# Influence of Elevated CO<sub>2</sub> on Growth of Soybean Plants<sup>1</sup>

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#### ABSTRACT

Continuing use of fossil fuels in industrialized countries has created a need to understand growth responses of major crop species to elevated concentrations of CO2. An analysis was therefore undertaken of the growth of soybeans [Glycine max (L.) Merr. 'Bragg'] grown in pots in open top field chambers at six CO<sub>2</sub> concentrations ranging from 332  $\mu$ L L<sup>-1</sup> (ambient) to 910 μL L-1. Major growth response occurred with the first increments of added CO<sub>2</sub> with a maximum 66% increase in total vegetative dry matter at the 910  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> level. Dry weight increases were proportionate among vegetative plant parts, although the harvest index was found to decrease slightly. Greater absolute growth rates in elevated CO2 treatments were associated with greater rates of branch and internode elongation, leaf initiation, and leaf expansion. Yield increases represented greater seed numbers per plant rather than larger seeds. Percentage protein of seed decreased with CO2 enrichment. In the interval from day 5 to 2 weeks after planting, mean relative growth rate (RGR) increased asymptotically with CO<sub>2</sub> concentration. Of the two components of RGR, the mean net assimilation rate (NAR) increased dramatically and mean leaf area ratio (LAR) decreased. In the intervals from week 2 to 7 and from week 7 to 12, RGR became constant across CO2 treatments as the positive response of NAR and the negative response of LAR became less pronounced. Both RGR and NAR fell through the vegetative growth phase at each CO2 level. The adjustment in LAR resulted from a decrease in specific leaf area while leaf weight ratio remained unaffected by CO2.

Additional index words: Growth analysis, Growth characteristics, Morphology.

INCREASING concentrations of global atmospheric CO<sub>2</sub> resulting from massive combustion of fossil fuels (Stuiver, 1978) has generated interest in evaluating the long term implications for the growth and yield of crop plants. The CO<sub>2</sub> concentration influences the rate of dry matter production through both its role as regulator of stomatal aperture and its role as biochemical substrate for photosynthesis. But the relationship between photosynthesis, growth, and yield is not straightforward (Evans, 1975; Kramer, 1981); hence the need for evaluating the influence of CO<sub>2</sub> concentration on growth itself.

The purpose of this experiment was to characterize the growth and development responses of soybean [Glycine max (L.) Merr. 'Bragg'] to increasing atmospheric CO2 concentration in open top chambers (Rogers et al., 1983a, 1983b) under field conditions.

# MATERIALS AND METHODS

## **Exposure System**

The open top chamber (Rogers et al., 1983a, b) was a cylindrical (3 m diam × 2.4 m high) aluminum frame covered with PVC film "Roll-A Glass" with a 45 degree frustum (2.1m diam opening) attached at the top. The bottom half of the cylinder was double-walled, the inner wall perforated for uniform air distribution into the chamber. A 0.75 HP axial fan mounted in a sheet metal plenum box with a particulate filter supplied air at 1.06 m<sup>5</sup> s<sup>-1</sup>. Pure CO2 was injected into the air stream ahead of the fan to assist thorough mixing. A time-shared monitoring system sequentially sampled each of the 14 test plot atmospheres

for 50 s (30 s settling time and 20 s of rapid sampling). Air samples entered the 0.63 cm O.D. polyallomer, black Impolene (Gould-Imperial Eastman) delivery lines through a 4 L damping vessel with a glass wool filter. A bank of threeway solenoids controlled by a microcomputer directed the flow streams, one at a time, to the CO2 analyzer (Horiba PIR2000). Data were automatically acquired and the sampling manifold controlled by a Digital Equipment Co. (DEC) LSI-11 microcomputer. The most recent scan of CO<sub>2</sub> values was available in digital form to assist adjustment of flow rates into the chambers.

