

Supporting Information

For

Efficient Chiral Monophosphorus Ligands for Asymmetric Suzuki-Miyaura Coupling Reactions

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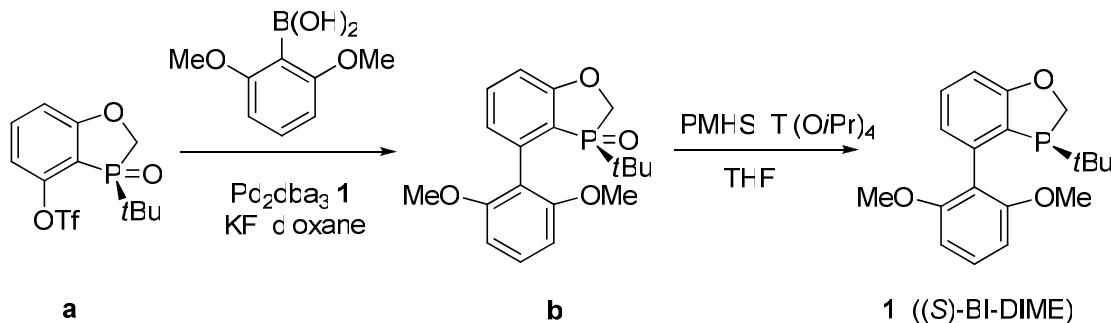
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1. General Methods. All reactions were carried out under a nitrogen atmosphere unless otherwise specified. THF (<0.02% water content), CH₂Cl₂, hexane, *n*-butanol, dioxane and toluene were purchased from J. T. Baker and used directly without further purifications. Commercialized reagents were used without further purifications. Chiral triflate **2** were prepared according to our reported procedure.^[1] Tf = trifluoromethylsulfonate, dba = dibenzylideneacetone, Cy = cyclohexyl, S-Phos = 2-dicyclohexylphosphino-2',6'-dimethoxybiphenyl, PMHS = polymethylhydrosiloxane,

¹H, ³¹P and ¹³C NMR data were recorded on a Bruker-Biospin DRX500 or DRX400 NMR Spectrometer with CDCl₃ or CD₂Cl₂ as the solvent. ¹H shifts were referenced to CDCl₃ at 7.26 ppm and CD₂Cl₂ at 5.32 ppm. ³¹P shifts were referenced to 85% H₃PO₄ in D₂O at 0.0 ppm as external standard and obtained with ¹H decoupling. ¹³C shifts were referenced to CDCl₃ at 77 ppm and CD₂Cl₂ at 54 ppm and obtained with ¹H decoupling. MS was measured on Agilent 1100 Series LC/MSD mass spectrometer. Chiral HPLC analyses were performed on a Hewlett-Packard 1100 system using a Chiralcel OD-H or Chiraldak AD-H column. Racemic compounds were prepared by reaction using S-Phos or racemic BI-DIME^[2] as the ligand at 65 °C. The optical rotations were recorded on a Rudolph Research Automatic Polarimeter.

2. Synthetic procedures for ligand 1a-d

a. Synthesis of ligand 1 ((S)-BI-DIME)

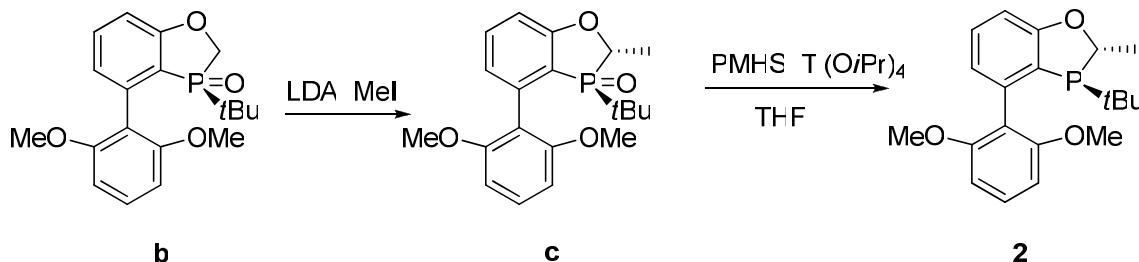


The synthesis of ligand **1 ((S)-BI-DIME)** was carried out from chiral triflate¹ **a** through a Suzuki coupling with 2,6-dimethoxyphenylboronic acid to form **b** followed by a stereospecific reduction of the phosphane oxide **b** mediated by PMHS/Ti(O*i*Pr)₄. The synthetic procedures were similar to those for racemic BI-DIME published in our previous paper.²

Phosphane oxide **b**: white solid; >99% ee; $[\alpha]_D^{20} = -66.4^\circ$ (c = 0.64, CHCl₃); Chiral HPLC conditions: Chiralcel AD-H, heptane/*n*-propanol = 90:10, 1 mL/min, 25 °C, 4.93 min (*S*-enantiomer), 10.22 min (*R*-enantiomer, **b**).

1 ((S)-BI-DIME): white solid; >99% ee; $[\alpha]_D^{20} = -175.3^\circ$ (c = 0.3, CHCl₃); The enantiomeric excess was determined by converting to its phosphine oxide via stereospecific oxidation with H₂O₂ as the oxidant.

b. Synthesis of ligand 2

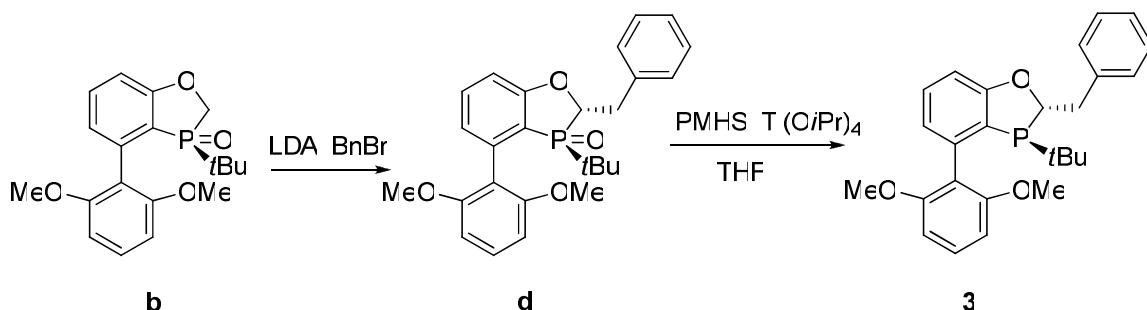


The synthesis of ligand **2** was carried out from chiral phosphane oxide **b** through methylation to form **c** followed by a stereospecific reduction of the phosphane oxide **c** under conditions of PMHS/Ti(O*i*Pr)₄. The synthetic procedures were similar to those for its racemic form published in our previous paper.³ The relative configuration between P chirality and C chirality was determined by NOE experiments.

Phosphane oxide **c**: white solid; $[\alpha]_D^{20} = -12.8^\circ$ ($c = 0.53$, CHCl₃).

Phosphane **2**: white solid; $[\alpha]_D^{20} = -121.6^\circ$ ($c = 0.55$, CHCl₃).

c: Synthesis of ligand 3

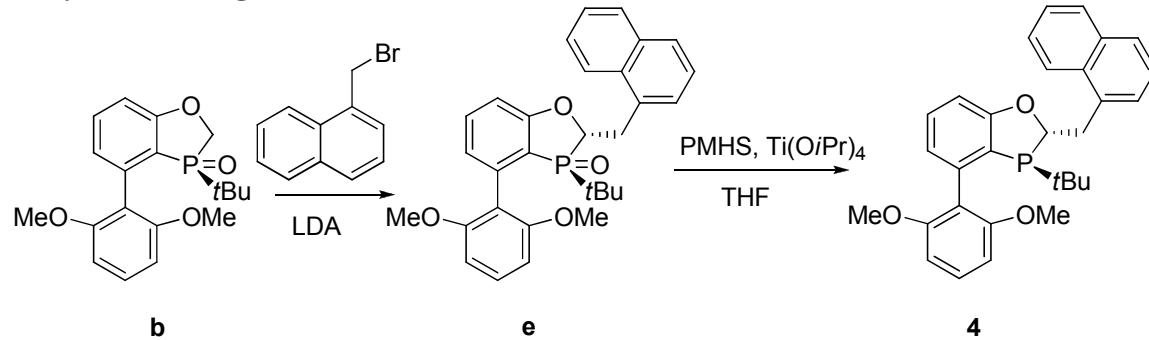


To a solution of (*R*)-3-*tert*-butyl-4-(2,6-dimethoxyphenyl)-2,3-dihydrobenzo[*d*][1,3]oxaphosphole oxide (**b**, 5.0 g, 14.4 mmol) in THF (50 mL) at -70 °C was charged LDA (9.6 mL, 1.8 M, 17.3 mmol, 1.2 equiv) over 5 min. The mixture was kept at -70 °C for 1 h. To the mixture was charged BnBr (2.96 g, 17.3 mmol, 1.2 equiv) over 5 min while controlling the reaction temperature below -60 °C. The resulting mixture was stirred at -70 °C for additional 1 h and then allowed to warm to rt over 2 h. Water (50 mL) and dichloromethane (50 mL) was charged. The dichloromethane layer was separated, washed with brine (50 mL), dried over sodium sulfate, and concentrated. The residue was purified by silica gel column chromatography (eluent: hexane to EtOAc) to provide the desired product **d** as white solid (5.0 g, 11.5 mmol, 80% yield). **d**: $[\alpha]_D^{20} = +55.9^\circ$ ($c = 0.42$, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.48 (t, $J = 7.8$ Hz, 1H), 7.20-7.40 (m, 6H), 6.90 (m, 2H), 6.67 (d, $J = 8.4$ Hz, 1H), 6.57 (d, $J = 8.3$ Hz, 1H), 4.54 (m, 1H), 2.91 (s, 3H), 2.95 (s, 3H), 3.10-3.30 (m, 2H), 0.83 (d, $J = 15.6$ Hz, 9H); ³¹P NMR (162 MHz, CDCl₃) δ 59.9; ¹³C NMR (100 MHz, CDCl₃) δ 163.5 (d, $J = 18.6$ Hz), 158.7, 157.3, 138.4 (d, $J = 4.7$ Hz), 138.3 (d, $J = 9.4$ Hz), 134.1 (d, $J = 1.6$ Hz), 129.8, 129.4,

128.5, 126.6, 124.9 (d, J = 8.3 Hz), 117.5, 114.5 (d, J = 89.9 Hz), 112.6 (d, J = 5.5 Hz), 104.5, 103.1, 75.0 (d, J = 60.4 Hz), 55.1, 56.4, 35.8, 33.3 (d, J = 70.8 Hz), 23.7; ESI-MS: m/z 437 [M + H]⁺.

To a solution of (2*S*,3*R*)-2-benzyl-3-*tert*-butyl-4-(2,6-dimethoxyphenyl)-2,3-dihydrobenzo[d][1,3]oxaphosphole oxide (**d**, 3.0 g, 6.87 mmol, 1.0 equiv) in THF (30 mL) at rt was added PMHS (3 g) and Ti(O*i*Pr)₄ (1.95 g, 6.87 mmol, 1.0 equiv). The mixture was stirred at reflux for 24 h and then distilled under reduced pressure to remove most THF. To the residue was charging carfully 30% NaOH solution (30 mL). Gas was generated during the addition. To the mixture at rt was charged Me-THF (30 mL). The resulting mixture was stirred at 60 °C for 0.5 h. The organic layer was separated and the most of the aqueous slurry was discarded. To the organic layer was charged water (30 mL). The aqueous layer was discarded. The organic layer was further washed with water (30 mL X2) and 5% NaCl solution (30 mL), and then concentrated at rt. The residue was purified by passing through a neutral alumina plug to provide **3** as white solid (2.5 g, 5.95 mmol, 87% yield). **3**: $[\alpha]_D^{20} = -16.2^\circ$ (c = 0.49, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.10-7.30 (m, 7H), 6.79 (m, 2H), 6.56 (d, J = 8.4 Hz, 1H), 6.49 (d, J = 8.4 Hz, 1H), 4.87 (t, J = 7.7 Hz, 1H), 3.71 (s, 3H), 3.61 (s, 3H), 3.00 (m, 1H), 2.77 (m, 1H), 0.58 (d, J = 12.0 Hz, 9H); ³¹P NMR (202 MHz, CDCl₃) δ 5.84; ¹³C NMR (125 MHz, CDCl₃) δ 162.6, 157.8, 157.0, 138.8 (d, J = 17.2 Hz), 138.5 (d, J = 10.0 Hz), 130.6, 129.6, 129.5, 129.0, 128.3, 126.4, 124.2 (d, J = 14.9 Hz), 123.8 (d, J = 4.1 Hz), 119.6, 109.9, 104.4, 103.6, 84.2 (d, J = 26.4 Hz), 55.8 (d, J = 1.8 Hz), 53.4, 41.4 (d, J = 28.1 Hz), 31.0 (d, J = 18.4 Hz), 26.6 (d, d, J = 14.3 Hz); ESI-MS: m/z 421 [M + H]⁺.

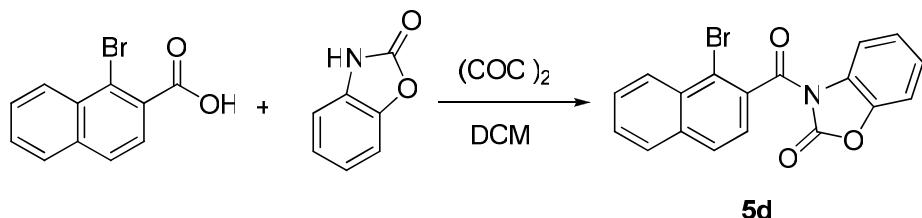
d: Synthesis of ligand 4



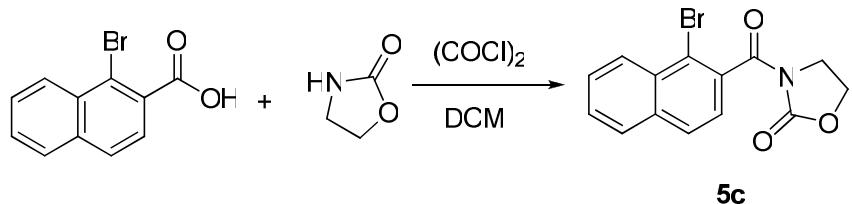
The synthesis of compound **e** from compound **b** was carried out following a procedure similar to the preparation of compound **d** for the synthesis of ligand **3**. **e**: 85% yield (white solid); $[\alpha]_D^{20} = +43.4^\circ$ (c = 0.35, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.06 (d, J = 7.8 Hz, 1H), 7.89 (d, J = 7.7 Hz, 1H), 7.79 (d, J = 8.2 Hz, 1H), 7.40-7.60 (m, 5H), 7.32 (t, J = 8.2 Hz, 1H), 6.90 (m, 2H), 6.69 (d, J = 8.4 Hz, 1H), 6.58 (d, J = 8.4 Hz, 1H), 4.68 (m, 1H), 3.87 (s, 3H), 3.82 (m, 1H), 3.71 (s, 3H), 3.55 (m, 1H), 0.82 (d, J = 16.0 Hz, 9H); ³¹P NMR (162 MHz, CDCl₃) δ 60.5; ¹³C NMR (100 MHz, CDCl₃) δ 163.5 (d, J = 18.8 Hz), 158.7, 157.4, 138.5 (d, J = 5.1 Hz), 134.2 (d, J = 1.5 Hz), 133.9, 131.9, 129.9, 128.9, 128.0, 127.5, 126.0, 125.6, 125.5, 125.0 (d, J = 8.3 Hz), 123.6, 117.4, 114.2 (d, J = 89.5 Hz), 112.7 (d, J = 5.5 Hz), 104.5, 103.0, 74.3 (d, J = 59.7 Hz), 56.1, 55.4, 35.4, 33.4 (d, J = 70.6 Hz), 23.6; ESI-MS: m/z 487 [M + H]⁺.

The synthesis of ligand **4** from compound **e** was carried out following a procedure similar to the preparation of ligand **3** from compound **d**. **4**: 83% yield (white solid); $[\alpha]_D^{20} = -28.6^\circ$ ($c = 0.29$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.94 (d, $J = 8.1$ Hz, 1H), 7.77 (d, $J = 7.7$ Hz, 1H), 7.67 (d, $J = 7.4$ Hz, 1H), 7.12-7.50 (m, 6H), 6.80 (m, 2H), 6.59 (d, $J = 8.2$ Hz, 1H), 6.50 (d, $J = 8.4$ Hz, 1H), 5.05 (t, $J = 7.6$ Hz, 1H), 3.76 (s, 3H), 3.62 (s, 3H), 3.45 (m, 1H), 3.25 (m, 1H), 0.55 (d, $J = 12.0$ Hz, 9H); ^{31}P NMR (162 MHz, CDCl_3) δ 7.5; ^{13}C NMR (100 MHz, CDCl_3) δ 162.6, 157.8, 157.0, 138.8 (d, $J = 17.0$ Hz), 134.6 (d, $J = 10.7$ Hz), 133.9, 132.1, 130.6, 129.1, 128.9, 127.9 (d, $J = 1.8$ Hz), 127.3, 125.9, 125.4, 125.3, 123.8 (d, $J = 4.1$ Hz), 123.8, 119.6, 110.1, 104.4, 103.7, 83.5 (d, $J = 26.2$ Hz), 55.9 (d, $J = 1.5$ Hz), 55.4, 38.5 (d, $J = 29.3$ Hz), 31.0 (d, $J = 18.4$ Hz), 26.6 (d, $J = 14.3$ Hz); ESI-MS: m/z 471 [M + H]⁺.

3. Procedures for substrate preparation



To a suspension of 1-bromo-2-naphthoic acid (1.0g, 4.0 mmol) in DCM (10 mL) was charged DMF (14.5 mg, 0.2 mmol, 0.05 equiv) followed by slow addition of oxaly chloride (0.56 g, 4.38 mmol, 1.1 equiv). Gas was generated during the addition and the resulting mixture was stirred at rt for 1 h. In a separated flask was charged benzo[d]oxazol-2(3H)-one (0.54 g, 3.98 mmol, 1.0 equiv), triethylamine (1.21 g, 11.9 mmol, 3.0 equiv) and DCM (10 mL). To the mixture at rt was charged the aforementioned solution. The resulting mixture was stirred at rt overnight. Water (20 mL) was charged to quench the reaction. The DCM layer was separated, washed with water (25 mL X 2) and brine (25 mL), dried over sodium sulfate, concentrated and solvent-switched to EtOAc (10 mL). The slurry was heated to 55 °C for 10 min and then cooled to rt. Filtration of the slurry provided the desired product **5d** as yellow solid (1.2 g, 3.3 mmol, 82% yield). **5d**: ^1H NMR (400 MHz, CD_2Cl_2) δ 8.39 (d, $J = 8.3$ Hz, 1H), 8.23 (m, 1H), 8.04 (d, $J = 8.4$ Hz, 1H), 8.00 (d, $J = 7.9$ Hz, 1H), 7.74 (m, 2H), 7.52 (d, $J = 8.4$ Hz, 1H), 7.41 (m, 2H), 7.35 (m, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 167.2, 150.5, 143.2, 135.4, 134.4, 132.0, 129.2, 129.1, 128.9, 128.8, 128.0, 127.9, 126.4, 125.5, 124.2, 120.5, 116.1, 110.7; ESI-MS: m/z 390 [M + Na]⁺.



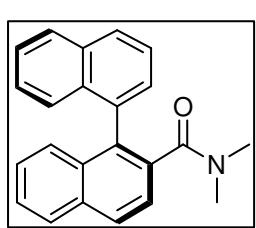
Preparation of **5c** was carried out according to a procedure similar to that for the synthesis of **5d**. **5c**: 85% yield; ^1H NMR (500 MHz, CDCl_3) δ 8.30 (d, $J = 8.5$ Hz, 1H),

7.89 (d, J = 8.4 Hz, 1H), 7.86 (d, J = 8.1 Hz, 1H), 7.65 (m, 1H), 7.58 (m, 1H), 7.34 (d, J = 8.4 Hz, 1H), 4.53 (t, J = 7.8 Hz, 2H), 4.28 (t, J = 8.3 Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.0, 152.1, 134.6, 134.1, 131.6, 128.4, 128.3, 128.2, 127.8, 127.5, 123.6, 119.8, 62.2, 42.5; ESI-MS: m/z 342 [M +Na] $^+$.

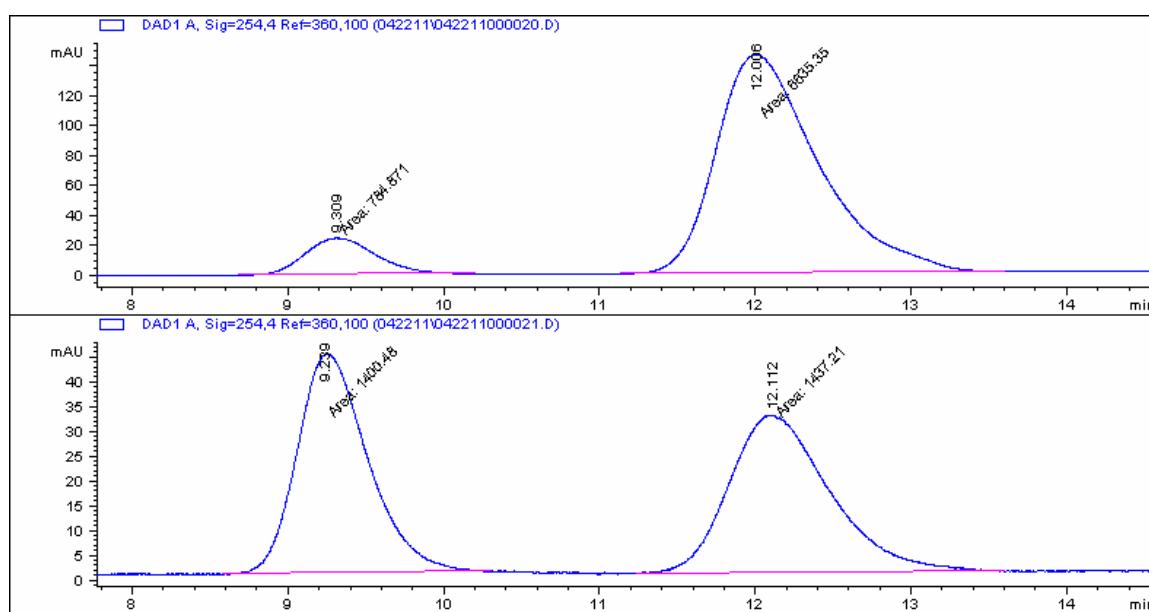
4. General procedure for asymmetric Suzuki-Miyaura couplings

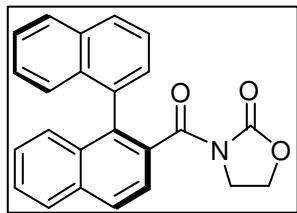
To a mixture of aryl bromide (1.0 mmol), arylboronic acid (2.0 mmol), potassium phosphate (3 mmol, 3 equiv), $\text{Pd}(\text{OAc})_2$ (0.01-0.05 mmol), and ligand **2** or **4** (0.012-0.06 mmol, Pd/ligand ratio = 1/1.2) was charged dry THF (2 mL). The mixture was stirred at room temperature or 40 °C under nitrogen for 4-36 h and then quenched by addition of 10% citric acid solution (4 mL). EtOAc (4 mL) was added and the EtOAc layer was separated, washed sequentially with water (4 mL) and brine (4 mL), dried over sodium sulfate, concentrated, and purified by silica gel column chromatography to provide biaryl products. Their ee's were determined by chiral HPLC on a chiralcel OD-H or chiralpak AD-H column.

5. Analytical data of asymmetric Suzuki-Miyaura coupling products

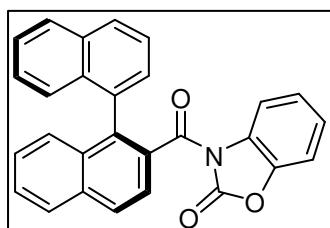
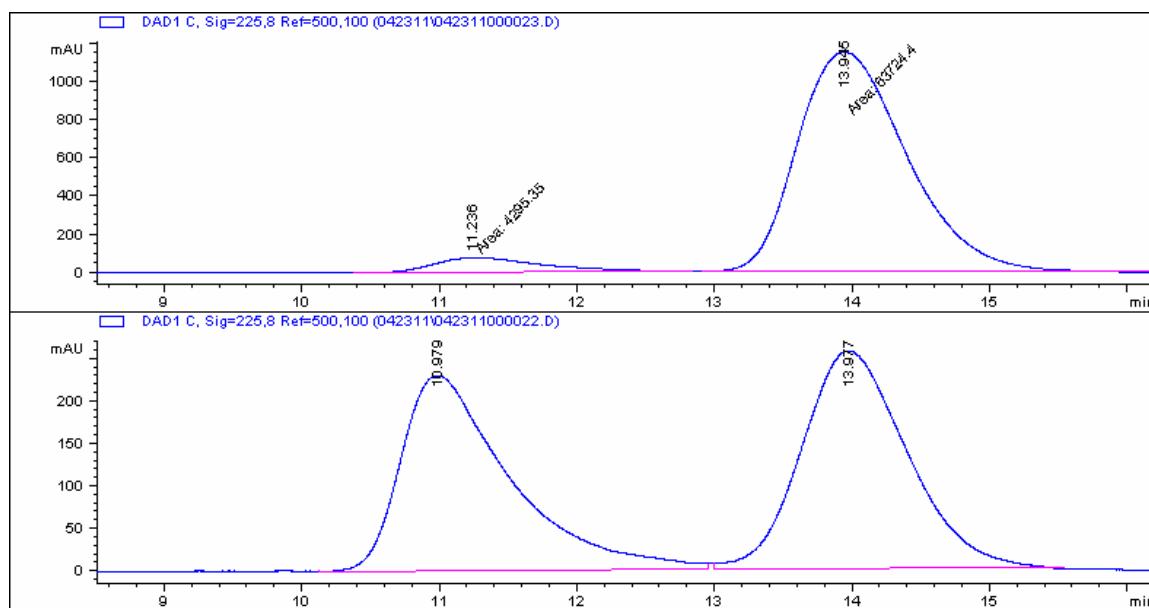


(*R*)-*N,N*-Dimethyl-1,1'-binaphthyl-2-carboxamide: white solid (95% yield); 79% ee; $[\alpha]_D^{20} = +66.0^\circ$ ($c = 0.80, \text{CHCl}_3$); ^1H NMR (500 MHz, CDCl_3) δ 8.02 (d, J = 7.3 Hz, 1H), 7.95 (br m, 3H), 7.40-7.75 (m, 5H), 7.29 (m, 4H), 2.20-3.10 (br m, 4H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.7, 135.1, 134.4, 133.4, 133.2, 132.8, 128.6, 128.4, 128.1, 127.1, 126.7, 126.4, 125.8, 123.8, 38.8, 34.3. ESI-MS: m/z 326 [M +H] $^+$. Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 254 nm, 9.30 min (*S*), 12.00 min (*R*).

