is a measure of the distance between a single observation and a larger distribution and is commonly used in the identification of outliers in a multivariate space (Des Maesschalck et al, 2000). The `FilterSpectra()` function in the R package `waves` calculates the Mahalanobis distance of each observation in a given spectral matrix using the `stats:::mahalanobis()` function. Observations are identified as outliers if the squared distance is greater than the 95th percentile of a χ^2 -distribution with p degrees of freedom, where p is the number of columns (wavelengths) in the spectral matrix (Johnson and Wichern, 2007). In Breedbase, this procedure is applied on a per-dataset basis on upload and outliers are given binary tags “Outlier.”

24.3 Plot Spectra

After outlier identification, a plot is generated using the `PlotSpectra()` function in `waves`. This function uses the filtered spectra and `ggplot2::ggplot()` to

create a line plot with outliers highlighted by color. A list of rows identified as outliers are shown beneath the plot. Plots are saved as .png files and linked to the original input datasets. Plot image files can be downloaded via the “Download Plot” button in the upload workflow.



24.4 Aggregate Spectra

To obtain a stable and reliable spectral profile, most spectrometer manufacturers recommend that multiple spectral scans are captured for each sample. While some spectrometers aggregate these scans internally, many do not, requiring the user to do so before analysis can take place. Breedbase handles these cases upon data upload following filtering steps by calling the *AggregateSpectra()* function from *waves*, saving the aggregated scans for future access through the search wizard feature. Scans are aggregated by sample mean (e.g. plot-level basis) according to the provided observationUnitName field. After aggregation, the user exits the upload workflow and the raw data file is saved in the upload archive.

24.5 References

- De Maesschalck, R., Jouan-Rimbaud, D., and Massart, D. L. (2000). The Mahalanobis distance. Chemom. Intell. Lab. Syst. 50(1): 1-18.
- Johnson, R. A. & Wichern, D. W. (2007). Applied Multivariate Statistical Analysis (6th Edition). p 773.
- Mahalanobis, P. C. (1936). On the generalized distance in statistics. National Institute of Science of India.

Analysis tool documentation

Chapter 25

Managing Sequence Metadata

Manage Sequence Metadata

Sequence Metadata

Upload Sequence Metadata Search Sequence Metadata

Sequence Metadata Protocols View and query existing sequence metadata

GWAS Results Report of quantitative trait loci (QTLs) identified by running rrBLUP analysis on phenotype trials and genotype trials within the T3 database.

Show 10 entries Search:

Protocol Name	Description	Properties																				
Akhunov eQTL Analysis	eQTL analysis performed by the Akhunov lab.	<p>Data Type: GWAS Results Reference Genome: RefSeq_v1 Score: effect size Attributes:</p> <table border="1"><thead><tr><th>Key</th><th>Description</th></tr></thead><tbody><tr><td>effect</td><td>effect size</td></tr><tr><td>r2</td><td>coefficient of determination</td></tr><tr><td>gene</td><td>gene name</td></tr><tr><td>t</td><td>t-statistic</td></tr><tr><td>p</td><td>p-value</td></tr><tr><td>fdr</td><td>false discovery rate</td></tr><tr><td>tissue</td><td>tissue sampled, either 'seedling' or 'spike'</td></tr></tbody></table> <p>Links:</p> <table border="1"><thead><tr><th>Title</th><th>URL Template</th></tr></thead><tbody><tr><td>JBrowse - eQTL SNP</td><td>https://graingenomes.org/jb/?data=/ggds/whe-lwgsc2018&loc=chr{{feature}}:{{start}}..{{end}}&tracks=eQTL-annot,eQTL-seedling,eQTL-spike</td></tr></tbody></table>	Key	Description	effect	effect size	r2	coefficient of determination	gene	gene name	t	t-statistic	p	p-value	fdr	false discovery rate	tissue	tissue sampled, either 'seedling' or 'spike'	Title	URL Template	JBrowse - eQTL SNP	https://graingenomes.org/jb/?data=/ggds/whe-lwgsc2018&loc=chr{{feature}}:{{start}}..{{end}}&tracks=eQTL-annot,eQTL-seedling,eQTL-spike
Key	Description																					
effect	effect size																					
r2	coefficient of determination																					
gene	gene name																					
t	t-statistic																					
p	p-value																					
fdr	false discovery rate																					
tissue	tissue sampled, either 'seedling' or 'spike'																					
Title	URL Template																					
JBrowse - eQTL SNP	https://graingenomes.org/jb/?data=/ggds/whe-lwgsc2018&loc=chr{{feature}}:{{start}}..{{end}}&tracks=eQTL-annot,eQTL-seedling,eQTL-spike																					

25.1 What is Sequence Metadata?

Sequence Metadata is a feature that allows for the efficient storage and retrieval of sequence annotations for a specific region along a reference genome. The annotation data can contain a primary “score” value and any number of secondary key/value attribute data. For example, Sequence Metatadata can store MNase open chromatin scores for every 10 basepairs along the reference genome as well as genome-wide association study (GWAS) statistics, including the trait information associated with the result. This data can then be filtered by position and/or scores/attribute values and even cross-referenced with markers stored in the database.

25.2 Loading Sequence Metadata

Sequence Metadata can be loaded into the database using a gff3-formatted file. The following columns are used to load the data:

- **#1 / seqid:** The name of the database feature (ie chromosome) the metadata is associated with (The feature name must already exist as a feature in the database)
- **#4 / start:** The metadata’s start position
- **#5 / end:** The metadata’s end position
- **#6 / score:** (optional) The primary score attribute of the metadata
- **#9 / attributes:** (optional) Secondary key//value attributes to be saved with the score. These should be formatted using the gff3 standard (key1=value1;key2=value2). The attribute key cannot be either score, start, or end.

To upload the gff3 file:

1. Go to the **Manage > Sequence Metadata** page
2. Click the **Upload Sequence Metadata** button
3. On Step 2 of the Wizard, select the Type of data to be uploaded
 - This groups similar datasets together in the same Data Type category
4. On Step 3 of the Wizard, select an existing Protocol or create a new one
 - The Protocol is used to describe how the data was generated and define the score value and any secondary attributes. Adding the

attributes (and their descriptions) to the Protocol will allow the Sequence Metadata queries to filter the data based on the value of one or more of these attributes. Attributes not defined in the Protocol will still be stored and displayed on retrieval, but will not be able to be used in a search filter.

5. Finally, select and upload your gff3 file to the database. The database will verify the format of the file before its contents are stored.

25.3 Searching Sequence Metadata

To retrieve stored Sequence Metadata, go to the **Search > Sequence Metadata** page.

25.3.1 Basic Search

The basic Sequence Metadata search options include selecting the reference genome and species, the chromosome, and (optionally) the start and/or end position(s) along the reference genome. In addition, one or more specific protocols can be selected to limit the results.

Search Sequence Metadata

Filter the sequence metadata by position, sequence metadata type and/or protocol, and/or by protocol attribute value(s).

Query Range

Reference Genome:	RefSeq_v1 (Triticum aestivum)
Feature:	1A
Start:	1200000
	End: 1300000

Protocol

Protocol:	Gene Annotation IWGSC Assembly Variant Effect Predictor GWAS Results Akhunov eQTL Analysis T3 Automated GWAS MNase MNase Open Chromatin
-----------	---

Advanced Search

Filter by attribute values

Search

The Sequence Metadata search results are returned as a table, including the chromosome and start/stop positions of the annotation, along with the primary score value and any additional key/value attributes. The markers column will include a list of marker names of any stored markers that are found within the start/stop positions of the Sequence Metadata. The data can be downloaded as a table in an Excel or CSV file or a machine-readable (code-friendly) JSON file. If the Sequence Metadata JBrowse configuration is set, the filtered results can be displayed as a dynamic JBrowse track.

Protocol	Feature	Start	End	Score	Attributes	External Links	Markers
T3 Automated GWAS	1A	1207522	1207522	0.0454952857260828	ID: RAC875_c20883_801 Locus: TraeCS1A02G002300 Population: TCAP90K_SpringAM_panel x SW-AMPanel_2012_Saskatoon Trait: SDS sedimentation Variables: CO_321:0001138 pvalue: 0.00080725065468678 qvalue: 0.0454952857260828 zvalue: 3.350296620481	EnsemblPlants - Gen Summary GrainGenes - Probe Report JBrowse - Gene Annotations, Variants, and GWAS Knetminer - Gene Network	1 marker found: 1A @ 1207522 (T/C) • RAC875_c20883_801 (Infinium 90K)

25.3.2 Advanced Search

Any number of advanced search filters can be applied to the query. The advanced filters can limit the search results by the value of the primary score and/or any of the secondary attribute values.

The screenshot shows the 'Advanced Search' interface. At the top, there is a note: 'Return only sequence metadata features that have attribute values that match the added comparisons. If more than one attribute filter is added, the sequence metadata feature must match all of the filters.' Below this, there are two main sections for defining filters:

- Score:** A dropdown menu set to 'Protocol' with 'T3 Automated GWAS' selected, a dropdown menu for 'Comparison' set to 'Greater Than or Equal', and a text input field for 'Value'. A blue 'Add' button is to the right.
- Attribute:** A dropdown menu for 'Protocol' set to 'T3 Automated GWAS', a dropdown menu for 'Key' set to 'Trait', a dropdown menu for 'Comparison' set to 'Equal', and a text input field for 'Value'. A blue 'Add' button is to the right.

Below these sections is a table titled 'Attribute Filters' containing the following data:

Protocol	Attribute	Comparison	Value
T3 Automated GWAS	score	Greater Than or Equal	0.04 X
T3 Automated GWAS	Trait	Equal	grain yield X

25.4 Marker Integration

A table of Sequence Metadata annotations are embedded on the Marker/Variant detail page. The table will include any annotations that span the position of the marker (for data of the same reference genome and species).

25.5 Sequence Metadata API

A publicly accessible RESTful API (Application Programming Interface) is available to query the database for Sequence Metadata directly from your programming environment (R, python, etc) to be used in analysis. The data is returned in a JSON format. Documentation for the API can be found on the **Manage > Sequence Metadata** page

Chapter 26

Managing Outliers in Dataset

26.1 What is Outliers Functionality in Dataset ?



As in step [The Search Wizard](#) we can create a dataset.

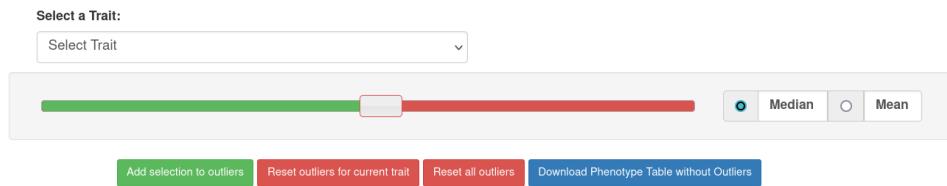
The dataset incorporates a feature to identify outlier points, which we may choose to exclude from a specific dataset. It's important to note that these exclusions only apply at the dataset level, and no data is permanently removed from the database. Additionally, outlier categorization can be modified at any time, and these changes are visible to all other functionalities within the system.

Each dataset stores a wholly unique set of outlier points, completely independent of any other dataset in the database. Outliers are specifically designated for traits within datasets, exclusively encompassing phenotype data. If a particular dataset lacks traits as a part of wizard selection, this functionality is not available.

Each trait has its own set of defined outliers.

26.2 Accessing Trait Visualization

Once you've selected a specific trait, the web application provides access to a visualization of the data points associated with that trait.



26.3 Interpreting Visual Elements

Once you've selected a specific trait, the web application provides access to a visualization of the data points associated with that trait.

- **Green Points:** As per the legend, represent values for the selected trait that fall below the cut-off point set by the slider. (non-outliers)
- **Black Outlined Points:** These data points are outlined with black borders, indicating that they are currently designated as outliers in the database.
- **Red Points:** The red data points denote the cut-off points established by the slider for the allowable deviation value.



26.4 Choosing Cut-Off Values

You have two fundamental options for setting cut-off points:

- **Median with MAD:** This option involves using the median (middle value) along with the Mean Absolute Deviation (MAD) as a reference point for determining cut-off values.
- **Mean with Standard Deviation:** Alternatively, you can choose to use the mean (average) in conjunction with the Standard Deviation to set cut-off points.

26.5 Setting Deviation Multiplier

The slider allows you to specify the deviation multiplier from a central point, which influences the cut-off values.

26.6 Utilizing Graph Controls

Beneath the graph, you'll find four buttons, each serving a distinct function:

- **Add selection to outliers:** This button enables you to save the current cut-off points to the database for future reference.
- **Reset outliers for current trait:** You can use this option to reset outliers for the selected trait.
- **Reset all outliers:** This button allows you to reset outliers for the entire dataset.
- **Download Phenotype Table without outliers:** You can download the phenotype data table in a comma-separated value format file, using this feature, with outliers excluded for selected dataset.



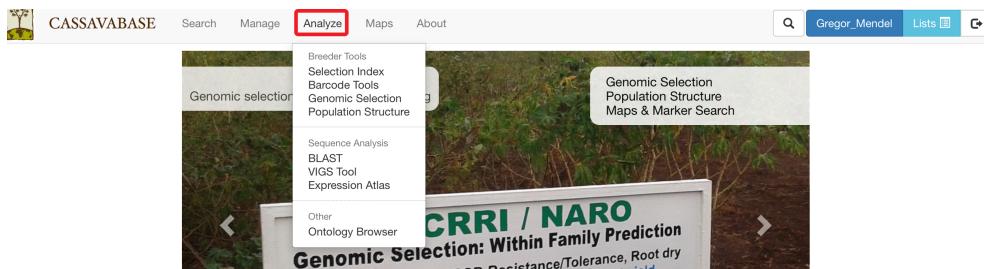
Add selection to outliers Reset outliers for current trait Reset all outliers Download Phenotype Table without Outliers

These tools and functions are designed to provide you with control and insights when working with data visualization and outliers.

Chapter 27

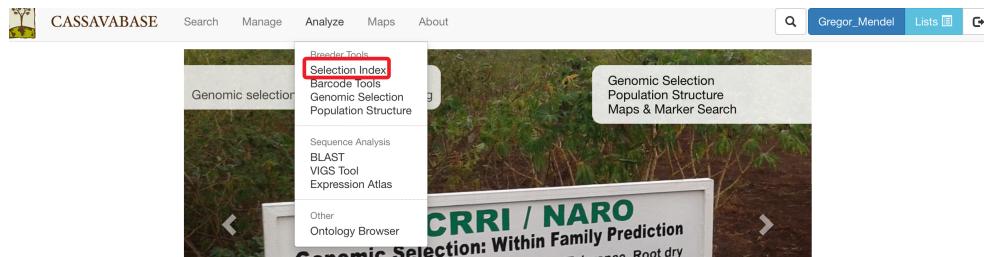
Data Analysis Tools

SGN databases provides several tools for phenotype data analysis, marker-assisted selection, sequence and expression analyses, as well as ontology browser. These tools can be found in the “Analyze” menu.



27.1 Selection Index

To determine rankings of accessions based on more than one desirable trait, SGN databases provide a “Selection Index” tool that allows you to specify a weighting on each trait. To access the tool, clicking on “Selection Index” in the “Analyze” menu.



On the Selection Index page, selecting a trial that you want to analyze.

Build a Selection Index

Parameters				
Trial select:	Trait select:			
<input type="text" value="Please select a trial"/>				
Traits and weights:				
Trait name	Trait CO id	Trait synonym	Weight	Remove?

After you selected a trial, you can find traits that were assayed in that trial in the “Trait” box.

Build a Selection Index

Parameters				
Trial select:	Trait select:			
<input type="text" value="06uyt25Ncmdlk"/>	<input checked="" type="checkbox"/> Select a trait boiled tuberous root color visual 1-3 cassava bacterial blight severity 3-month evaluation cassava bacterial blight severity 9-month evaluation cassava mosaic disease incidence 1-month evaluation cassava mosaic disease incidence 3-month evaluation cassava mosaic disease severity 1-month evaluation cassava mosaic disease severity 3-month evaluation dry matter content percentage dry yield ease of peeling root cortex visual rating 1-7 fibre content estimation in percentage fresh root weight fresh root yield fresh shoot weight measurement in kg harvest index variable initial vigor assessment 1-7 plant stands harvested counting poundability assessment 0-4 root neck length visual rating 0-7 root number counting			
Traits and weights:				
Trait name	Trait CO id	Trait synonym	Weight	Remove?
Additional options:				
<input type="checkbox"/> Include accessions with missing phenotypes <input type="checkbox"/> Scale values to a reference accession Select a reference accession				
Rankings				
Selection Index Raw Averages				

Selecting a trait that you want to include in the analysis will open a new dialogue showing the selected trait and a box that you can assign a “Weight” of that trait. After you are done, you can continue by selecting another trait by clicking on “Add another trait” link.

