



02- Occurrence Record Cleaning

Shelly Gaynor
University of Florida



Occurrence data cleaning

0. Exploring the dataset
1. Resolve taxon names
2. Clean localities
 1. Precision
 2. Remove impossible points
 - Most common coordinate: 0.00, 0.00
 - Cultivated zones, botanical gardens, etc
3. Remove duplicates
4. Spatial correction
5. Visualize
6. Save cleaned.csv
- REPEAT -

Load Packages

```
library(gatoRs)  
library(fields)  
library(sf)  
library(ggplot2)  
library(ggspatial)  
library(leaflet)
```

- **gatoRs** custom package for processing biodiversity data
- Geospatial packages include **sf** and **fields**
- **ggplot2**, **ggspatial**, and **leaflet** are for visualization

Load Data File

```
rawdf <- read.csv("data/download/raw/Shortia_galacifolia_  
raw_20230605.csv")
```

- **read.csv** is a base function of R, it reads csv files

Look at the Data File

How many observations do we start with?

```
nrow(rawdf)
```

```
## [1] 1219
```

- **nrow** print the **number of rows**

1. Resolve taxon names

```
unique(rawdf$scientificName)
```

```
## [1] "Shortia galacifolia Torr. & A.Gray"  
## [2] "Sherwoodia galacifolia (Torr. & A.Gray) House"  
## [3] "Shortia galacifolia"  
## [4] "Shortia galacifolia Torr. & A. Gray"  
## [5] "Shortia galacifolia Torrey & A. Gray"  
## [6] "Shortia galacifolia var. galacifolia"  
## [7] "Shortia galacifolia var. brevistyla"  
## [8] "Shortia galacifolia var. brevistyla P. A. Davies"  
## [9] "Sherwoodia galacifolia"
```

Create a list of accepted names based on the name column in your data frame

```
search <- c("Shortia galacifolia", "Sherwoodia galacifolia")
```

- **unique** prints unique values in the column ***name*** from the dataframe ***rawdf***.
- **c** creates a list stored in the object ***search*** that contains “***Shortia galacifolia***” and “***Sherwoodia galacifolia***”

1. Resolve taxon names

Filter to only include accepted name:

```
df <- taxa_clean(df = rawdf,  
                 synonyms.list = search,  
                 taxa.filter = "fuzzy",  
                 accepted.name = "Shortia galacifolia")
```

- Here we are going to harmonize taxonomy using **taxa_clean()**.
- This function has three filter options: exact, fuzzy, or interactive.

2. Clean localities

Here we remove any records with missing coordinates, impossible coordinates, coordinates at (0,0), and any that are flagged as skewed. We also round the provided latitude and longitude values to a specified number of decimal places.

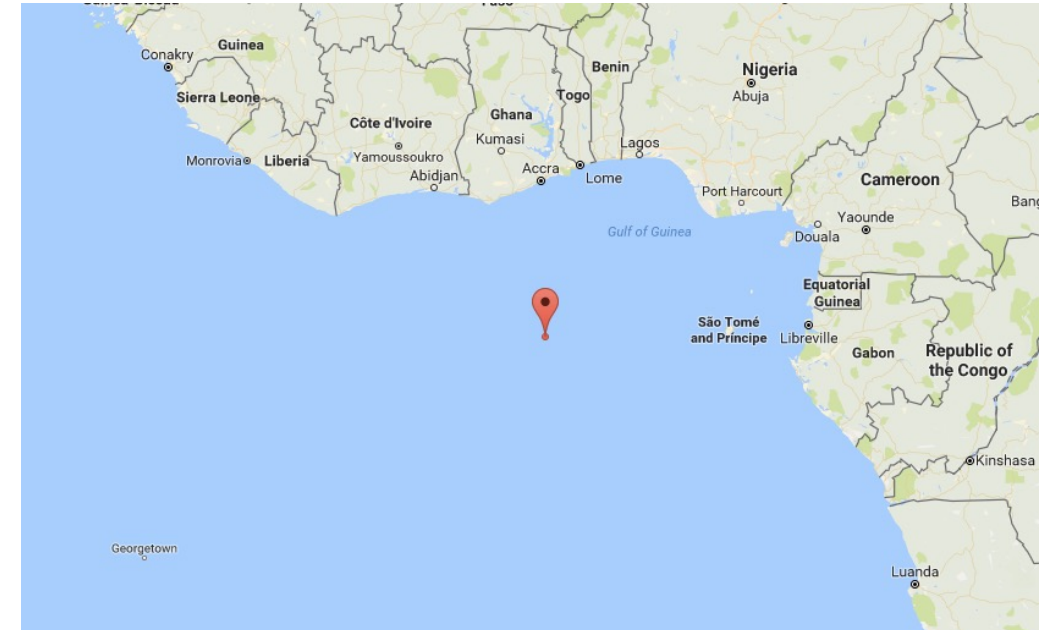
```
df <- basic_locality_clean(df = df,  
                           remove.zero = TRUE, # Records at (0,0) are removed  
                           precision = TRUE, # latitude and longitude are rounded  
                           digits = 2, # round to 2 decimal places  
                           remove.skewed = TRUE)
```