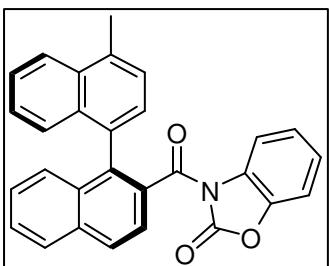
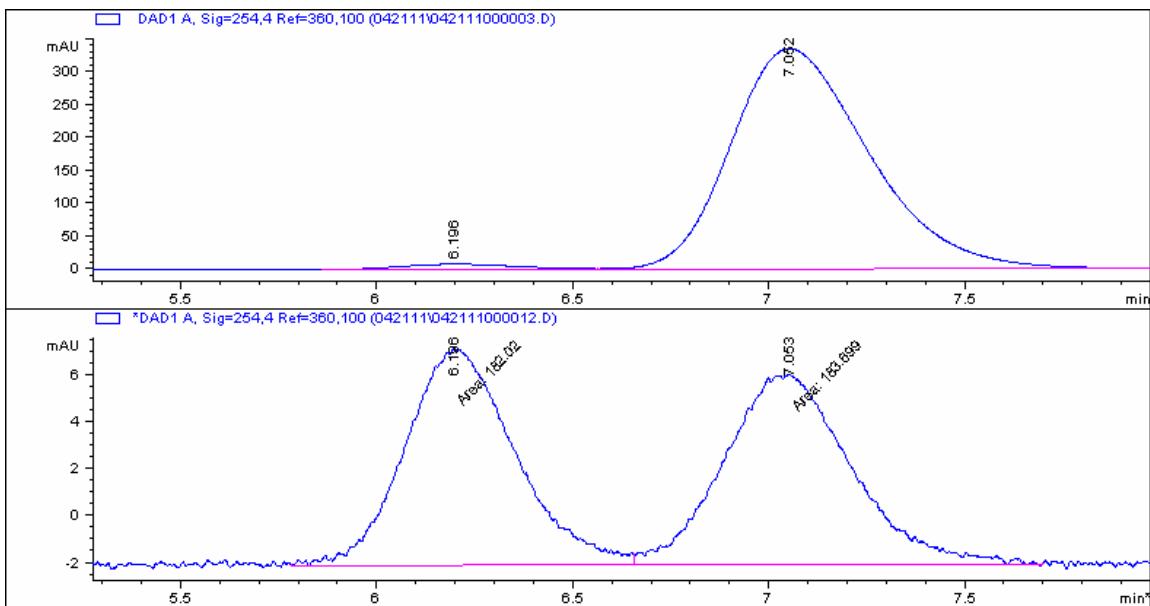




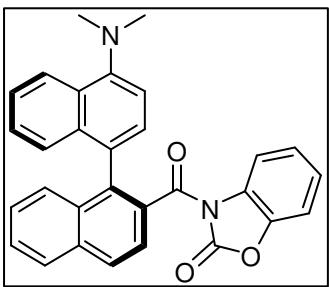
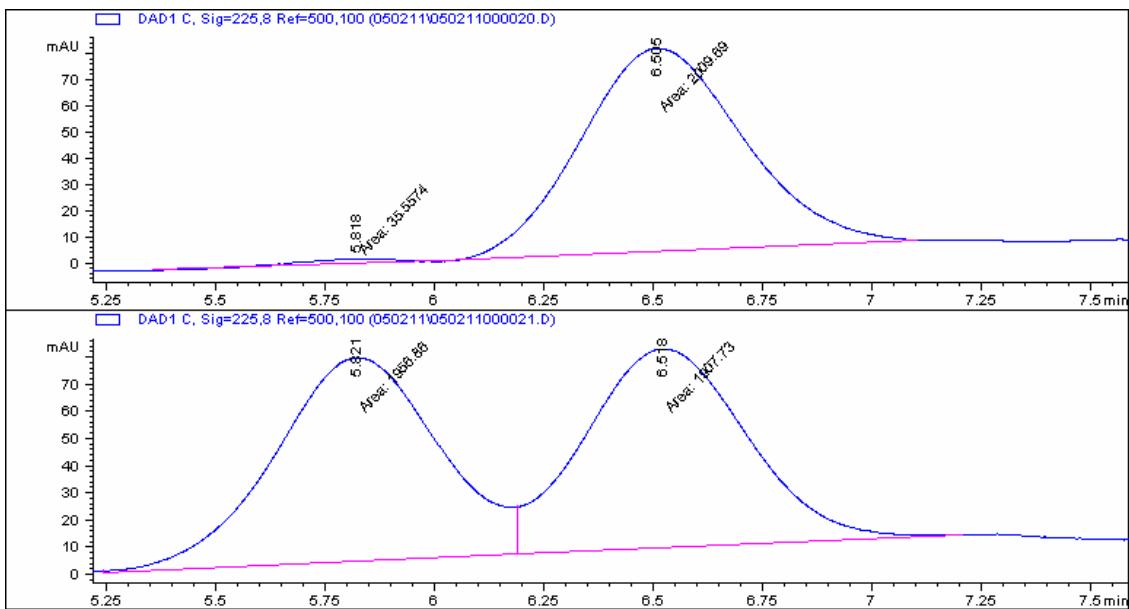
(R)-3-(1,1'-binaphthyl-2-carbonyl)oxazolidin-2-one: yellowish solid (96% yield). 87% ee; $[\alpha]_D^{20} = -61.6^\circ$ ($c = 0.55$, CHCl_3); ^1H NMR (400 MHz, CD_2Cl_2) δ 8.10 (d, $J = 8.4$ Hz, 1H), 7.95-8.07 (m, 3H), 7.55-7.70 (m, 3H), 7.52 (m, 2H), 7.38 (m, 4H), 3.99 (m, 2H), 3.63 (m, 1H), 3.46 (m, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 169.9, 153.2, 136.3, 135.6, 134.6, 133.9, 133.7, 133.2, 133.1, 129.0, 128.8, 128.5, 127.7, 127.6, 127.4, 127.3, 126.7, 125.7, 124.1, 62.9, 43.0; ESI-MS: m/z 390 [$M + \text{Na}$] $^+$. Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 85/15, 225 nm, 11.00 min (S), 13.98 min (R).



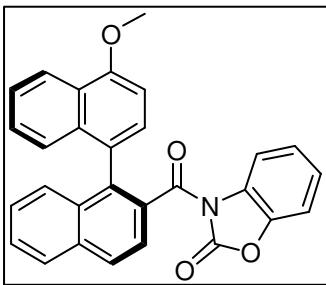
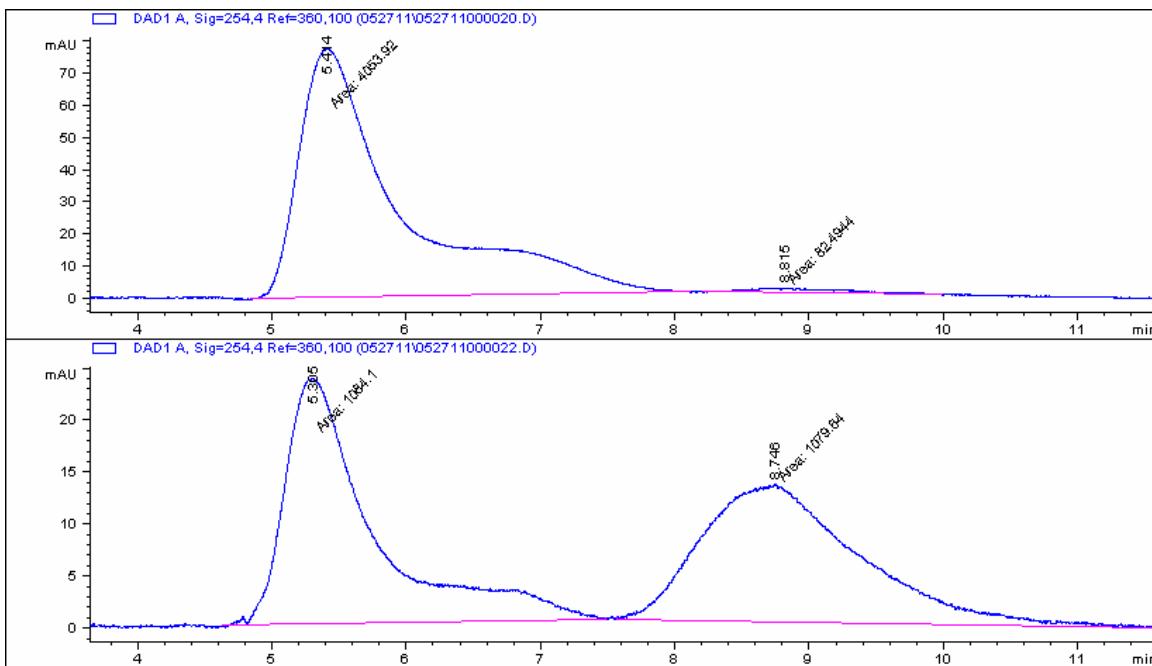
(R)-3-(1,1'-binaphthyl-2-carbonyl)benzo[d]oxazol-2(3H)-one: white solid (96% yield); 96% ee; $[\alpha]_D^{20} = -126.1^\circ$ ($c = 1.33$, CHCl_3); ^1H NMR (400 MHz, CD_2Cl_2) δ 8.11 (d, $J = 6.7$ Hz, 1H), 8.04 (d, $J = 6.4$ Hz, 1H), 7.79 (m, 2H), 7.72 (d, $J = 6.7$ Hz, 1H), 7.60 (m, 1H), 7.25-7.50 (m, 8H), 7.14 (t, $J = 6.0$ Hz, 1H), 7.04 (m, 2H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 168.5, 151.0, 142.9, 137.7, 135.1, 135.0, 133.9, 133.3, 133.2, 132.8, 129.0, 128.9, 128.8, 128.7, 128.6, 128.2, 128.0, 127.9, 127.6, 127.0, 126.8, 126.6, 125.6, 125.5, 124.9, 124.1, 114.8, 110.1. ESI-MS: m/z 438 [$M + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 254 nm, 6.19 min (S), 7.05 min (R).



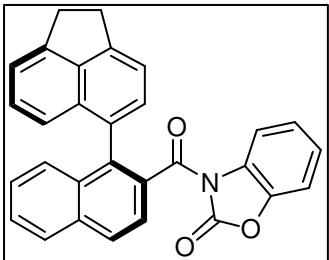
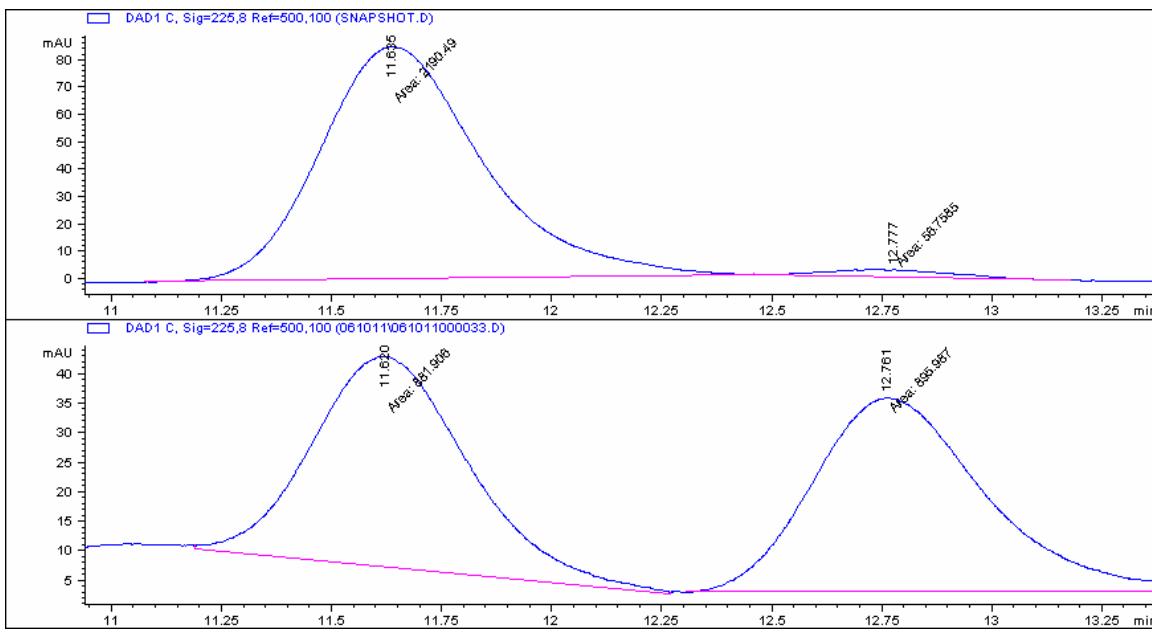
(*R*)-3-(4'-Methyl-1,1'-binaphthyl-2-carbonyl)benzo[d]oxazol-2(3H)-one: white solid (95% yield); 96% ee; $[\alpha]_D^{20} = -128.8^\circ$ ($c = 0.49$, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 8.07 (d, $J = 8.5$ Hz, 1H), 7.98 (d, $J = 8.3$ Hz, 1H), 7.87 (d, $J = 8.4$ Hz, 1H), 7.70 (d, $J = 8.4$ Hz, 1H), 7.56 (m, 1H), 7.37-7.48 (m, 4H), 7.32 (m, 3H), 7.25 (d, $J = 7.0$ Hz, 1H), 7.06 (m, 1H), 6.97 (m, 2H), 2.56 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.3, 150.4, 142.3, 137.5, 134.9, 134.6, 132.8, 132.5, 132.4, 132.3, 132.1, 128.4, 128.2, 127.9, 127.6, 127.5, 127.3, 127.1, 126.9, 125.9, 125.8, 125.7, 124.7, 124.2, 124.1, 123.6, 114.1, 109.4, 19.4; ESI-MS: m/z 452 [$M + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 254 nm, 5.82 min (S), 6.52 min (R).



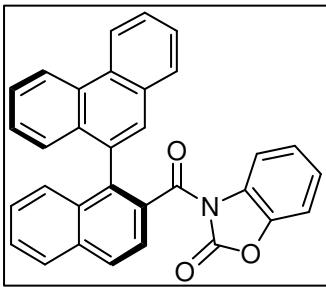
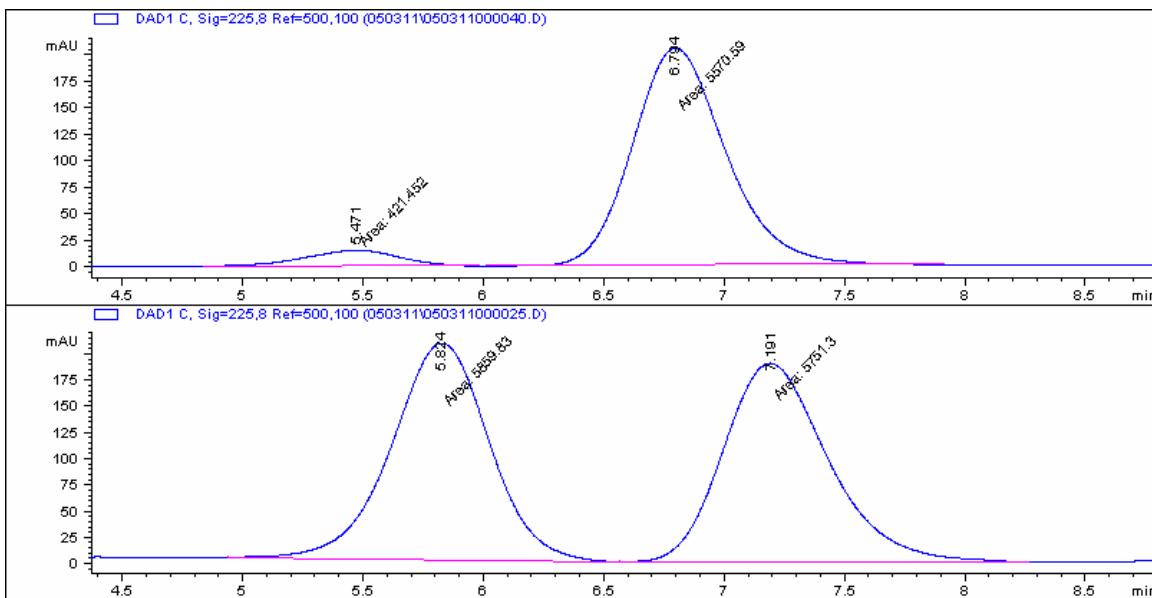
(*R*)-3-(4'-(dimethylamino)-1,1'-binaphthyl-2-carbonyl)benzo[d]oxazol-2(3H)-one: 95% yield; 96% ee; $[\alpha]_D^{20} = -175.2^\circ$ ($c = 0.65$, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 8.09 (d, $J = 8.4$ Hz, 1H), 8.07 (d, $J = 8.3$ Hz, 1H), 7.99 (d, $J = 8.2$ Hz, 1H), 7.73 (d, $J = 8.4$ Hz, 1H), 7.56 (m, 2H), 7.38 (m, 4H), 7.30 (m, 1H), 7.23 (m, 1H), 7.04 (m, 1H), 7.94 (m, 3H), 2.71 (s, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.6, 151.2, 150.3, 142.2, 137.5, 134.8, 133.6, 132.9, 132.3, 128.5, 128.45, 128.4, 128.3, 128.2, 127.6, 127.5, 127.3, 127.1, 127.0, 126.9, 126.0, 125.2, 124.5, 124.2, 124.1, 123.8, 113.8, 113.0, 109.3, 45.0; ESI-MS: m/z 481 [$\text{M} + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel AD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 97/3, 254 nm, 5.30 min (*R*), 8.75 min (*S*).



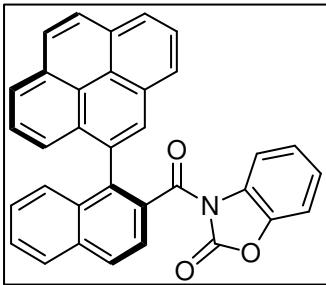
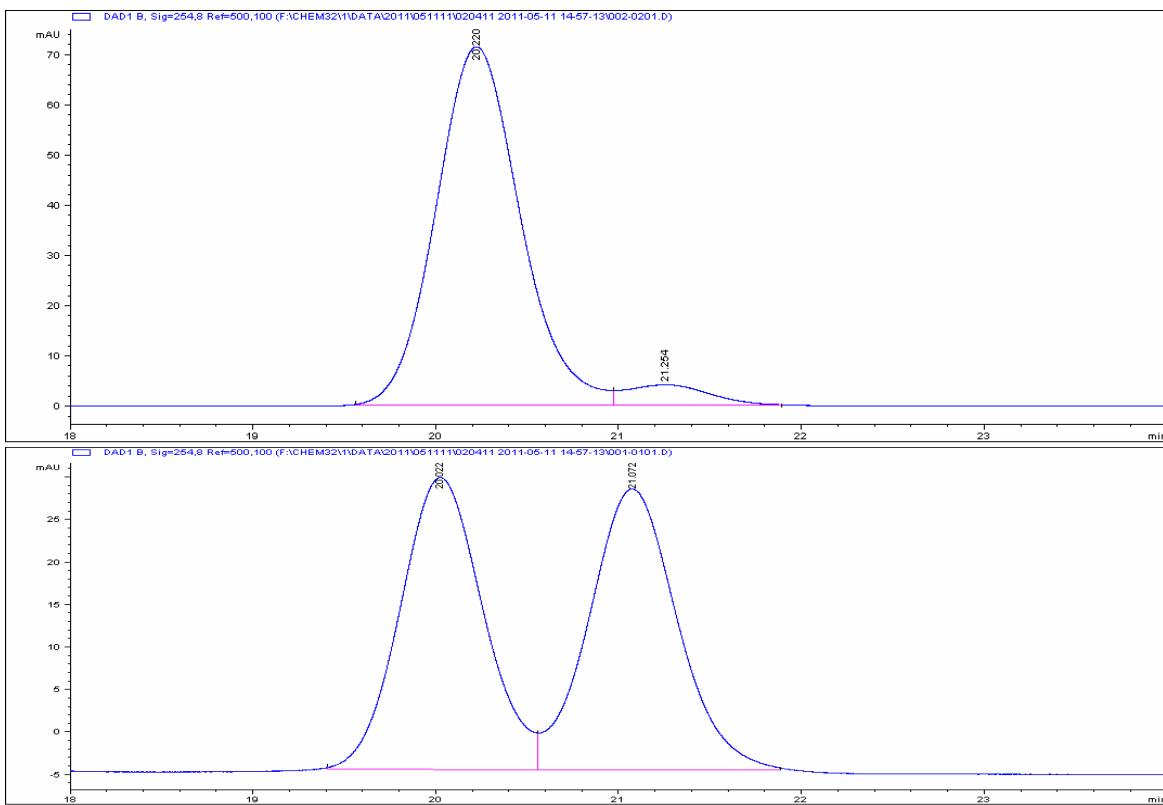
(R)-3-(4'-methoxy-1,1'-binaphthyl-2-carbonyl)benzo[d]oxazol-2(3H)-one: white solid (95% yield); 94% ee; $[\alpha]_D^{20} = -70.6^\circ$ ($c = 0.17$, CHCl_3); ^1H NMR (400 MHz, CD_2Cl_2) δ 8.19 (d, $J = 8.3$ Hz, 1H), 8.10 (d, $J = 8.5$ Hz, 1H), 8.03 (d, $J = 8.3$ Hz, 1H), 7.70 (d, $J = 8.5$ Hz, 1H), 7.59 (t, $J = 7.2$ Hz, 1H), 7.28-7.50 (m, 7H), 7.15 (t, $J = 7.4$ Hz, 1H), 7.06 (m, 2H), 6.80 (d, $J = 7.8$ Hz, 1H), 3.94 (s, 3H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 168.7, 156.2, 151.0, 142.9, 137.8, 135.1, 134.1, 133.6, 133.1, 128.9, 128.8, 128.7, 128.1, 128.0, 127.5, 127.2, 126.9, 125.9, 125.5, 124.9, 124.1, 122.5, 114.8, 110.1, 103.8, 56.1; ESI-MS: m/z 468 [$\text{M} + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel AD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 11.53 min (R), 12.78 min (S).



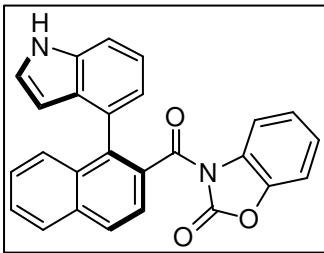
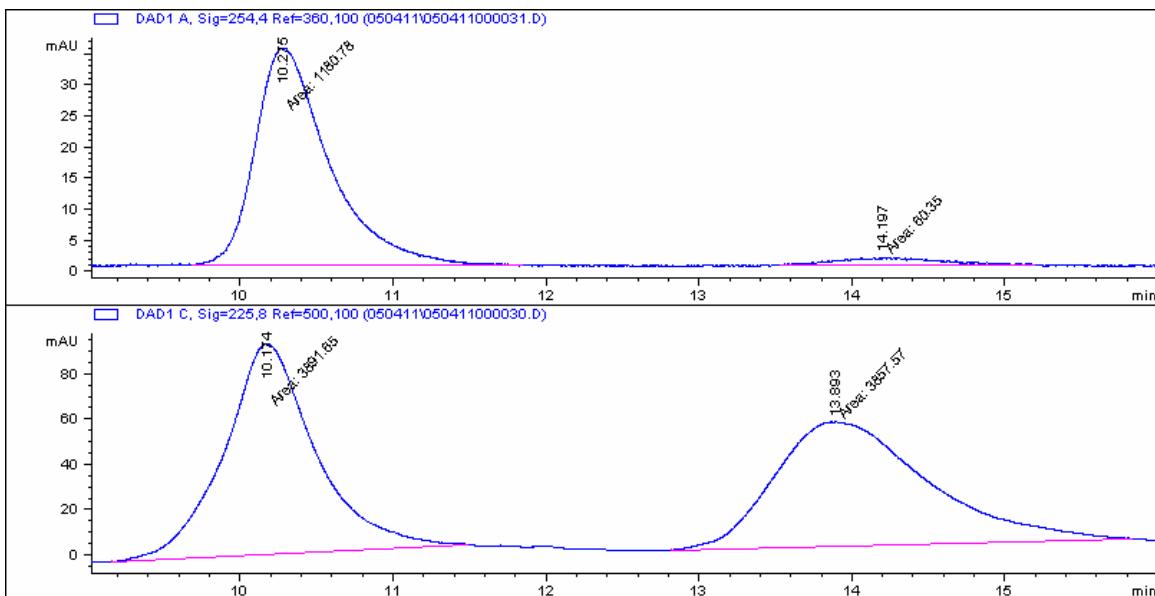
(*R*)-3-(1-(1,2-dihydroacenaphthylen-5-yl)-2-naphthoyl)benzo[d]oxazol-2(3H)-one: white solid (96% yield); 88% ee; $[\alpha]_D^{20} = -96.8^\circ$ ($c = 0.41$, CHCl₃); ¹H NMR (400 MHz, CD₂Cl₂) δ 8.10 (d, $J = 8.5$ Hz, 1H), 8.03 (d, $J = 8.2$ Hz, 1H), 7.73 (d, $J = 8.5$ Hz, 1H), 7.61 (m, 1H), 7.54 (d, $J = 8.6$ Hz, 1H), 7.40 (m, 2H), 2.31 (m, 2H), 7.23 (m, 2H), 6.95-7.13 (m, 4H), 3.13-3.42 (m, 4H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 168.9, 151.0, 147.4, 146.8, 142.9, 139.6, 137.7, 135.2, 133.2, 132.7, 131.4, 130.4, 130.1, 128.82, 128.8, 128.7, 128.2, 127.9, 127.5, 125.2, 124.7, 124.3, 121.8, 120.1, 119.3, 114.4, 109.9, 30.8, 30.6; ESI-MS: m/z 464 [M + Na]⁺; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 5.47 min (*S*), 6.80 min (*R*).



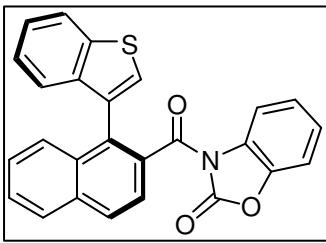
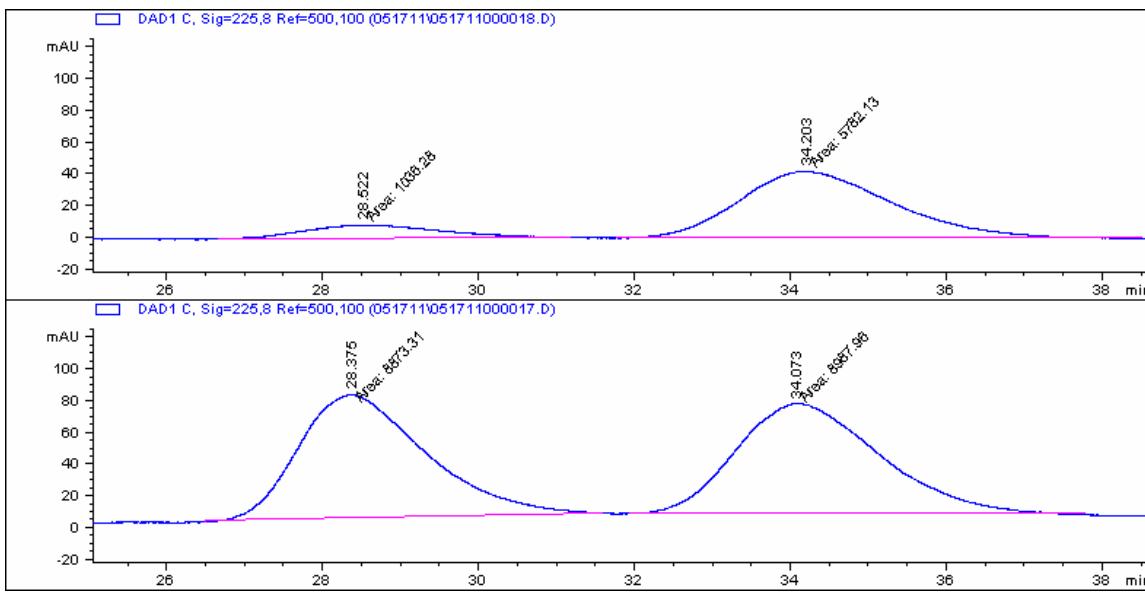
(R)-3-(1-(phenanthren-9-yl)-2-naphthoyl)benzo[d]oxazol-2(3H)-one: white solid (96% yield); 90% ee; $[\alpha]_D^{20} = -91.7^\circ$ ($c = 0.36$, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 8.61 (d, $J = 8.3$ Hz, 1H), 8.57 (d, $J = 8.2$ Hz, 1H), 8.11 (d, $J = 8.5$ Hz, 1H), 8.01 (d, $J = 8.3$ Hz, 1H), 7.83 (d, $J = 7.3$ Hz, 1H), 7.78 (s, 1H), 7.74 (d, $J = 8.6$ Hz, 1H), 7.53-7.65 (m, 4H), 7.40-7.53 (m, 3H), 7.34 (t, $J = 7.3$ Hz, 1H), 7.29 (d, $J = 8.0$ Hz, 1H), 6.96 (m, 1H), 6.85 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.1, 150.6, 142.1, 137.1, 134.6, 133.2, 132.8, 132.2, 131.5, 131.0, 130.2, 130.1, 129.1, 128.8, 128.6, 128.3, 127.7, 127.6, 127.5, 127.2, 127.1, 126.9, 126.8, 126.78, 126.77, 124.7, 124.2, 123.6, 122.6, 122.4, 114.0, 109.4; ESI-MS: m/z 488 [$\text{M} + \text{Na}^+$]; Chiral HPLC conditions: Chiraldak AD-3, 4.6 mm X 250 mm, 25 °C, flow rate: 1.2 mL/min, heptane/isopropanol: 85/15, 254 nm, 20.0 min (*R*), 21.1 min (*S*).