Build a Selection Index

Parameters

Trial select: 06uyt25Ncmdlk

Traits and weights:				
Trait name	Trait CO id	Trait synonym	Weight	Remove?
fresh root weight	CO:0000012	RtWt_Wgh_kg	Must be a number (+ or -, d)	X

Add another trait

Additional options:

- Include accessions with missing phenotypes
- Scale values to a reference accession:

SIN formula:
SIN = 1 * (fresh root weight)

Select a reference accession

Calculate Rankings

After you selected another trait, this page will automatically update information for you by showing all of the traits that you selected for the analysis.

Build a Selection Index

Parameters

Trial select: 06uyt25Ncmdlk

Traits and weights:				
Trait name	Trait CO id	Trait synonym	Weight	Remove?
fresh root weight	CO:0000012	RtWt_Wgh_kg	7	X
initial vigor assessment 1-7	CO:0000009	IVig_IITAVisScg_1to7	3	X

Add another trait

Additional options:

- Include accessions with missing phenotypes
- Scale values to a reference accession:

SIN formula:
SIN = 7 * (fresh root weight) + 3 * (initial vigor assessment 1-7)

Select a reference accession

Calculate Rankings

You also have options to choose a reference accession, choose to include accessions with missing phenotypes, scaling values to a reference accession. After you complete your setting, clicking on “Calculate Rankings”

Build a Selection Index

Parameters				
Trial select: 06uyt25Ncmdlk				
Traits and weights:				
Trait name	Trait CO id	Trait synonym	Weight	Remove?
fresh root weight	CO:0000012	RtWt_Wgh_kg	7	X
initial vigor assessment 1-7	CO:0000009	IV(g_IITAVisScg_1to7)	3	X
Add another trait				
<input type="checkbox"/> Include accessions with missing phenotypes <input type="checkbox"/> Scale values to a reference accession: Select a reference accession			SIN formula: $SIN = 7 * (\text{fresh root weight}) + 3 * (\text{initial vigor assessment 1-7})$ Calculate Rankings	

The Selection Index tool will generate rankings of accessions based on the information that you specified. You can copy the results to your system clipboard, convert the table data to CSV format, or print the data.

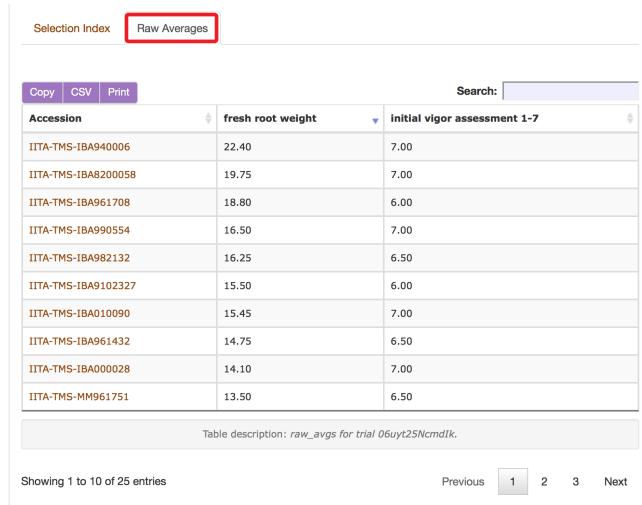
Rankings

Selection Index				
Raw Averages				
Copy CSV Print <input type="text" value="Search:"/> Search				
Accession	7 * (fresh root weight)	3 * (initial vigor assessment 1-7)	SIN	SIN Rank
IITA-TMS-IBA940006	156.8	21	177.80	1
IITA-TMS-IBA8200058	138.25	21	159.25	2
IITA-TMS-IBA961708	131.6	18	149.60	3
IITA-TMS-IBA990554	115.5	21	136.50	4
IITA-TMS-IBA982132	113.75	19.5	133.25	5
IITA-TMS-IBA010090	108.15	21	129.15	6
IITA-TMS-IBA9102327	108.5	18	126.50	7
IITA-TMS-IBA961432	103.25	19.5	122.75	8
IITA-TMS-IBA000028	98.7	21	119.70	9
IITA-TMS-MM961751	94.5	19.5	114.00	10

Table description: weighted_values for trial 06uyt25Ncmdlk.

Showing 1 to 10 of 25 entries Previous [1](#) [2](#) [3](#) Next

Clicking on “Raw Average” will display average values of the phenotypes of those ranked accessions.



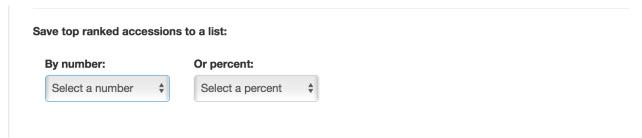
Accession	fresh root weight	initial vigor assessment 1-7
IITA-TMS-IBA940006	22.40	7.00
IITA-TMS-IBA820008	19.75	7.00
IITA-TMS-IBA961708	18.80	6.00
IITA-TMS-IBA990554	16.50	7.00
IITA-TMS-IBA982132	16.25	6.50
IITA-TMS-IBA9102327	15.50	6.00
IITA-TMS-IBA010090	15.45	7.00
IITA-TMS-IBA961432	14.75	6.50
IITA-TMS-IBA000028	14.10	7.00
IITA-TMS-MM961751	13.50	6.50

Table description: raw_avgs for trial 06uyt25NcmdIk.

Showing 1 to 10 of 25 entries

Previous 1 2 3 Next

Selection Index tool also allows you to save top ranked accessions directly to “Lists”. You can retrieve top ranked accessions by selecting a number or a percent.



Save top ranked accessions to a list:

By number:

Or percent:

27.2 Genomic Selection

The prediction of breeding values for a trait is a one step or two steps process, depending on what stage in your breeding cycle you are. The first step is to build a prediction model for a trait using a training population of clones with phenotype and genotype data. If you have yet to select parents for crossing for your first cycle of selection you can use the breeding values of the training population. If you are at later stages of your selection program, you need to do the second step which is applying the prediction model on your selection population. All clones in your training and selection populations must exist in the database.

To use the genomic selection tool, on cassavabase.org, select “Genomic Selection” from the “analyze” pull-down menu.



The screenshot shows the CASSAVABASE homepage with a navigation bar at the top. The navigation bar includes links for home, forum, contact, help, and wiki. Below the navigation bar, there are tabs for search, manage, analyze, maps, and a user account section with links for Isaak Tecle, lists, my account, and log out. A search bar with the placeholder "Search for a trait" and a "sol search" button are also present. The main content area displays the text "solGS: start building a GS model by searching for a trait or selecting a training population" followed by three options: "Search for a trait", "Use a trial as a training population", and "Create a training population".

There are three ways to build a model for a trait.

27.2.1 Building a Model - Method 1:

One way to build a model is, using a trait name, to search for trials in which the trait was phenotyped and use a trial or a combination of trials to build a model for the trait. For example, if you search for “mosaic disease severity”, you will get a list of trials you can use as training populations.



The screenshot shows the CASSAVABASE homepage with a search bar containing the trait name "mosaic". Below the search bar, a list of results is displayed, including "cassava mosaic disease incidence" and "cassava mosaic disease severity". At the bottom of the page, there are two buttons: "Use a trial as a training population" and "Create a training population".

You will get a list of trials (as shown below) in which the trait of your inter-

ested was phenotyped. From the list, you can use a single trial as a training population or combine several trials to form a training population for the prediction model of the trait. Let's say, you want to create a training population using individuals from trials "cassava ibadan 2001/02" and "cassava ibadan 02/03" and build a model for "cassava mosaic disease severity" using all clones from the training population.

GS populations evaluated for cassava mosaic disease severity

Select a training population or create a new one using one or more trials

Trial	Description	Location	Year	Tip(?)
<input checked="" type="checkbox"/> Cassava Ibadan 2002/03	Plants assayed at Ibadan in 2002/03	Ibadan	2002/03	
<input checked="" type="checkbox"/> Cassava Ibadan 2001/02	Plants assayed at Ibadan in 2001/02	Ibadan	2001/02	
<input type="checkbox"/> AYT 2011-2012	AYT 2011-2012 Trial NR09	Umudike	2011	
<input type="checkbox"/> Cassava Ibadan 2003/04	Plants assayed at Ibadan in 2003/04	Ibadan	2003/04	
<input type="checkbox"/> Cassava Ibadan 2004/05	Plants assayed at Ibadan in 2004/05	Ibadan	2004/05	
<input type="checkbox"/> Cassava Igbariam 2009	Plants assayed at Igbariam in 2009	Igbariam	2009	
<input type="checkbox"/> Cassava Ibadan 2005/06	Plants assayed at Ibadan in 2005/06	Ibadan	2005/06	
<input type="checkbox"/> Cassava Ibadan 2000/01	Plants assayed at Ibadan in 2000/01	Ibadan	2000/01	
<input type="checkbox"/> Cassava Ibadan 1999/00	Plants assayed at Ibadan in 1999/00	Ibadan	1999/00	
<input type="checkbox"/> Cassava Ibadan 2006/07	Plants assayed at Ibadan in 2006/07	Ibadan	2006/07	

1 2 3 4 5 >

Done selecting

Trials to combine

Trial	Description	Location	Year	
<input checked="" type="checkbox"/> Cassava Ibadan 2002/03	Plants assayed at Ibadan in 2002/03	Ibadan	2002/03	
<input checked="" type="checkbox"/> Cassava Ibadan 2001/02	Plants assayed at Ibadan in 2001/02	Ibadan	2001/02	

Combine trials & build model

Select the trials to combine (the same coloured), click 'done selecting', click the "combine trials and build model" button, and you will get a model and its output for the trait. On the model detail page, you can view the description of input data used in the model, output from the model and search interface for selection populations the model you can apply to predict their breeding values. The description of the input data for the model includes the number of phenotyped clones, and the number of markers, scatter and frequency distribution plots for the phenotype data, relationship between the phenotype

data and GEBVs, population structure. The model output includes model parameters, heritability of the trait , prediction accuracy, GEBVs of the individuals from the training population and marker effects.

**Genomic selection model for Cassava mosaic disease severity (CMDS) in Training population
2907368219**

(+) **Training population summary**

Name	Training population 2907368219	No. of lines	239
Description	This training population is a combination of Cassava Ibadan 2001/02 and Cassava Ibadan 2002/03 .	No. of markers	97337
Owner	Peter Kulakow	Genotyping version	GBS ApeKI Cassava genome v5

(+) **Trait phenotype data**

(+) **Model accuracy**

(+) **Population structure analysis - PCA**

(+) **Model Parameters**

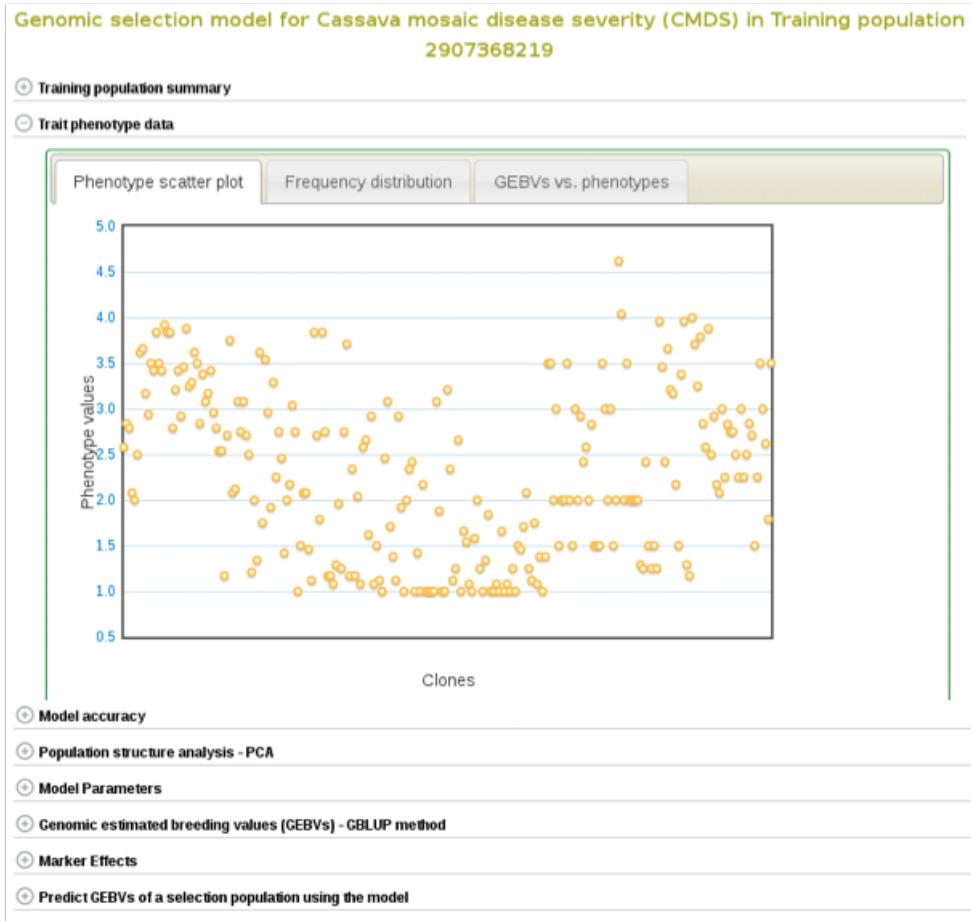
(+) **Genomic estimated breeding values (GEBVs) - GBLUP method**