- **basic_locality_clean()** is from the package **gatoRs**

2. Clean localities

```
df <- basic_locality_clean(df = df,  
                           remove.zero = TRUE, # Records at (0,0) are removed
```

- Only retains records in the column long/lat that are not (\neq) equal to 0.00
- "There's no place like 0,0"



2. Clean localities

```
df <- basic_locality_clean(df = df,  
                           remove.zero = TRUE, # Records at (0,0) are removed  
                           precision = TRUE, # latitude and longitude are rounded  
                           digits = 2, # round to 2 decimal places
```

Precision	0 degrees Latitude	30 degrees Latitude	60 degrees Latitude	85 degrees Latitude
1.0 degrees	156904 m	146962 m	124605 m	112109 m
0.1 degrees	15691 m	14697 m	12461 m	11211 m
0.01 degrees	1570 m	1470 m	1247 m	1122m
0.001 degrees	157 m	147 m	125 m	113 m
0.0001 degrees	16 m	15 m	13 m	12 m
0.00001 degrees	2 m	2 m	2 m	2 m

- Our climate layers are 30 arc-sec resolution which is about 1000 meters

2. Clean localities

```
df <- basic_locality_clean(df = df,  
                           remove.zero = TRUE, # Records at (0,0) are removed  
                           precision = TRUE, # latitude and longitude are rounded  
                           digits = 2, # round to 2 decimal places  
                           remove.skewed = TRUE)
```

- The skewed records can be identified with based on the 'InformationWithheld' column.

```
remove_skewed <- function(df, info.withheld = "informationWithheld"){  
  df <- df[grepl("Coordinate uncertainty increased", df[[info.withheld]]) == FALSE, ]  
  return(df)  
}
```

2. Clean localities

Remove unlikely points

Removes points in a radius
around biodiversity institutions

Remove coordinates in cultivated zones, botanical gardens, and outside our desired range. Here we set `interactive = FALSE`, however you can visualize the outliers by setting `interactive = TRUE`.

```
df <- process_flagged(df, interactive = FALSE)
```

3. Remove Duplicates

Here we identify and remove both (1) specimen duplicates and (2) aggregator duplicates based on each specimens coordinates, occurrenceID, and eventDate. To leverage all date information available, set `remove.unparseable = FALSE` to manually populate the year, month, and day columns. Here, we also confirm all ID (UUID and key) are unique to remove any within-aggregator duplicates that may accumulate due to processing errors.