## **Experimental Procedure**

Soybean plants were grown from seed in pots in open top chambers (Rogers et al., 1983a, b) at six different atmospheric CO<sub>2</sub> concentrations: 332, 428, 534, 623, 772, and 910  $\mu$ L L<sup>-1</sup> (seasonal daytime mean). Plants were also grown in plots outside the chambers under ambient atmospheric conditions for comparison. There were two replicates of each treatment, or 14 plots in all. Plants were grown singly in 16.5 L pots containing 15 L of a 1:1:2 mixture by volume of sand, Metro-Mix 220 (W.R. Grace Co.), and a sandy clay loam soil. The 20 pots occupied the central portion of the 3m-diam chamber, leaving an access around the perimeter of the chamber area of about 44 cm. Therefore the plant density was approximately 5.7 plants m<sup>-2</sup>. The seeds were inoculated with a commercial Rhizobium preparation and were watered when tensiometer gauges (Irrometer Co., Riverside, Calif.) within pots reached 25 to 35 centibars (0.025 - 0.035 MPa). A N-free nutrient solution was applied weekly (Israel, 1981).

Destructive harvesting took place at 2, 7, 12, and 16 weeks after planting, corresponding to seedling, anthesis, early pod fill, and maturity stages. Leaf area was obtained photometrically. The plants were divided into roots, stems plus petioles, leaf blades, pod walls, and seeds. Branch length and branch number were recorded and the plants were dried at 55 ± 5°C for 72 h prior to weighing. Total weight in Table 1 represents vegetative growth, i.e., leaves, stems plus petioles, and roots. A more detailed breakdown of dry matter data is available (Rogers et al., 1982).

## Data Analysis

Growth characteristics were calculated for the intervals 5 days to 2 weeks, weeks 2 to 7 and weeks 7 to 12. For calculation of mean relative growth rate (RGR), mean net assimilation rate (NAR), and mean leaf area ratio (LAR) over the intervals, plants in each replicate plot at each CO2 level were ranked at the beginning and end of each interval in order of descending total dry weight and then paired,

<sup>1</sup> Joint contribution of the USDA-ARS and the North Carolina State Univ. Exp. Stn. Paper no. 8727 of the Journal Series of the North Carolina Agric. Res. Serv., Raleigh, NC 27650. Funding by the Dep. of Energy through interagency no. DEAI-01-81-ER 60001 to the USDA is acknowledged. Received 7 Mar. 1983.

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the largest with largest, etc. Calculations were then made for each pair of plants. These characteristics as well as leaf area ratio (LAR), specific leaf area (SLA), and leaf weight ratio (LWR) at the sampling points at weeks 5, 7, and 12 were calculated according to Kvet et al. (1971).

RGR = kg kg<sup>-1</sup> (plant) day<sup>-1</sup>
NAR = g m<sup>-2</sup> (leaf area) day<sup>-1</sup>
LAR = m<sup>2</sup> (leaf area) kg<sup>-1</sup> (plant);
LWR = kg (leaf) kg<sup>-1</sup> (plant);
SLA = m<sup>2</sup> (leaf area) kg<sup>-1</sup> (leaf).

For the growth characteristics in the first interval it was assumed that at 5 days all plants weighed 0.142 g dry weight (mean dry weight per seed at planting) and had cotyledonary photosynthetic area of 3.5 cm<sup>2</sup>.

Seeds were analyzed for protein, fiber and oil content by the Micro-Kjeldahl Method for protein N, Tecator Fibertec System for fiber and Tecator Refatec System for oil the (Assoc. of Official Anal. Chem., 1970).

The experimental design was a randomized complete block with two replicate blocks of seven CO<sub>2</sub> treatments. The experimental unit was the plot to which CO<sub>2</sub> levels were randomly assigned within each block. Either 10 or 20 plants were harvested at each sampling date, half from each

plot at each  $CO_2$  level. For the primary data in Tables 1 and 2 the treatment sum of squares was further partitioned to test for a chamber effect and linear and quadratic regressions on  $CO_2$  concentrations. Significance of the model terms ( $\alpha$ = 0.05) was tested with an F statistic from the analysis of variance with 1 and 6 df. The  $R^2$  measures the proportion of the treatment sum of squares accounted for by the chamber, linear, and quadratic effects.