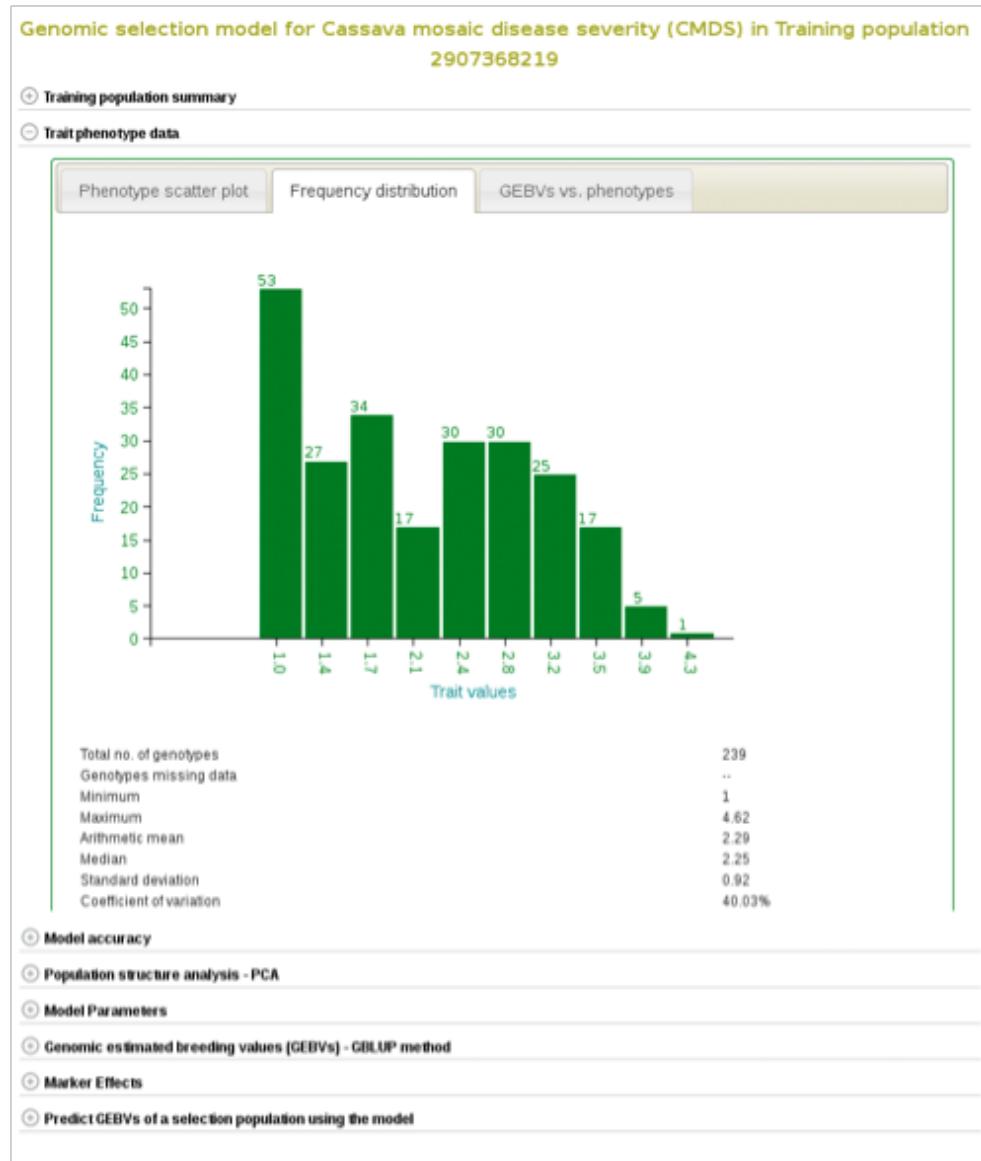
(+) **Marker Effects**

(+) **Predict GEBVs of a selection population using the model**

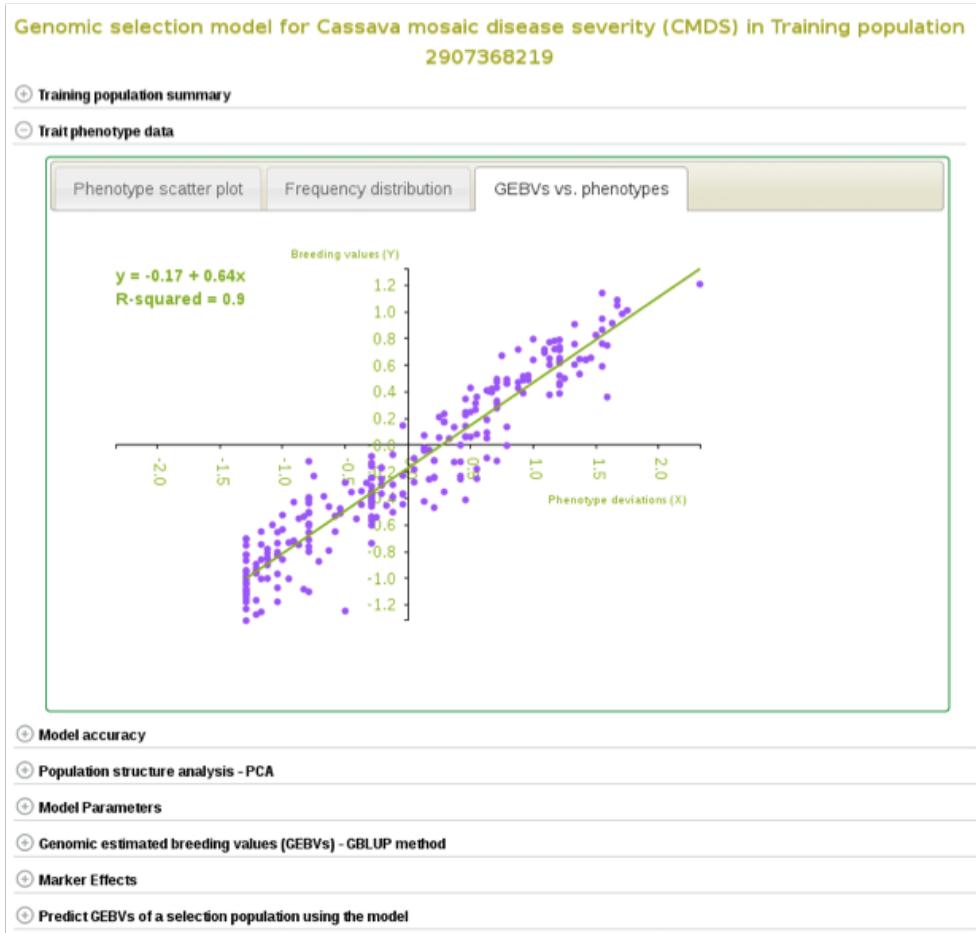
Expand each section to see detailed information.

If you expand the ‘Trait phenotype data’ section, you will find plots to explore the phenotype data used in the model. You can assess the phenotype data using a scatter and histogram plots and the descriptive statistics.

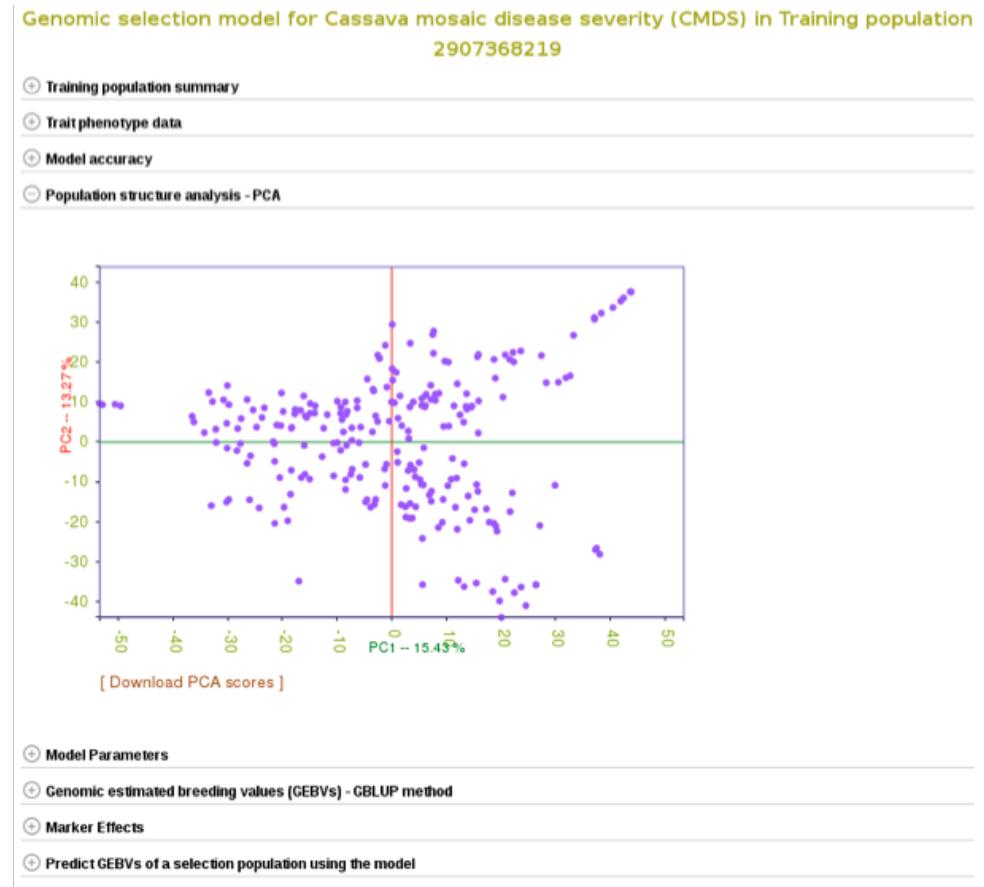




A regression line between observed phenotypes and GEBVs shows the relationship between the two.



You can also explore if there is any sub-clustering in the training population using PCA.



To check the model accuracy, a 10-fold cross-validation test, expand the ‘model accuracy’ section.

**Genomic selection model for Cassava mosaic disease severity (CMDS) in Training population
2907368219**

- [④ Training population summary](#)
- [④ Trait phenotype data](#)
- [④ Model accuracy](#)

Runs	Accuracy r)
Validation test 6	0.648
Validation test 3	0.612
Validation test 9	0.571
Validation test 4	0.556
Validation test 7	0.555
Validation test 5	0.478
Validation test 8	0.444
Validation test 2	0.422
Validation test 10	0.417
Validation test 1	0.335
Average	0.5

[Download model accuracy report]
- [④ Population structure analysis - PCA](#)
- [④ Model Parameters](#)
- [④ Genomic estimated breeding values \(GEBVs\) - GBLUP method](#)
- [④ Marker Effects](#)
- [④ Predict GEBVs of a selection population using the model](#)

Marker effects are also available for download. To do so, expand the ‘Marker Effects’ section and click the ‘Download all marker effects’ link and you will get a tab delimited output to save on your computer.

Genomic selection model for Cassava mosaic disease severity (CMDS) in Training population
2907368219

- [Training population summary](#)
- [Trait phenotype data](#)
- [Model accuracy](#)
- [Population structure analysis - PCA](#)
- [Model Parameters](#)
- [Genomic estimated breeding values \(GEBVs\) - GBLUP method](#)
- [Marker Effects](#)

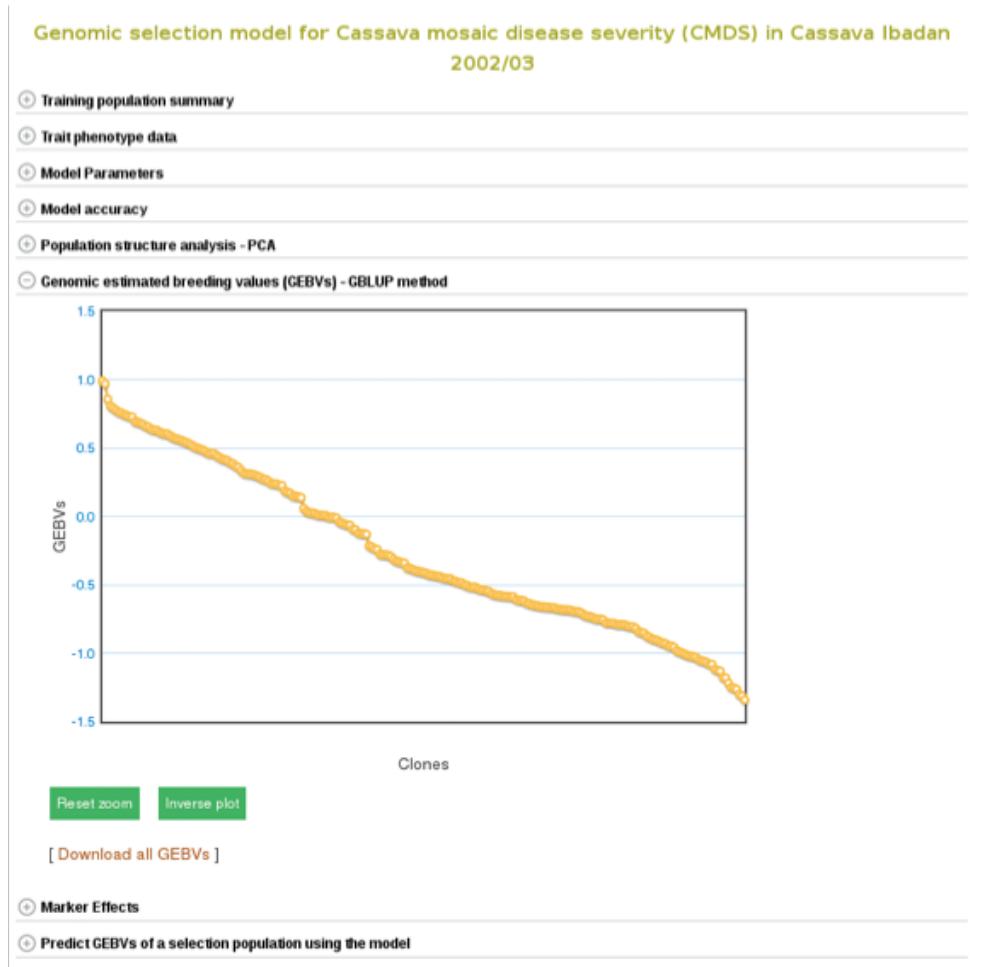
Top 10 markers:

Marker	Effects
S1_317473	0.00308
S9_16963656	0.00298
S14_18164196	0.00295
S19_117475272	0.00281
S19_113674348	0.00274
S8_472214	0.00256
S3_6218223	0.00256
S8_7605425	0.00253
S4_15467245	0.00249
S4_15467248	0.00249

[[Download all marker effects](#)]

- [Predict GEBVs of a selection population using the model](#)

The breeding values of the individuals used in the training population are displayed graphically. Mousing over each data point displays the clone and its breeding value. To examine better, you can zoom in into the plot by selecting an area on the plot. You can download them also by following the “Download all GEBVs” link.



Estimating breeding values in a selection population

If you already have a selection population (in the database), from the same model page, you can apply the model to the selection population and estimate breeding values for all the clones in the population. You can search for a selection population of clones in the database using the search interface or you can make a custom list of clones using the *list interface*. If you click the “search for all relevant selection populations”, you will see all relevant selection populations for that model. However, this option takes long time because of the large set of populations in the database and the filtering. Therefore, the fastest way is to search for each of your selection populations by name. If you are logged in to the website you will also see a list of your

custom set of genotyped clones.

Genomic selection model for Cassava mosaic disease severity (CMDS) in Training population
2907368219

- Training population summary**
- Trait phenotype data**
- Model accuracy**
- Population structure analysis - PCA**
- Model Parameters**
- Genomic estimated breeding values (GEBVs) - GBLUP method**
- Marker Effects**
- Predict GEBVs of a selection population using the model**

Go

Search for all relevant selection populations

Selection population	Description	Year	Predict GEBVs
Cassava Ibadan 2005/06	Plants assayed at Ibadan in 2005/06	2005	CMDS
Cassava Ibadan 2003/04	Plants assayed at Ibadan in 2003/04	2003	CMDS
Cassava Ibadan 2006/07	Plants assayed at Ibadan in 2006/07	2006	[Predict]

List-based selection population
Selection candidates list 2015
Go
+ Make a new list of clones

Predict GEBVs
[Predict]

To apply the model to a selection population, simply click your population name or “Predict Now” and you will get the predicted breeding values. When you see a name of (or acronym) of the trait, follow the link and you will see an interactive plot of the breeding values and a link to download the breeding values of your selection population.



27.2.2 Building a Model - Method 2

Another way to build a model is by selecting a trial, instead of selecting and searching for a specific trait. This approach is useful when you know a particular trial that is relevant to the environment you are targeting to breed material for. This method allows you to build models and predict genomic estimated breeding values (GEBVs) for several traits within a single trial at once. You can also calculate selection index for your clones when GEBVs are

estimated for multiple traits.

To do this select the “Genomic Selection” link found under the “analyze” menu. This will take you to the same home page as used with Method 1. However, instead of entering information to search for in “Search for a trait”, click on “Use a trait as a trial population”. This will expand a new menu that will show all available trials.

The screenshot shows the CASSAVABASE website interface. At the top, there is a navigation bar with links for home, forum, contact, help, and wiki. Below the navigation bar, there are links for search, manage, analyze, maps, and user accounts (Liana Acevedo, lists, my account, log out). A search bar with the placeholder "Search for a trait" is present. Below the search bar, a traits index is shown with links for B, C, D, E, F, H, I, L, P, R, S, and T. A search input field and a green "Search" button are located below the traits index. Further down, two options are listed: "Use a trial as a training population" and "Create a training population". A large red arrow points downwards from the "Use a trial as a training population" option towards the bottom of the page.

solGS: start building a GS model by searching for a trait or selecting a training population

Select a training population or create a new one using one or more trials

Trial	Description	Location	Year	Tip(?)
<input checked="" type="checkbox"/> Cassava Ibadan 2002/03	Plants assayed at Ibadan in 2002/03	Ibadan	2002/03	
<input type="checkbox"/> Cassava Ibadan 2001/02	Plants assayed at Ibadan in 2001/02	Ibadan	2001/02	
<input type="checkbox"/> AYT 2011-2012	AYT 2011-2012 Trial NR09	Umudike	2011	
<input type="checkbox"/> Cassava Ibadan 2003/04	Plants assayed at Ibadan in 2003/04	Ibadan	2003/04	
<input type="checkbox"/> Cassava Ibadan 2004/05	Plants assayed at Ibadan in 2004/05	Ibadan	2004/05	
<input type="checkbox"/> Cassava Igbariam 2009	Plants assayed at Igbariam in 2009	Igbariam	2009	
<input type="checkbox"/> Cassava Ibadan 2005/06	Plants assayed at Ibadan in 2005/06	Ibadan	2005/06	
<input type="checkbox"/> Cassava Ibadan 2000/01	Plants assayed at Ibadan in 2000/01	Ibadan	2000/01	
<input type="checkbox"/> Cassava Ibadan 1999/00	Plants assayed at Ibadan in 1999/00	Ibadan	1999/00	
<input type="checkbox"/> Cassava Ibadan 2006/07	Plants assayed at Ibadan in 2006/07	Ibadan	2006/07	

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 >

Select a list-based training population or create a new one

To begin creating the model, select the existing trial that you would like to use. In this example I will be using the trial and trait data from “Cassava Ibadan 2002/03” trial. Clicking on a trial will take you to a page where you can find information such as number of markers and number of phenotypes clones.