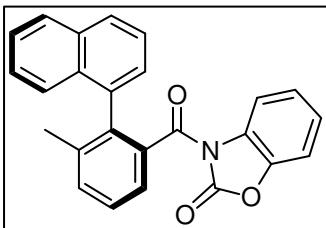
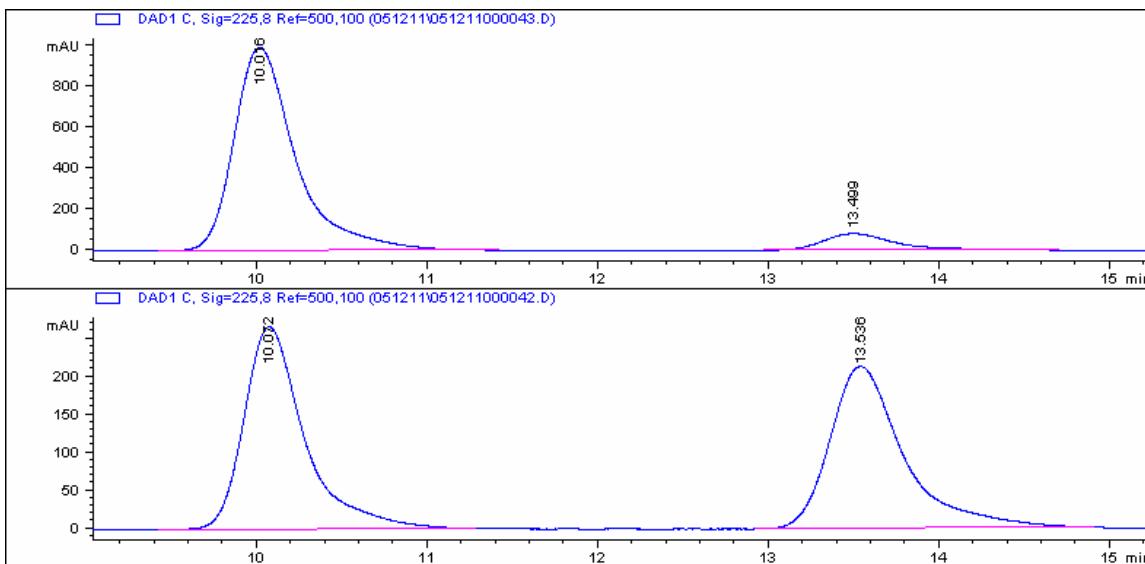
(*R*)-3-(1-(pyren-4-yl)-2-naphthoyl)benzo[d]oxazol-2(3H)-one: white solid (95% yield); 90% ee; $[\alpha]_D^{20} = -129.5^\circ$ ($c = 0.44$, CHCl_3); ^1H NMR (400 MHz, CD_2Cl_2) δ 8.13-8.25 (m, 4H), 8.00-8.12 (m, 5H), 7.94 (d, $J = 9.2$ Hz, 1H), 7.79 (d, $J = 8.5$ Hz, 1H), 7.63 (m, 2H), 7.36 (m, 2H), 7.29 (d, $J = 7.9$ Hz, 1H), 6.88 (m, 2H), 6.75 (t, $J = 7.8$ Hz, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 168.5, 151.0, 142.7, 138.2, 135.1, 133.5, 133.2, 132.1, 131.8, 131.7, 131.4, 130.8, 129.0, 128.9, 128.8, 128.32, 128.31, 128.29, 128.1, 127.8, 127.7, 126.8, 126.0, 125.9, 125.9, 125.4, 125.0, 124.93, 124.9, 124.6, 124.2, 114.6, 109.9; ESI-MS: m/z 512 [$\text{M} + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel AD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 254 nm, 10.17 min (*R*), 13.89 min (*S*).



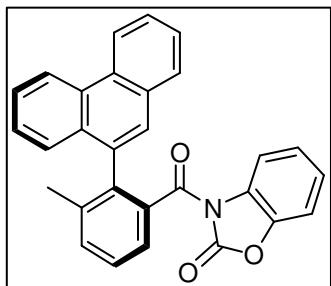
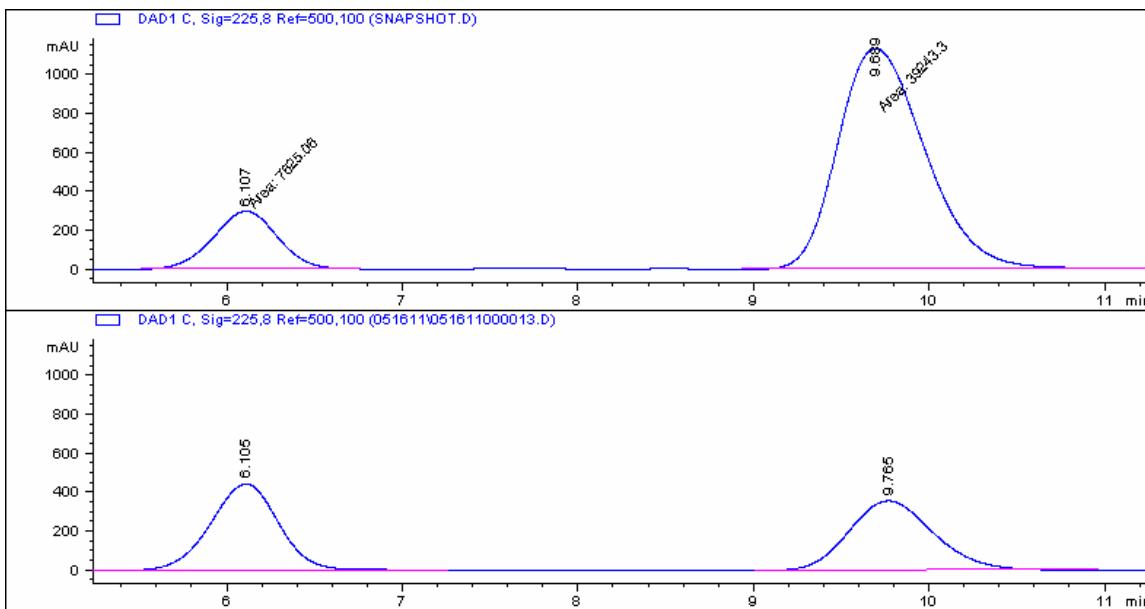
(R)-3-(1-(1H-indol-4-yl)-2-naphthoyl)benzo[d]oxazol-2(3H)-one: 95%; 68% ee; ^1H NMR (500 MHz, CDCl_3) δ 8.05 (d, $J = 8.4$ Hz, 1H), 8.04 (br, 1H), 7.98 (d, $J = 8.3$ Hz, 1H), 7.72 (d, $J = 8.4$ Hz, 2H), 7.57 (t, $J = 7.2$ Hz, 1H), 7.40 (m, 2H), 6.95-7.20 (m, 6H), 6.93 (d, $J = 7.9$ Hz, 1H), 6.14 (t, $J = 2.2$ Hz, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.9, 150.3, 142.3, 138.2, 135.5, 134.9, 132.0, 131.2, 128.9, 128.3, 128.2, 127.9, 127.7, 127.5, 126.7, 124.5, 124.4, 124.0, 123.9, 122.8, 121.9, 121.7, 113.9, 110.6, 109.2, 102.8; ESI-MS: m/z 427 [M + Na] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 93/7, 225 nm, 28.5 min (*S*), 34.2 min (*R*).



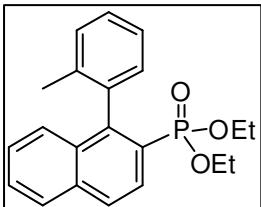
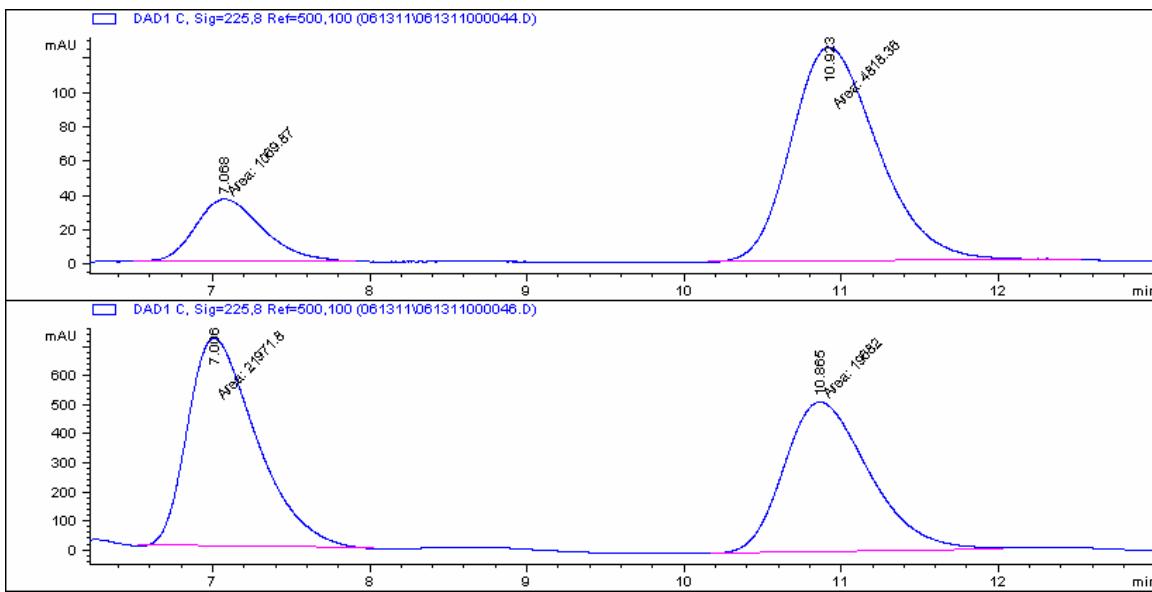
(S)-3-(1-(benzo[b]thiophen-3-yl)-2-naphthoyl)benzo[d]oxazol-2(3H)-one: 95% yield; 82% ee; $[\alpha]_D^{20} = -121.8^\circ$ ($c = 0.61$, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 8.09 (d, $J = 8.5$ Hz, 1H), 8.99 (d, $J = 8.3$ Hz, 1H), 7.71 (m, 2H), 7.64 (d, $J = 8.6$ Hz, 1H), 7.60 (t, $J = 8.0$ Hz, 1H), 7.54 (d, $J = 7.8$ Hz, 1H), 7.49 (s, 1H), 7.42 (m, 1H), 7.35 (m, 1H), 7.23-7.32 (m, 2H), 7.11 (m, 1H), 7.05 (m, 1H), 6.99 (d, $J = 8.0$ Hz, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 168.4, 150.4, 142.3, 139.5, 139.0, 134.7, 132.42, 132.38, 132.3, 129.1, 128.4, 127.7, 127.3, 127.2, 127.0, 126.5, 124.9, 124.7, 124.5, 124.4, 123.7, 123.6, 122.4, 114.2, 109.5; ESI-MS: m/z 431 [$\text{M} + \text{Na}]^+$; Chiral HPLC conditions: Chiralcel AD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 10.02 min (S), 13.50 min (R).



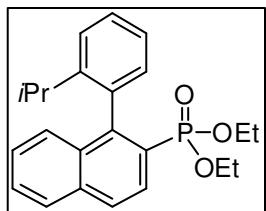
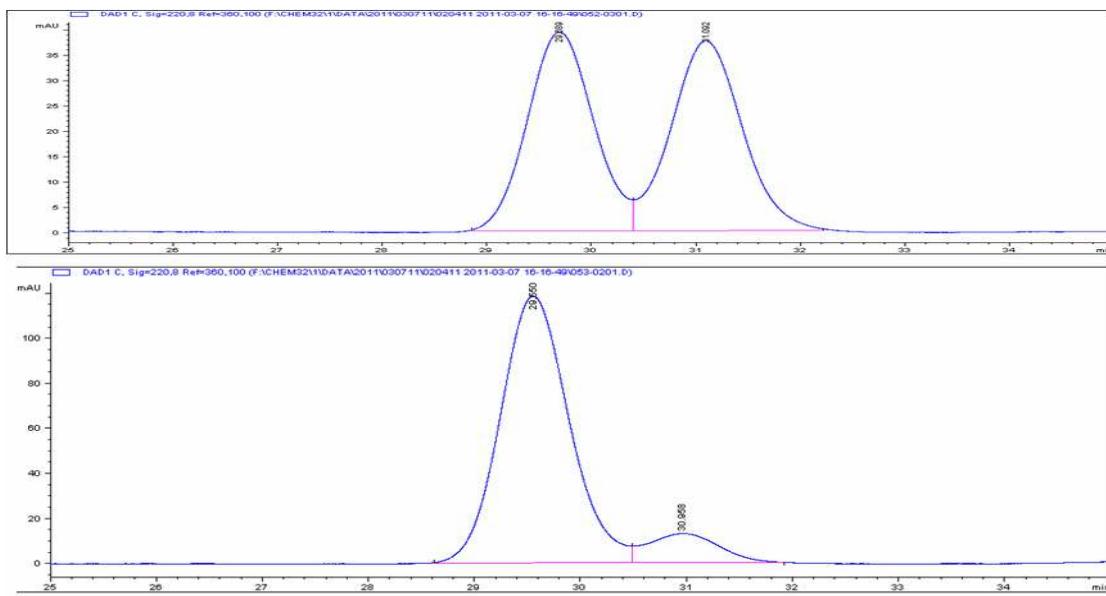
(*R*)-3-(3-methyl-2-(naphthalen-1-yl)benzoyl)benzo[d]oxazol-2(3H)-one: 85% yield; 70% ee; $[\alpha]_D^{20} = -121.8^\circ$ ($c = 0.66$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.70 (m, 1H), 7.66 (dd, $J = 7.4, 1.6$ Hz, 1H), 7.56 (m, 1H), 7.51 (m, 3H), 7.42 (m, 2H), 7.27-7.45 (m, 3H), 7.06 (m, 1H), 6.97 (m, 2H), 2.06 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.1, 150.4, 142.1, 138.2, 137.6, 135.5, 135.2, 133.3, 133.0, 131.5, 128.1, 128.0, 127.8, 127.3, 127.0, 126.3, 126.0, 125.9, 125.0, 124.99, 124.7, 124.2, 114.1, 109.4, 20.1; ESI-MS: m/z 402 [$\text{M} + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 6.11 min (*S*), 9.69 min (*R*).



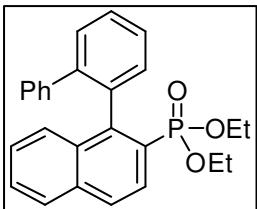
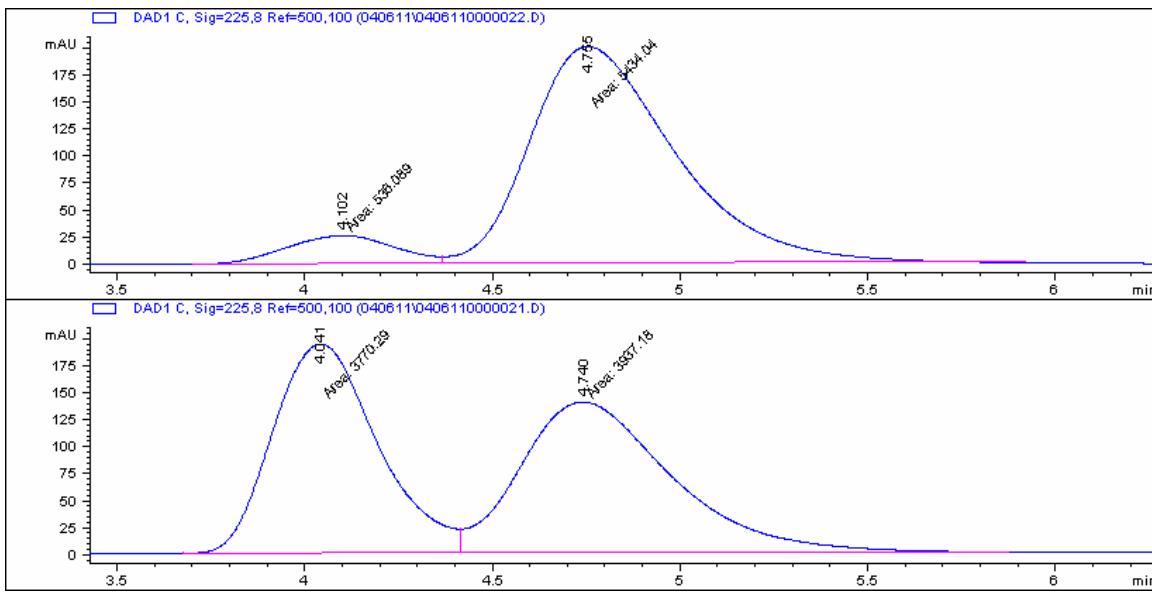
(*R*)-3-(3-Methyl-2-(phenanthren-9-yl)benzoyl)benzo[d]oxazol-2(3H)-one: white solid (90% yield); 64% ee; $[\alpha]_D^{20} = -17.8^\circ$ ($c = 0.45$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.57 (d, $J = 8.2$ Hz, 1H), 8.51 (d, $J = 7.4$ Hz, 1H), 7.80 (m, 1H), 7.50-7.65 (m, 9H), 7.22 (d, $J = 7.8$ Hz, 1H), 6.93 (t, $J = 7.9$ Hz, 1H), 6.81(m, 2H), 2.10 (s, 3H); ^1H NMR (100 MHz, CDCl_3) δ 168.1, 150.6, 142.0, 138.4, 137.6, 135.3, 134.4, 133.0, 131.1, 130.6, 130.2, 130.0, 128.7, 127.9, 127.8, 127.2, 126.9, 126.8, 126.72, 126.7, 125.1, 124.6, 124.1, 122.7, 122.3, 113.9, 109.3, 20.1; ESI-MS: m/z 452 [$\text{M} + \text{Na}$] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 94/6, 225 nm, 7.07 min (*S*), 10.92 min (*R*).



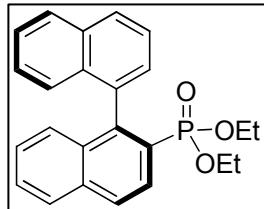
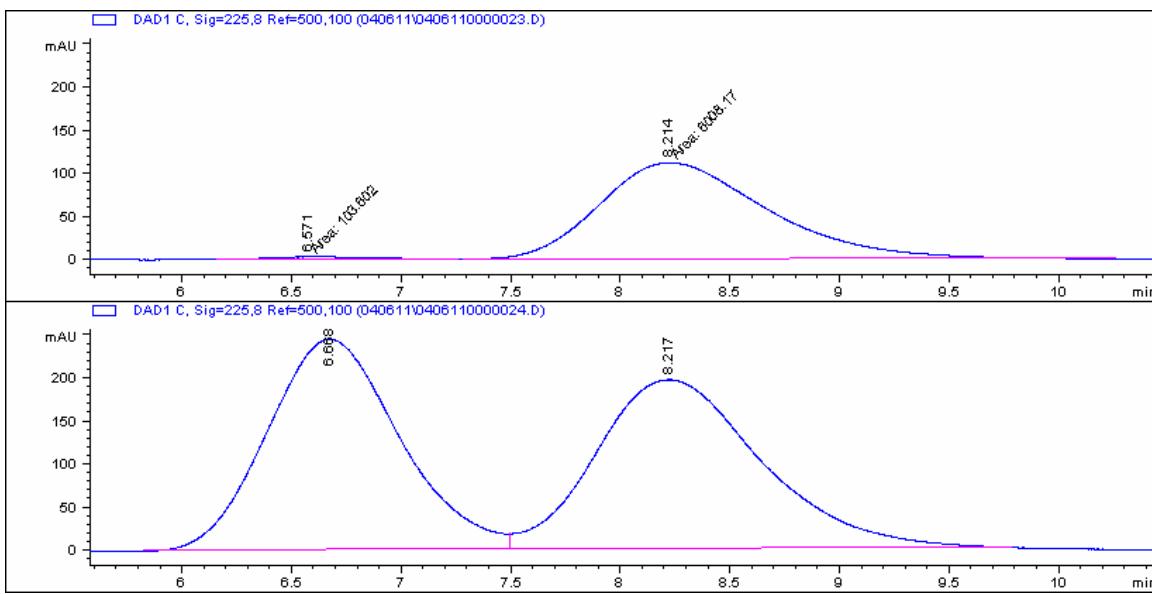
Diethyl 1-o-tolyl naphthalen-2-ylphosphonate: white solid (95% yield); 80% ee; $[\alpha]_D^{20} = -5.5^\circ$ ($c = 0.53, \text{CHCl}_3$); ^1H NMR (400 MHz, CDCl_3) δ 8.09 (dd, $J = 11.8, 8.8$ Hz, 1H), 7.92 (m, 2H), 7.55 (t, $J = 7.3$ Hz, 1H), 7.25-7.43 (m, 5H), 7.21 (d, $J = 7.3$ Hz, 1H), 3.82-4.00 (m, 3H), 3.70-3.82 (m, 1H), 1.92 (s, 3H), 1.85 (m, 6H); ^1H NMR (100 MHz, CDCl_3) δ 144.9 (d, $J = 10.0$ Hz), 137.8 (d, $J = 5.2$ Hz), 137.6, 134.9 (d, $J = 2.6$ Hz), 132.4 (d, $J = 16.1$ Hz), 130.7, 129.3, 128.3, 128.2, 128.0, 127.8, 127.3, 127.2, 127.0, 126.7 (d, $J = 1.2$ Hz), 124.9, 124.8 (d, $J = 129.0$ Hz), 61.7 (m), 20.0, 16.2 (m); ESI-MS: m/z 355 [$\text{M} + \text{H}$] $^+$; Chiral HPLC conditions: Chiralpak AD-3, 4.6 mm X 250 mm, 30 °C, flow rate: 1.0 mL/min, heptane/ethanol: 90/10, 220 nm, 29.6 min, 31.0 min;



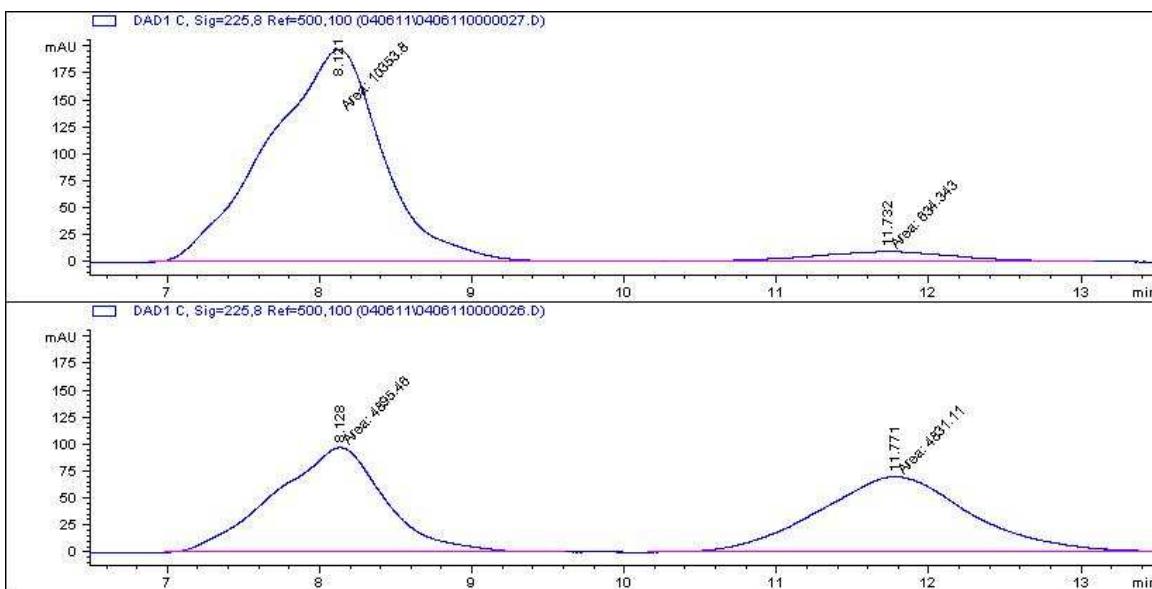
Diethyl 1-(2-isopropylphenyl)naphthalen-2-ylphosphonate: 75% yield; 80% ee; $[\alpha]_D^{20} = -19.8^\circ$ ($c = 0.4$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.06 (d, $J = 8.5$ Hz, 1H), 8.03 (d, $J = 8.7$ Hz, 1H), 7.92 (dd, $J = 8.5$, 3.4 Hz, 1H), 7.88 (d, $J = 8.2$ Hz, 1H), 7.54 (t, $J = 7.6$ Hz, 1H), 7.44 (m, 2H), 7.36 (m, 2H), 7.26 (m, 1H), 7.18 (d, $J = 7.4$ Hz, 1H), 3.98 (m, 1H), 3.88 (m, 2H), 3.84 (m, 1H), 2.37 (m 1H), 1.17 (m, 9H), 0.91 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 148.0, 144.7 (d, $J = 10.2$ Hz), 136.8 (d, $J = 5.1$ Hz), 134.8 (d, $J = 2.6$ Hz), 133.1 (d, $J = 16.4$ Hz), 130.4, 128.4, 128.0, 127.9, 127.86, 127.8, 127.3, 127.2, 126.3 (d, $J = 1.1$ Hz), 125.3, 124.8, 61.7 (t, $J = 6.0$ Hz), 30.5, 24.6, 23.5, 16.3 (t, $J = 6.4$ Hz); ESI-MS: m/z 383 [$\text{M} + \text{H}]^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 4.04 min, 4.74 min.

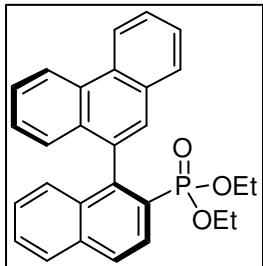


Diethyl 1-(biphenyl-2-yl)naphthalen-2-ylphosphonate: 74% yield; 96% ee; $[\alpha]_D^{20} = -4.7^\circ$ ($c = 0.6$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.98 (dd, $J = 12.0, 8.8$ Hz, 1H), 7.81 (dd, $J = 8.4, 3.6$ Hz, 1H), 7.73 (d, $J = 8.3$ Hz, 1H), 7.54 (m, 2H), 7.37-7.49 (m, 3H), 7.33 (d, $J = 8.6$ Hz, 1H), 7.25 (t, $J = 8.0$ Hz, 1H), 7.16 (m, 2H), 6.94 (m, 3H), 3.92 (m, 3H), 3.68 (m, 1H), 1.15 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 142.0, 141.2, 136.9, 134.6, 131.7, 129.7, 129.1, 128.3, 127.9, 127.8, 127.7, 127.68, 127.5, 127.4, 127.3, 127.26, 126.5, 126.3, 126.29, 61.9 (d, $J = 5.8$ Hz), 61.8 (d, $J = 6.6$ Hz), 16.3 (d, $J = 6.2$ Hz), 16.2 (d, $J = 6.5$ Hz); ESI-MS: m/z 417 [$\text{M} + \text{H}$] $^+$; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 6.57 min, 8.21 min.