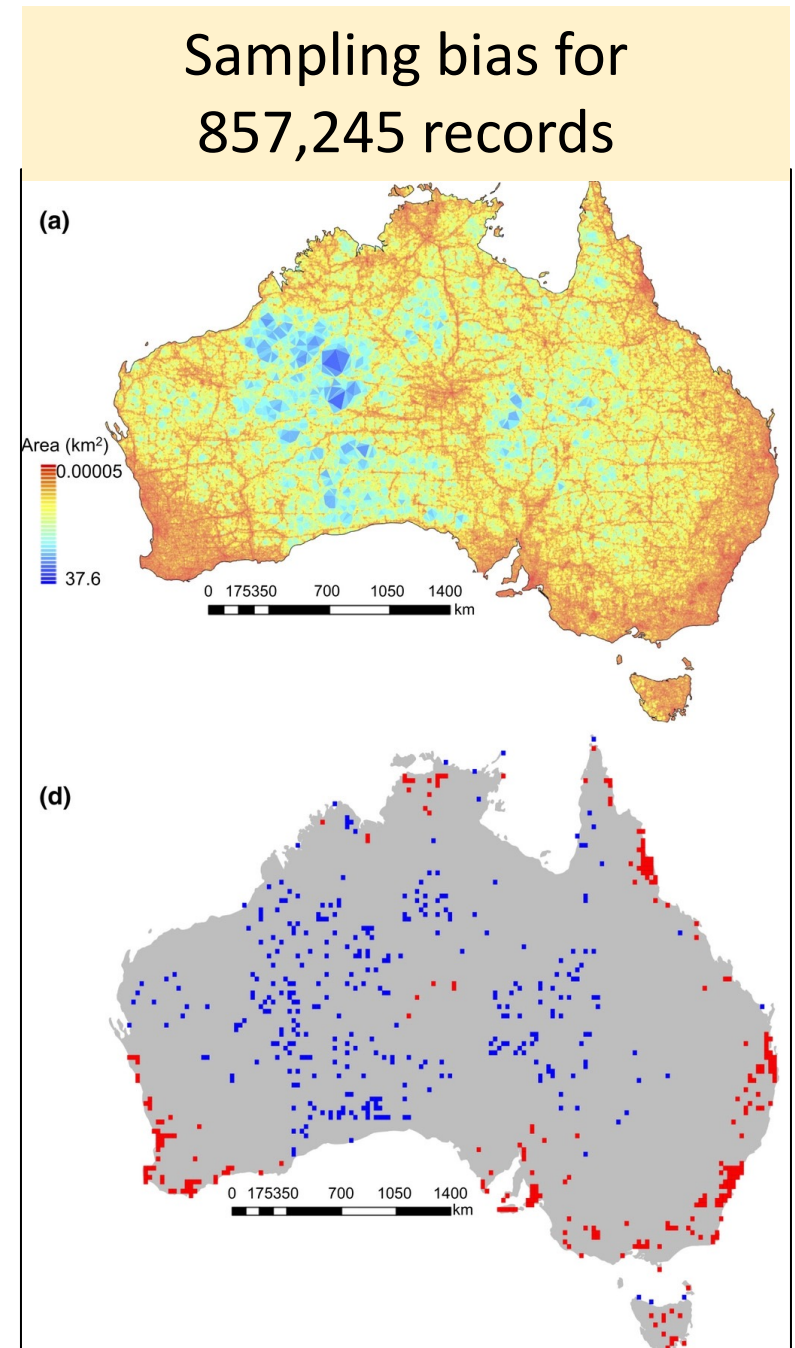
```
df <- remove_duplicates(df, remove.unparseable = TRUE)
```

- **parsedate** is a beautiful package that parses dates – However, it only parses dates after 1970!
 - Try **remove.unparable = FALSE**

4. Spatial Correction

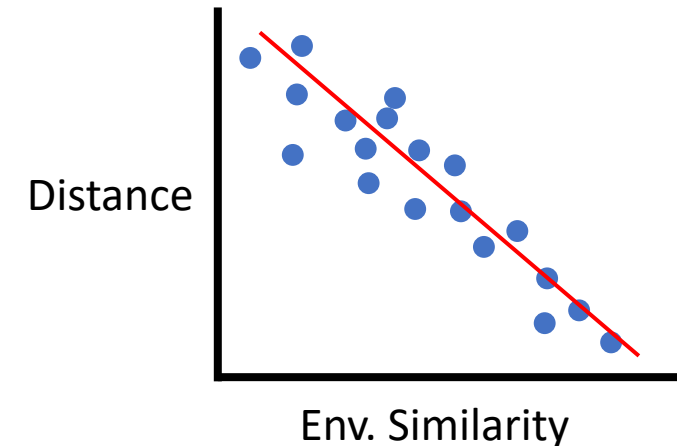
- Filtering is a procedure to reduce the clustering of species records
 - Collection efforts can lead to clustering of points

Daru et al. 2017. Widespread sampling biases in herbaria revealed from large-scale digitization. *New Phytologist*.