Select one or more traits from training population "Cassava Ibadan 2002/03" to build a GS model and predict GEBVs for:

Training population summary

Name	Cassava Ibadan 2002/03	No. of lines	237
Description	Plants assayed at ibadan in 2002/03	No. of traits	20
Owner	Peter Kulakow	No. of markers	97337
		Genotyping version	GBS ApeKI Cassava genome v5

Traits

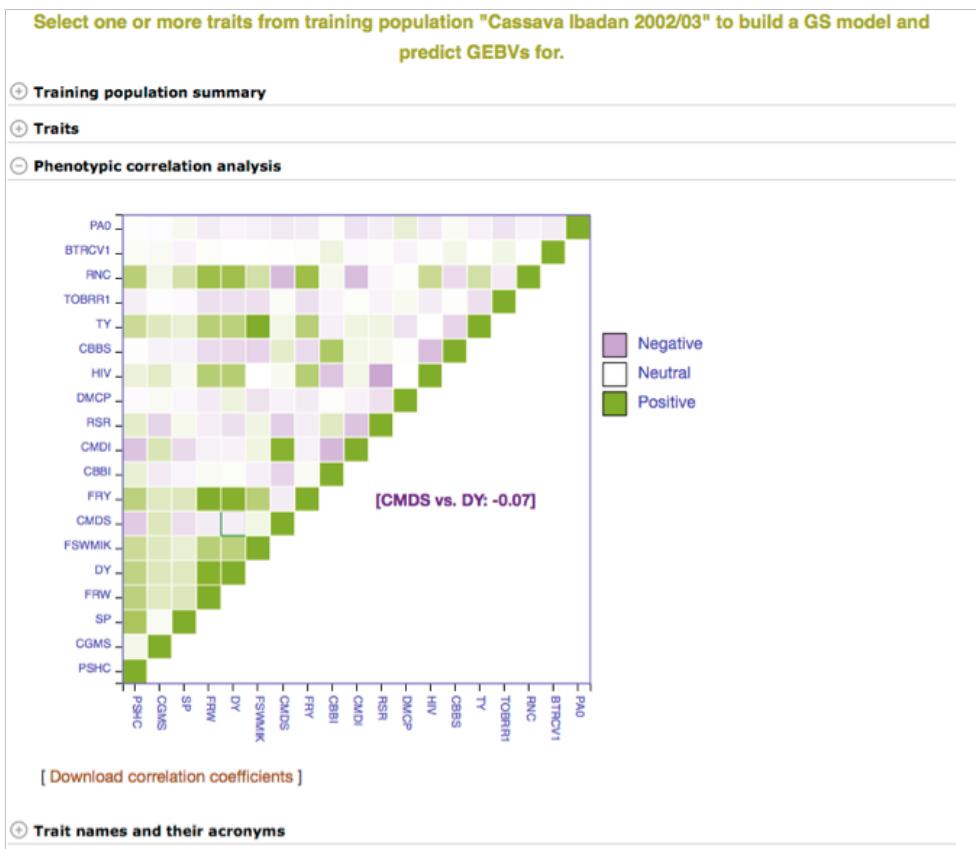
- boiled tuberous root color visual 1-3
- Cassava bacterial blight incidence
- Cassava bacterial blight severity
- Cassava green mite severity
- Cassava mosaic disease incidence
- Cassava mosaic disease severity
- dry matter content percentage
- dry yield
- top yield

Build model

Phenotypic correlation analysis

Trait names and their acronyms

In addition to the number of phenotype clones and number of markers, the main page for the trial selected also has information and graphs on phenotypic correlation for all of the traits. By moving your cursor over the graph you can read the different values for correlation between two traits. A key with all of the trait names of the acronyms used can be found in the tab below the graph.



Below the “Training population summary” there is a tab for “Traits”. Clicking on this tab will show all available traits for the specific trial. You can create a model by choosing one or multiple traits in the trial and clicking “Build Model”. In this example, the traits for “cassava bacterial blight severity” and “cassava mosaic disease severity” have been selected.

Select one or more traits from training population "Cassava Ibadan 2002/03" to build a GS model and predict GEBVs for:

Training population summary

Name	Cassava Ibadan 2002/03	No. of lines	237
Description	Plants assayed at Ibadan in 2002/03	No. of traits	20
Owner	Peter Kulakow	No. of markers	97337
		Genotyping version	GBS ApeKI Cassava genome v5

Traits

- boiled tuberous root color visual 1-3
- Cassava bacterial blight incidence
- Cassava bacterial blight severity
- Cassava green mite severity
- Cassava mosaic disease incidence
- Cassava mosaic disease severity
- dry matter content percentage
- dry yield
- top yield

Build model

Phenotypic correlation analysis

Trait names and their acronyms

Clicking on “Build Model” will take you to a new page with the models outputs for the traits. Under the “Genomic Selection Model Output” tab you can view the model output and the model accuracy. Clicking on any of the traits will take you to a page with information about the model output on that individual trait within the trial. There you can view all of the trait information that was seen in more detail in *Method 1*.

Prediction models from Cassava Ibadan 2002/03

Models summary

Training population	Description	Models						
Cassava Ibadan 2002/03	Plants assayed at Ibadan in 2002/03	<table border="1"> <thead> <tr> <th>Trait</th> <th>Model accuracy</th> </tr> </thead> <tbody> <tr> <td>DY</td> <td>0.46</td> </tr> <tr> <td>CMDS</td> <td>0.46</td> </tr> </tbody> </table>	Trait	Model accuracy	DY	0.46	CMDS	0.46
Trait	Model accuracy							
DY	0.46							
CMDS	0.46							

Predict GEBVs of a selection population using the models

Genetic correlation analysis

Calculate selection index

Trait names and their acronyms

You can apply the models to simultaneously predict GEBVs for respective traits in a selection population by clicking on “Predict Now” or the name of the selection population. You can also apply the models to any set of genotyped clones that you can create using the “lists” feature. For more information on lists, click [here](#). Follow the link to the trait name to view and download the predicted GEBVs for the trait in a selection population.

Prediction models from Cassava Ibadan 2002/03

Models summary			
Training population	Description	Models	
Cassava Ibadan 2002/03	Plants assayed at Ibadan in 2002/03	Trait	Model accuracy
		DY	0.46
		CMDS	0.46

Predict GEBVs of a selection population using the models			
search for a selection population	<input type="button" value="Go"/>	Search for all relevant selection populations	
Selection population	Description	Year	Predict GEBVs
Cassava Ibadan 2005/06	Plants assayed at Ibadan in 2005/06	2005	DY CMDS
Cassava Ibadan 2006/07	Plants assayed at Ibadan in 2006/07	2006	[Predict]

List-based selection population	Selection candidates list 2015	<input type="button" value="Go"/>	+ Make a new list of clones
List-based selection population	Selection candidates list 2015	<input type="button" value="Go"/>	Predict GEBVs [Predict]

- Genetic correlation analysis
- Calculate selection index
- Trait names and their acronyms

To compare clones based on their performance on multiple traits, you can calculate selection indices using the form below. Choose from the pulldown menu the population with predicted GEBVs for the traits and assign relative weights for each trait. The relative weight of each trait must be between 0 - 1. 0 being of least weight and importance, not wanting to consider that particular trait in selecting a genotype and 1 being a trait that you give highest importance.

In this example we will be using the “Cassava Ibadan 2002/03” population and assigning values to each of the traits. Remember that there is a list of acronyms and trait names at the bottom of the page for reference. After entering whatever values you would like for each trait click on the “Calculate”

button to generate results. This will create a list of the top 10 genotypes that most closely match the criteria that you entered. The list will be displayed right below the “selection index” tab. This information can also be downloaded onto your computer by clicking on the “Download selection indices” link underneath the listed genotypes and selection indices.

Prediction models from Cassava Ibadan 2002/03

- Models summary**
- Predict GEBVs of a selection population using the models**
- Genetic correlation analysis**
- Calculate selection index**

Cassava Ibadan 2002/03 And assign relative weights to traits.

DY: CMDS:

Calculate

Genotype ranking based on multiple traits performance [selection index]

Top 10 genotypes:

Genotypes	Selection indices
IITA-TMS-IBA974763	3.37
IITA-TMS-IBA974779	2.91
IITA-TMS-IBA930266	2.88
IITA-TMS-IBA950061	2.71
IITA-TMS-IBA940239	2.6
IITA-TMS-ZAP940153	2.49
IITA-TMS-IBA972205	2.36
IITA-TMS-IBA996016	2.11
IITA-TMS-IBA930098	1.87
IITA-TMS-ZAP930151	1.76

[Download selection indices]
Relative weights: DY: 0.6 CMDS: 0.4
Name: Cassava Ibadan 2002/03

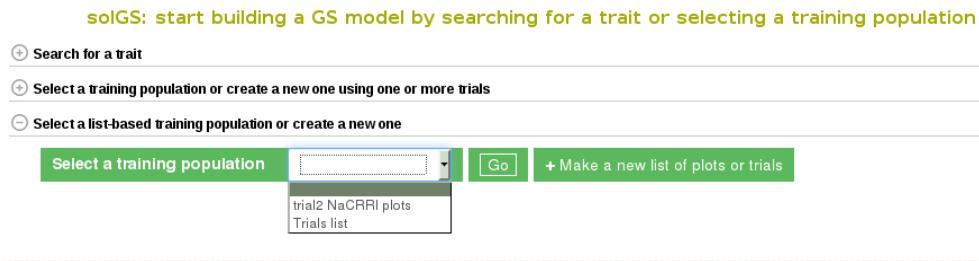
Correlation between selection index and GEBVs of traits

	DY	CMDS	Index
DY	Positive	Neutral	Negative
CMDS	Neutral	Positive	Neutral
Index	Negative	Neutral	Positive

27.2.3 Building a Model - Method 3

In addition to creating a model by searching for pre-existing traits or by preexisting trial name, models can also be created by using your own list of clones. This creates a model by using or creating a training population.

The page to use the third Method for creating a population model is the same as for the other two models. Select “Genomic Selection” from under the “analyze” menu of the main toolbar. This will take you to the Genomic Selection homepage and show you all three available methods to create a model. To see and use Method 3 scroll down and click on the tab labeled “Create a Training Population”. This will open a set of tools that will allow you to use pre-existing lists or to create a new list.



Once the “Create a Training Population” tab is opened you have the option to use a pre-existing list or create new one. To learn how to create a list, click [here](#). The “Make a new list of plots” link will take you directly to the Search Wizard that is usually used to create lists.

Please note: the only lists that can be used in Method 3 to create a model are lists of plots and trials. If the pre-existing list is not of plots or trials (for example, traits, or locations) it will not show up and cannot be used as a training population. When you create you use a list of trials, the trials data will be combined to create a training data set.

To use your custom list of plots or trials as a training population, select the list and click “Go”. This will take you to a detail page for the training population.

Select one or more traits from training population "Training population 1" to build a GS model and predict GEBVs for.

Training population summary

Name	Training population 1	No. of lines	195
Description	Uploaded on: Wed Jan 6 14:46 2016	No. of traits	26
Owner	isaaktecle	No. of markers	97337
		Genotyping version	GBS ApeKI Cassava genome v5

Traits

- Cassava anthracnose disease incidence
- Cassava anthracnose disease severity
- Cassava bacterial blight incidence
- Cassava bacterial blight severity
- Cassava green mite severity
- Cassava mosaic disease incidence
- Cassava mosaic disease severity
- dry matter content percentage
- dry yield
- top yield

Build model

Phenotypic correlation analysis

Run correlation

Trait names and their acronyms

From here on you can build models and predict breeding values as described in *Method 2*.

27.3 Genome Browsing

There are two ways to evaluate genotype information within the browser, from an accession detail page or a trial detail page.

27.3.1 Browsing Genotype data by Accession

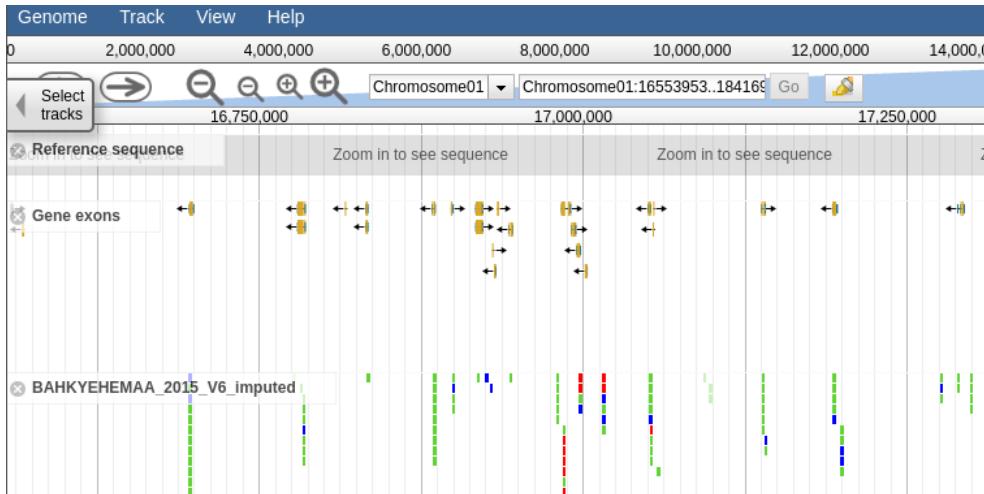
If you are interested in browsing genotype information for a single accession, for example 'BAHKEYHEMAA', navigate to the accession detail page.

Search Results	
Show	10 entries
Stock Name	Stock Type
BAHKYEHHEMAA	accession
Showing 1 to 1 of 1 entries	

Near the bottom of the detail page is a collapsible section called “Accession Jbrowse”.

+ Genotype data
- Accession JBrowse
View the tracks for this accession in JBrowse

This section will contain a link to the accession jbrowse page if the necessary genotype data is available. Clicking the link should take you to a page that looks like this, at which point you can browse the genotype data in the form of a vcf track aligned to the latest build of the genome.



27.3.2 Browsing Genotype data by Trial

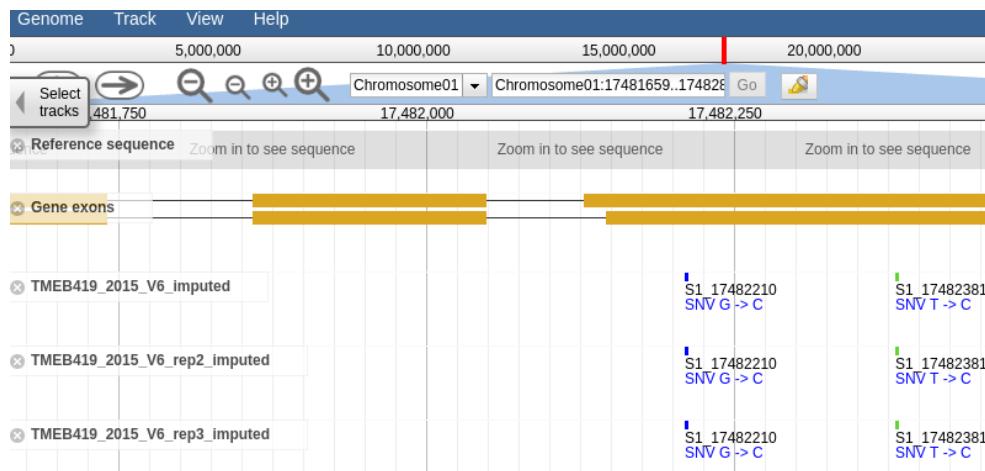
If you are interested in browsing genotype information for the accessions within a given trial, navigate to the trial detail page.

Trial Search			
Show 10 entries			
Trial name	Description	Breeding program	Folder
12ayt30whrmK	Assessment of Varieties of Cassava for high yield, high dry matter and disease resistance using Advance Yield Trial (30 clones) in Mokwa 2012/2013 Breeding Season	IITA	12_Mokwa

Halfway down the page is a collapsible section called “Trial Jbrowse”. This section will contain a link to the trial jbrowse page if the necessary genotype data for at least two accessions planted in the trial is available.