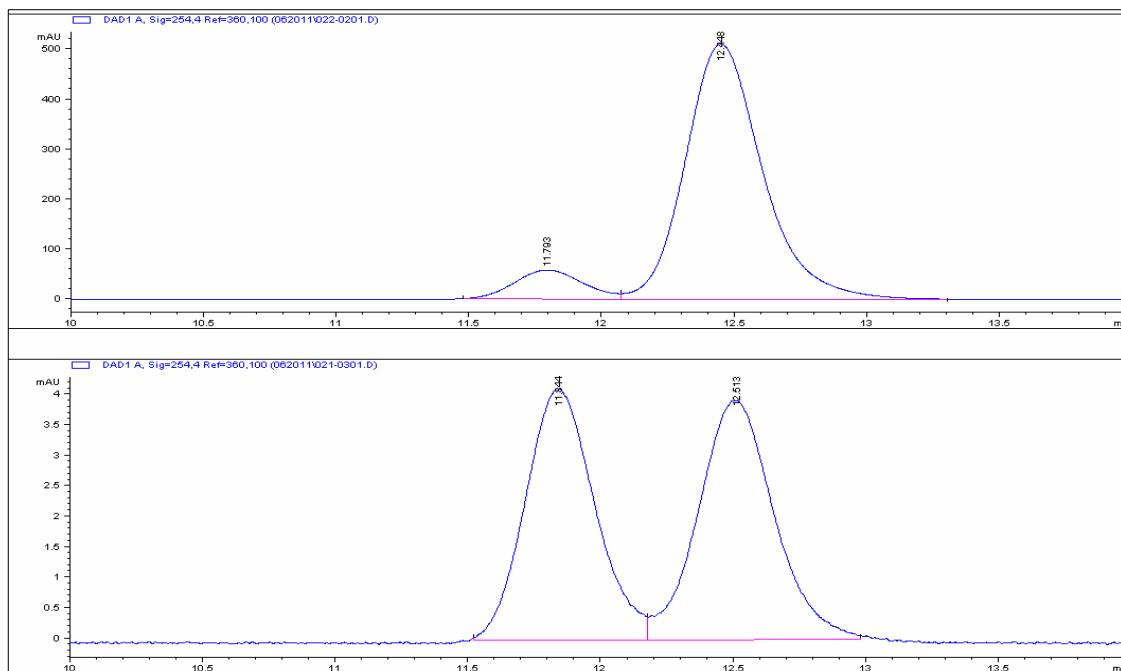


(R)-Diethyl 1,1'-binaphthyl-2-ylphosphonate:⁵ 88% yield; 90% ee; $[\alpha]_D^{20} = +46.4^\circ$ ($c = 0.29$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.21 (dd, $J = 12.0, 8.7$ Hz, 1H), 8.02 (dd, $J = 8.6, 3.7$ Hz, 1H), 7.94 (m, 3H), 7.60 (t, $J = 7.4$ Hz, 1H), 7.53 (t, $J = 7.3$ Hz, 1H), 7.49 (d, $J = 7.0$ Hz, 1H), 7.44 (t, $J = 7.4$ Hz, 1H), 7.15-7.30 (m, 3H), 7.08 (d, $J = 8.4$ Hz, 1H), 3.80 (m, 1H), 3.69 (m, 1H), 3.58 (m 2H), 0.97 (t, $J = 7.0$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 135.9, 134.9, 133.4, 133.2, 128.73, 128.71, 128.6, 128.3, 128.0, 127.9, 127.8, 127.7, 127.6, 126.72, 126.7, 125.9, 125.6, 124.9, 61.8 (d, $J = 6.0$ Hz), 61.6 (d, $J = 6.0$ Hz), 15.9 (d, $J = 6.7$ Hz), 15.4 (d, $J = 7.2$ Hz); ESI-MS: m/z 391 [$\text{M} + \text{H}]^+$; Chiral HPLC conditions: Chiralcel AD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 225 nm, 8.13 min (*R*), 11.7 min (*S*).

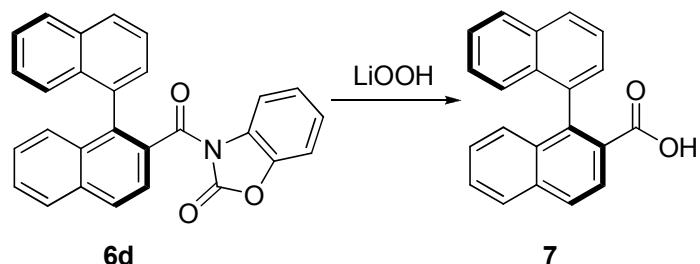




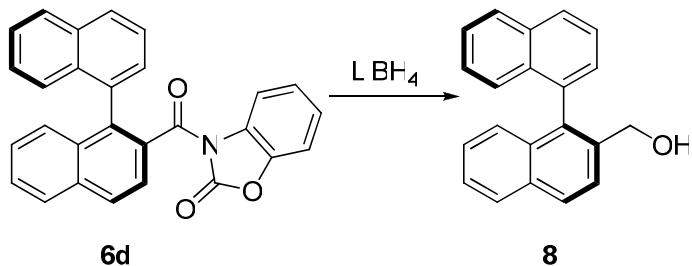
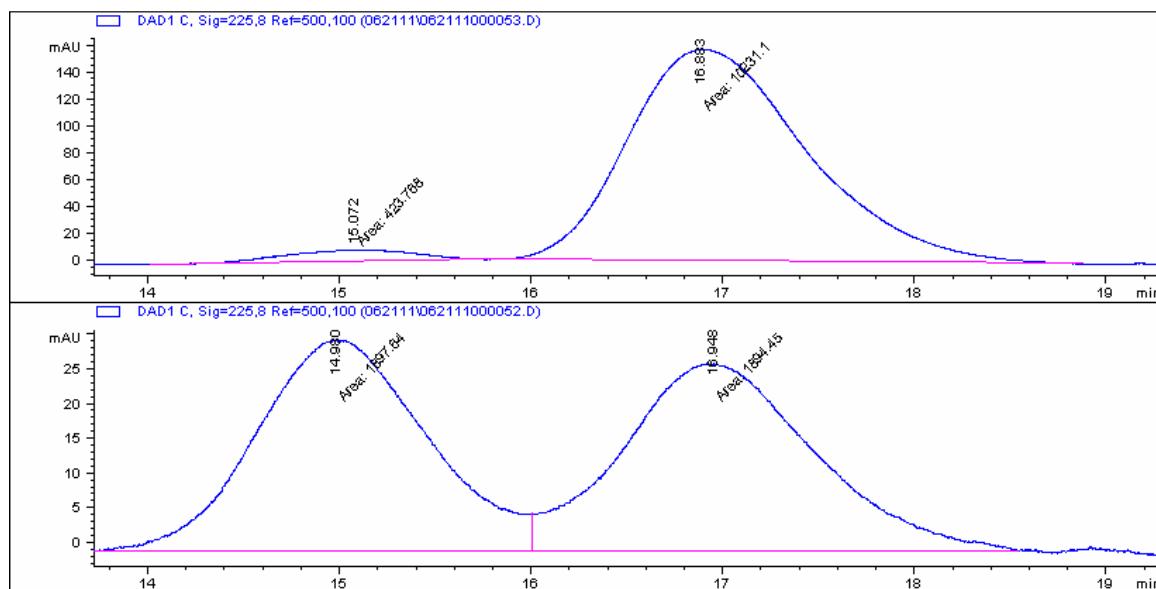
(R)-diethyl 1-(phenanthren-9-yl)naphthalen-2-ylphosphonate: 90% yield; 82% ee; $[\alpha]_D^{20} = +30.4^\circ$ ($c = 0.25$, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 8.79 (d, $J = 8.3$ Hz, 2H), 8.26 (dd, $J = 12.2$, 8.6 Hz, 1H), 8.06 (dd, $J = 8.6$, 3.7 Hz, 1H), 8.96 (d, $J = 8.3$ Hz, 1H), 7.89 (d, $J = 7.8$ Hz, 1H), 7.77 (s, 1H), 7.74 (m, 1H), 7.65 (m, 2H), 7.54 (t, $J = 7.0$ Hz, 1H), 7.34 (m, 2H), 7.23 (m, 1H), 7.15 (d, $J = 7.7$ Hz, 1H), 3.77 (m, 1H), 3.64 (m, 2H), 3.54 (m, 1H), 0.95 (t, $J = 7.1$ Hz, 3H), 0.61 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 143.3 (d, $J = 9.3$ Hz), 135.0 (d, $J = 2.6$ Hz), 134.5 (d, $J = 5.3$ Hz), 133.3 (d, $J = 15.7$ Hz), 132.6, 131.3, 130.3, 129.9, 129.63, 129.62, 128.9, 128.8, 128.79, 128.0 (d, $J = 1.0$ Hz), 127.9 (d, $J = 1.0$ Hz), 127.87, 127.8, 127.7, 127.1, 127.6, 126.9, 126.7 (d, $J = 1.4$ Hz), 126.5, 126.3, 125.6, 122.63, 122.61, 61.8 (d, $J = 6.1$ Hz), 61.5 (d, $J = 6.1$ Hz), 15.9 (d, $J = 7.1$ Hz), 15.3 (d, $J = 7.2$ Hz); ESI-MS: m/z 441 [$\text{M} + \text{H}]^+$; Chiral HPLC conditions: Chiracel OD-H, 4.6 X150 mm, flow rate: 2 mL/min, Water/Acetonitrile: 50/50; 225 nm, 11.70 min (*S*), 12.50 min (*R*).



6. Derivatization of chiral biaryl coupling product 6d

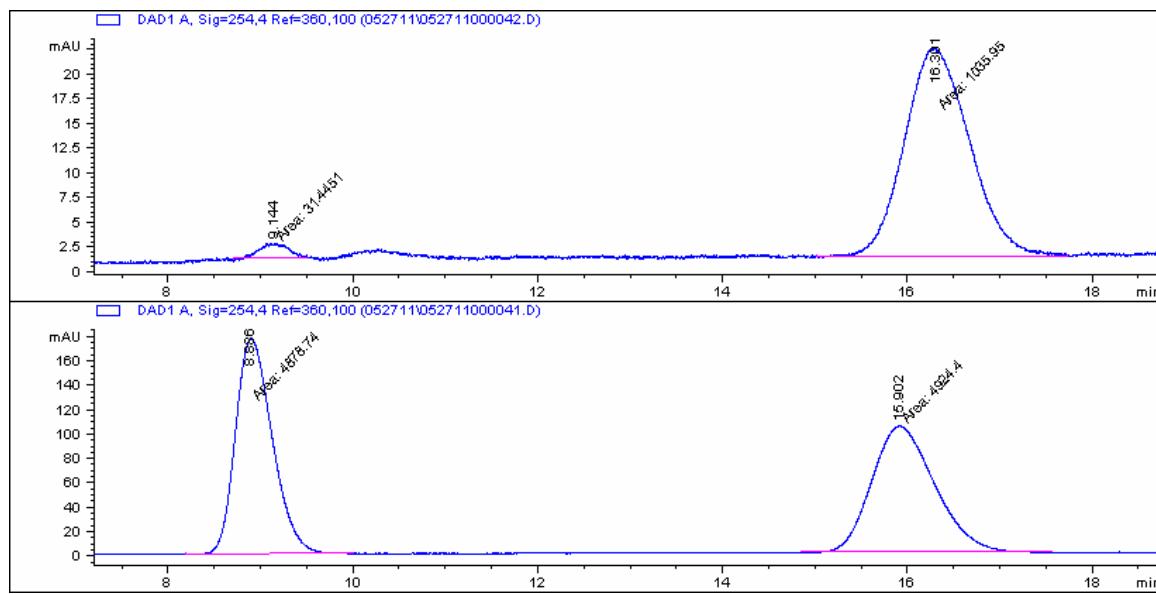


To a solution of **6d** (100 mg, 0.24 mmol, 92% ee) in THF/water (4 mL, 3/1 v/v) at 0 °C was charged 30% H₂O₂ solution (109 mg, 0.96 mmol, 3 equiv) followed by lithium hydroxide (11.5 mg, 0.48 mmol, 2 equiv). The resulting mixture was stirred at 0–25 °C for ~ 0.5 h. The starting material was totally consumed. To the mixture was charged 10 % sodium bisulfite solution (4 mL) and dichloromethane (4 mL). The DCM layer was separated, washed with brine, concentrated, and purified by column chromatography to provide **7** as white solid (71 mg, 0.238 mmol, 99% yield). **7**: 92% ee; $[\alpha]_D^{20} = +23.6^\circ$ (c = 0.70, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 8.7 Hz, 1H), 7.95 (m, 4H), 7.54 (m, 2H), 7.44 (t, *J* = 7.3 Hz, 1H), 7.31 (d, *J* = 6.8 Hz, 1H), 7.20–7.28 (m, 3H), 7.13 (d, *J* = 8.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 171.5, 141.2, 136.5, 135.2, 133.2, 133.1, 132.9, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.2, 127.0, 126.7, 126.1, 126.0, 125.7, 125.2; ESI-MS: m/z 299[M + H]⁺; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 94/6, 225 nm, 15.07 min (*S*), 16.88 min (*R*).



To a solution of **6d** (100 mg, 0.24 mmol, 94% ee) in THF (5 mL) at 0 °C was charged LiBH₄ (8 mg, 0.36 mmol, 1.5 equiv) in one portion. The mixture was then stirred at 0–25 °C for 1 h and then quenched by addition of water (5 mL) and EtOAc (5 mL). The organic layer was separated, washed with brine, dried over sodium sulfate, concentrated, and purified by silica gel column chromatography to provide alcohol **8** as white solid (63 mg, 0.22 mmol, 94% ee). **8**: 94% ee; $[\alpha]_D^{20} = +46.3^\circ$ (c = 1.0, CHCl₃); ¹H NMR (400

MHz, CDCl₃) δ 7.85-8.05 (m, 4H), 7.78 (d, *J* = 8.3 Hz, 1H), 7.59 (t, *J* = 7.7 Hz, 1H), 7.33-7.50 (m, 3H), 7.12-7.30 (m, 4H), 4.4 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 136.7, 135.8, 135.7, 133.6, 133.1, 132.9, 132.8, 128.3, 128.1, 127.9, 127.8, 126.6, 126.4, 126.2, 126.1, 125.8, 125.7, 125.5, 63.4; ESI-MS: m/z 285[M +H]⁺; Chiral HPLC conditions: Chiralcel OD-H, 4.6 mm X 250 mm, 30 °C, flow rate: 2 mL/min, heptane/isopropanol: 96/4, 254 nm, 9.14 min (*S*), 16.30 min (*R*).



7. Computational studies

Both Buchwald⁶ and Baudoin⁷ have made a reasonable hypothesis that the enantioselectivities in the Suzuki-Miyaura coupling reactions are determined in the transition state of reductive elimination. The occurrence of a particular conformation of reaction intermediate is determined by its energy. The chirality of the product follows the shortest path from the conformation of the transition state. To determine the most probable conformation of the intermediates, we first sampled all possible conformations of the palladium complex at the reductive elimination step in the reaction of **5d** and 1-naphthylboronic acid with the Pd-**2** catalyst. This was accomplished by fixing the conformation of the ligand and systematically rotating the bonds between the Pd center and the two reactants in 30° intervals. Two differently probable coordination configurations were considered as shown in Figure 1S. After eliminating conformations that have obvious steric clashes, 28 unique conformations were obtained. The geometry of each of these conformations was then optimized by energy minimized. All calculations were carried out with Gaussian 03 package.⁸ The DFT/B3LYP method was used in the optimization. Because our system consists of bulky aromatic system, non-bonded van der Walls interactions are expected to make a significant contribution to the relative energy of different conformations. We therefore employed the MIDI basis set developed by

Truhlar and co-workers⁹, supplied by the Gaussian 03 package (MidiX) for the first-row atoms (H, C, N and O). This functional was reported to produce good results for non-bonded interactions comparable with higher-order theories. For phosphorus, the 6-31G(d) basis set was employed; and for palladium, the all-electron basis set from optimization of fcc Pd bulk¹⁰ was employed.

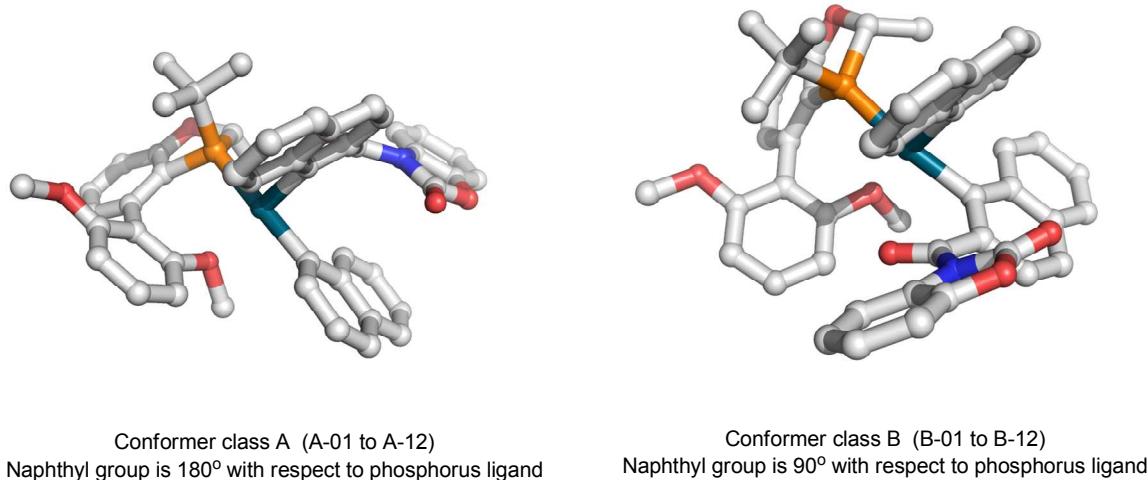


Figure S1: Two different conformation classes that were sampled

Although there were 28 unique starting conformations, four pairs of conformations were found to be almost duplicates after full energy minimization for each conformation. The coordinates of the 24 unique energy-minimized conformations are provided in the enclosed minimized-conformation.cif file. Figure S2 shows the relative energy of the 24 conformations. Three lowest-energy conformations were found with relative energy of 0, 0.51 and 0.78 kcal/mol, respectively. All other conformations have energy > 2 kcal/mol or higher than the lowest-energy state.

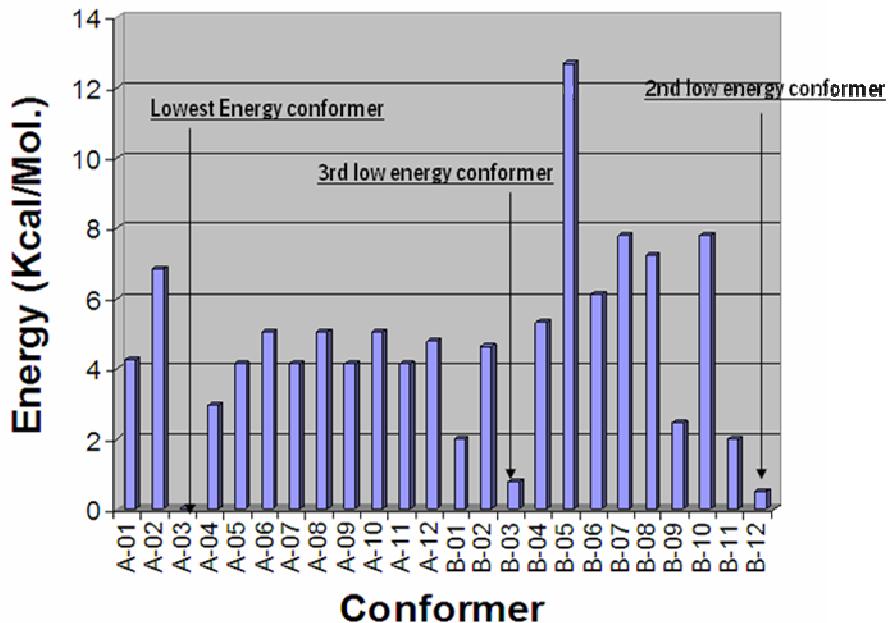


Figure S2: The relative energy of all the sample conformations in the energy-minimized state

There are two main interesting observations from calculation results. First, all three lowest-energy conformations turn out to lead to the same chirality in the coupling product that is in agreement with the inferred axial chirality of **6d**. This result suggests that the energetics of non-bonded π -interaction of the aromatic systems indeed dictate the conformation of the intermediate and the chirality of the product. Secondly, the origin of the π -stacking interaction in determining the energetics of this system is most clearly seen in the lowest and 2nd lowest-energy conformations. Figure S3 and S4 compare the conformations of these two conformations. In each case, there is a π -stacking interaction between the naphthal group and the benzooxazolidinone; and more revealing, in each case, there is a related energy-minimum conformation in which the naphthal group rotates by 180 that reverses the chirality of the system and that has higher energy by ~ 2 kcal/mol.

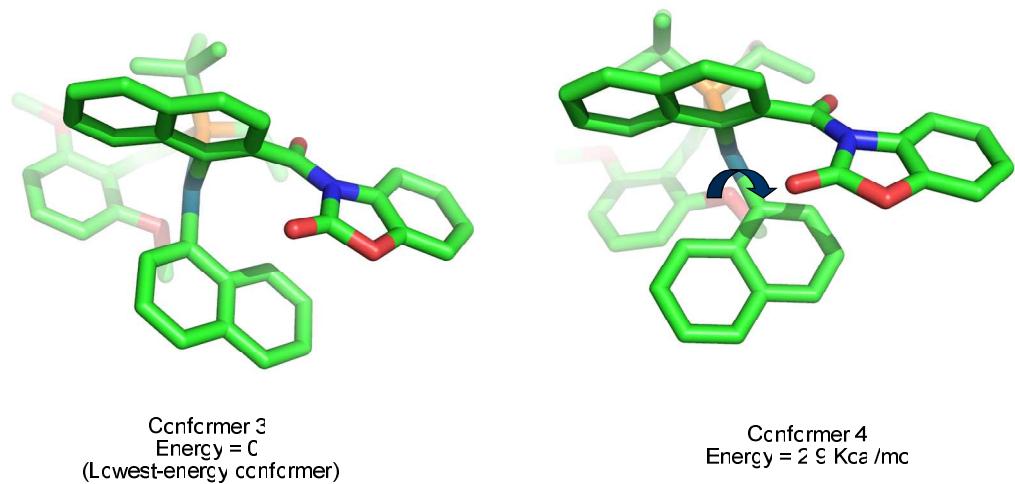


Figure S3: Comparison of the lowest-energy conformation and the local energy minimum in which the naphthyl group approximately flaps by 180° with respect to the first conformation

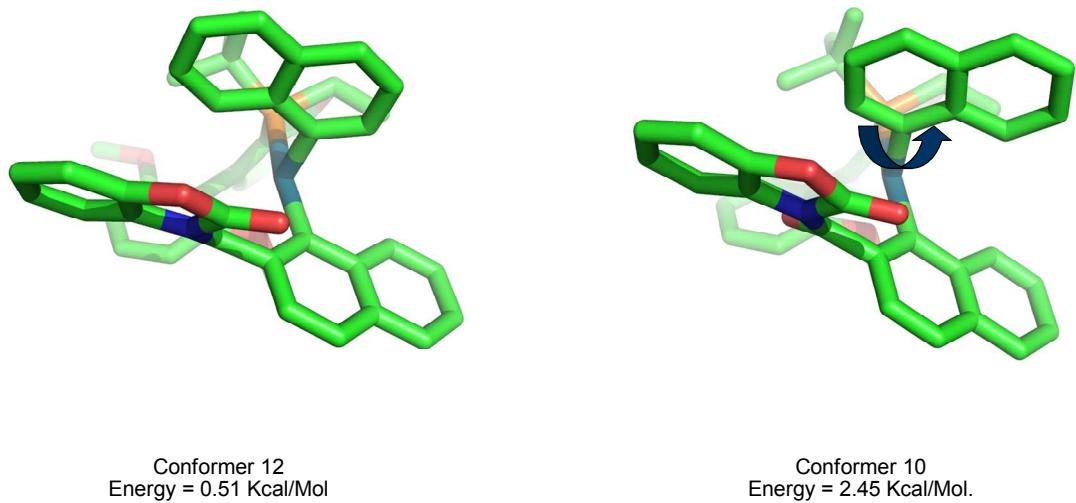
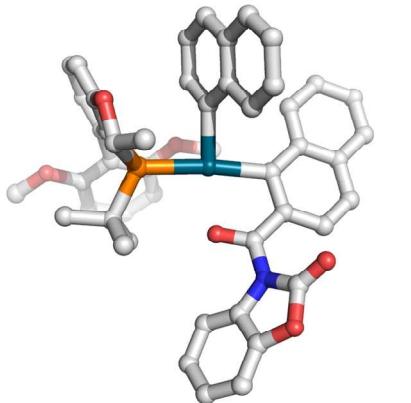


Figure S4: Comparison of the 2nd lowest-energy conformation and the local energy minimum in which the naphthyl group approximately flaps by 180° with respect to the first conformation



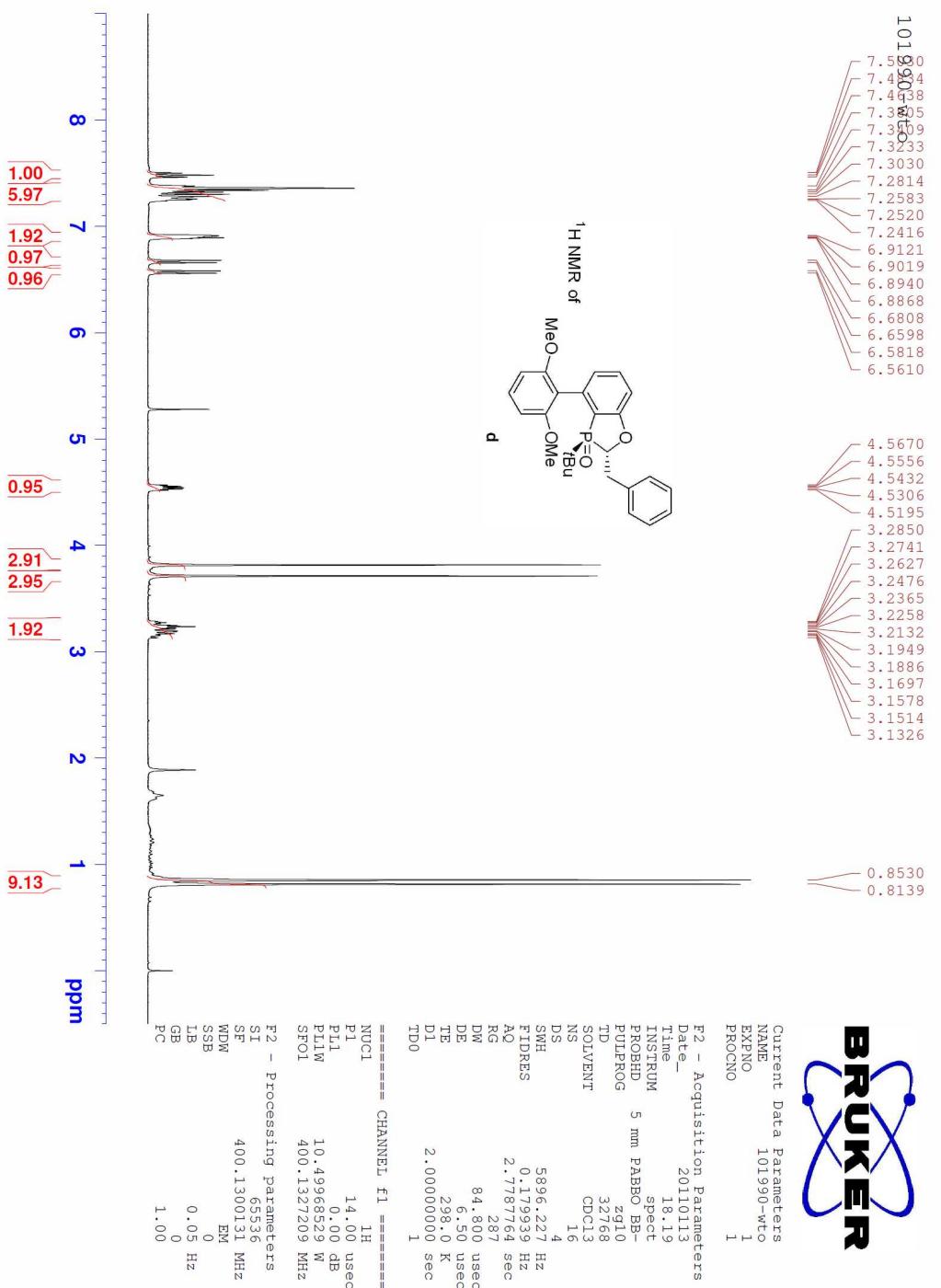
Ccnfcrmer B-C5 Energy = C 78 Kca /Nc

Figure S5: The conformation of the 3rd lowest-energy conformation. Interestingly, the chirality of the coupling product from this intermediate conformation is the same as the 1st and 2nd lowest energy conformations.