4. Spatial Correction

- Filtering is a procedure to reduce the clustering of species records
 - Collection efforts can lead to clustering of points
- After filtering, there may still be spatial autocorrelation
 - This can be accounted for by data partitioning



Sillero N. and A. M. Barbosa. 2020. Common mistakes in ecological niche models. International Journal of Geographical Information Science.

4. Spatial Correction

Maxent will only retain one point per pixel. To make the ecological niche analysis comparable, we will retain only one point per pixel. Note: Default is a raster with 30 arc sec resolution.

One point per pixel

Maxent will only retain one point per pixel. To make the ecological niche analysis comparable, we will retain only one point per pixel. Note: Default is a raster with 30 arc sec resolution.

```
df <- one_point_per_pixel(df)
```

How many observations do we have now?

```
nrow(df)
```

```
## [1] 21
```


4. Spatial Correction

Step 1: What should your minimum distance be?

First, let's assess the current minimum distance among your occurrence records. Here, we calculate minimum nearest neighbor distance in km.

```
nnDm <- rdist.earth(as.matrix(data.frame(lon = df$long, lat = df$lat)), m
iles = FALSE, R = NULL)
nnDmin <- do.call(rbind, lapply(1:5, function(i) sort(nnDm[,i])[2]))
min(nnDmin)
```

```
## [1] 2.226477
```

Here the current minimum distance is 2.22 km. Based on literature, we find a 2 meters (or 0.002 km) distance was enough to collect unique genets, so we do not need to thin our points.

- **rdist.earth** can be used to look at nearest neighbor distance in kilometers
- **do.call** applies to a list, **rbind** binds rows, and the **sort** is set up to select the lowest value.
- We want to look at the **min** distance between points remaining

4. Spatial Correction

- **spThin** function *thin*
- Set thin.par = minimum nearest neighbor distance
- Next, the function calculates pairwise distance between all records
- Then:
 - 1) For each record, IDs the # of occurrence records within distance thin.par
 - 2) One record of those which share the greatest # from (1) is removed at random.
 - 3) Repeat 1 – 2 till no record has a nearest neighbor closer than thin.par

Aiello-Lammens et al. 2015. spThin: an R package for spatial thinning of species occurrence records for use in ecological niche models. Ecography.

4. Spatial Correction

Step 2: Thin occurrence records using spThin through gatoRs.

When you do need to thin your records, here is a great function to do so!

```
df <- thin_points(df,  
                  distance = 0.002, # in km  
                  reps = 100)
```

5. Plot Cleaned Records

Make points spatial

```
df_fixed <- st_as_sf(df, coords = c("longitude", "latitude"), crs = 4326)
```

Set basemap

```
USA <- borders(database = "usa", colour = "gray50", fill = "gray50")  
state <- borders(database = "state", colour = "black", fill = NA)
```

- crs = 4326 is WGS84
- Using the function **borders** from the package **ggplot2**, we download basemaps to plot our points on.

5. Plot Cleaned Records

```
simple_map <-ggplot() +  
  USA +  
  state +  
  geom_sf(df_fixed,  
    mapping = aes(col = name),  
    col = "blue") +  
  coord_sf(xlim = c(min(df$longitude) - 3,  
    max(df$longitude) + 3),  
    ylim = c(min(df$latitude) - 3,  
    max(df$latitude) + 3)) +  
  xlab("Longitude") +  
  ylab("Latitude") +  
  annotation_scale(plot_unit = "km") +  
  annotation_north_arrow(height = unit(1, "cm"),  
    width = unit(1, "cm"),  
    location = "t1")  
  
simple_map
```