#### RESULTS

# Growth, Morphology, and Yield.

Growth enhancement was observed for the soybean plants grown under elevated CO<sub>2</sub> as early as the first sampling at 2 weeks (Table 1). Although the first increments of CO<sub>2</sub> added to the ambient concentration seemed to result in greater increases in growth than subsequent CO<sub>2</sub> increments at all three sampling dates, the quadratic terms in the regression models for total dry weight or leaf area against CO<sub>2</sub> concentration were not statistically significant.

Plants harvested at 12 weeks after planting had

Table 1. Vegetative growth of Bragg soybeans grown in ambient plots and in open top field chambers at various CO2 concentrations.

co,	Week 2		Week 7		Week 12				
	Total dry wt	Leaf area	Total dry wt	Leaf area	Total dry wt	Leaf area	Height	Branch length	Branch no.
μL L <sup>*1</sup>	g	cm²	g·	cm <sup>2</sup>	g	cm²		cm ——	
331† 332 428 534 623 772 910	0.40 0.36 0.47 0.51 0.52 0.56 0.61	69 69 86 85 82 83	29.7 24.0 27.6 32.2 34.0 31.4 36.6	1 941 2 870 2 909 3 378 3 189 2 969 3 615	70.0 82.0 86.4 118.2 133.5 124.3 140.2	6 261 7 273 7 835 8 750 8 930 9 447 10 419	66 74 80 90 89 89	308 349 453 598 581 665 706	18.7 21.0 23.2 27.6 26.2 31.6 34.0
n S±‡ CV bo§ bchamber blinear bquadratic R <sup>2</sup>	10 0.02 14.0 0.40 ± 0.02 0.009 ± 0.03 0.37 ± 0.04 NS 0.96	10 4 15 69 ± 4 4.8 ± 5.2 35 ± 9 NS 0.74	10 2.2 15.6 19.7 ± 2.2 6.3 ± 2.7 18.0 ± 4.6 NS 0.88	10 227.35 17.5 2 982 NS NS NS NS	20 9.64 14.2 70.6 ± 7.0 17.3 ± 8.5 100 ± 10 NS 0.87	20 179.9 14.8 6 261 ± 181.6 1 132.8 ± 220.8¶ 5 151 ± 377 NS 0.99	10 2.7 9.9 64.3 ± 2.0 12.3 ± 2.3 31.0 ± 3.9 NS 0.95	10 39.0 16.2 308.1 ± 27.3 51.9 ± 41.2 1 140 ± 240 0.96 ± 0.37 0.99	10 1.75 14.8 18.7 ± 1.3 2.7 ± 1.6 22.0 ± 2.9 NS 0.98

<sup>†</sup> Ambient plot (no chamber).

<sup>¶</sup> Significant F (0.95 level) for chamber effect.

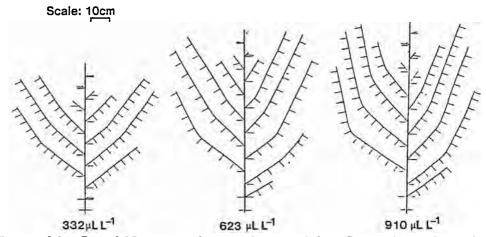


Fig. 1. Schematic diagram of the effects of CO<sub>2</sub> concentration on soybean morphology. Center axis = mainstem; horizontal lines = cotyledonary, primary and mainstem leaves; diagonal lines = secondary branches; lines normal to these = leaves.

<sup>‡</sup> Sz and CV from ANOVA.

<sup>§</sup> Curves were fitted against CO<sub>2</sub> added to ambient concentration (332 ppm). Note: linear and quadratic coefficients and their standard errors should be multiplied by 10<sup>-3</sup>.

Table 2. Reproductive growth and seed composition for Bragg soybeans grown in ambient plots and in open top field chambers at various CO<sub>2</sub> concentrations.