8. References:

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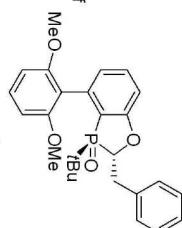
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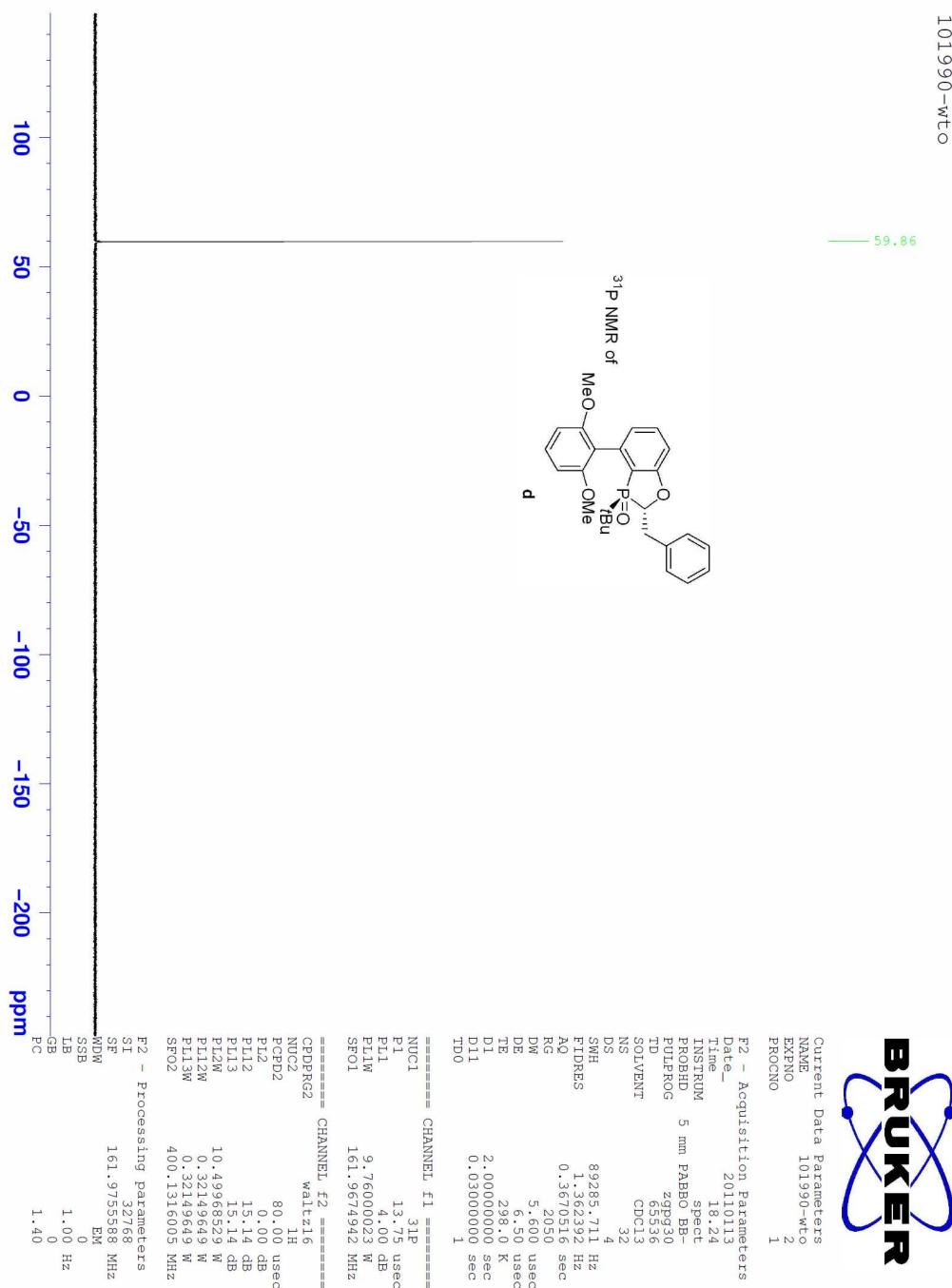
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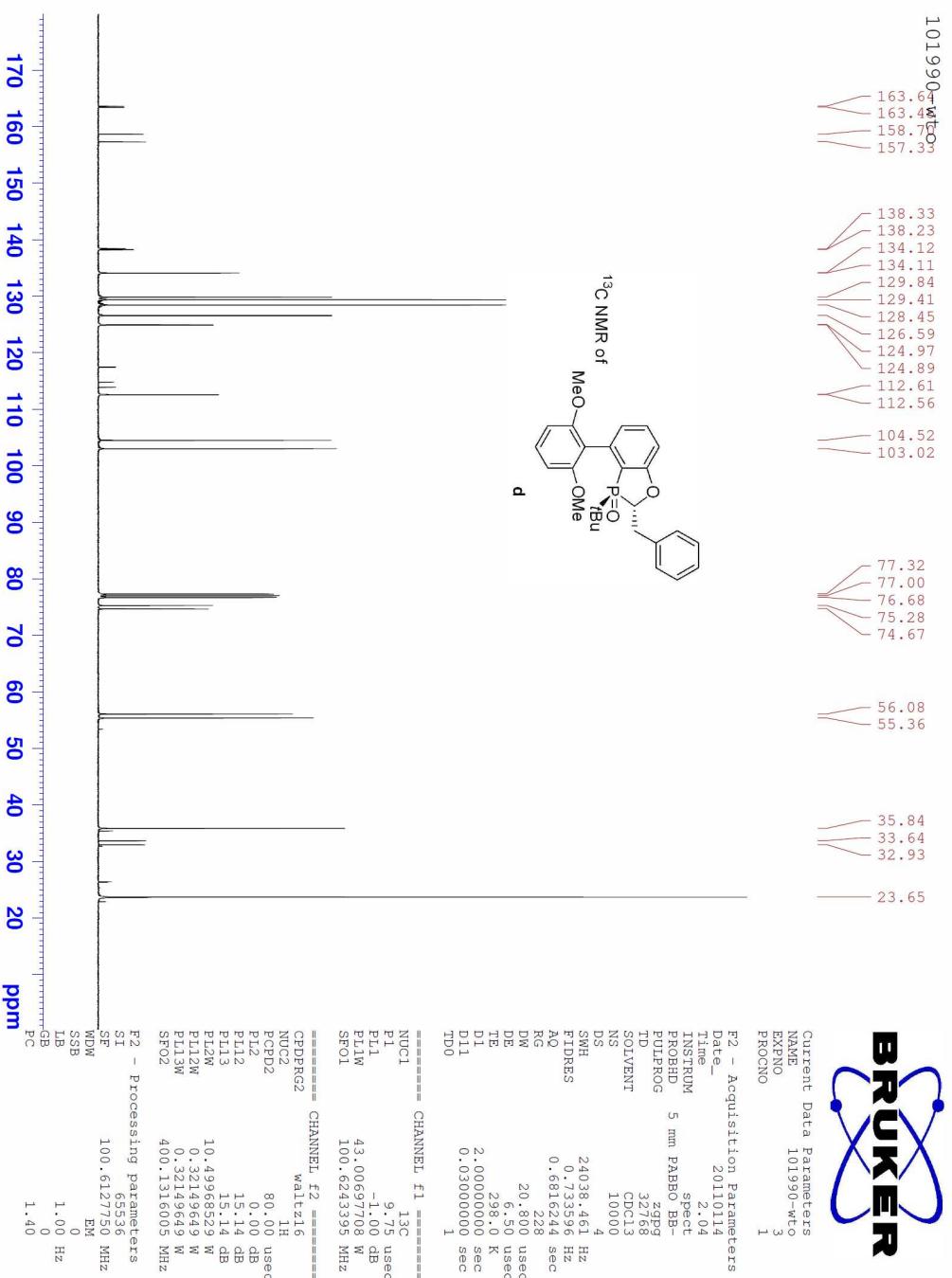
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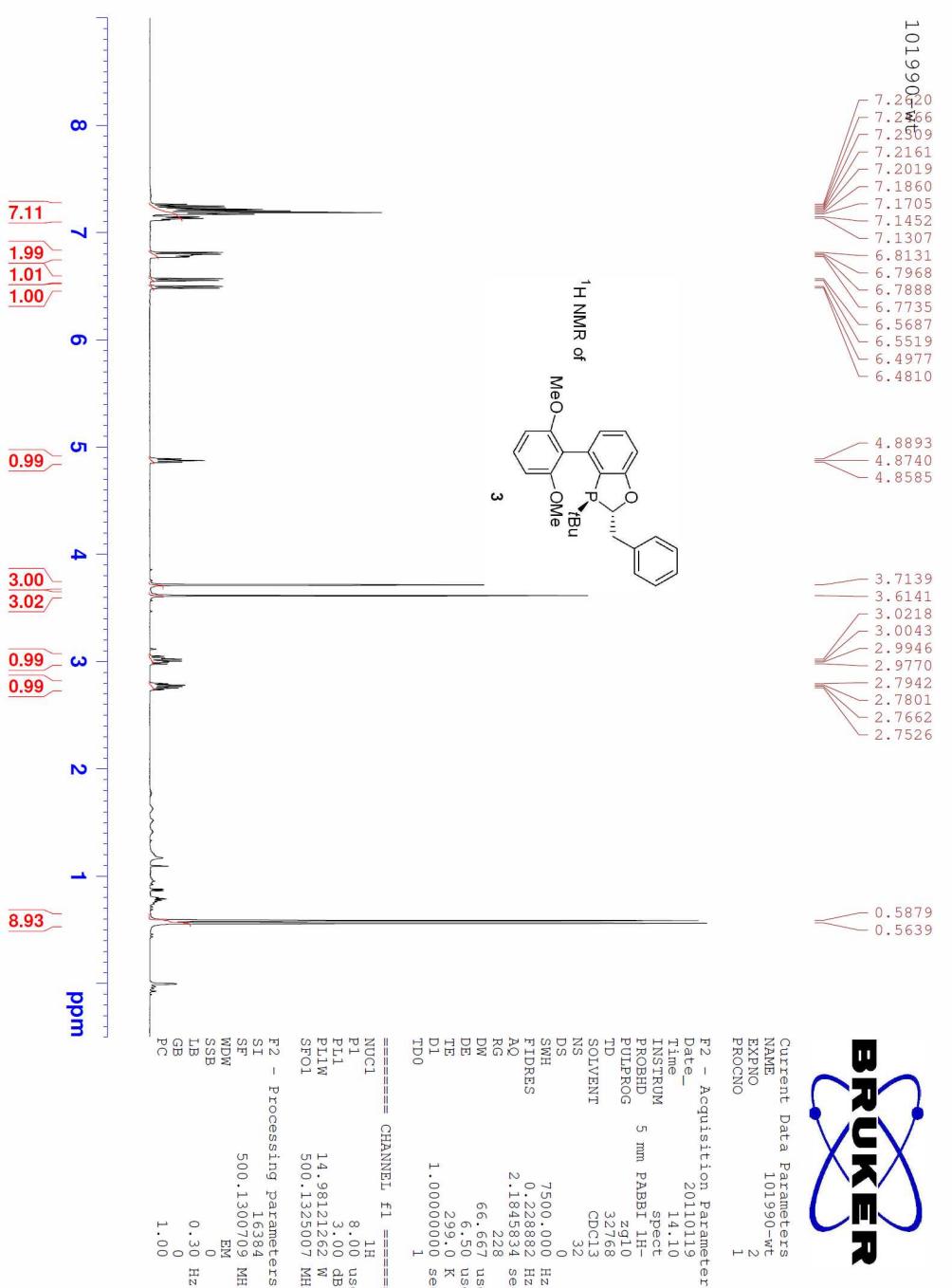
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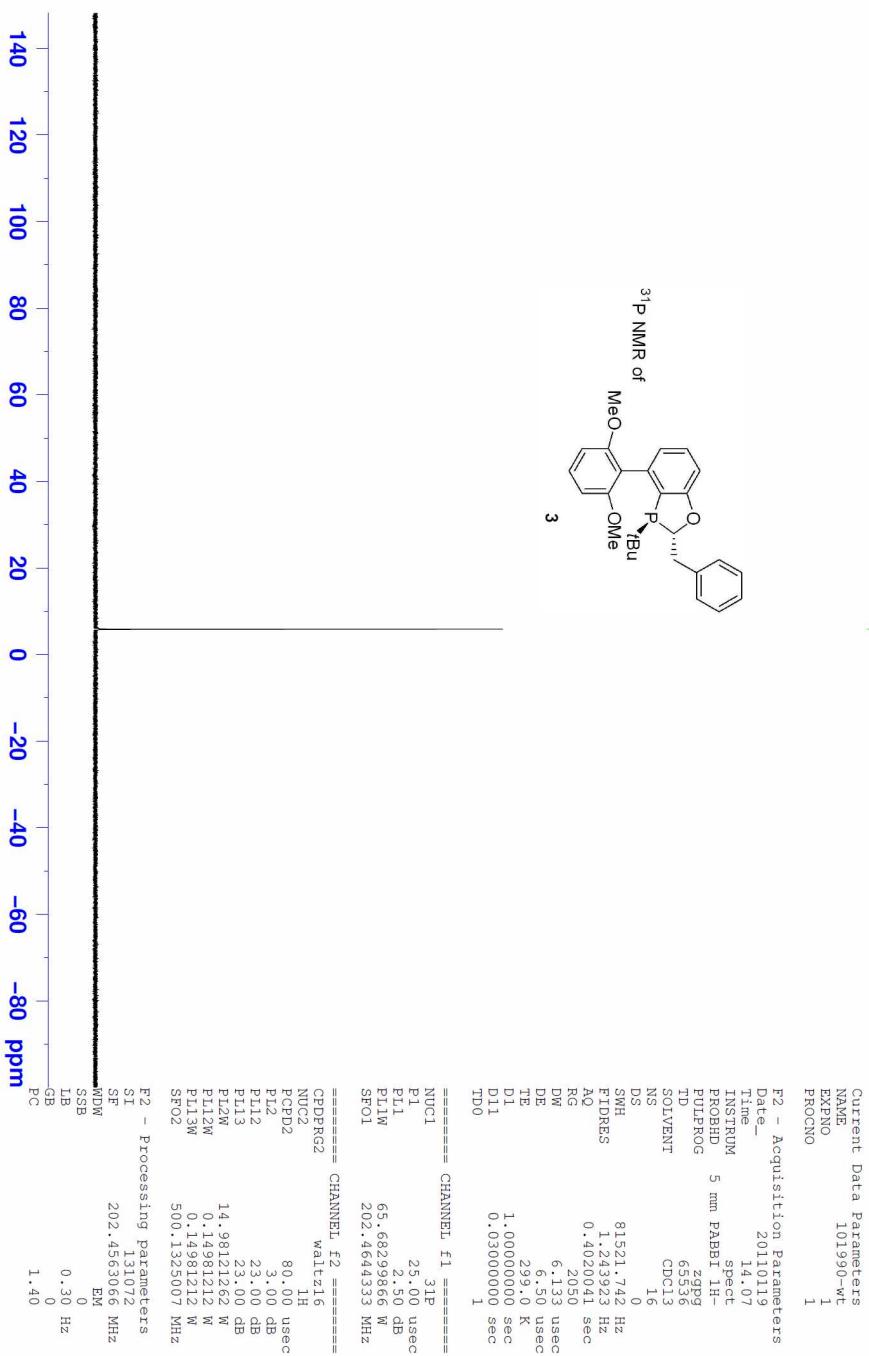
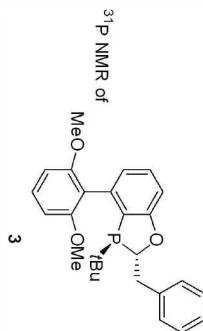


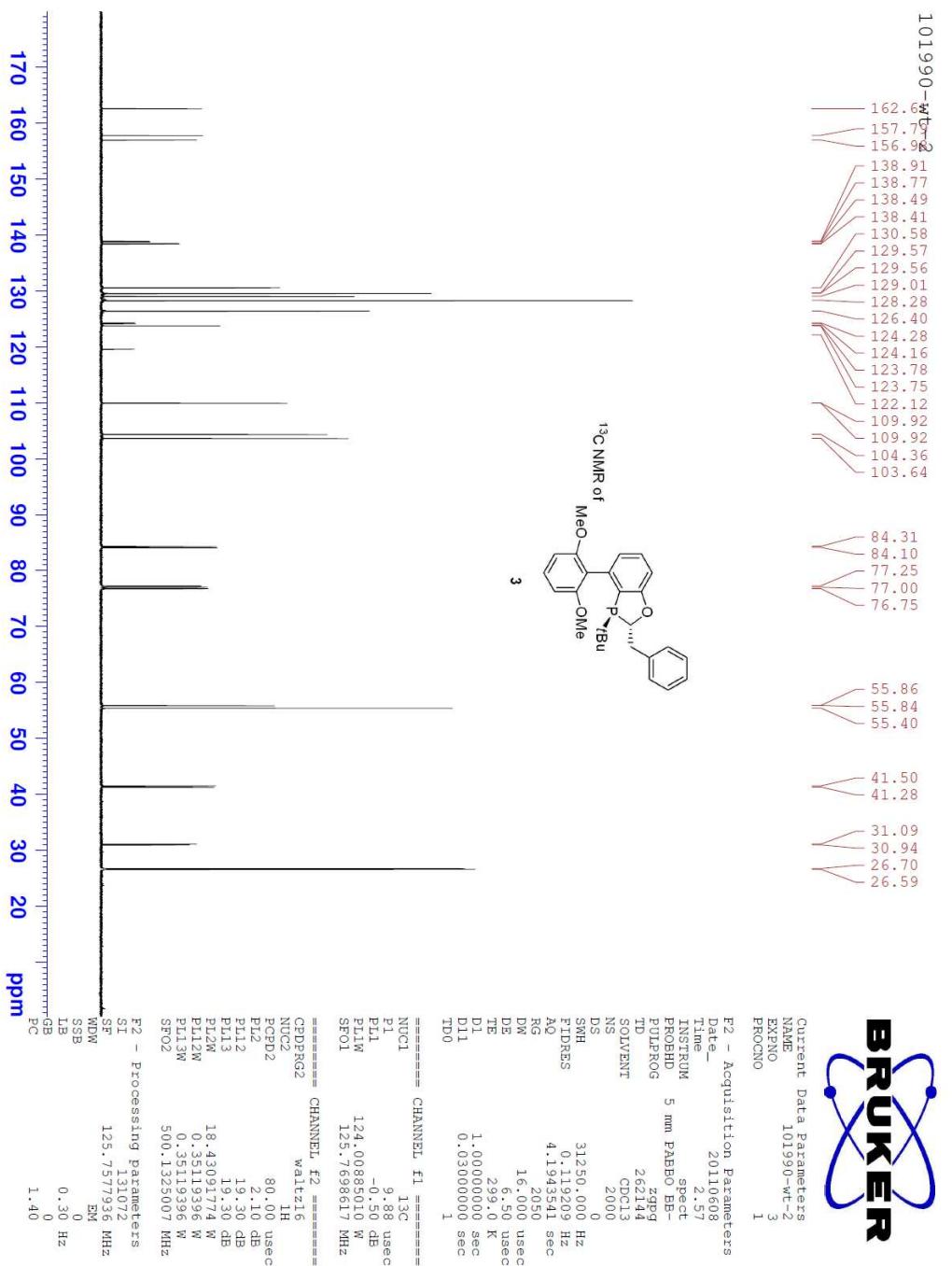


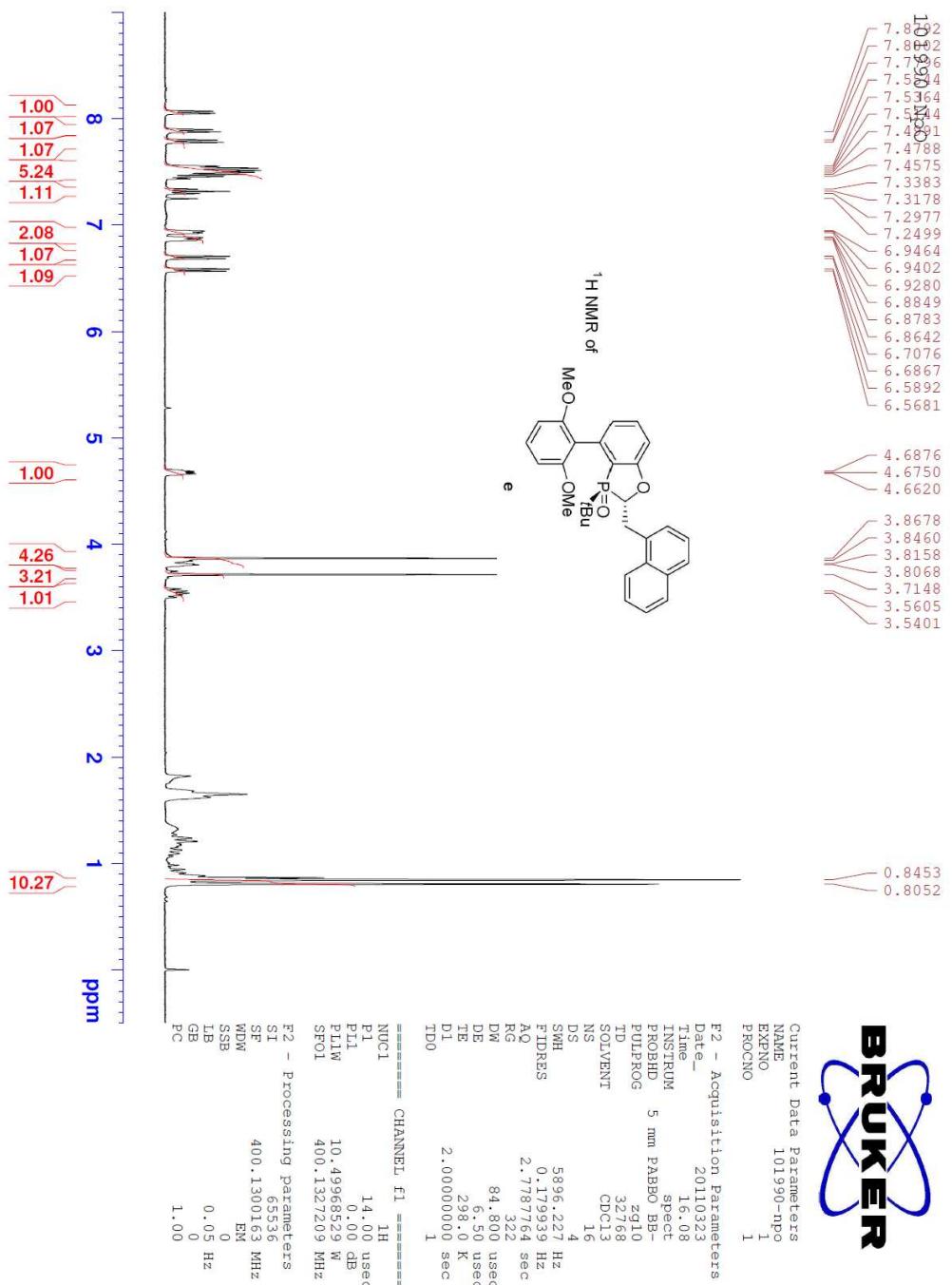


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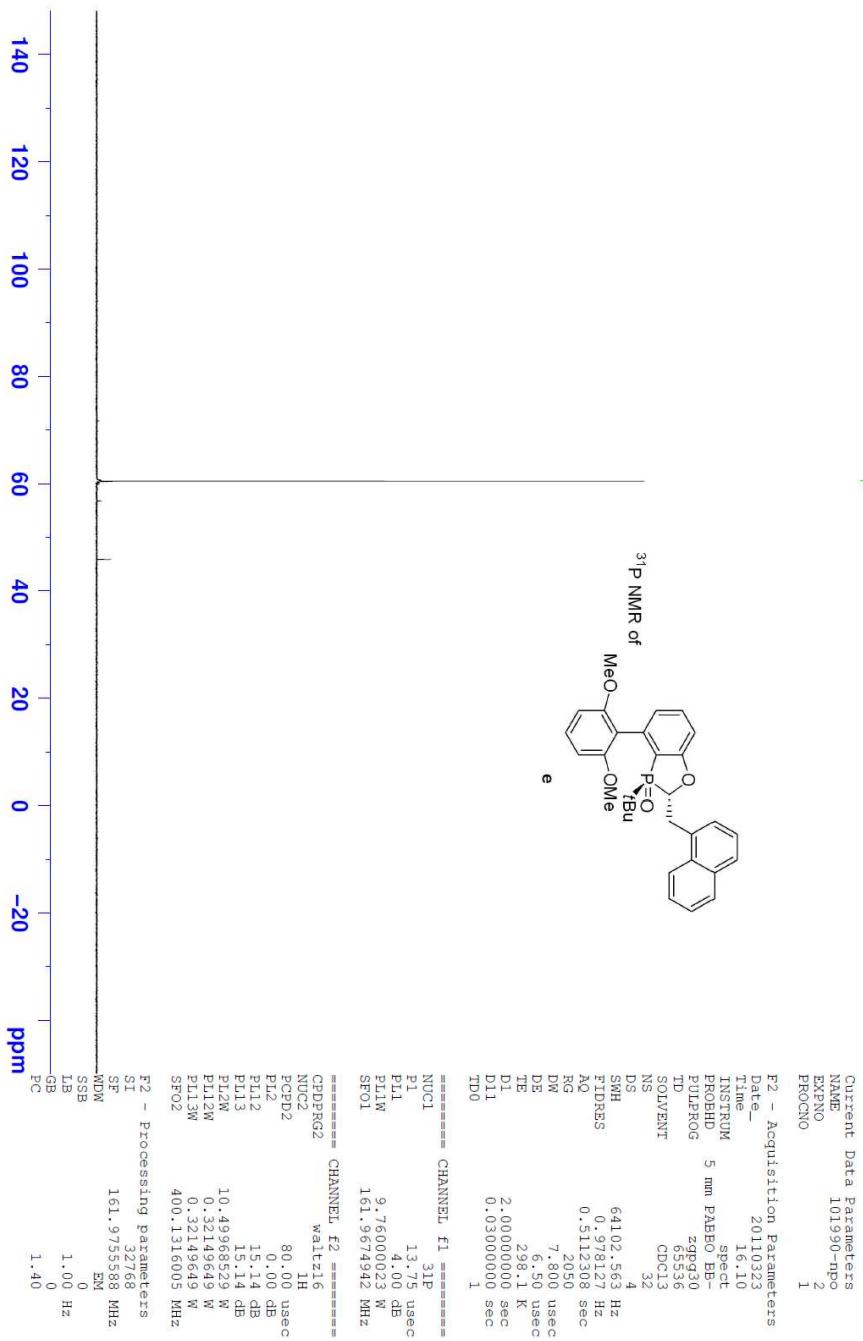
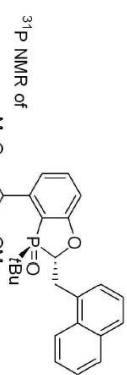