1. Add basemaps

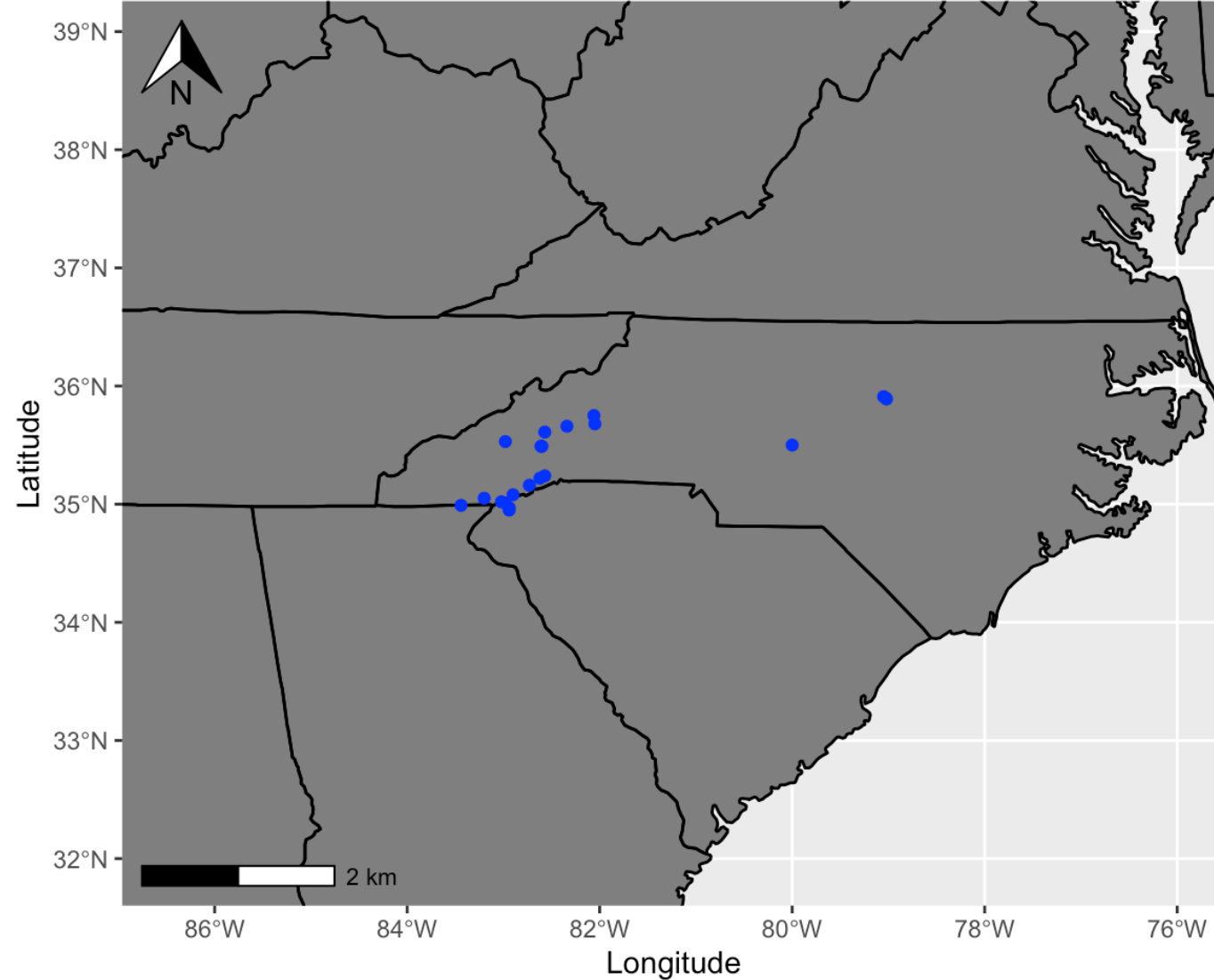
2. Add points

3. Zoom in

4. Fix axis labels

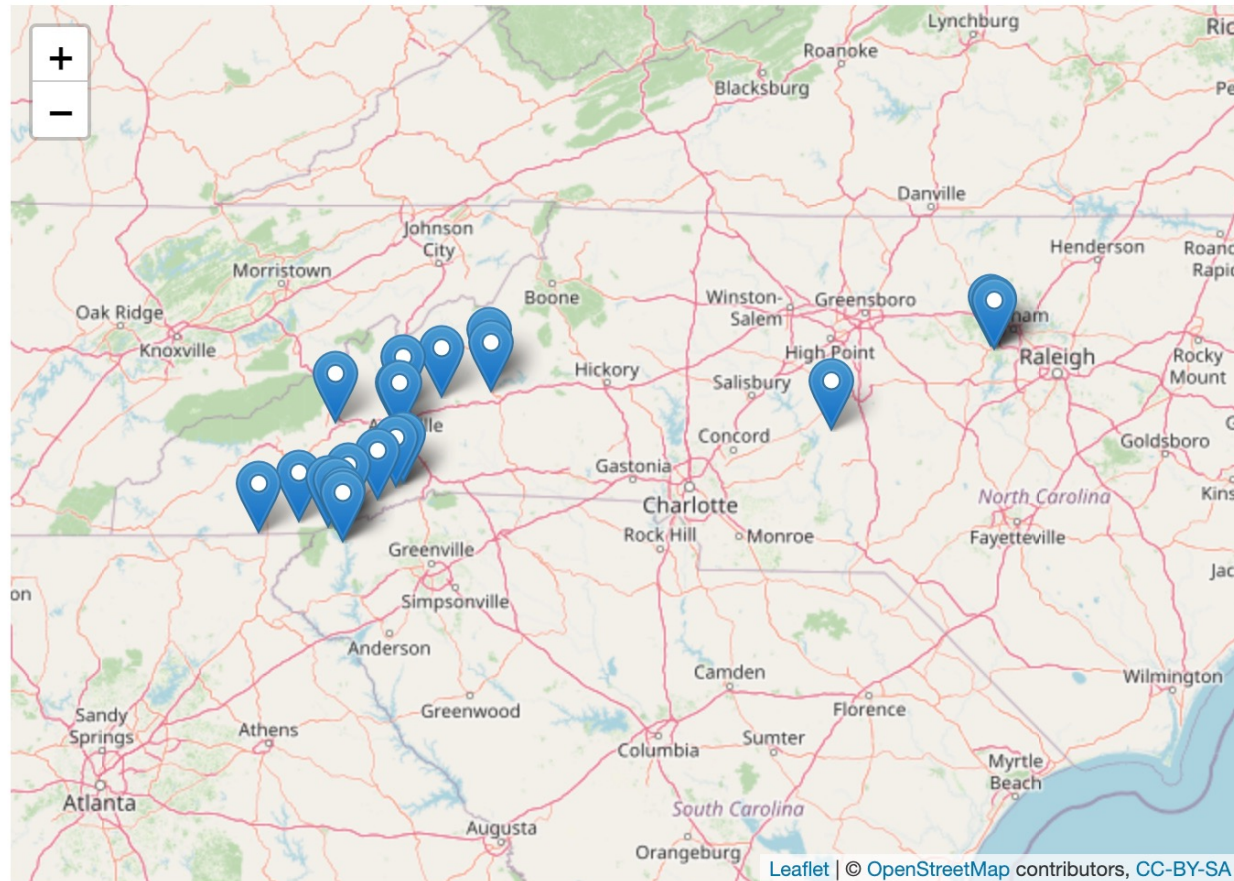
5. Add scale and North arrow

5. Plot Cleaned Records



5. Plot Cleaned Records

```
leaflet(df_fixed) %>%  
  addMarkers(label = paste0(df$longitude, ", ", df$latitude)) %>%  
  addTiles()
```



6. Saved Cleaned.csv

```
write.csv(df, "data/cleaning_demo/Shortia_galacifolia_20230605-cleaned.csv", row.names  
= FALSE)
```