CO,	Pod dry wt per plant	Pod no. per plant .	Harvest index	Seed oil	Seed protein	Seed fiber
μLL-'	g					
331†	55.6	117	0.54	16.5	34.2	9.4
332	73.2	126	0.52	16.8	34.0	9.2
428	79.9	138	0.49	17.5	32.6	8.6
534	93.6	169	0.48	16.8	31.8	10.3
623	95.0	170	0.47	18.1	32.1	9.3
772	96.7	168	0.46	18.0	30.7	9.6
910	101.1	185	0	17.9	30.4	9.4
n	20	20	20	5	4	4
S <sub>∓</sub> ‡ CV	6.3	18.7	0.01	0.75	0.38	1.02
CV	11.1	11.6	10.30	0.32	0.76	1.78
B <sub>o</sub> §	$59.5 \pm 4.6$	$117.0 \pm 6.7$	$0.54 \pm 0.011$	17.34	$34.2 \pm 0.4$	9.34
bchamber	$21.9 \pm 5.4$	$13.6 \pm 16.0$	$-0.032 \pm 0.012$	NS	$-0.71 \pm 0.5$	NS
blinear	$46.1 \pm 9.2$	$121.3 \pm 27.4$	$-0.10 \pm 0.02$	NS	$-5.8 \pm 0.8$	NS
bquadratic	NS	NS	NS	NS	NS	NS
Ri	0.97	0.95	0.97	NS	0.96	NS

† Ambient plot (no chamber). ‡ S<sub>x</sub> and CV from ANOVA.

¶ Significant F (0.95 level) for chamber effect.

attained their maximum values for height and branch elongation and had ceased to initiate any new leaves or branches. Comparison of gross morphologies of 12 week-old plants grown in ambient, 623 and 910  $\mu L L^{-1} CO_2$  (Fig. 1 and Table 1) illustrates the impact of elevated CO2 on the form of the mature determinate soybean plant. Plants grown in 910 μL L-1 CO<sub>2</sub> initiated at least 2 more mainstem nodes, 30 more leaves, and 13 more (primary plus secondary) branches than did plants in unenriched treatments (Fig. 1 and Table 1). However, mainstem height increments in higher CO2 concentrations (Table 1) resulted from stimulation of internode elongation as well as from greater node numbers. The mean internode length of the 910  $\mu$ L L<sup>-1</sup> plants was 6.3 cm compared with 5.5 cm in 332  $\mu$ L L<sup>-1</sup> plants. Individual leaf expansion was slightly more rapid in the 3 highest CO2 concentrations, requiring 6 to 7 days compared with 7 to 8 days in the lower CO<sub>2</sub> atmospheres (data not shown; see Rogers et al., 1982).

Reproductive yield per plant was also enhanced in high CO<sub>2</sub> atmospheres (Table 2). The increases in seed yield per plant with CO<sub>2</sub> enrichment were due to greater numbers of pods rather than greater weight per pod. Harvest index (seed dry weight/total top dry weight) decreased steadily with increasing CO<sub>2</sub>. Seed protein as a percentage of seed dry weight decreased with increasing CO<sub>2</sub>, although total seed protein per plant increased from 16.2 g at 332 µL L<sup>-1</sup> to 18.8 g at 910 µL L<sup>-1</sup> (Table 2). A trend towards increased oil concentration with increasing CO<sub>2</sub> was not statistically significant, and seed fiber was apparently unaffected (Table 2). Germination tests revealed no significant differences in germination or early seedling growth due to mother plant pretreatment with CO<sub>2</sub> enrichment.