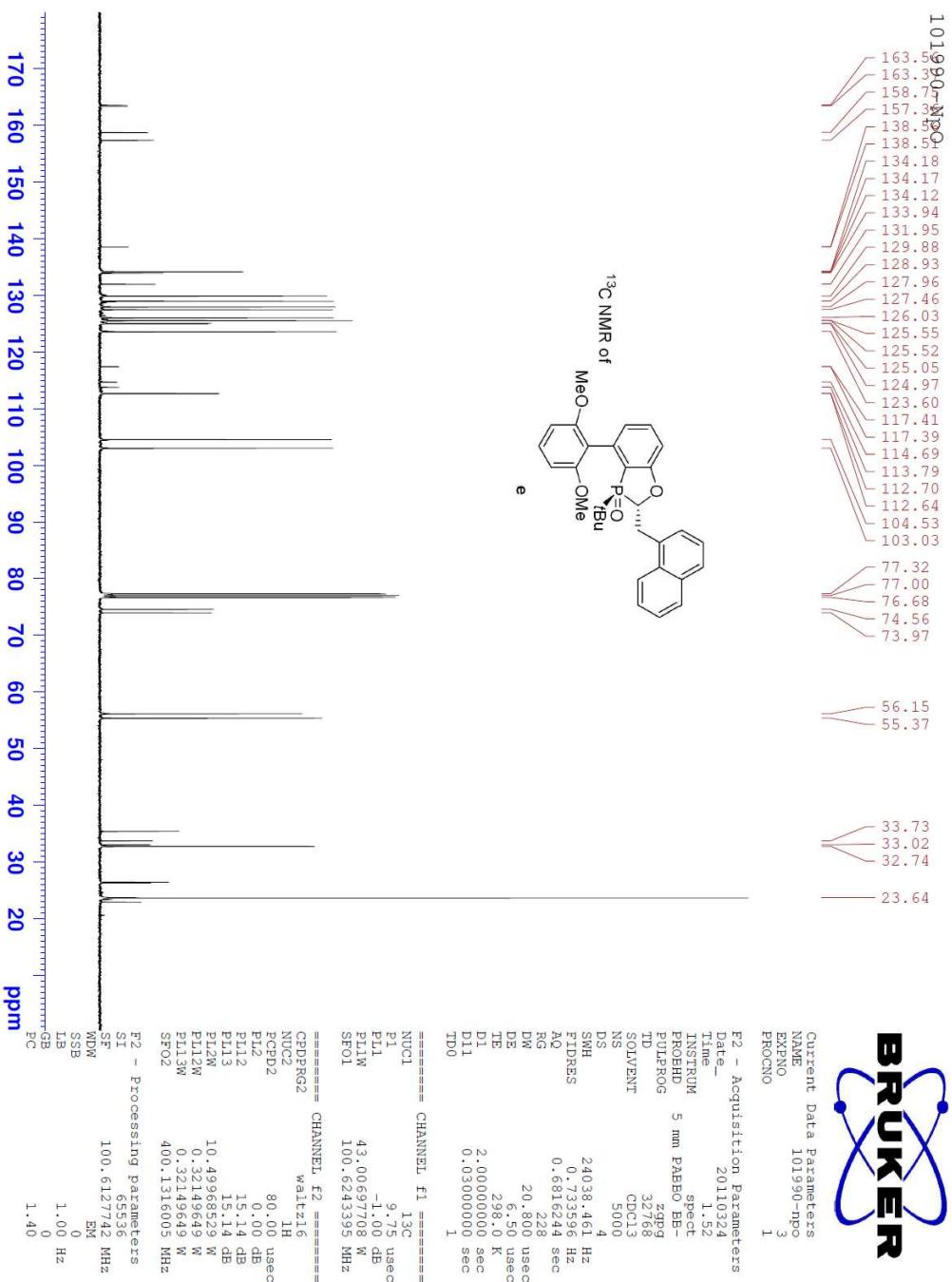


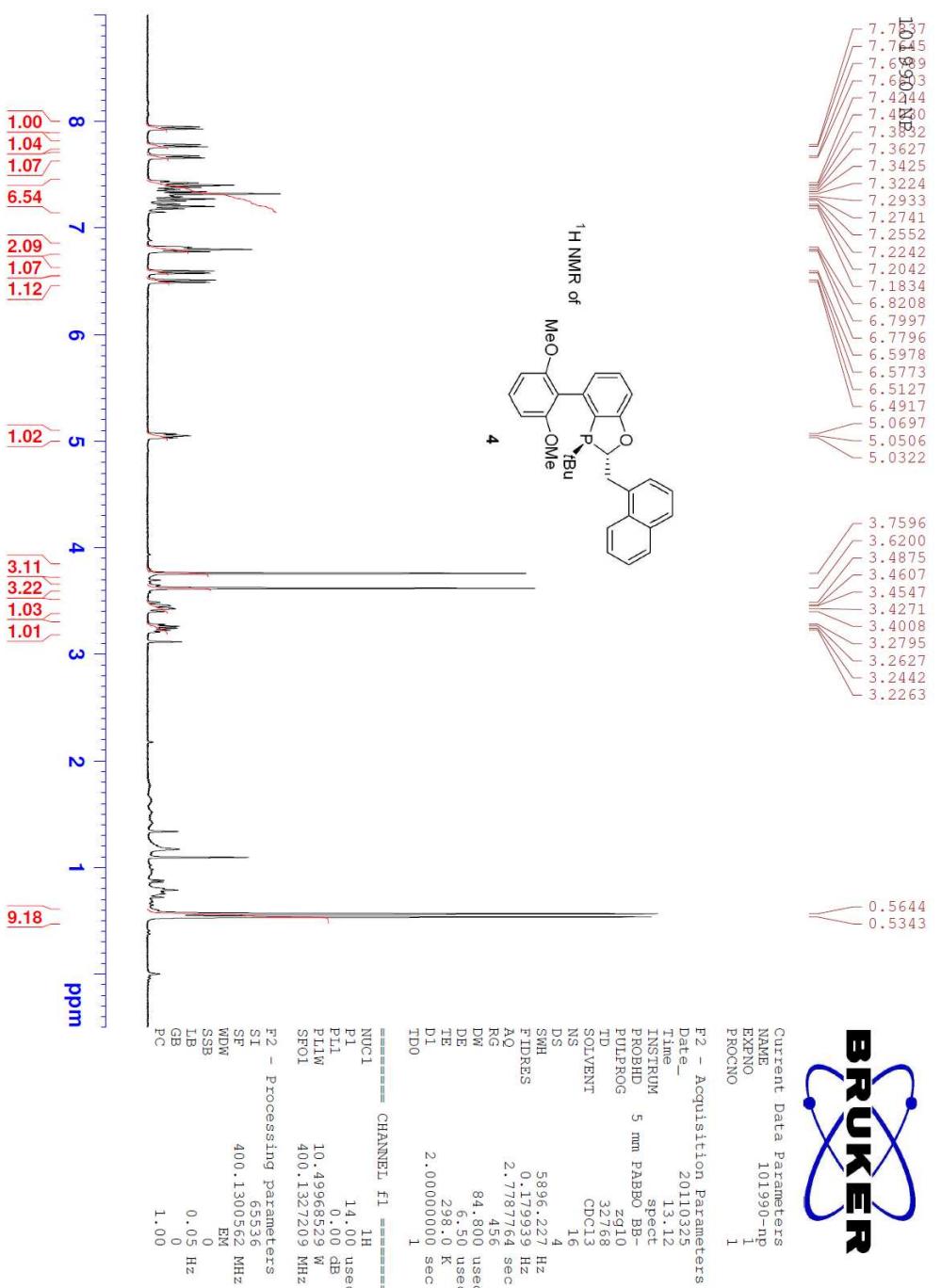


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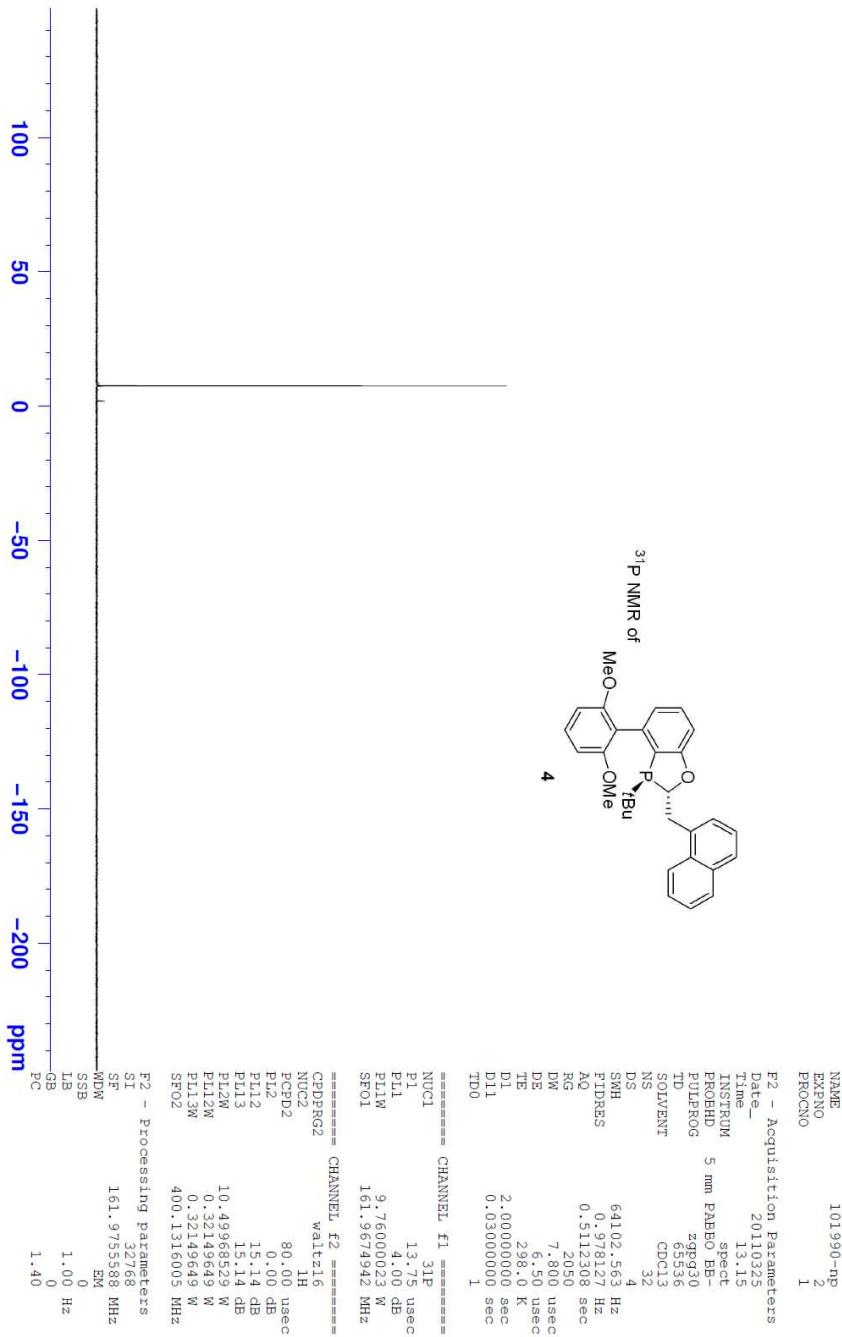
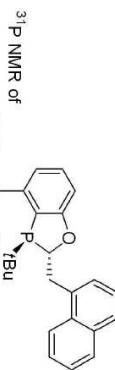


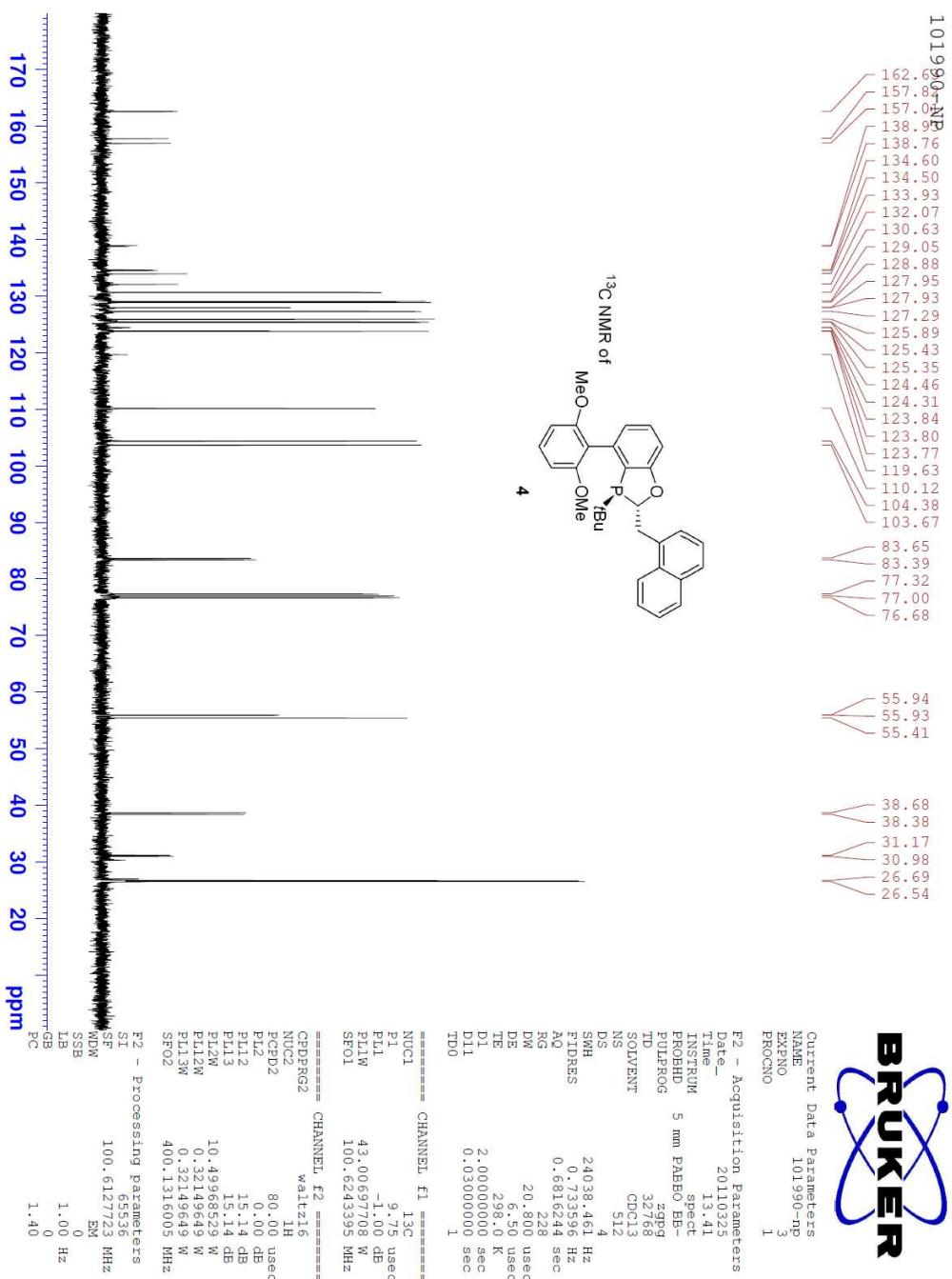






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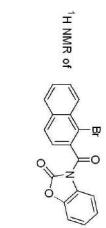


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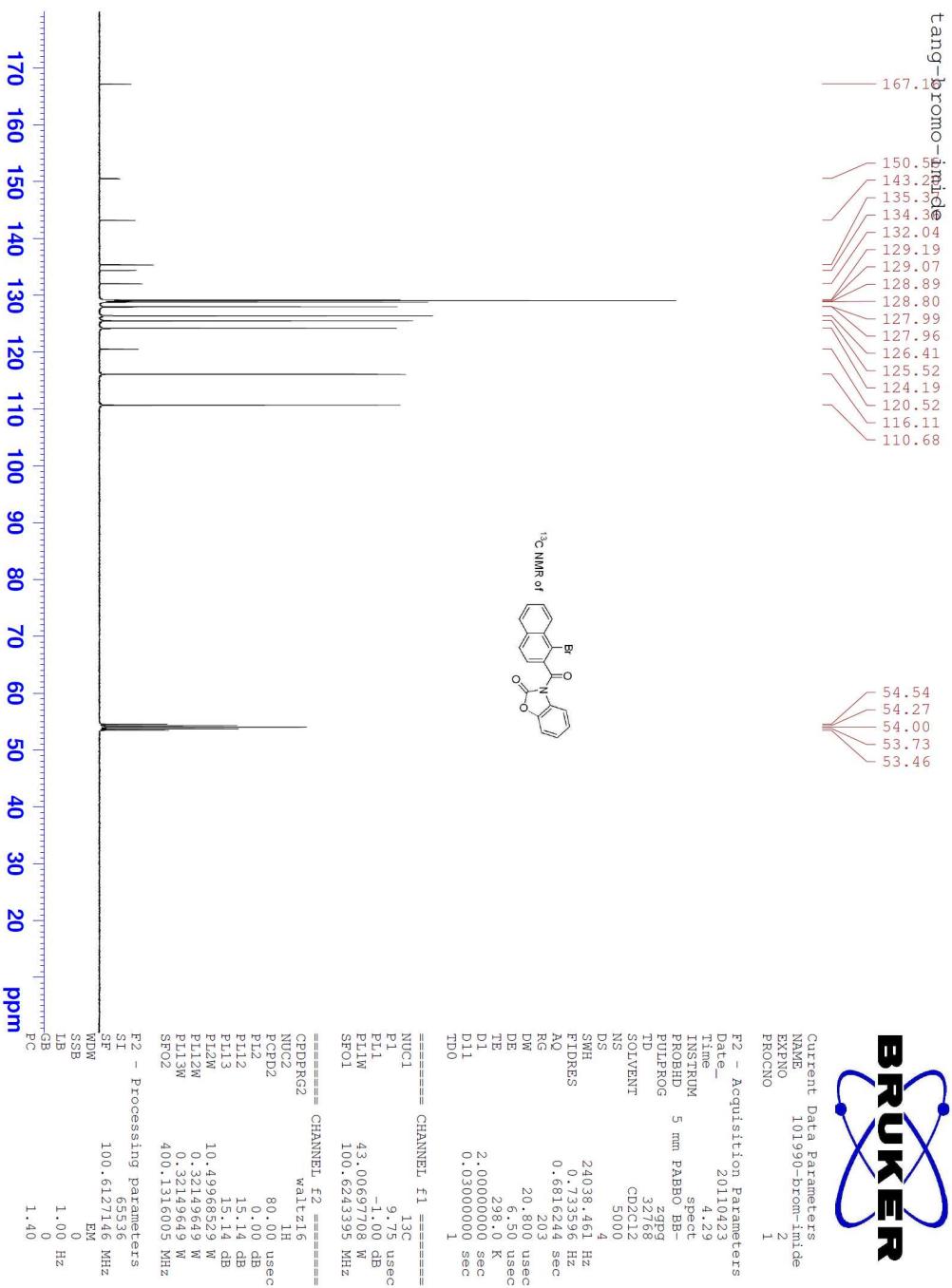


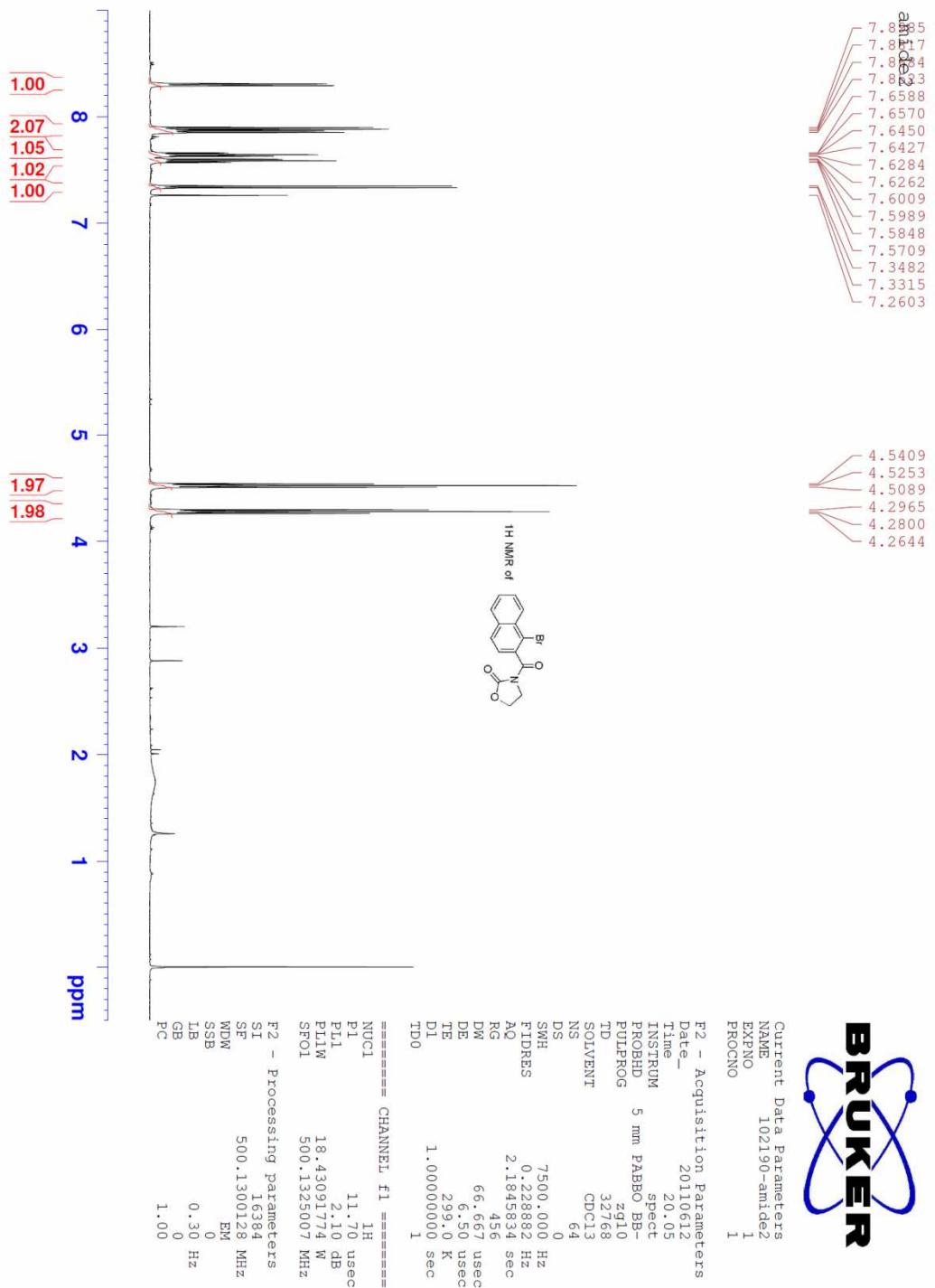
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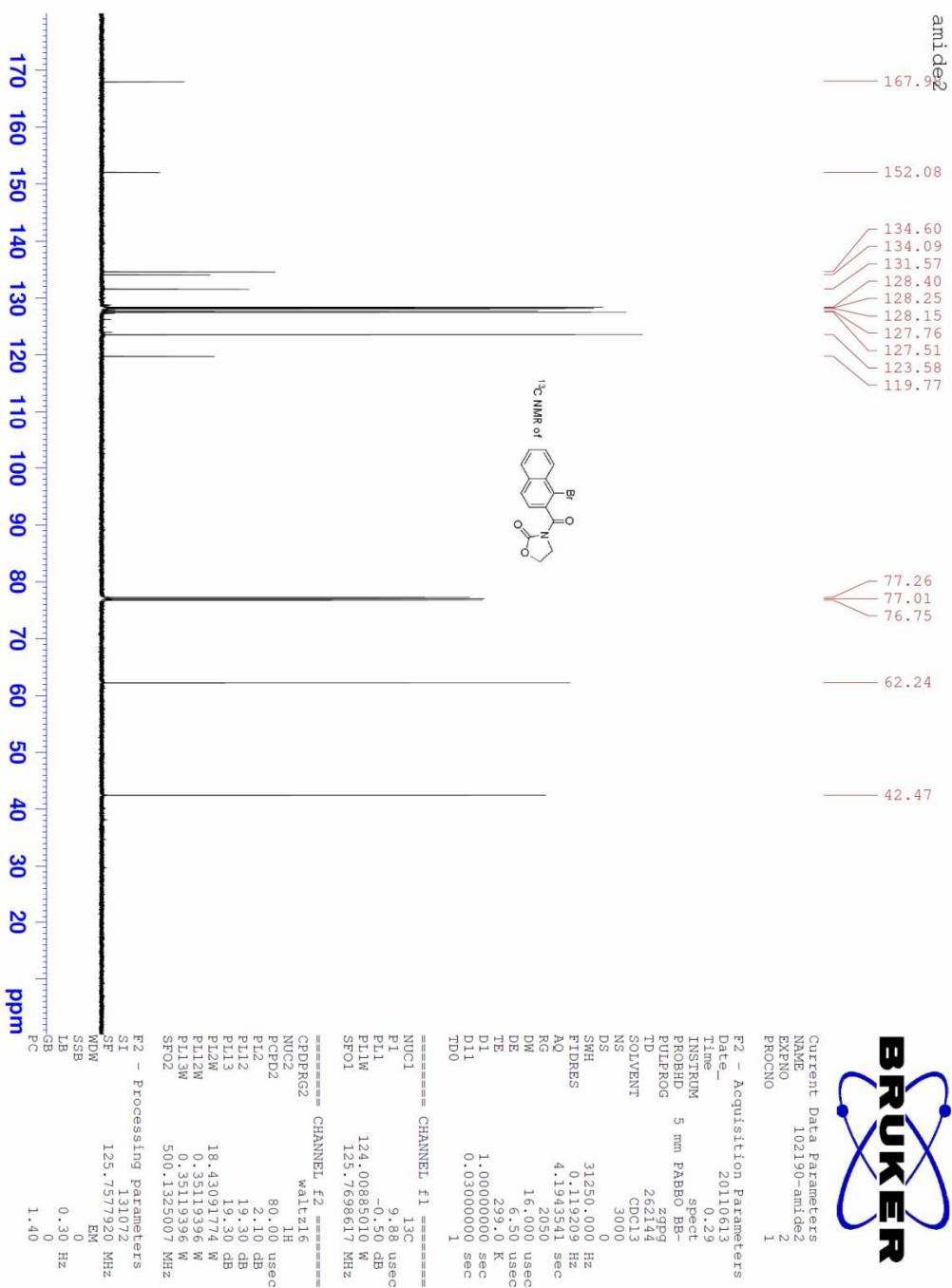
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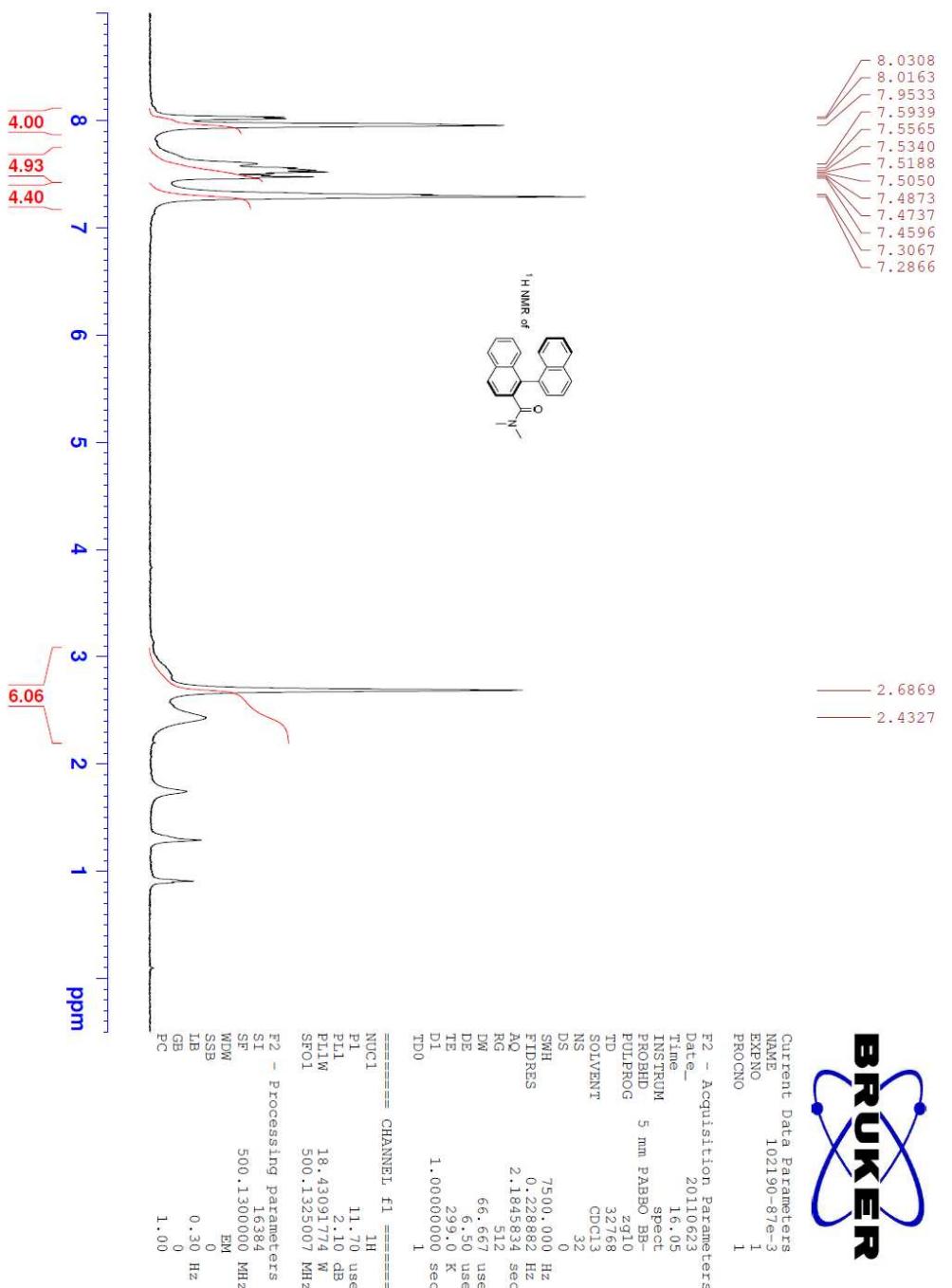