- [Compute Trait Phenotypes](#)
- [Trial JBrowse](#)
- [View the dataset for this trial in JBrowse](#)
- [Files](#)

Clicking the link should take you to a page that looks like this, at which point you can browse the genotype data in the form of vcf tracks aligned to the latest build of the genome.

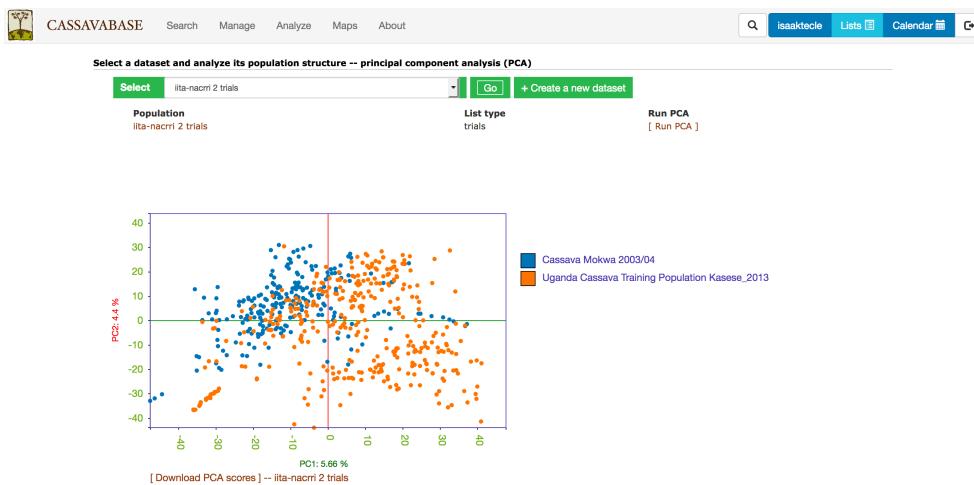


27.4 Principal Component Analysis (PCA)

Principal component analysis helps estimate and visualize if there is subgrouping of individuals within a dataset based on a number of variables. Currently, you can use marker data to run PCA on datasets.

You can run PCA from multiple places on the website. To do PCA on

- (1) individuals from a trial, go to the trial detail page and find the PCA tool under the “Analysis tools” section.
- (2) individuals from a training population you used in a GS modeling, do your modeling and find the PCA tool in the model output page.
- (3) individuals in a training population and selection population you applied the training model, do your modeling, apply the model on the selection population and find the PCA tool on the selection population prediction output page.
- (4) individuals in a list of accessions you created, for example using the search wizard, go to the “Analyze” menu and select the “Population Structure”, select your list of individuals and run PCA.
- (5) individuals from multiple trials, create a list of the trials using the search wizard, go to the “Analyze” menu and select the “Population Structure”, select your list of trials and run PCA.



With all the options, you will get a interactive plot of the two PCs (shown below) that explain the largest variance. Point the cursor at any data point and you will see the individual name with its corresponding PCs scores. By clicking the ‘Download all PCs’, you can also download the 10 PCs scores in the text format.

27.5 ANOVA

Currently, ANOVA is implemented for a single trial (single year and single location). You can do ANOVA for RCBD, CRD, Alpha and Augmented trial designs. ANOVA is done using linear mixed effects model, where the genotypes is fixed effect and the replications and blocks are random effects. Fixed effect significance level is computed using “lmer” from “lmeTest” R package.

You can do ANOVA from two places: trial detail and training population detail. In both cases, if the phenotype data was from the supported trial designs,

- Go to the ANOVA section down in the trial or training population page
- Select the trait of you want to perform ANOVA
- Click the “Run ANOVA” and wait for the result

The screenshot shows a user interface for running an ANOVA analysis. At the top, there is a header with a circular icon and the text "ANOVA". Below this, a search bar contains the text "dry matter content percentage". To the right of the search bar is a green button labeled "Run ANOVA" with a dropdown arrow icon. The main area displays the "ANOVA result: DMCP" table. The table has columns for Sum Sq, Mean Sq, NumDF, DenDF, F.value, and Pr(> F). A single row is shown for "genotypes", with values 7,266.18, 24.38, 298, 102.95, 1.69, and 0.001 respectively. At the bottom of the table, there is a link "Download: [Anova table] | [Model Summary] | [Adjusted Means] | [Model Diagnostics]".

	Sum Sq	Mean Sq	NumDF	DenDF	F.value	Pr(> F)
genotypes	7,266.18	24.38	298	102.95	1.69	0.001

Download: [\[Anova table\]](#) | [\[Model Summary\]](#) | [\[Adjusted Means\]](#) | [\[Model Diagnostics\]](#)

27.6 Clustering (K-Means, Hierarchical)

The K-Means method allows you to partition a dataset into groups (K number). The hierarchical clustering, agglomerative, allows you to explore underlying similarity and visualize in a tree structure (dendrogram) the different levels of similarities (clusters) among samples. You can do clustering based on marker data, phenotype data and GEBVs. When you use phenotype data, first clone averages for each trait are calculated. Both methods use Euclidean distance as a measure of similarity. For the hierarchical clustering, the complete-linkage (farthest neighbour) method is used to link up clusters.

There are three pathways to using this tool.

(1) When you have data in the form of a list or dataset from the search wizard:

- (A) – go to the “Analyze” menu and select the clustering option
- (B) – make sure you are logged in
- (C) – Select the relevant genotyping protocol, if you are clustering using genotype data
- (D) – select your list or dataset, click “Go”
- (E) – select clustering type
- (F) – select the data type to use
- (G) – If you are running K-Means clustering, provide the number of partitions (K). If left blank it will partition the data set into optimal numbers for the dataset.
- (H) – click the “Run Cluster” and wait for the analysis to finish or queue the request and wait for an email with the analysis result.
- (I) – You can download the outputs following the download links.

(2) From the trial detail page:

- (A) – Go to the “Analysis Tools” section
- (B) – Follow steps D to G in (1)

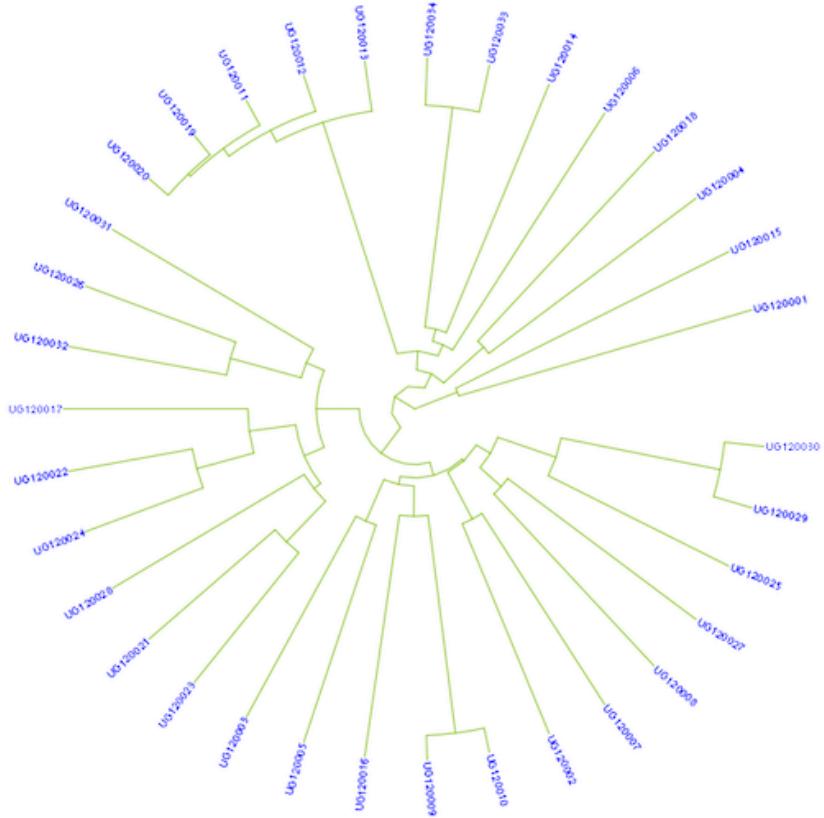
(3) In the solGS pipeline:

- (A) – Once you are in a model output put page, you will see a section where you can do clustering in the same way as above (option 2).

K-Means clustering:



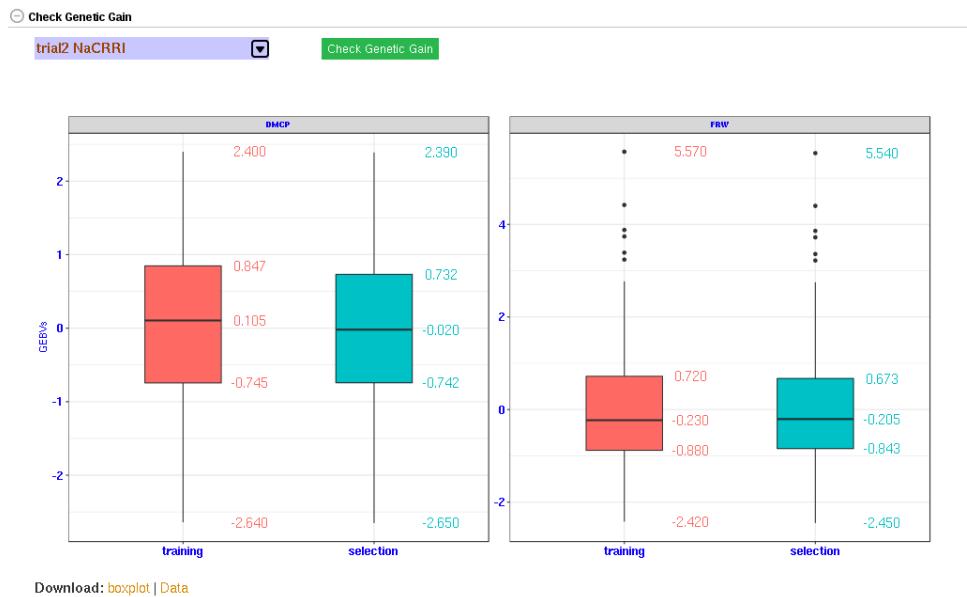
Hierarchical clustering:



[Download 34 clones](#) : [Dendrogram](#) | [Newick tree format](#) | [Analysis Report](#)

27.7 Genetic Gain

You can check for genetic gain by comparing the GEBVs of a training and a selection population. You can do this in the solGS pipeline once you build a model and apply the model to predict the GEBVs of a selection population. Once at that stage, you will see a section “Check Genetic Gain”. Select a selection population to compare with the training population and click the “Check Genetic Gain” button. The genetic gain will be visualized in boxplots. You can download the boxplot(s) as well as the GEBVs data used for the plot(s).



27.8 Kinship and Inbreeding Coefficients

This tool allows you to estimate genetic relatedness between a pair of individuals (kinship), homozygosity across loci in an individual (inbreeding coefficient), and genetic similarity of an individual relative to the rest of the population (average kinship).

There are three pathways to using this tool.

- (1) When you have a list or dataset clones, created from the search wizard:
 - (A) – go to the “Analyze” menu and select the kinship and inbreeding
 - (B) – make sure you are logged in
 - (C) – Select the genotypic protocol for the marker data
 - (D) – select your list or dataset of clones, click “Go”
 - (E) – click the “Run Kinship” and wait for the analysis to finish, depending on the data size this may take minutes. You can choose to submit the analysis and wait for an email notice to view the results or wait for it to complete.

(F) – You can download the output following the download links.

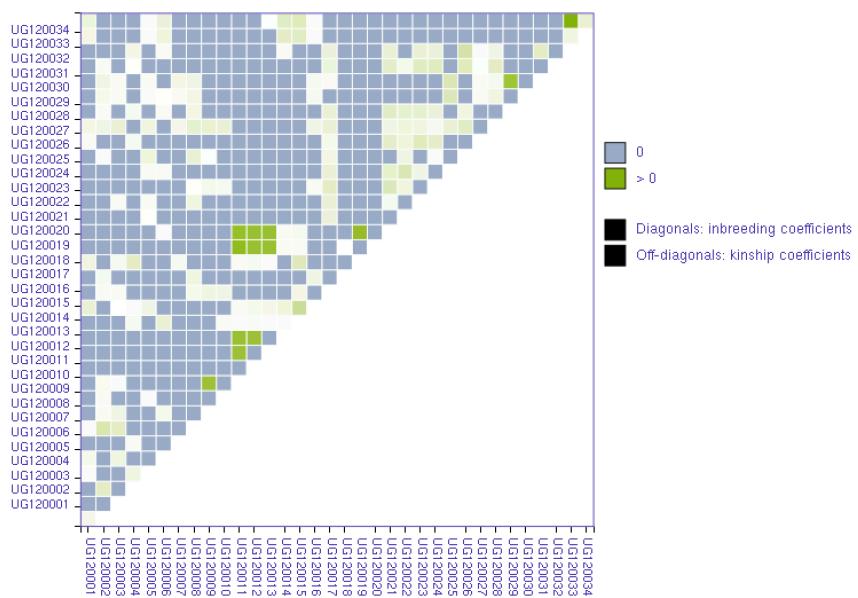
(2) From the trial detail page:

(A) – Go to the “Analysis Tools” section

(B) – Follow steps C to G in (1)

(3) In the solGS pipeline:

(A) – Once you are in a model output put page, scroll down to the “Kinship and Inbreeding” section and run kinship.



[Download: 34 clones Kinship matrix | Average kinship | Inbreeding coefficients](#)

27.9 Creating Crossing Groups

If you calculate selection index based on GEBVs of multiple traits, and you want to select a certain proportion of the indexed individuals (e.g. top 10%, or bottom 10%) and then you want to partition the selected individuals into a number of groups based on their genotypes, you can use the k-means clustering method.

The procedure is:

- (1) predict GEBVs for 2 or more traits
- (2) In the models output page, calculate selection indices. Note the name of the selection index data.
- (3) Go to the clustering section,
 - select the selection index data,
 - select “K-means”,
 - select “Genotype”,
 - in the K-numbers textbox, fill in the number of groups you want to create,
 - in the selection proportion textbox, fill in the proportion of the indexed individuals you want to select, e.g. for the top 15 percent, 15. if you wish to select bottom performing, prefix the number with minus sign (e.g. -15)
 - then run cluster and wait for the result.



27.10 Search Wizard Genomic Relationship Matrix (GRM) Download

The genomic relationship matrix (GRM) is useful for understanding underlying structure in your population. Breedbase can compute the GRM using rrBLUP. First, select accessions in the search wizard and optionally select a genotyping protocol. If no genotyping protocol is selected, the default genotyping protocol in your system is used (as defined in sgn_local.conf). Specify the minor allele frequency, missing marker data, and missing individuals data filters to apply. The GRM can be returned in a matrix format (.tsv) which shows all pairwise relationships between the selected accessions and is useful for visualization; alternatively, the GRM can be returned in a three-column format (.tsv) which is useful for programs like ASReml outside of Breedbase. The GRM can also be returned as a simple correlation heatmap image (.pdf). The GRM can be computed from parents of the selected accessions granted the parents were genotyped, by clicking the checkbox “compute from parents”; this is useful for programs where parental lines are genotyped and then hybrids are created and evaluated in the field.