### Analysis of Growth

The RGR and its two component growth characteristics, the NAR and the LAR were plotted for the three vegetative growth intervals in Fig. 2 and 3. By

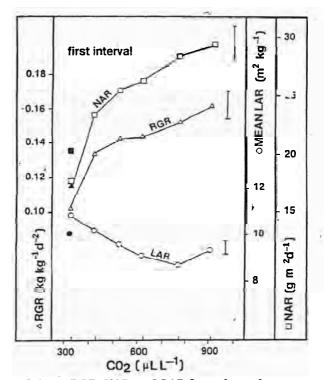


Fig. 2. Mean RGR, NAR, and LAR for soybean plants grown in ambient plots (closed symbols) and open top field chambers (open symbols) at various CO<sub>2</sub> concentrations during the interval day 5 to 2 weeks. Vertical bars denote one standard error.

2 weeks LAR had increased much more in the low  $CO_2$  treatments than in the high  $CO_2$  treatments so that the arithmetic average LAR for the interval was inversely related to  $CO_2$  concentration (Fig. 2). The LAR increased rapidly in the second interval (weeks 2–7) and then decreased in the third (weeks 7–12) but the inverse relationship to  $CO_2$  concentration was maintained, becoming less pronounced with time. The NAR increased significantly with  $CO_2$  concentration

Curves were fitted against CO, added to ambient concentration (332 ppm). Note: linear and quadratic coefficients and their standard errors should be multiplied by 10-3.

in the first interval (note the difference in scale for NAR between Fig. 2 and 3), increasing from 17.6 g m<sup>-2</sup>day<sup>-1</sup> at 332  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> to 29.4 g m<sup>-2</sup>day<sup>-1</sup> at 910  $\mu$ L L<sup>-1</sup>, a 73% increase. The mean RGR, the product of NAR and LAR, increased from 0.10 to 0.16 or 58%, over the same CO<sub>2</sub> concentration range. While NAR at all CO<sub>2</sub> levels adjusted downwards in the second and third intervals, the decrease with time was greater in the high CO<sub>2</sub> atmospheres so that by

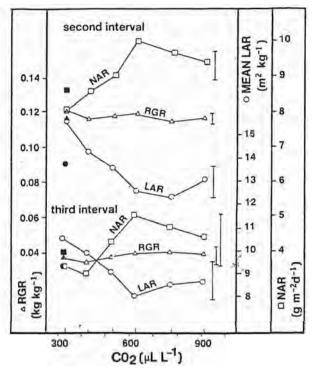


Fig. 3. Mean RGR, NAR, and LAR for soybean plants grown in ambient plots (closed symbols) and open top field chambers (open symbols) at various CO<sub>2</sub> concentrations during the intervals weeks 2 to 7 and weeks 7 to 12. Vertical bars denote one standard error.

the third interval the CO<sub>2</sub> effect was no longer statistically significant. Thus the RGR became constant across CO<sub>2</sub> treatments for most of the vegetative growth phase due to changes in both LAR and NAR.

The adjustment in LAR occurred through a change in SLA rather than through a change in relative allocation of dry matter to leaves, or LWR. In Fig. 4, data for weeks 2, 7, and 12 show the LWR, although falling with time, remained essentially constant with CO<sub>2</sub> concentration, whereas SLA fell with both time and CO<sub>2</sub> concentration. Since LAR is the product of the other 2 characteristics, the LAR curve assumed the shape of the SLA curve.

In general, the effect of CO<sub>2</sub> on all the growth characteristics diminished as CO<sub>2</sub> concentration increased.

#### DISCUSSION

We observed a 66% increase in total vegetative dry matter at the 910  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> level based on regression values (Table 1). This stimulation of growth after 12 weeks in elevated CO2 atmospheres is in general agreement with results from other CO2 enrichment studies with soybeans. Patterson and Flint (1980) grew soybeans for 45 days in growth cabinets and reported 24 and 74% increases at 600 and 1,000  $\mu$ L L<sup>-1</sup> CO<sub>2</sub>, respectively. Cooper and Brun (1967) obtained 61 and 71% increases for two varieties after 38 days in 350 and 1350  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> in growth cabinets within a greenhouse. Hardy and Havelka (1975) reported a 49% increase in total dry matter of soybeans grown in CO<sub>2</sub> enriched atmospheres (800-1200  $\mu$ L L<sup>-1</sup>) during reproductive phase. Also Hardman and Brun (1971), who provide the only other season-long CO<sub>2</sub> enrichment study, show a 64% increase in total dry weight of plants grown in the ground at 1200  $\mu$ L L<sup>-1</sup> vs. 425  $\mu$ L L<sup>-1</sup>. Of the published studies of soybean growth under CO2 enrichment, our results depart substantially only from those of Mauney et al. (1978) who report a 380% stimulation of total dry matter production at 630  $\mu$ L L<sup>-1</sup> vs. 330  $\mu$ L L<sup>-1</sup>.