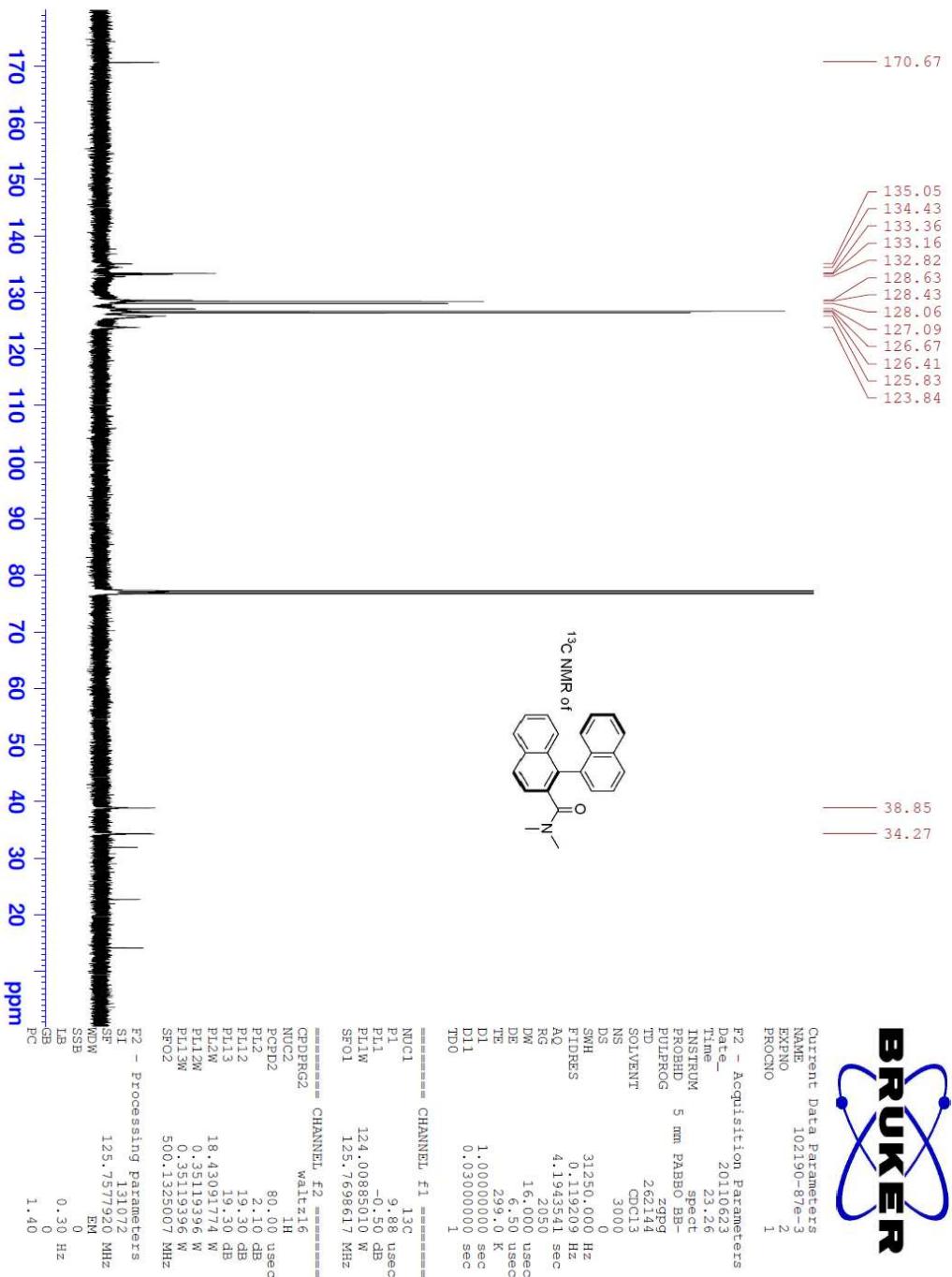
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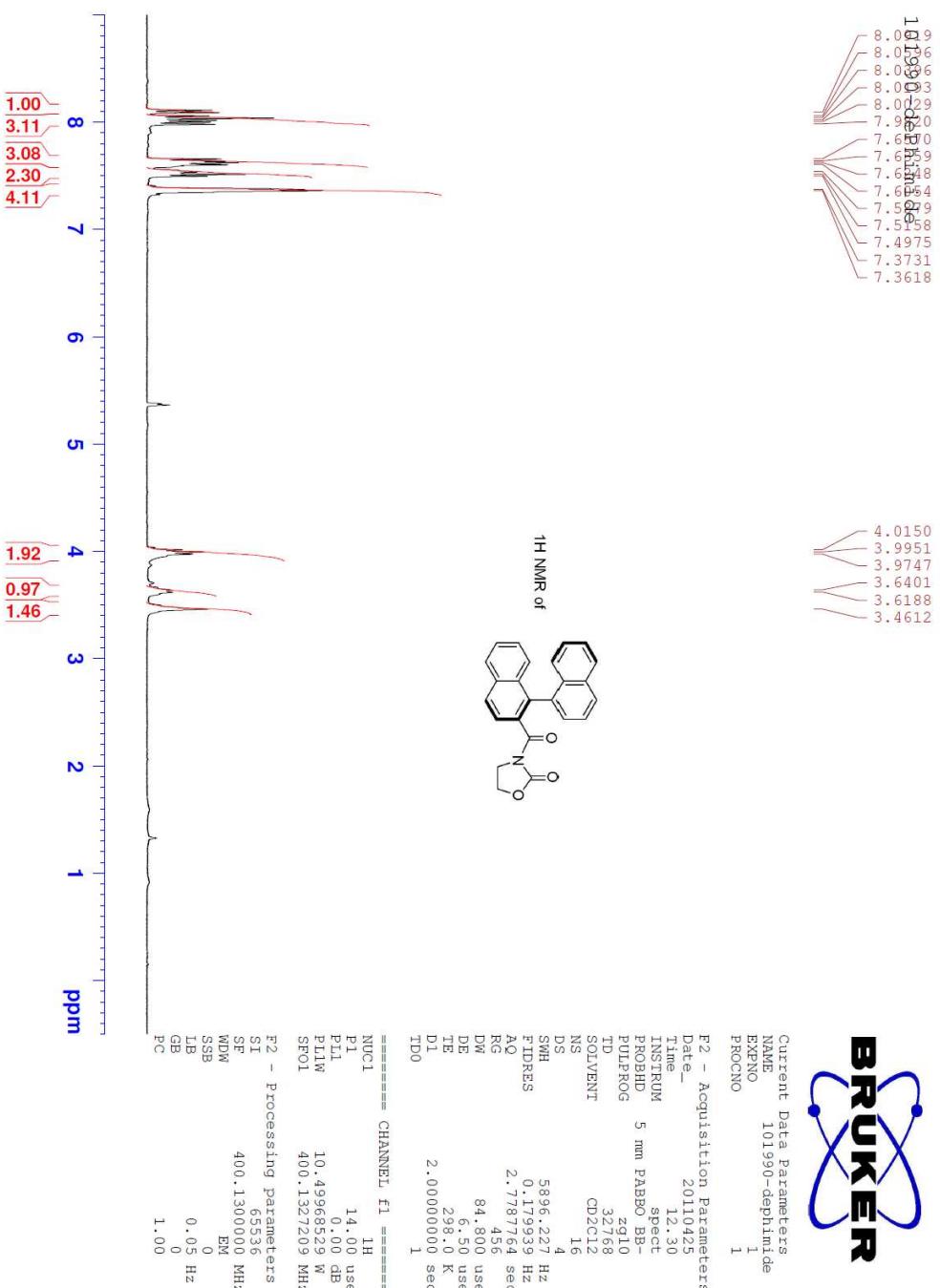