7. Repeat for all taxa

```
## Set up for loop
files <- list.files("data/download/raw", full.names = TRUE)[1:3]
synonymns <- list(Galax_urceolata = c("Galax urceolata",
                                       "Galax urceolata (Poir.) Brummitt",
                                       "Galax urceolata (Poiret) Brummitt",
                                       "Galax urceolaa",
                                       "Galax aphylla L.",
                                       "Galax aphylla"),
                  Pyxidanthera_barbulata = c("Pyxidanthera barbulata",
                                              "Pyxidanthera barbulata Michx.",
                                              "Pyxidanthera barbulata var. barbulata" ,
                                              "Pyxidanthera barbulata"),
                  Pyxidanthera_brevifolia = c("Pyxidanthera brevifolia",
                                              "Pyxidanthera brevifolia Wells",
                                              "Pyxidanthera barbulata var. brevifolia (We
lls) H.E.Ahles",
                                              "Pyxidanthera barbulata var. brevifolia"
))
```

1. Create list of files with paths

2. Create list of synonyms

7. Repeat for all taxa

```
## Repeat cleaning steps for the remaining taxa
for(i in 1:3){
  df <- read.csv(files[i])
  # Taxa clean
  search <- synonymns[[i]]
  df <- taxa_clean(df = df, synonyms.list = search,
                  taxa.filter = "exact",
                  accepted.name = search[1])

  # Locality clean
  df <- basic_locality_clean(df = df, remove.zero = TRUE,
                           precision = TRUE, digits = 2,
                           remove.skewed = TRUE)

  df <- process_flagged(df, interactive = FALSE)
  # Remove duplicates
  df <- remove_duplicates(df, remove.unparseable = TRUE)
  # Spatial correct
  df <- one_point_per_pixel(df)
  df <- thin_points(df, distance = 0.002, reps = 100)
  # Save file
  outfile <- paste0("data/cleaning_demo/",
                   gsub(" ", "_", search[1]),
                   "_20230605-cleaned.csv")

  write.csv(df, outfile,
            row.names = FALSE)
  rm(df, search, outfile)
}
```

0. Read in downloaded data
1. Resolve taxon names
2. Clean localities
 1. Precision
 2. Remove impossible points
 - Most common coordinate: 0.00, 0.00
 - Cultivated zones, botanical gardens, ect.
3. Remove duplicates
4. Spatial correction
5. Visualize
6. Save cleaned.csv

8. Make Maxent Ready

Read in all cleaned files

```
alldf <- list.files("data/cleaning_demo/", full.names = TRUE,  
                  recursive = FALSE, include.dirs = FALSE, pattern = "*.csv")  
alldf <- lapply(alldf, read.csv)  
alldf <- do.call(rbind, alldf)
```

8. Make Maxent Ready

Visually inspect records to verify no additional records should be removed.

