## Manual

#### Part I: Installation

Dependent tools:

- ✓ MCL: https://micans.org/mcl/
- ✓ BLAST: http://blast.ncbi.nlm.nih.gov
- ✓ QGRS: http://bioinformatics.ramapo.edu/QGRS/downloads.php
- ✓ Mfold: http://unafold.rna.albany.edu/?q=mfold Or ViennaRNA: http://www.tbi.univie.ac.at/RNA/

## Part II: Inputs

Library sequences: at least 1 initial library and 2 enriched libraries are needed, and the sequenced sequences (without primers) are formatted in .txt files. One .txt file for each round.

Example: R2.txt

. . . . . .

Primers: .txt file with primers sequences (first line: the forward primer; second line:

the backward primer)
Example: Primer.txt

TTCAGCACTCCACGCATAGC CCTATGCGTGCTACCGTGAA

List file: which contain the round name of used libraries

Example: Rlist

R4

R6

R9

R12

### Part III: Run

The whole procedure of SMART-Aptamer is consisting of 4 steps as follow:

✓ Step 1: Calculate distributions of k-mer frequencies.

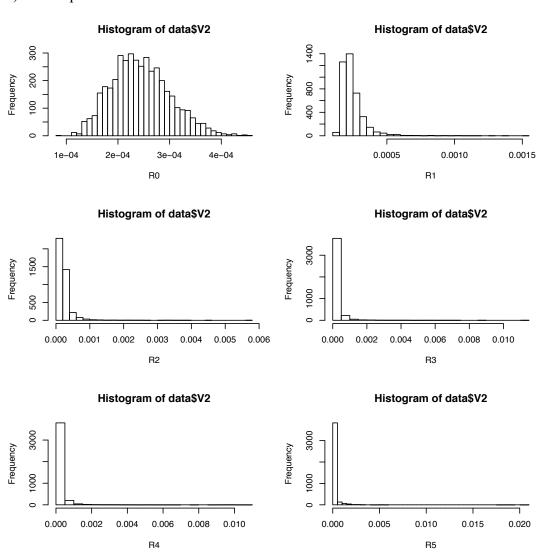
Example: Find\_score -k 6 -t 35 -q 0.995 -c 'R4' -f input/Rlist -i input -d /home/songjiajia/test\_data/WenKu/my\_software/SMART-Apta -o output

usages: Find\_score ...

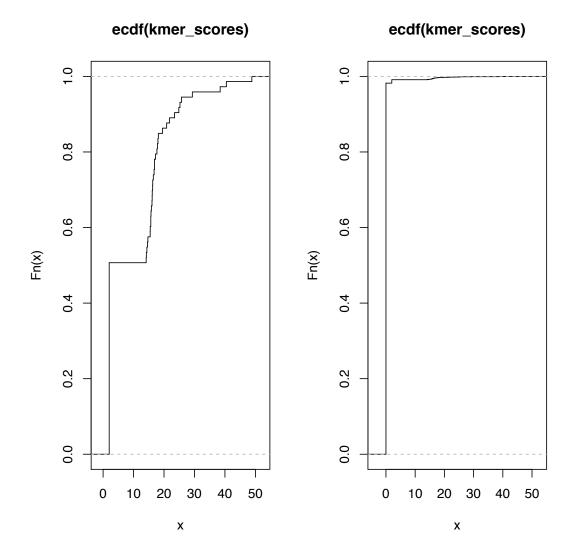
- -k, the predefined length of k-mers (default: 6)
- -t, threads (default: 1)
- -q, the quantile that used to define the enriched k-mers (default: 0.995)
- -c, the control round (default R0)
- -f, library list (default Rlist)
- -i, input directory where the sequenced library txt files located (default: input
- -o, output directory (default: result)
- -d, the SMART-Apta directory

Important output files:

1) Kmer.pdf: can be used to understand the enrich status of each round



2) ecdf.pdf: used to find out the filter score cutoff value for example: the following ecdf plots show that 10 can be a suitable cutoff value



✓ Step 2: find out the enriched motifs and filter data based on filter score

Example: SMART-motif -k 6 -t 35 -f input/Rlist -p 0.6 -s 2 -n 150000 -o output -i
input

usages: SMART-motif ...

- -k, the predefined length of k-mers (default: 6)
- -t, threads (default: 1)
- -f, library list (default Rlist)
- -p, sequences with T bases > this cutoff are considered as t-rich sequences (0-1, default 0.6)
- -s, the filter scores cutoff value used to filter sequences based on motif enrichment status (only need to set one between -s and -n, default 10)
- -n, the total unique sequences should be left after filtering (set one between -s and -n, default 150000)
  - -o, output directory (default result)
  - -i, input directory (default input)

Important output files:

1) score kmers.txt: the enriched motifs and their kscore

- 2) scores.txt: the calculated filter score for each aptamer
- 3) all used uniq.fasta: the sequences that kept after data filtering
- 4) all\_used\_info.txt: the frequency information of the kept sequences
- ✓ Step 3: cluster aptamers based on BLAST-MCL strategy

**Example: SMART-cluster -t 35 -o output -i 1.5 -e 0.05 -p input/primer.fa** usages: SMART-cluster ...

- -t, threads (default: 1)
- -o, output directory (default result)
- -i, inflation value for mcl algorithm (default 1.5)
- -e, cutoff e-value after blast results (default 0.05)
- -p, the primer file

Important output files:

- 1) aptamer\_clusters: the aptamer families
- ✓ Step 4: Multidimensional assessment including the calculation of Kscore, Sscore and Fscore

Example 1: SMART-MDA-RNAfold -k 6 -t 35 -c 25 -o output -i input -r 'R6:R9' -d '+:+' -f input/Rlist

Example 2: SMART-MDA-RNAfold -k 6 -t 35 -c 25 -o output -i input -r 'R12' -d '+' -f input/Rlist

# Example 3: SMART-MDA-RNAfold -k 6 -t 35 -c 25 -o output -i input -f input/Rlist

usages: SMART-MDA-RNAfold ...

- -k, the predefined length of k-mers (default: 6)
- -t, threads (default: 1)
- -c, rescale energy parameters to a temperature of temp C. (default

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- -o, output directory (default result)
- -i input directory (default input)
- -r rounds where the selection pressure has changed lead to a fixed family expansion/reduction trend; In case of multiple rounds, use ':' separate; For example, in this study ,we changed the selection pressure from 12<sup>th</sup> round SELEX, thus we set -r '12'
- -d Use the  $\pm$ -symbol to mark family expansion/reduction; In case of multiple rounds, use ':' separate
  - -f library list (default Rlist)

Important output files:

- 1) **result\_aptamer.txt:** the final results, which contain the kscore, fscore, sscore and MDA-score and the representative sequence of each aptamer families. The outputs have been ranked by the MDA-score.
- 2) family size rounds.txt: Contains the size/per round of each aptamer family.

3) conflict\_trends.txt: contain the kscore, fscore, sscore, MDA-score and the representative sequence of aptamer families (whose family expansion/ reduction trends conflict with the experimental design).