27.11. SEARCH WIZARD GENOME WIDE ASSOCIATION STUDY (GWAS)283

Search Wizard

Don't see your data? Refresh Lists Update Wizard

Trials

Search

- CassavaTrial
- GenoTestCassava
- GenoTestMaize
- GenoTestMusa
- MaizeTrial
- MaizeInbredTrial

Match ANY ALL

Add to List... Create New List... Create

Accessions

Search

- 554353-1-1-B
- 554360-1-1-B
- 554363-1-1-B
- 554371-1-1-B
- 554372-1-1-B

Match ANY ALL

Add to List... Create New List... Create

Traits

Search

- Plot Weight [lbs]G2F-0000011
- Pollen DAP [days]G2F-0000013
- Root Lodging [plants]G2F-0000015
- Silk DAP [days]G2F-0000017
- Other traits: G2F-0000019
- Grain Moisture [percent]G2F-0000005
- Grain Yield [bu/acre]G2F-0000007
- Plant Height [cm]G2F-0000009

Match ANY ALL

Add to List... Create New List... Create

Select Column Type

Search

- Select All 0/0 Clear

Related Genotype Data

Download Genotype Data Compute From Parents
135 accessions; default protocol

Chromosome Start Position End Position

All

Genotypes Download Format Marker Set Filter

VCF File Format Select a marker set

Download Genotypes Download Genotypes

Marker Set Filter Manage Marker Sets

Minor Allele Frequency Marker Filter Individuals Filter

0.05 0.60 0.80

Genomic Relationship Matrix (GRM) Download Format

Matrix (.tsv)

Download GRM Download GRM

Genome Wide Association Study (GWAS) Download Format

Manhattan + QQ Plots (.pdf)

Selected Traits Are All Repeated Measurements

No

Run GWAS Download GWAS

Related Trial Metadata

27.11 Search Wizard Genome Wide Association Study (GWAS)

Performing a genome wide association study (GWAS) can determine genotypic markers which are significantly correlated to phenotypic traits. Breedbase can compute GWAS using rrBLUP. First, select accessions and trait(s)

in the search wizard, and optionally select a genotyping protocol. If no genotyping protocol is selected, the default genotyping protocol in your system is used (as defined in sgn_local.conf). Several traits can be selected in the search wizard; if the traits are not to be treated as repeated measurements then select “no” in the select box and this will tell Breedbase to return GWAS results independently for the selected traits. If the selected traits are indeed all repeated measurements then select “yes” in the select box and Breedbase will return a single GWAS analysis across all the phenotypic records. Specify the minor allele frequency, missing marker data, and missing individuals data filters to apply. GWAS results can be returned in a tabular format (.tsv) where the -log10(p-values) for the selected traits are returned; alternatively, the GWAS results can be returned as Manhattan and QQ plots for the selected traits. The GWAS can be computed from parents of the selected accessions granted the parents were genotyped, by clicking the checkbox “compute from parents”; this is useful for programs where parental lines are genotyped and then hybrids are created and evaluated in the field.

The GWAS will filter the data by the input MAF and missing data filters provided. After filtering the data is imputed using an “EM” method in rrBLUP. The Kinship matrix (GRM) is computed from the imputed genotypic data and used in the GWAS model. The GWAS uses fixed effects for different field trials and replicates in the phenotypic data.

27.11. SEARCH WIZARD GENOME WIDE ASSOCIATION STUDY (GWAS)285

Search Wizard

Don't see your data? Refresh Lists Update Wizard

Trials

Search

- CassavaTrial
- GenoTestCassava
- GenoTestMaize
- GenoTestMusa
- MaizeTrial
- MaizeInbredTrial

Match ANY ALL

Add to List... Create New List... Create

Accessions

Search

- 554353-1-1-B
- 554360-1-1-B
- 554363-1-1-B
- 554371-1-1-B
- 554372-1-1-B

Match ANY ALL

Add to List... Create New List... Create

Traits

Search

- Plot Weight [lbs]G2F:0000011
- Pollen DAP [days]G2F:0000013
- Root Lodging [plants]G2F:0000015
- Silk DAP [days]G2F:0000017
- Other Trait Codes: G2F:0000019
- Grain Moisture [percent]G2F:0000005
- Grain Yield [bu/acre]G2F:0000007
- Plant Height [cm]G2F:0000009

Match ANY ALL

Add to List... Create New List... Create

Select Column Type

Search

- Select All 3/14 Clear

Match ANY ALL

Add to List... Create New List... Create

Load/Create Datasets using Match Columns

Load Dataset Create New Dataset Load Create

Related Genotype Data

Download Genotype Data Compute From Parents
135 accessions; default protocol

Chromosome Start Position End Position

All VCF File Format Marker Set Filter

Download Genotypes Select a marker set

Download Genotypes Marker Set Filter

Download Genotypes Manage Marker Sets

Minor Allele Frequency	Marker Filter	Individuals Filter
0.05	0.60	0.80

Genomic Relationship Matrix (GRM) Download Format

Matrix (.tsv)

Download GRM

Download GRM

Genome Wide Association Study (GWAS) Download Format

Manhattan + QQ Plots (.pdf)

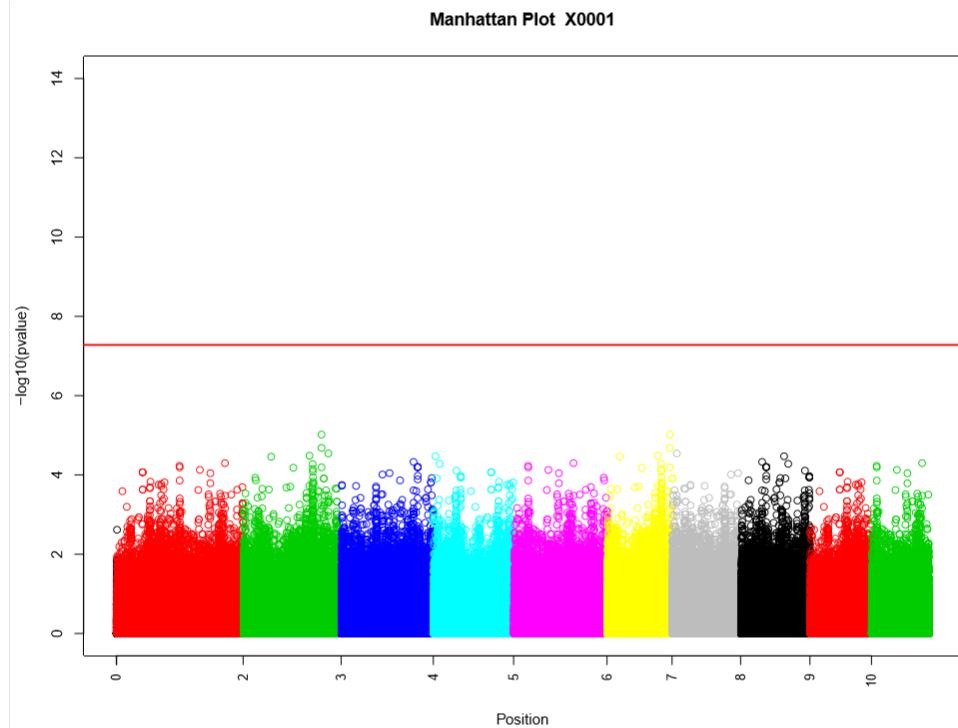
Selected Traits Are All Repeated Measurements

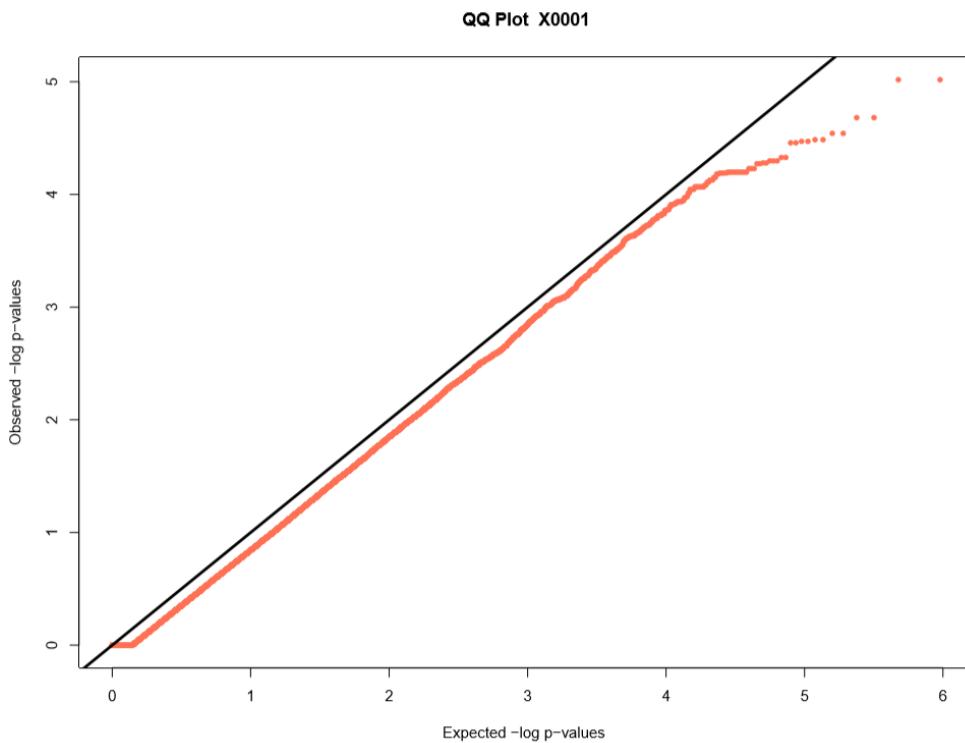
No

Run GWAS

Download GWAS

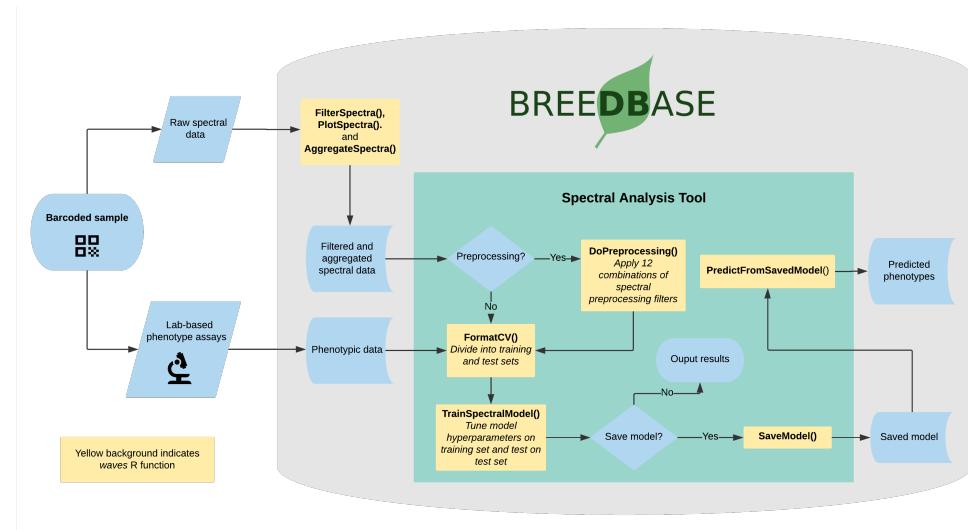
Related Trial Metadata





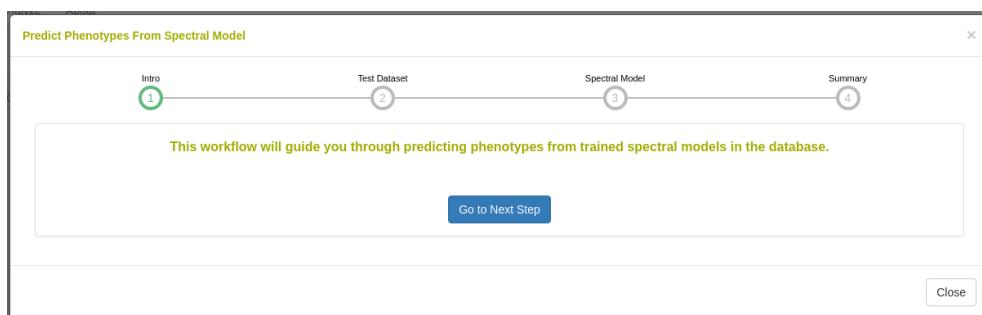
27.12 Spectral Analysis

Visible and near-infrared spectroscopy (vis-NIRS) can be related to reference phenotypes through statistical models to produce accurate phenotypic predictions for unobserved samples, increasing phenotyping throughput. This technique is commonly used for predicting traits such as total starch, protein, carotenoid, and water content in many plant breeding programs. Breedbase implements the R package [waves](#) to offer training, evaluation, storage, and use of vis-NIRS prediction models for a wide range of spectrometers and phenotypes.



27.12.1 Dataset selection

In order to initiate an analysis, the user must select one or more datasets using 2.1. A dataset in Breedbase can contain observationUnit-level (plot-, plant-, or sample-level) trial metadata and phenotypic data from one or more trials. After navigating to the “NIRS” webpage under the “Manage” tab in Breedbase, the user can initiate an analysis and select one of these datasets as input for model training. An optional test dataset can be selected in the second step of the workflow.



Predict Phenotypes From Spectral Model

Intro **1** Test Dataset **2** Spectral Model **3** Summary **4**

Select the dataset you are interested in predicting phenotypes for (the accessions or plots or tissues samples in the dataset need to have spectra uploaded):

Dataset: Show [2] entries Search:

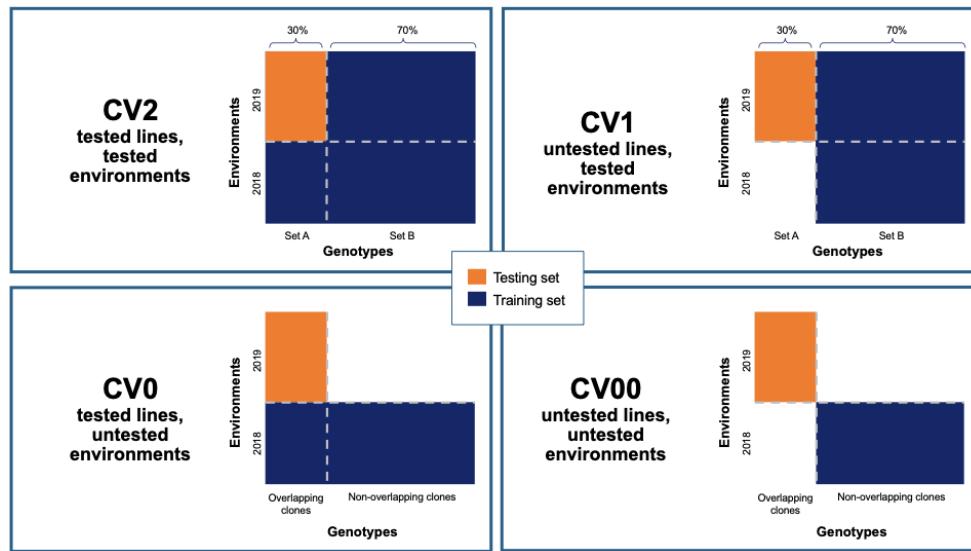
Select	Dataset Name	Contents						
<input type="checkbox"/>	dataset1	<table border="1"> <thead> <tr> <th>Trials</th> <th>Accessions</th> <th>Traits</th> </tr> </thead> <tbody> <tr> <td>field_tr</td> <td>test_acces[↑] test_acces[↓] test_acces[↑] test_acces[↓]</td> <td>Mean Pixel Value NIR (780-3000nm) Thresholded NIR Denoised Original Image [↑] Mean Pixel Value Red (600-590nm) Red Denoised Original Image day 2.541666 [↑]</td> </tr> </tbody> </table>	Trials	Accessions	Traits	field_tr	test_acces [↑] test_acces [↓] test_acces [↑] test_acces [↓]	Mean Pixel Value NIR (780-3000nm) Thresholded NIR Denoised Original Image [↑] Mean Pixel Value Red (600-590nm) Red Denoised Original Image day 2.541666 [↑]
Trials	Accessions	Traits						
field_tr	test_acces [↑] test_acces [↓] test_acces [↑] test_acces [↓]	Mean Pixel Value NIR (780-3000nm) Thresholded NIR Denoised Original Image [↑] Mean Pixel Value Red (600-590nm) Red Denoised Original Image day 2.541666 [↑]						
<input checked="" type="checkbox"/>	nirs_dataset1	<table border="1"> <thead> <tr> <th>Trials</th> <th>Accessions</th> </tr> </thead> <tbody> <tr> <td>nirsFieldTrial</td> <td>IBA011368 IBA011371 IBA141092 IBA30572 IBA30573</td> </tr> </tbody> </table>	Trials	Accessions	nirsFieldTrial	IBA011368 IBA011371 IBA141092 IBA30572 IBA30573		
Trials	Accessions							
nirsFieldTrial	IBA011368 IBA011371 IBA141092 IBA30572 IBA30573							

Showing 1 to 2 of 3 entries Previous **1** **2** Next Go to Next Step Close

27.12.2 Cross-validation

Five cross-validation schemes that represent scenarios common in plant breeding are available for this analysis. These include CV1, CV2, CV0, and CV00 as outlined below and described in depth by Jarquín et al. (2017) as well as random and stratified random sampling with a 70% training and 30% validation split. For those schemes from Jarquín et al. (2017), specific input datasets must be chosen based on genotype and environment relatedness. Cross-validation choices:

