Manual

Part I: Installation

Dependent tools:

- ✓ MCL: https://micans.org/mcl/
- ✓ BLAST: http://blast.ncbi.nlm.nih.gov
- ✓ QGRS: http://bioinformatics.ramapo.edu/QGRS/downloads.php
- ✓ Mfold: http://unafold.rna.albany.edu/?q=mfold Or ViennaRNA: http://www.tbi.univie.ac.at/RNA/

Part II: Inputs

Library sequences: at least 1 initial library and 2 enriched libraries are needed, and the sequenced sequences (without primers) are formatted in .txt files. One .txt file for each round.

Example: R2.txt

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Primers: .txt file with primers sequences (first line: the forward primer; second line: the

backward primer) Example: Primer.txt

TTCAGCACTCCACGCATAGC CCTATGCGTGCTACCGTGAA

List file: which contain the round name of used libraries

Example: Rlist

R2

R3

R4

R6

R7

R8

Part III: Run

The whole procedure of SMART-Aptamer is consisting of 4 steps as follow:

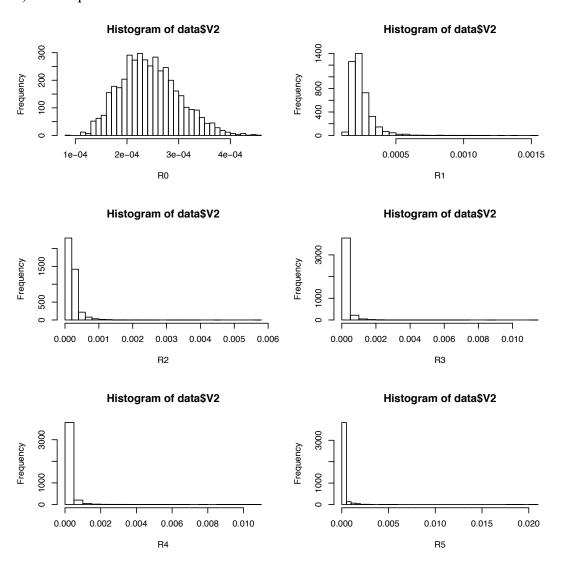
✓ Step 1: Calculate distributions of k-mer frequencies.

Example: Find_score -k 6 -t 35 -q 0.995 -c 'R0' -f icsv/list -i icsv -d /home/songjiajia/test_data/WenKu/my_software/SMART-Apta -o ocsv usages: Find score ...

- -k, the predefined length of k-mers (default: 6)
- -t, threads (default: 1)
- -q, the quantile that used to define the enriched k-mers (default: 0.995)
- -c, the control round (default R0)
- -f, library list (default Rlist)
- -i, input directory where the sequenced library txt files located (default: input
- -o, output directory (default: result)
- -d, the SMART-Apta directory

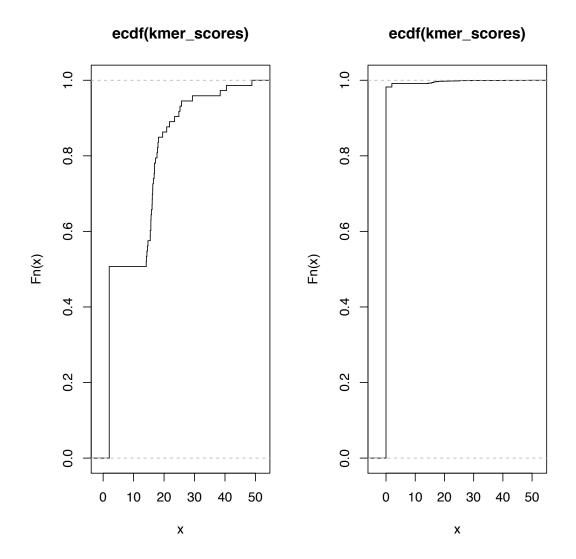
Important output files:

1) Kmer.pdf: can be used to understand the enrich status of each round



2) ecdf.pdf: used to find out the filter score cutoff value

for example: the following ecdf plots show that 10 can be a suitable cutoff value



✓ Step 2: find out the enriched motifs and filter data based on filter score Example: SMART-motif -k 6 -t 35 -f icsv/list -p 0.6 -s 2 -n 150000 -o ocsv2 -i icsv usages: SMART-motif ...

- -k, the predefined length of k-mers (default: 6)
- -t, threads (default: 1)
- -f, library list (default Rlist)
- -p, sequences with T bases > this cutoff are considered as t-rich sequences (0-1, default 0.6)
- -s, the filter scores cutoff value used to filter sequences based on motif enrichment status (only need to set one between -s and -n, default 10)
- -n, the total unique sequences should be left after filtering (set one between -f and -n, default 150000)
 - -o, output directory (default result)
 - -i, input directory (default input)

Important output files:

1) score kmers.txt: the enriched motifs and their kscore

- 2) scores.txt: the calculated filter score for each aptamer
- 3) all used uniq.fasta: the sequences that kept after data filtering
- 4) all_used_info.txt: the frequency information of the kept sequences
- ✓ Step 3: cluster aptamers based on BLAST-MCL strategy Example: SMART-cluster -t 35 -o ocsv -i 1.5 -e 0.05 -p primer.fa usages: SMART-cluster ...
 - -t, threads (default: 1)
 - -o, output directory (default result)
 - -i, inflation value for mcl algorithm (default 1.5)
 - -e, cutoff e-value after blast results (default 0.05)
 - -p, the primer file

Important output files:

- 1) aptamer clusters: the aptamer families
- ✓ Step 4: Multidimensional assessment including the calculation of Kscore, Sscore and Fscore
 - 1) Example: SMART-MDA-mfold -k 6 -t 35 -c 24 -n 0.15 -m 0.005 -o result usages: SMART-MDA-mfold ...
 - -k, the predefined length of k-mers (default: 6)
 - -t, threads (default: 1)
 - -c, rescale energy parameters to a temperature of temp C. (default 37)
 - -n, Na+ molar concentration (default 1.0)
 - -m, Mg++ molar concentration (default 0.0)
 - -o, output directory (default result)

Or

- 2) Example: SMART-MDA-RNAfold -k 6 -t 35 -c 37 -o ocsv usages: SMART-MDA-RNAfold ...
 - -k, the predefined length of k-mers (default: 6)
 - -t, threads (default: 1)
 - -c, rescale energy parameters to a temperature of temp C. (default 37)
 - -o, output directory (default result)

Important output files:

1) **result_aptamer.txt:** the final results, which contain the kscore, fscore, sscore and MDA-score and the representative sequence of each aptamer families. The outputs have been ranked by the MDA-score.