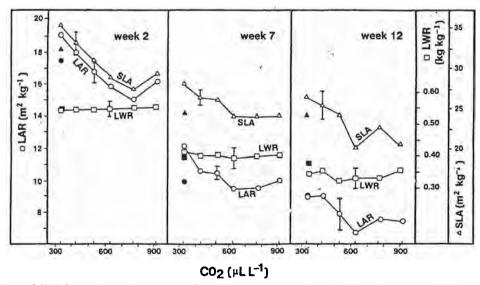


Fig. 4. LAR, LWR, and SLA for soybean plants grown in ambient plots (closed symbols) and open top field chambers (open symbols) at various CO<sub>2</sub> concentrations at 2, 7, and 12 weeks. Vertical bars denote one standard error.

The increased size of the vegetative plant in high CO<sub>2</sub> atmospheres was primarily due to higher net assimilation rate (see below) and higher absolute growth rate. The higher growth rate was due to higher rate of leaf initiation, also observed for soybeans in high CO<sub>2</sub> growth cabinets by Hofstra and Hesketh (1975). Greater node numbers represent both additional sites for pod initiation and additional leaf area to support pod development. However, a slight delay in anthesis and a slower transition from vegetative to reproductive phase were also observed in higher CO<sub>2</sub> treatments. Since flowering puts an upper limit on vegetative growth of determinate soy-beans, potential capacity for reproductive development (i.e., leaf area) was increased by the slower transition from vegetative to reproductive growth as well (Tables 1 and 2). Hesketh and Hellmers (1973) have also reported delayed flowering in four other crop species under CO2-enriched growth chamber conditions. Although Cooper and Brun (1967) did not observe any effect of CO<sub>2</sub> enrichment on flowering of soybean, the varieties used in their study were indeterminate.

Seed protein values in our experiment, which decreased with increasing atmospheric CO2 (Table 2), are at variance with those of Hardy and Havelka (1975) who reported that CO<sub>2</sub> enrichment had no effect on pod N concentration. The N concentration of leaves sampled at week 12 decreased with increasing CO2 (data not shown), a finding in agreement with observations of Hardy and Havelka (1975). Concentrations of P, K, and Ca also decreased in this foliar tissue, but Mg, Fe, Mn, Zn, Cu, and B concentrations were unaffected (see Rogers et al., 1982).

In this study of soybean growth in pots under field chamber conditions the effectiveness of successive increments of added  $CO_2$  on the growth characteristics (RGR, NAR, LAR, and SLA) at any interval appeared to diminish at the higher CO<sub>2</sub> concentrations (Fig. 2, 3, and 4). In other growth chamber studies where the effects of multiple CO<sub>2</sub> concentrations on growth characteristics were observed, asymptotic responses to CO<sub>2</sub> enrichment were reported for wheat (Triticum aestivum L.) (Neales and Nicholls, 1978) and tomato (Lycopersicon esculentum; Hurd and Thornley, 1974), and NAR and RGR were smaller at 900  $\mu$ L  $L^{-1}$  than at 600  $\mu$ L  $L^{-1}$  for asters Callistephus chinensis (Hughes and Freeman, 1967; Hughes and Cockshull, 1968). Likewise, the influence of elevated CO2 on the changes with time of these characteristics resembles the results of the above studies for wheat, tomato, and asters as well as for soybeans under green-house conditions (Mauney et al., 1978) and growth cabinet conditions (Patterson and Flint, 1980). As in these other studies, the highest values for the growth characteristics were obtained during early development, followed by a gradual decline termed ontogenetic drift (Kvet et al., 1971).