- * **Random sampling** (70% training / 30% validation)
- * **Stratified random sampling**, stratified based on phenotype (70% training / 30% validation)
- * **CV1**, untested lines in tested environments
- * **CV2**, tested lines in tested environments
- * **CV0**, tested lines in untested environments
- * **CV00**, untested lines in untested environments

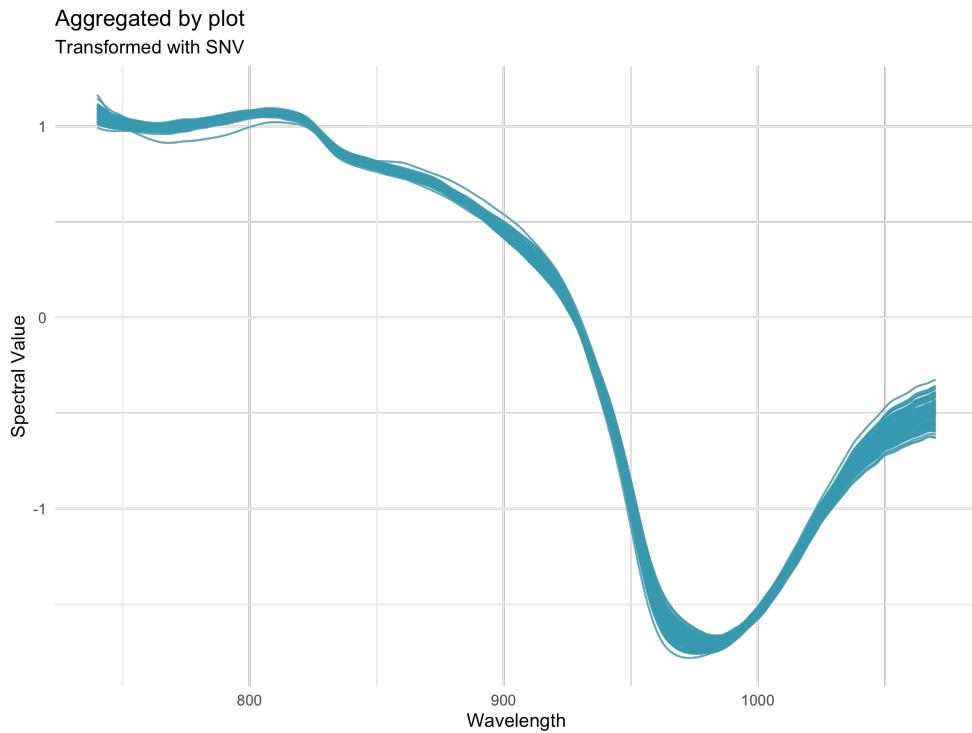


27.12.3 Preprocessing

Preprocessing, also known as pretreatment, is often used to increase the signal to noise ratio in vis-NIR datasets. The *waves* function *DoPreprocessing()* applies functions from the *stats* and *prospectr* packages for common spectral preprocessing methods with the following options:

- * Raw data (default)
- * First derivative
- * Second derivative
- * Gap segment derivative
- * Standard normal variate (SNV; Barnes et al., 1989)
- * Savitzky-Golay polynomial smoothing (Savitzky and Golay, 1964)

For more information on preprocessing methods and implementation, see the *waves* manual, available through CRAN: [waves.pdf](#)



27.12.4 Algorithms

Several algorithms are available for calibration model development in Breedbase via the `waves` package. The `TrainSpectralModel()` function in `waves` performs hyperparameter tuning as applicable using these algorithms in combination with cross validation and train functions from the package `caret`. Currently, only regression algorithms are available, but classification algorithms such as PLS-DA and SVM classification are under development. *

Partial least squares regression (PLSR; Wold et al., 1982; Wold et al., 1984) is a popular method for spectral calibrations, as it can handle datasets with high levels of collinearity, reducing the dimensionality of these data into orthogonal latent variables (components) that are then related to the response variable through a linear model (reviewed in Wold et al., 2001). To avoid overfitting, the number of these components included in the final model must be tuned for each use case. The PLSR algorithm from the `pls` package is implemented by `waves`. * **Random Forest regression** (RF; Ho, 1995) is a machine learning algorithm based on a series of decision trees. The num-

ber of trees and decisions at each junction are hyperparameters that must be tuned for each model. Another feature of this algorithm is the ability to extract variable importance measures from a fitted model (Breiman, 2001). In Breedbase, this option is made available through implementation of the RF algorithm from the package `randomForest` in the `waves` function `TrainSpectralModel()`. This function outputs both model performance statistics and a downloadable table of importance values for each wavelength. It is worth noting that this algorithm is computationally intensive, so the user should not be alarmed if results do not come right away. Breedbase will continue to work in the background and will display results when the analysis is finished. * **Support vector machine regression** (SVM; Vapnik, 2000) is another useful algorithm for working with high-dimension datasets consisting of non-linear data, with applications in both classification and regression. The package `waves` implements SVM with both linear and radial basis function kernels using the `kernlab` package.

27.12.5 Output: common model summary statistics

After training, model performance statistics are both displayed on a results webpage and made available for download in .csv format. These statistics are calculated by the `TrainSpectralModel()` function in `waves` using the `caret` and `spectacles` packages. Reported statistics include:

- * Tuned parameters depending on the model algorithm
- * **Best.n.comp**, the best number of components to be included in a PLSR model
- * **Best.ntree**, the best number of trees in an RF model
- * **Best.mtry**, the best number of variables to include at every decision point in an RF model
- * **RMSECV**, the root mean squared error of cross-validation
- * **R2cv**, the coefficient of multiple determination of cross-validation for PLSR models
- * **RMSEP**, the root mean squared error of prediction
- * **R2p**, the squared Pearson's correlation between predicted and observed test set values
- * **RPD**, the ratio of standard deviation of observed test set values to RMSEP
- * **RPIQ**, the ratio of performance to interquartile distance
- * **CCC**, the concordance correlation coefficient
- * **Bias**, the average difference between the predicted and observed values
- * **SEP**, the standard error of prediction
- * **R2sp**, the squared Spearman's rank correlation between predicted and observed test set values

27.12.6 Export model for later use

Once a model has been trained, it can be stored for later use. This action calls the `SaveModel()` function from [waves](#). Metadata regarding the training dataset and other parameters specified by the user upon training initialization are stored alongside the model object itself in the database.

The screenshot shows a web-based application interface for managing analysis details. At the top, a header reads "Analysis NIRS_MODEL_1_PREDICTION". Below this, a section titled "Analysis Details" contains a table with the following data:

Analysis Details	
Analysis Name	NIRS_MODEL_1_PREDICTION
Breeding Program	Breedbase
Year	2020
Description	Testing predicting phenotypes from saved trained NIRS model
Protocol	<pre>waves::SaveModel(df = train.ready, save.model = FALSE, autoselect.preprocessing = FALSE, preprocessing.method = pls, model.save.folder = NULL, model.name = 'PredictionModel', best.model.metric = 'RMSE', tune.length = 10, model.method = model.method, num.iterations = 10, wavelengths = wls, stratified.sampling = stratified.sampling, cv.scheme = random, trial1 = NULL, trial2 = NULL, trial3 = NULL)</pre>
Dataset ID	2
Created	2020-08-10 20:33:58
Result Summary	

To the right of the table is a QR code with the text "NIRS_MODEL_1_PREDICTION BB240" below it. A small link "View basic information about the analysis" is also present.

27.12.7 Predict phenotypes from an exported model (routine use)

For phenotype predictions, users select a dataset and can then choose from models in the database that were trained using the same spectrometer type as the spectral data in the chosen dataset. Predicted phenotypes are stored as such in the database and are tagged with an ontology term specifying that they are predicted and not directly measured. Metadata regarding the model used for prediction is stored alongside the predicted value in the database. Predicted phenotypes can then be used as normal in other Breedbase analysis tools such as the Selection Index and GWAS.

Screenshot 1: Spectral Model Step

The interface shows a navigation bar with four steps: Intro (1), Test Dataset (2), Spectral Model (3), and Summary (4). The current step is Spectral Model (3). A table lists prediction models:

Select	Model Name	Description	Format	Trait	Algorithm
<input type="checkbox"/>	nir_model1	asd	SCIO	dry matter content percentage CO_334:00000092	pls
<input checked="" type="checkbox"/>	NIRS_MODEL_1	NIRS to predict dry matter content	SCIO	dry matter content percentage CO_334:00000092	pls

Show 10 entries | Search: [] | Previous 1 2 Next | Predict | Close

Screenshot 2: Summary Step

The interface shows a navigation bar with four steps: Intro (1), Test Dataset (2), Spectral Model (3), and Summary (4). The current step is Summary (4). A summary table displays prediction results:

Stock	Prediction
SCIOTest_CASS_IBA011368_1	27.2301403275642
SCIOTest_CASS_IBA011368_2	27.9752347564317
SCIOTest_CASS_IBA011368_3	29.194847396204
SCIOTest_CASS_IBA011368_4	28.1528775118183
SCIOTest_CASS_IBA011368_5	29.0267489566395
SCIOTest_CASS_IBA011371_1	24.3428851923192
SCIOTest_CASS_IBA011371_2	26.196242114604
SCIOTest_CASS_IBA011371_3	26.031275629321
SCIOTest_CASS_IBA011371_4	23.3548384379248
SCIOTest_CASS_IBA011371_5	23.1089990379728
SCIOTest_CASS_IBA141092_1	30.2819198542414
SCIOTest_CASS_IBA141092_2	32.7734677979137

Do you want to save the prediction results?: | Close

27.12.8 FAQ

The Breedbase Spectral Analysis Tool does not allow for prediction models involving data from multiple spectrometer types at once.

References * Barnes, R.J., M.S. Dhanoa, and S.J. Lister. 1989. Standard normal variate transformation and de-trending of near-infrared diffuse reflectance spectra. Appl. Spectrosc. 43(5): 772-777. doi:

10.1366/0003702894202201. * Breiman, L. 2001. Random forests. *Mach. Learn.* 45: 5-32. doi: 10.1201/9780429469275-8. * Ho, T.K. 1995. Random decision forests. *Proc. Int. Conf. Doc. Anal. Recognition, ICDAR* 1: 278-282. doi: 10.1109/ICDAR.1995.598994. * Jarquín, D., C. Lemes da Silva, R.C. Gaynor, J. Poland, A. Fritz, et al. 2017. Increasing Genomic-Enabled Prediction Accuracy by Modeling Genotype x Environment Interactions in Kansas Wheat. *Plant Genome* 10(2): plantgenome2016.12.0130. doi: 10.3835/plantgenome2016.12.0130. * Johnson, R.A., and D.W. Wichern. 2007. *Applied Multivariate Statistical Analysis* (6th Edition). De Maesschalck, R., D. Jouan-Rimbaud, and D.L. Massart. 2000. The Mahalanobis distance. *Chemom. Intell. Lab. Syst.* 50(1): 1-18. doi: 10.1016/S0169-7439(99)00047-7. * Mahalanobis, P.C. 1936. On the generalized distance in statistics. *Natl. Inst. Sci. India.* * Savitzky, A., and M.J.E. Golay. 1964. Smoothing and Differentiation of Data by Simplified Least Squares Procedures. *Anal. Chem.* 36(8): 1627-1639. doi: 10.1021/ac60214a047. * Shrestha, R., L. Matteis, M. Skofic, A. Portugal, G. McLaren, et al. 2012. Bridging the phenotypic and genetic data useful for integrated breeding through a data annotation using the Crop Ontology developed by the crop communities of practice. *Front. Physiol.* 3 AUG(August): 1-10. doi: 10.3389/fphys.2012.00326. * Vapnik, V.N. 2000. *The Nature of Statistical Learning Theory*. Springer New York, New York, NY. * Wold, S., A. Ruhe, H. Wold, and W.J. Dunn, III. 1984. The Collinearity Problem in Linear Regression. The Partial Least Squares (PLS) Approach to Generalized Inverses. *SIAM J. Sci. Stat. Comput.* 5(3): 735-743. doi: 10.1137/0905052. * Wold, S., M. Sjöström, and L. Eriksson. 2001. PLS-regression: a basic tool of chemometrics. *Chemom. Intell. Lab. Syst.* 58(2): 109-130. doi: 10.1016/S0169-7439(01)00155-1.

27.13 General Mixed Model Tool

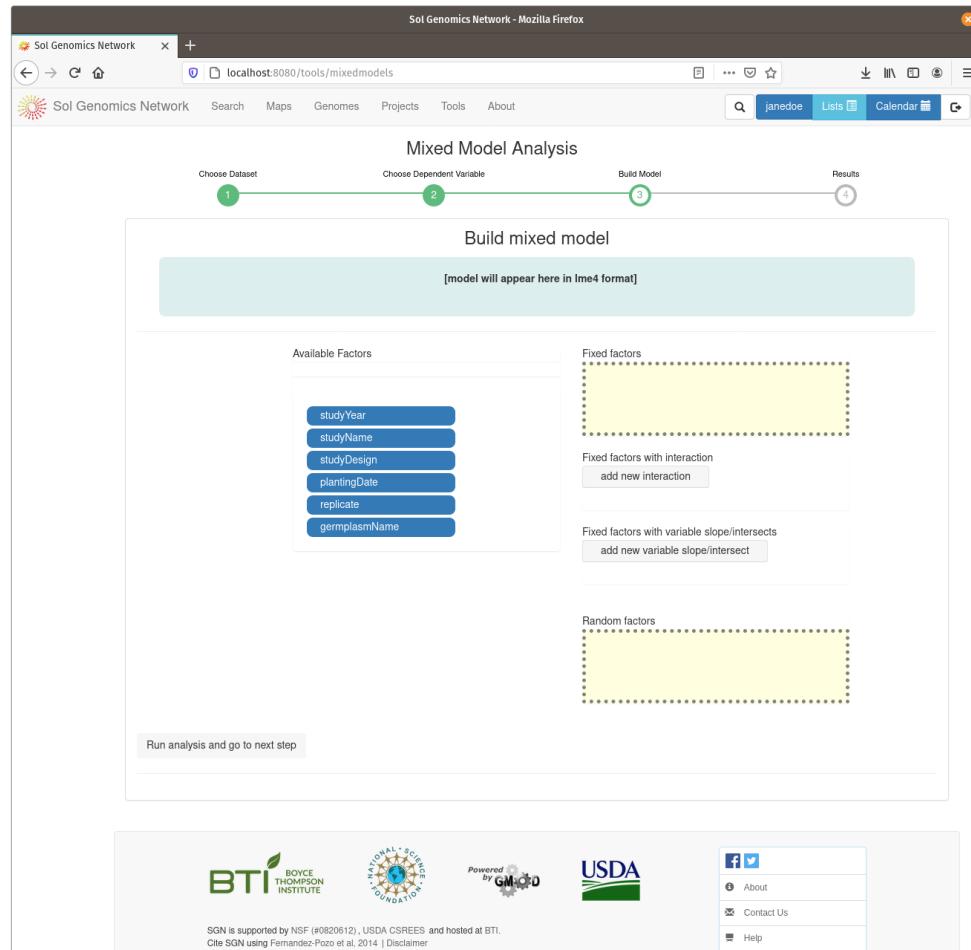
The general mixed model tool is available at /tools/mixedmodels and a link is provided from the Analyze menu.

To use the mixed model tool, first create dataset using the Wizard containing the data that you would like to analyze.

Select the Mixed Model tool from the Analyze menu.

You are presented with a workflow. On the first step of the workflow, select the dataset that you wish to analyze, click on “Choose dataset” to continue.

The second part of the workflow presents you with the traits in the dataset; you can select one or more traits from the lists using the select buttons. If you selected one trait, a bargraph of the trait distribution will be shown. Click the “Next step” button to move to the next screen.



On the model build screen, all the factors are displayed that are contained within the dataset. The factors are presented as a list of blue buttons that can be dragged using the mouse to areas on the screen which build a mixed model equation. The areas correspond to fixed factors, random factors, and optionally to more complex factors, such as fixed factors with interaction

and fixe factors with vriable slope/intersects. Drag the available factors to the corresponding area. To calculate BLUPs for germplasm, drag the germplasmName button to the “Random factors” area. To calculate BLUEs, drag it to the “Fixed factors” area. The factors need to have different levels contained within them, for example, if there is only one trial in the dataset, it cannot be used as one of the factors. Click on “Run analysis and got to next step” to run the mixed model and display the results.