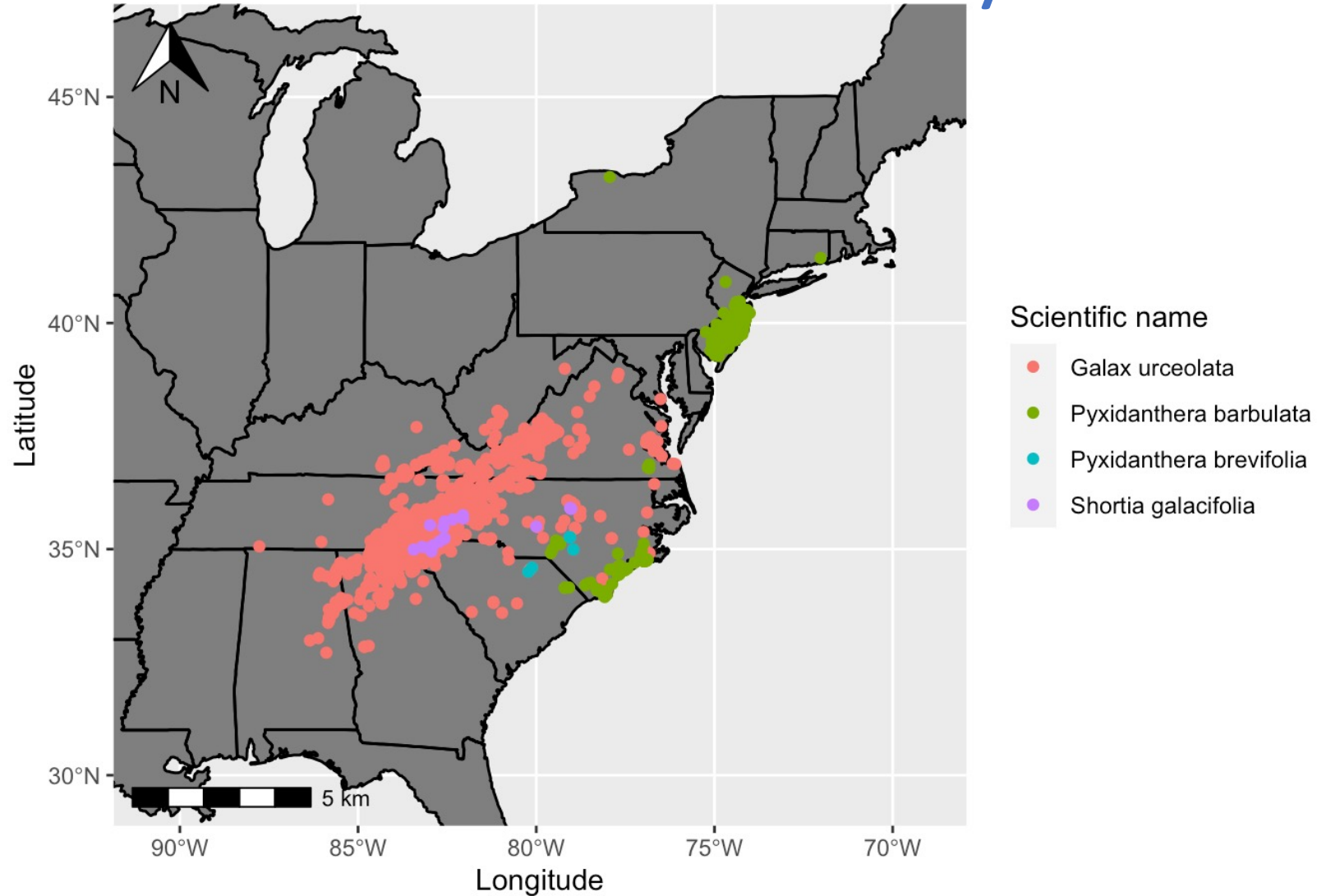
```
### Make points spatial
alldf_fixed <- st_as_sf(alldf, coords = c("longitude", "latitude"),
                      crs = 4326)

### Set basemap
USA <- borders(database = "usa", colour = "gray50", fill = "gray50")
state <- borders(database = "state", colour = "black", fill = NA)

### Plot
all_map <- ggplot() +
  USA +
  state +
  geom_sf(alldf_fixed,
          mapping = aes(col = factor(accepted_name))) +
  coord_sf(xlim = c(min(alldf$longitude) - 3, max(alldf$longitude) + 3),
           ylim = c(min(alldf$latitude) - 3, max(alldf$latitude) + 3)) +
  xlab("Longitude") +
  ylab("Latitude") +
  labs(color = "Scientific name") +
  annotation_scale(plot_unit = "km") +
  annotation_north_arrow(height = unit(1, "cm"),
                        width = unit(1, "cm"),
                        location = "tl")

all_map
```

8. Make Maxent Ready



8. Make Maxent Ready

Select needed columns

```
alldfchomp <- c()
species <- unique(alldf$accepted_name)
for(i in 1:4){
  alldfchomp[[i]] <- data_chomp(alldf[alldf$accepted_name ==species[i], ],
                                accepted.name = species[i])
}
alldf <- do.call(rbind, alldfchomp)
```

Save Maxent.csv

```
write.csv(alldf, "data/cleaning_demo/maxent_ready/diapensiaceae_maxentready_20230605.csv", row.names = FALSE)
```

Minimum occurrence records

- Your points should represent the env. variability true to your species.
 - Too few points may not classify your species suitable conditions properly.
- It is not clear the minimum number of points needed.
 - Might be dependent on the extent of the study area
- 3, 4, or 5 have been suggested
 - 5 per env. variable has also been suggested
 - More env. variables may not significantly change the minimum sample size needed (see study below).

Proosdij et al. 2016. Minimum required number of specimen records to develop accurate species distribution models. *Ecography*.