Very early in development, when differences in LAR across CO<sub>2</sub> treatments were relatively small, changes in RGR were determined largely by its other component, NAR, which was initially greatly stimulated in high CO<sub>2</sub> concentrations (Fig. 1). In the second and third intervals, however, RGR appeared unaffected by CO<sub>2</sub>. In studies where multiple harvests have permitted estimation of instantaneous values for

the growth characteristics, RGR and/or NAR have been shown to be increased by high CO2 initially, only to decrease as fast or faster than for control plants. Such a pattern was described by Neales and Nicholls (1978) for wheat, by Hurd (1968) for tomato, by Hughes and Cockshull (1969) for aster and by Thomas et al. (1975) for tobacco (Nicotiana tobaccum L.). In the wheat study (Neales and Nicholls, 1978), the analysis of variance for all data showed no CO2 effect per se, but a highly significant  $CO_2 \times$  time interaction. Such a pattern may underlie the apparently constant RGR calculated over rather long intervals (35 days) in this study. The initial increase and subsequent steeper decline in RGR and NAR observed for high CO<sub>2</sub> plants have also been observed for plants treated with high light intensity (Hurd, 1968) or greater daily light integral (Hurd and Thornley, 1974). thus the adjustments in growth characteristics have tended to equalize RGR, at least for CO2 and light treatments,

as growth diminishes with time.

This equalizing of RGR occurred through adjustment of both NAR and LAR. The steeper drop in NAR with time in CO<sub>2</sub>-enriched treatments may have been due to a shading effect, since total leaf area per plant was greater in high CO<sub>2</sub> grown plants than in low CO<sub>2</sub> grown plants (Table 1). Possibly there were adjustments in CO<sub>2</sub> uptake rates in the upper canopy as well, an area which has not been sufficiently investigated. The adjustment in leaf area ratio has been reported for the tomato, wheat, and aster studies referenced above and appears to be a widespread response of plants, raising questions as to the partitioning of assimilate towards leaf area vs. leaf thickness. The decrease in SLA of soybeans with increasing atmospheric CO<sub>2</sub> concentration has been related to additional cell layers of the leaf (Hofstra and Hesketh, 1975). This has also been observed for soybean, sweetgum (Liquidambar styraciflua L.), and pine (Pinus taeda L.) by Thomas and Harvey (1983). These results suggest that as soybean plants develop, two important mechanisms serve to modify or buffer an initially striking growth response to elévated CO<sub>2</sub>, 1) a dampening of the stimulation of NAR through self shading (and perhaps through a more direct effect on photosynthesis) and 2) a downshift in the LAR resulting from reduced SLA and altered leaf anatomy.

#### **ACKNOWLEDGMENTS**

Acknowledgments are gratefully extended to D. T. Patterson and Nasser Sionit for helpful discussion, to J C. Rawlings for statistical support, and to Donna Li of the Virginia State Univ., Food Quality and Safety Lab. for need analyses.

# REFERENCES

- 1. Association of Official Analytical Chemists. 1970. Official methods of analysis. 11th Ed. Assoc. of Official Anal. Chem., Washington, D.C.
- 2. Cooper, R.L., and W.A. Brun. 1967. Response of soybeans to a carbon dioxide enriched atmosphere. Crop Sci. 7:455-457. Evans, L.T. (ed.) 1975. Crop physiology. Cambridge Univ.

Press, London, England.