The result view contains two tabs, one with the raw data, either BLUPS or BLUEs, and the other the adjusted means from the raw data.

The results can be stored in the database as an analysis, by clicking the button provided on the top of the data.

27.14 Genomic Prediction of Cross Performance (GPCP)

The GPCP tool is available at /tools/gpcp and a link is provided from the Analyze menu. The GCPG tool implements genomic prediction with additive and directional dominance in the linear mixed model to predict for cross performance.

Before using the tool, first create a dataset using the Wizard containing the data that you would like to analyze. (The dataset should have genotyping_protocols). Second, create Selection Indices for your traits using Selection Index in Analyze Menu.

To use the tool, Select the GPCP tool from the Analyze menu.

Then, select the dataset with genotyping_protocols that you wish to analyze, click on “Proceed to Factor Selection” to load available factors that can be included in the model.

Select the factors you wish to include in the model either as Fixed or Random. Click “None” for factors that you don’t want to include in the model. Note that the “germplasmName” is factored as Random by default.

The next step is to select the selection index for your traits on the dropdown menu.

Once you are through, click “Run GPCP” to run the model. The output will be presented in form of a table with “ID”, “Parent1”, “Parent2” and their cross prediction merit organized in descending order. The results will also have sex information based on whether the dataset has plant sexes available in the database.

27.15 Tool Compatibility

The dataset definition enables one to predict whether the dataset can be used in various analysis tools.

Upon creating a dataset, the site will automatically predict its compatibility with the available analysis tools and report these values on the dataset details page.

Tool Compatibility	Correlation ✓	Stability ✗	Mixed Models ✓	Population Structure ▲	Clustering ▲	Heritability ✗	Boxplotter ✓
	traits fresh root weight harvest index variable dry matter content percentage fresh shoot weight measurement in kg		traits fresh root weight harvest index variable dry matter content percentage fresh shoot weight measurement in kg	traits Genotype Phenotype	types Phenotype Genotype		traits fresh root weight harvest index variable dry matter content percentage fresh shoot weight measurement in kg

In the table, each tool will report to the user which traits are available to be analyzed based on phenotype data, and if different types of analyses are available, these will also be reported to the user. Some tools may give a warning sign to indicate that this dataset is compatible, but with potentially low sample sizes. Hover over the warning symbol to get a readout of the reason for the warning.

Below the table, there is a button that enables the user to re-calculate tool compatibility. This can be useful if a dataset is created before phenotypes are uploaded to a trial, since phenotype data is used in determining dataset compatibility. Even if the page appears to hang, do not worry; the compatibility check will continue in the background, and you can check later.

Tool Compatibility		Data Summary					
markers per genotyping protocol		number of genotyped accessions per protocol					
GBS ApeKI genotyping v4 : 14522		GBS ApeKI genotyping v4 : 300					
		trait observations per location					
		test_location [Computation]					
		fresh root weight : 549 harvest index variable : 268					
		dry matter content percentage : 531					
		fresh shoot weight measurement in kg : 575					
		fresh root weight : 549 harvest index variable : 268					
		dry matter content percentage : 531					
		fresh shoot weight measurement in kg : 575					
		number of observations per trait					
		fresh root weight : 549 harvest index variable : 268					
		dry matter content percentage : 531					
		fresh shoot weight measurement in kg : 575					
		number of accessions per trial					
		Shared across all trials : 0					
		number of phenotyped accessions per trait					
		fresh root weight : 272 harvest index variable : 268					
		dry matter content percentage : 266					
		fresh shoot weight measurement in kg : 280					
Check Tool Compatibility							

Below the tool compatibilities, there is also a summary of the data encompassed by the dataset and the criteria used for determining tool compatibility. Those criteria are used in the following way:

- Correlation: A dataset can be used in a correlation analysis if there are many phenotype measurements for different traits made on the same accession.
- Population Structure (PCA): A genotype PCA can be run if there are many accessions all genotyped with the same protocol. A phenotype PCA can be run if many accessions all have measurements on many traits.
- Clustering: Like a PCA, clustering can be done in both phenotype and genotype modes. They have the same requirements as PCA.
- Kinship & Inbreeding: A dataset with many accessions genotyped with the same protocol can be used for kinship analyses.
- Stability: A dataset containing many accessions with the same trait measured across multiple locations can be used in stability analyses.
- Heritability: This requires one or more trials with the same trait measured on the same accession across those trial(s).
- Mixed Models: This requires sufficient accession numbers, trait measurements, and trial designs.
- GWAS: A dataset is compatible with GWAS if there are many accessions genotyped for the same genotyping protocol, and the genotyping protocol has enough markers to run a GWAS. In addition, each accession needs to be phenotyped for a trait.
- Boxplotter: There must be sufficient trait measurements to make a boxplot of the trait.

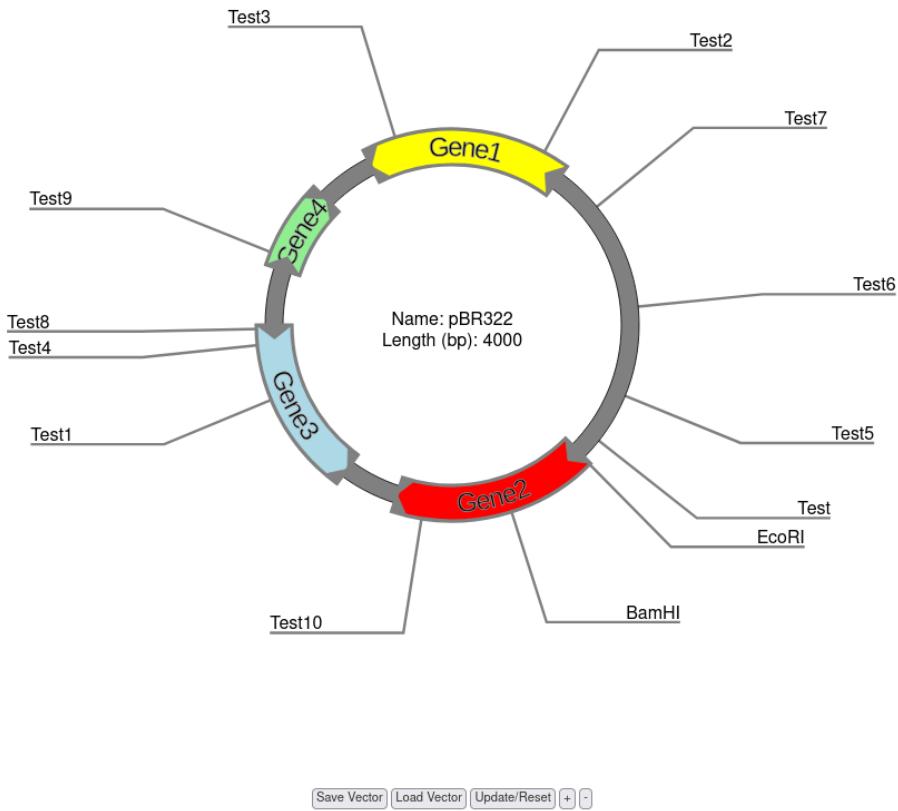
In addition to being on the dataset details page, tool compatibilities may be listed on the dataset selection screens for analysis tools. The compatibilities are non-blocking; you may always try using a dataset in an analysis even if there are warnings or if it is deemed non-compatible. As before, you can hover over the warning symbols to see why a dataset may not have statistical

power. For analyses with multiple modes, such as clustering and PCA, you can also hover over the compatibility checkmark to see what types (phenotype or genotype) the dataset is compatible with.

Chapter 28

Vector Viewer

BreedBase provides a vector viewer tool that can generate, save, or upload vectors onto BreedBase.

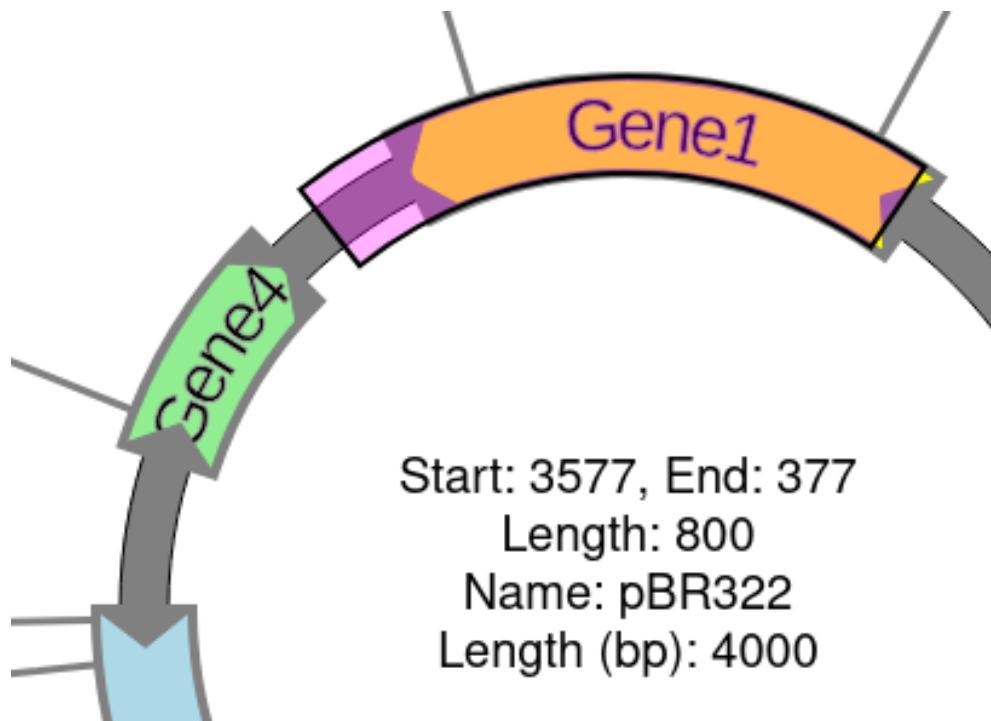


The vector viewer tool comes with many interactive features:

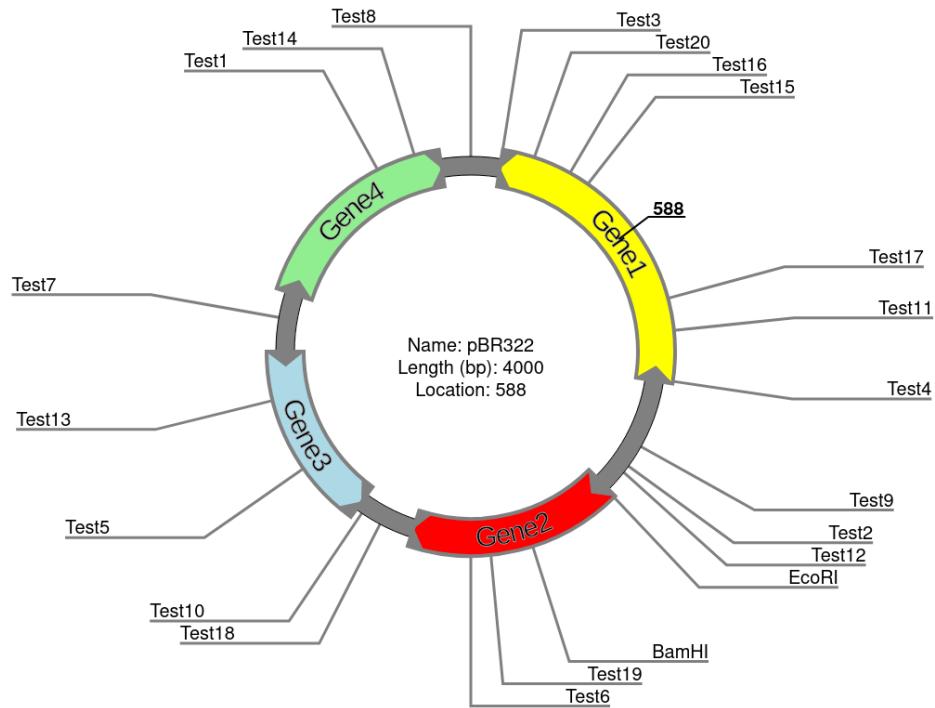
- Highlighting
- Location display
- Zooming in / panning out
- A feature and restriction enzyme table that support real time editing, deleting, or adding

To highlight a region of the vector viewer, click and drag your cursor clockwise. To delete the new highlighted segment, either start a new highlighted segment by clicking on the vector, or click the “Update/Reset” button.

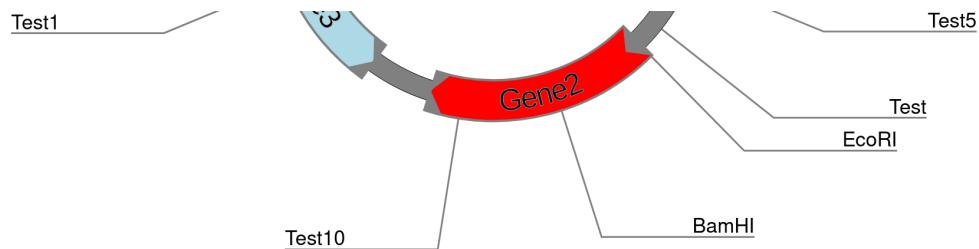
303

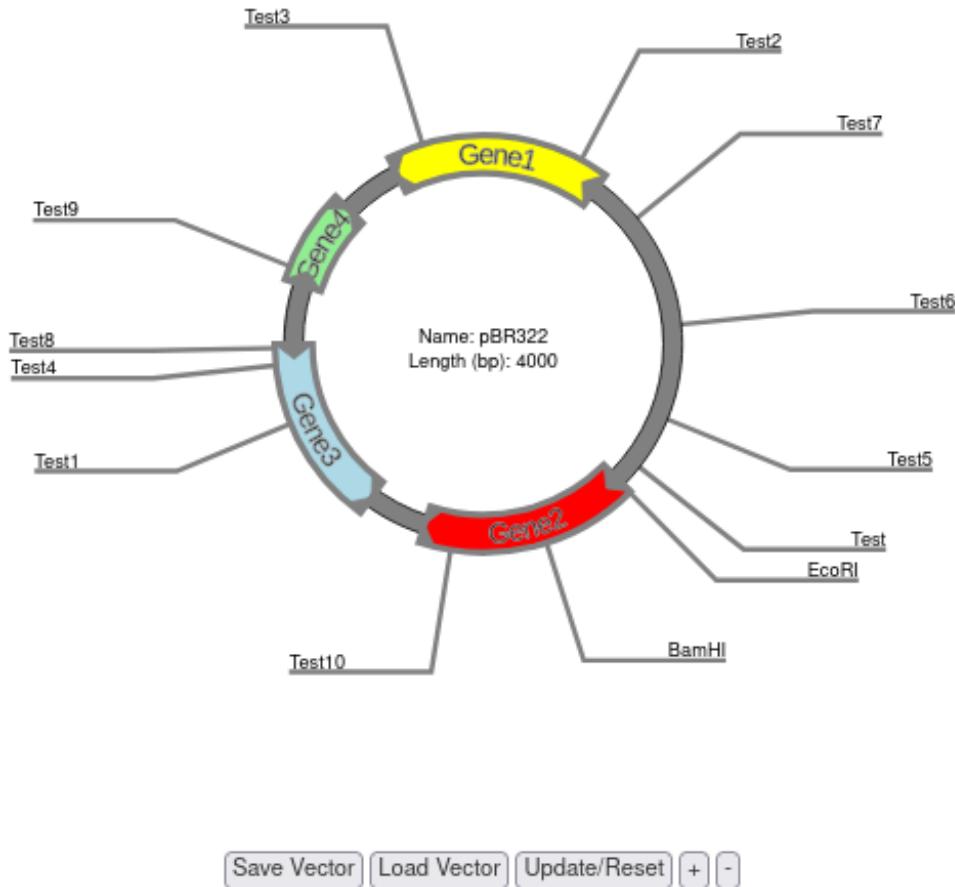


To find the location of a spot on the vector, hover your cursor over the vector and a number will appear. That is the location, in base pairs, of your cursor.



To zoom in and pan out, click the “+” or the “-” button underneath the vector.





To add a feature or restriction site, click “Add”. To edit a feature or restriction site, select the feature or restriction site of choice, click “Edit”, and make any wanted changes. To delete a feature or restriction site, selected the feature or restriction site that you want to delete and then click “Delete”.

<input type="button" value="Add"/>	<input type="button" value="Edit"/>	<input type="button" value="Delete"/>
Search: <input type="text"/>		