4. Hardman, L.L., and W.A. Brun. 1971. Effect of atmospheric carbon dioxide enrichment at different developmental stages

- on growth and yield components of soybeans. Crop Sci. 11:886–888.
- 5. Hardy, R.W.F., and U.D. Havelka. 1975. Photosynthate as a major factor limiting nitrogen fixation by field-grown legumes with emphasis on soybeans. p. 421-439. In P. Nutman (ed.) Symbiotic nitrogen fixation in plants. Int. Biological Programme Series, vol. 7. Cambridge Univ. Press, London, England.
- 6. Hesketh, J.D., and H. Hellmers. 1973. Floral initiation in four plant species growing in CO2-enriched air. Environ. Control in Biol. 11:51–53.
- 7. Hofstra, G., and J.D. Hesketh. 1975. The effects of temperature and CO2 enrichment on photosynthesis in soybean. p. 71-80. In R. Marcelle (ed.) Environmental and biological control of photosynthesis. Dr. W. Junk, The Hague, The Netherlands.
- 8. Hughes, A.P., and K.E. Cockshull. 1968. Effects of carbon dioxide concentration on the growth of Callistephus chinensis
- cultivar Johannistag. Ann. Bot. 33:351-365.

  —, and P.R. Freeman. 1967. Growth analysis using fre-
- quent small harvests. J. Appl. Ecol. 4:553-560.

  10. Hurd, R.G. 1968. Effects of CO<sub>2</sub> enrichment on the growth of the young tomato plants in low light. Ann. Bot. 32:531-
- , and J.H.M. Thornley. 1974. An analysis of the growth of young tomato plants in water culture at different light integrals and CO2 concentrations. I. Physiological aspects. Ann. Bot. 38:375-388.
- 12. Israel, D.W. 1981. Cultivar and Rhizobium strain effects on nitrogen fixation and remobilization by soybeans. Agron. J.
- 13. Kramer, P.J. 1981. Carbon dioxide concentration, photosynthesis, and dry matter production. BioScience 31:29-33.

- 14. Kvet, J., J.P. Ondok, J. Necas, and P.G. Jarvis. Methods of growth analysis. 1971. Catsky and P.G. Jarvis (eds.) Plant photosynthetic production manual of methods. Dr. W. Junk, The Haue, The Netherlands.
- 15. Mauney, J.R., K.E. Fry, and G. Guinn. 1978. Relationship of photosynthetic rate to growth and fruiting of cotton, soybean, sorghum, and sunflower. Crop Sci. 18:259-263.
- 16. Neales, T.F., and A.O. Nicholls. 1978. Growth responses of young wheat plants to a range of ambient CO2 levels. Aust. . Plant Physiol. 5:45-59.
- 17. Patterson, D.T., and E.P. Flint. 1980. Potential effects of global atmospheric CO2 enrichment on the growth and competitiveness of C<sub>3</sub> and C<sub>4</sub> weed and crop plants. Weed Sci. 28:71-
- Rogers, H.H., R.D. Beck, G.E. Bingham, J.D. Cure, J.M. Davis, W.W. Heck, J.O. Rawlings, A.J. Riordan, N. Sionit, J.M. Smith, and J.F. Thomas. 1982. 1981 Report: Field studies of plant responses to elevated carbon dioxide levels. Prepared for Dep.
- of Energy, Washington, D.C.

  —, G.E. Bingham, J.D. Cure, J.M. Smith, and K.A. Surano.

  1983a. Responses of selected plant species to elevated CO<sub>2</sub> in the field. J. Environ. Qual. 12:569-574.

  ——, W.W. Heck, and A.S. Heagle. 1983b. A field technique
- for the study of plant responses to elevated carbon dioxide concentration. J. Air Pollut. Control Assoc. 33:42-44.
  21. Stuiver, M. 1978. Atmospheric carbon dioxide and carbon
- reservoir changes. Science 199:253-258.
- 22. Thomas, J.F., and C.N. Harvey. 1983. Leaf anatomy of four species grown under continuous long-term CO2 enrichment. Bot. Gaz. 144:303-309.
- ----, C.D. Raper, Jr., C.E. Anderson, and R. J. Downs. 1975. Growth of young tobacco plants as affected by carbon dioxide and nutrient variables. Agron. J. 67:685-689.