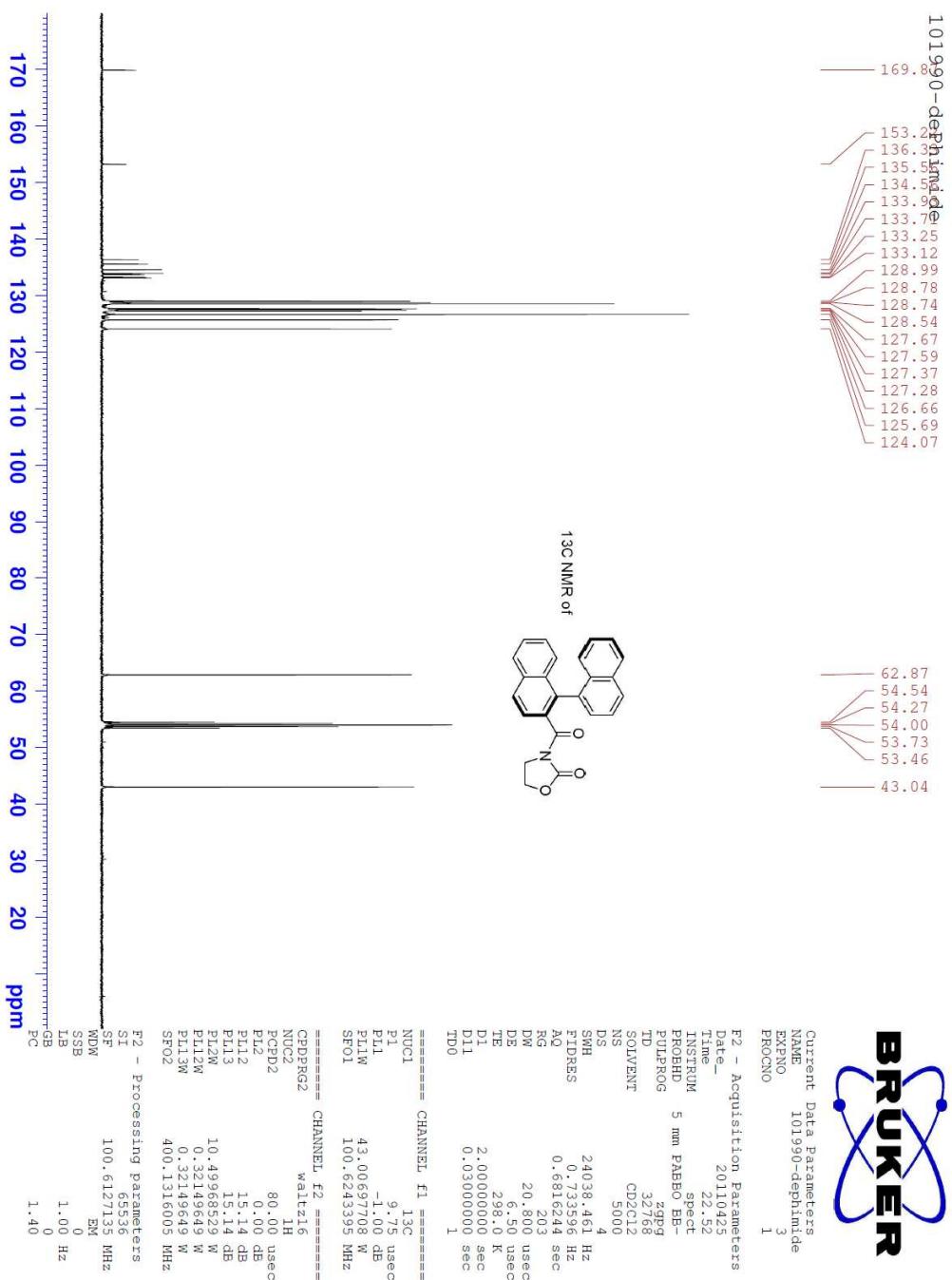


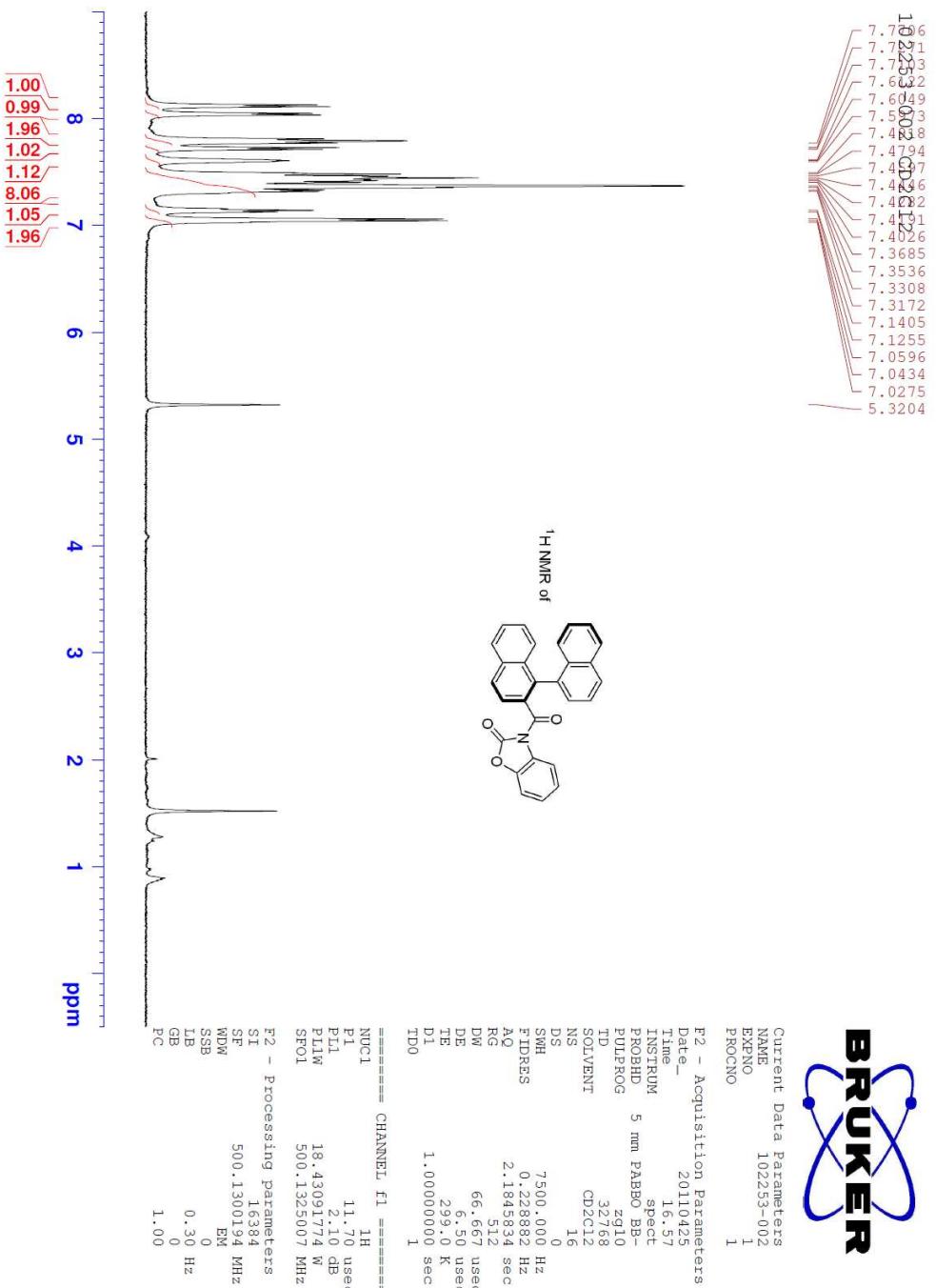


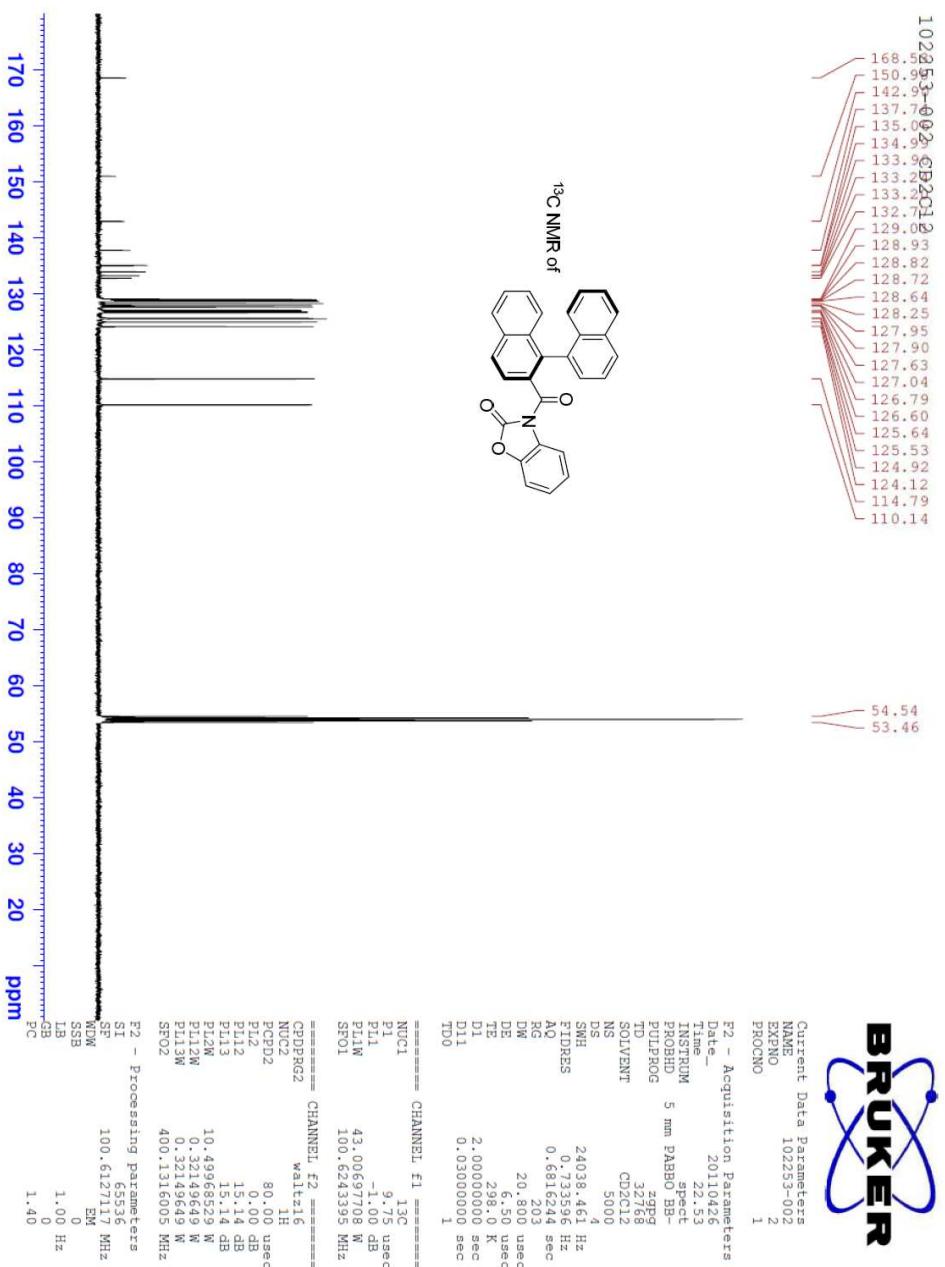




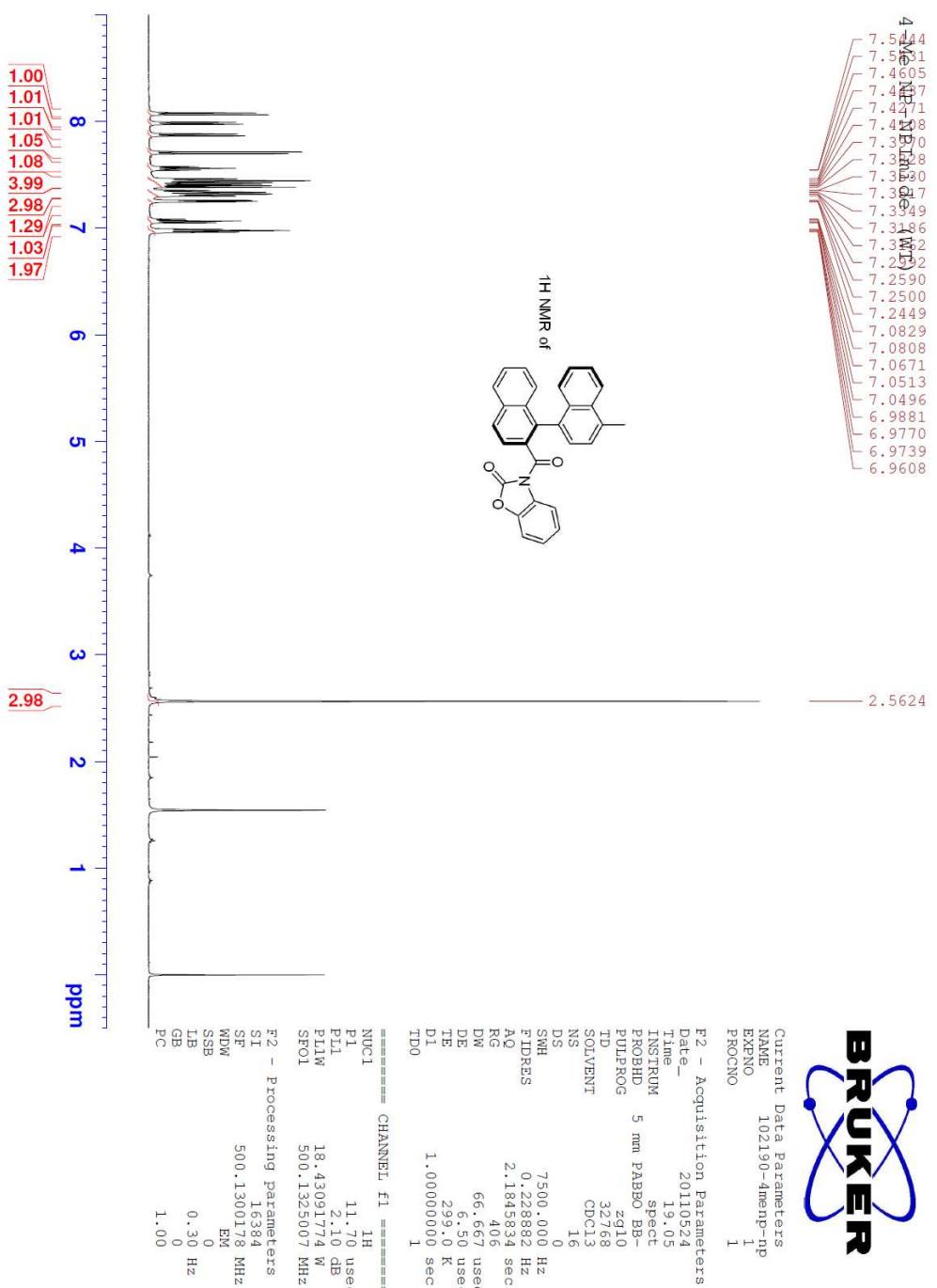


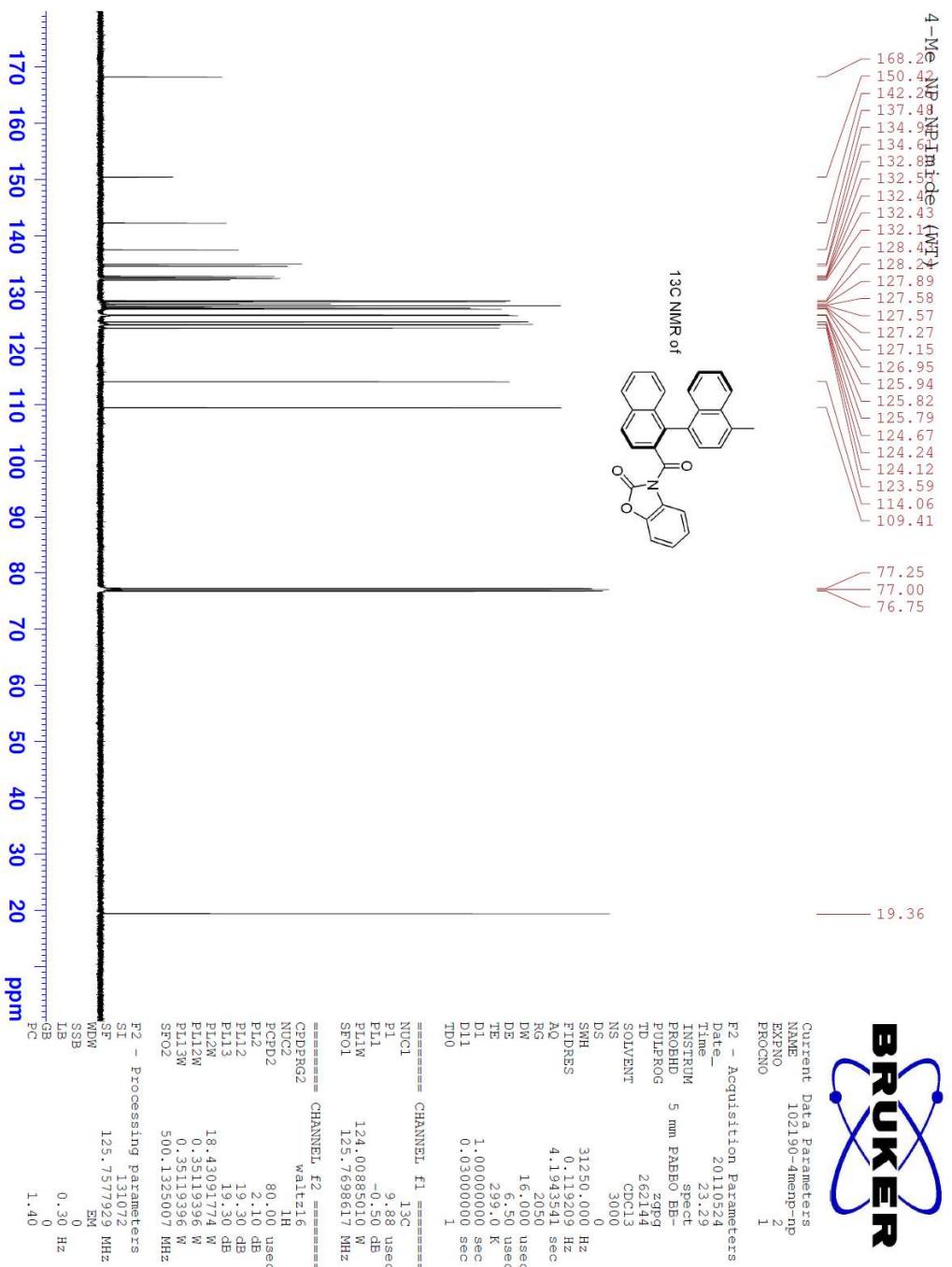


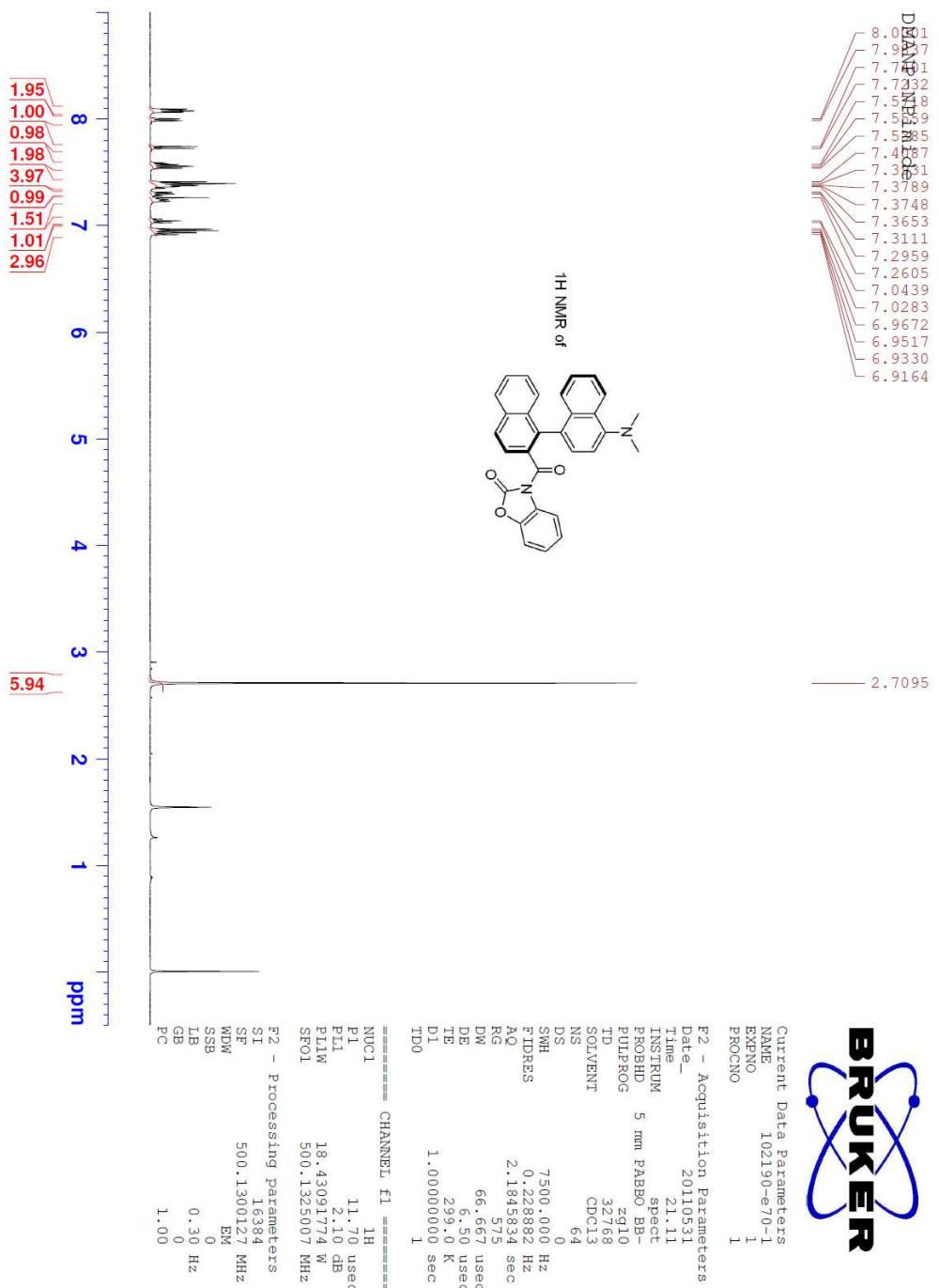


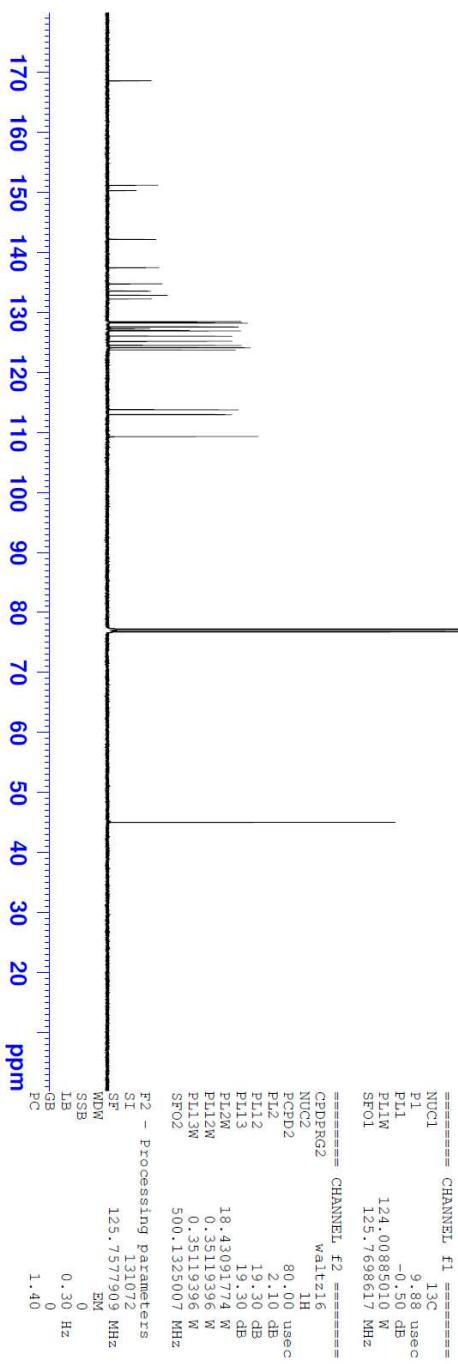
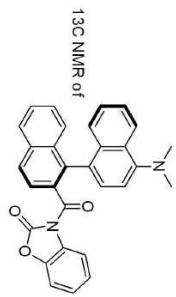
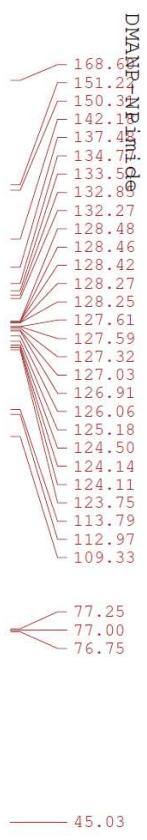


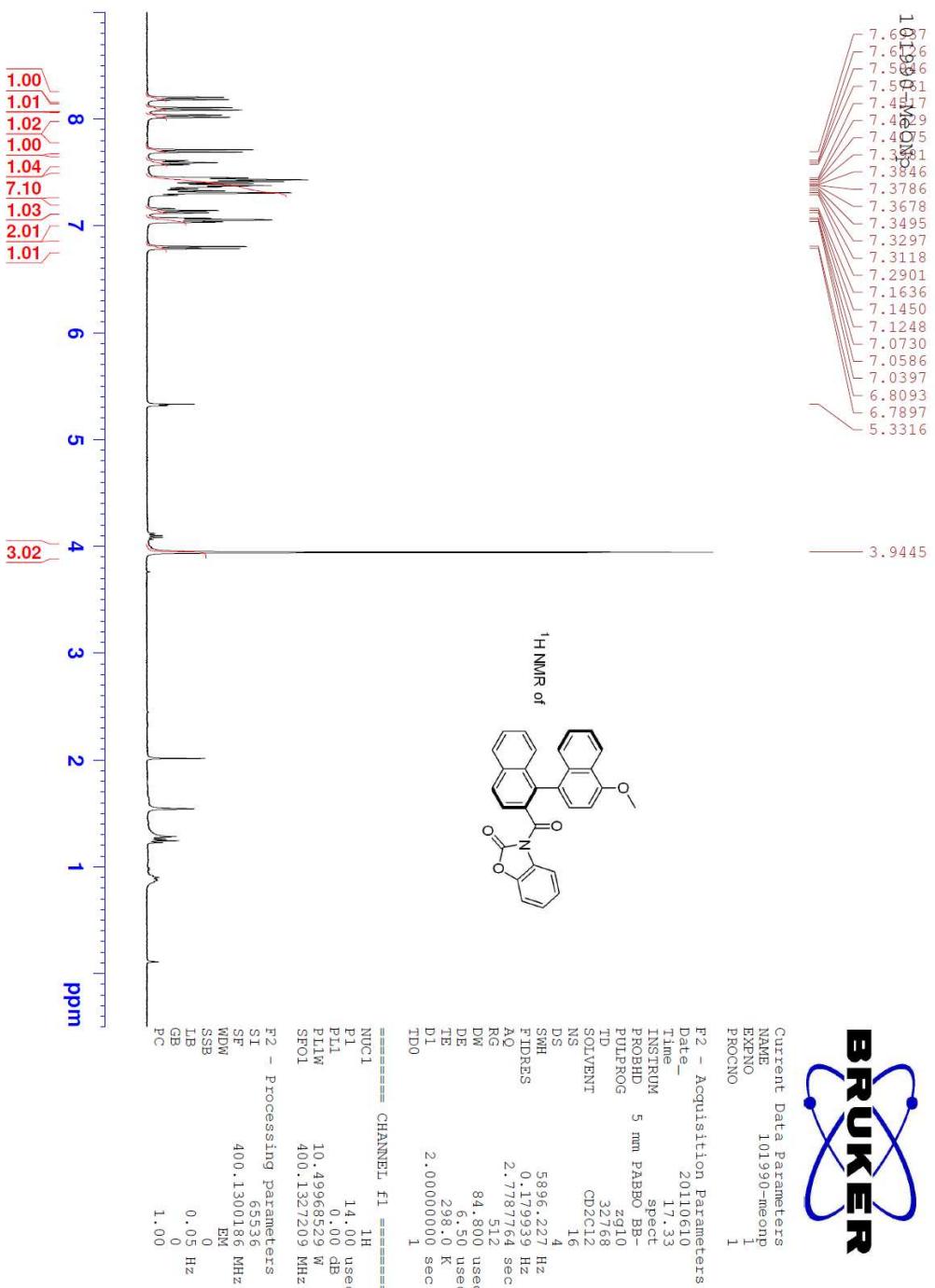
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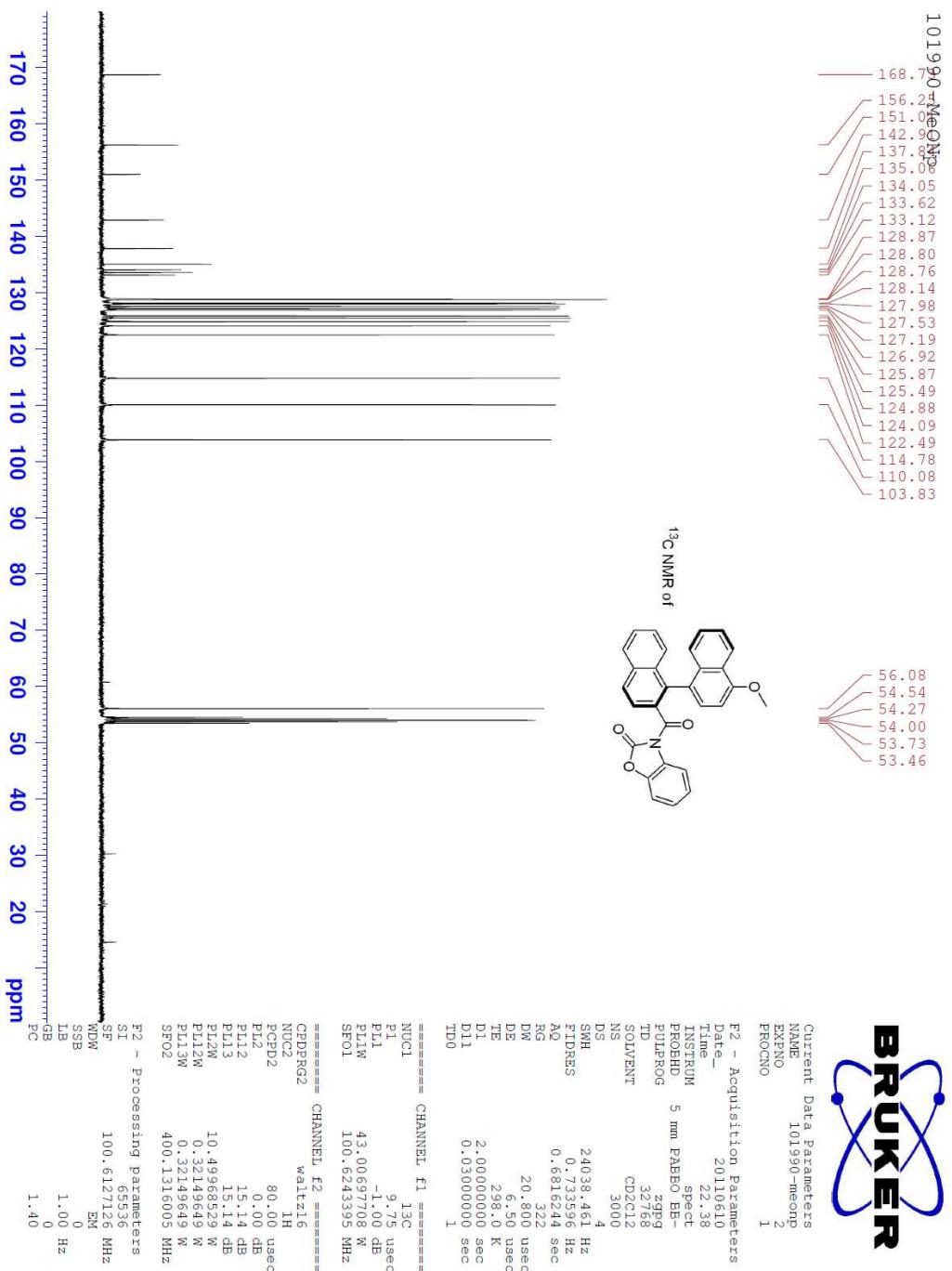


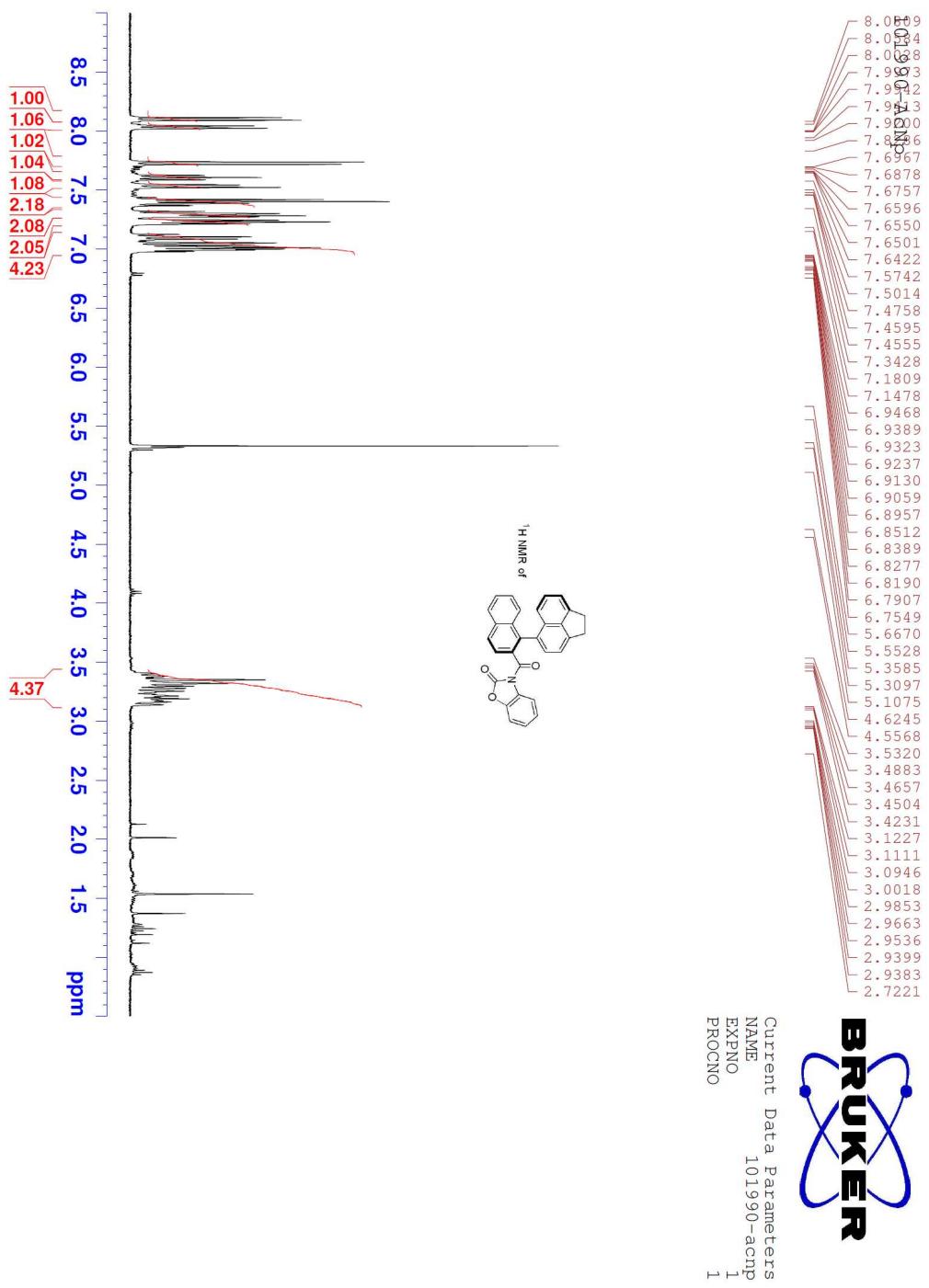


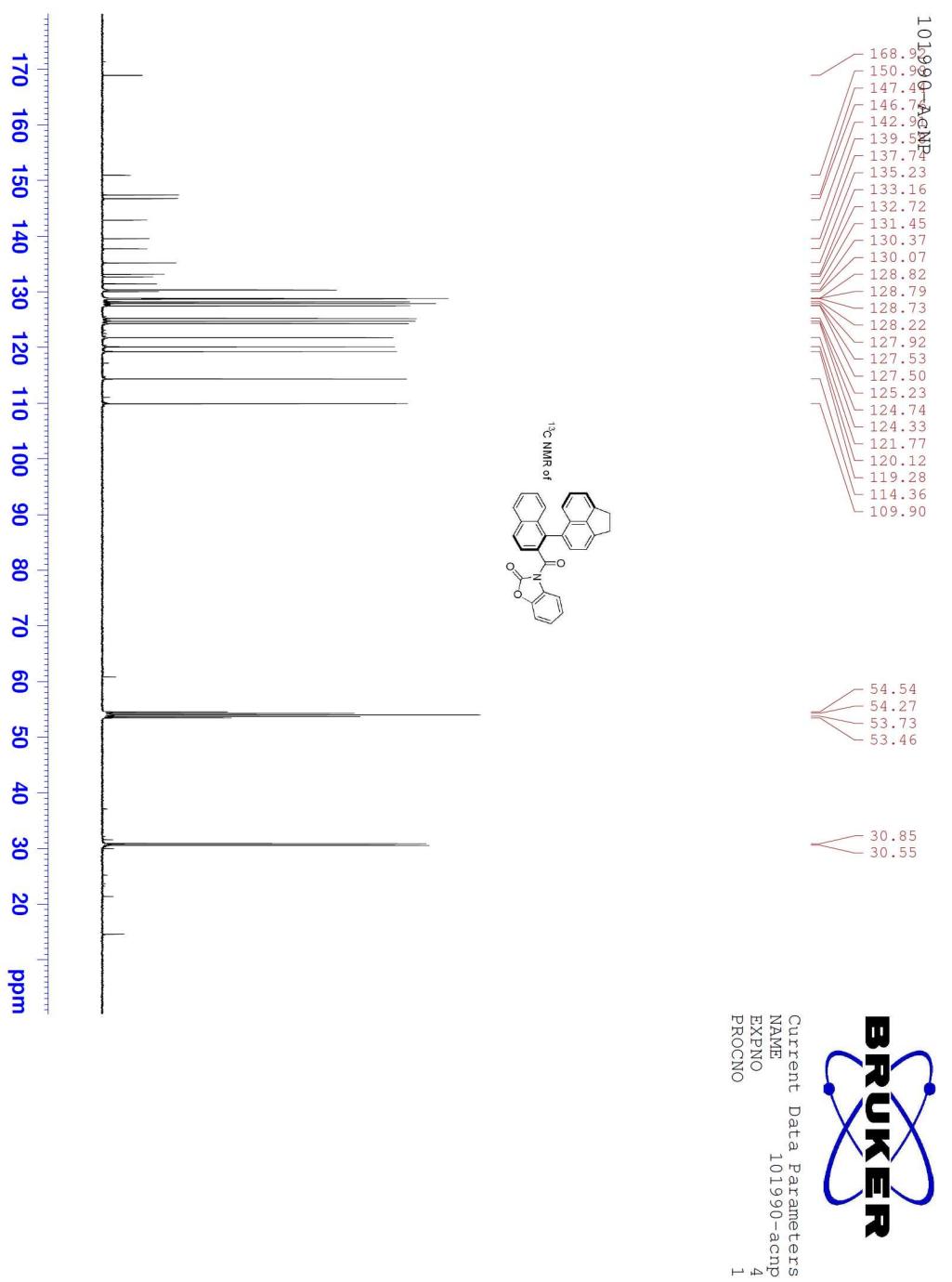




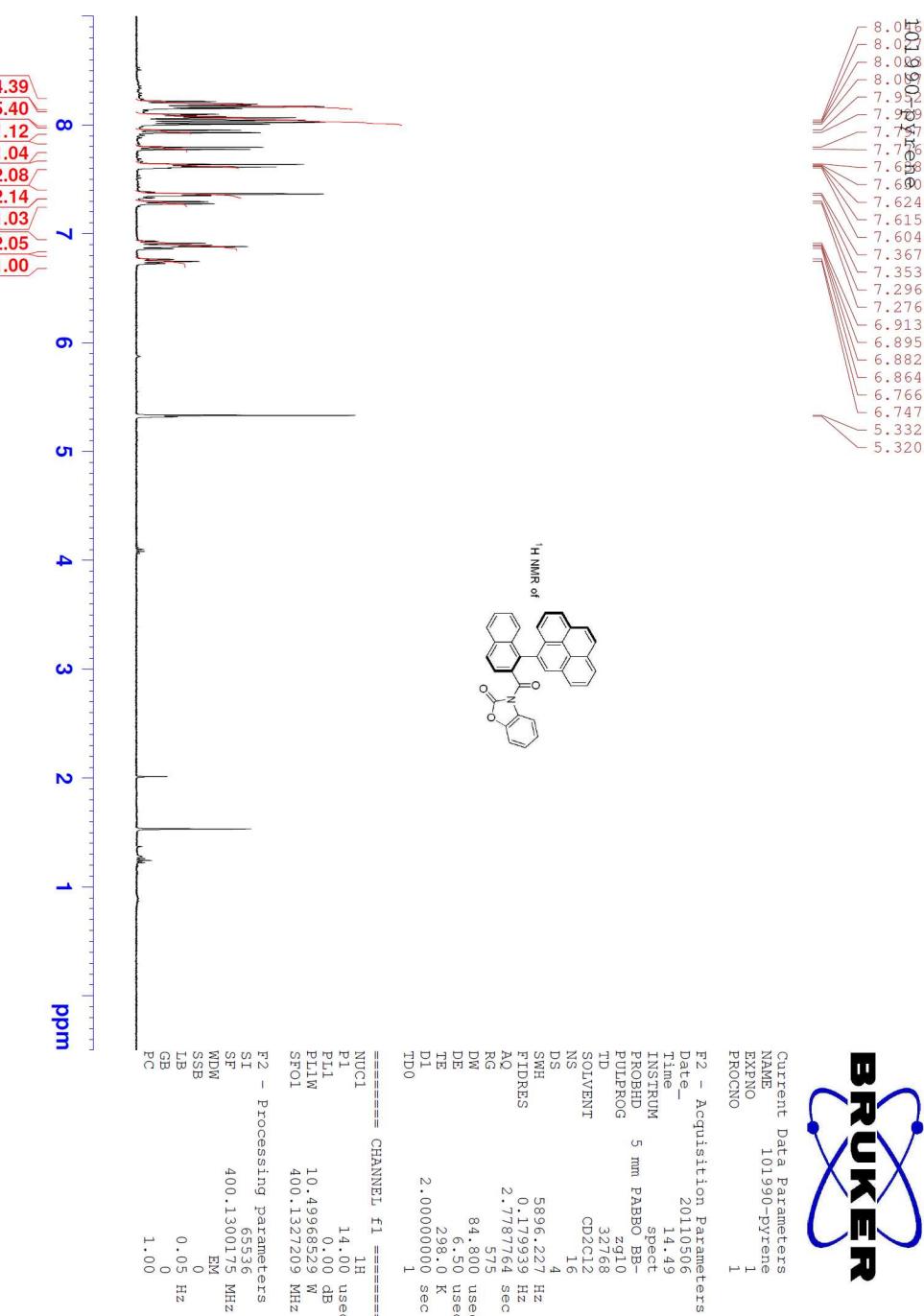
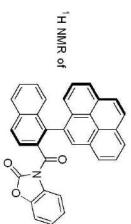


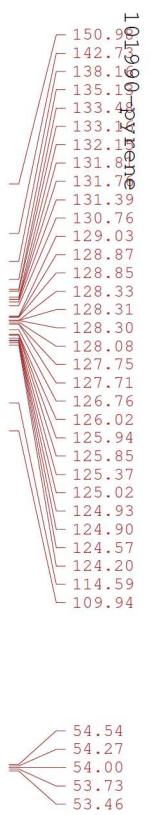






160
 8.012
 8.005
 8.009
 8.025
 7.995
 7.773
 7.761
 7.687
 7.606
 7.6240
 7.6159
 7.6044
 7.3676
 7.3534
 7.2966
 7.2768
 6.9138
 6.8950
 6.8828
 6.8644
 6.7666
 6.7471
 5.3323
 5.3202





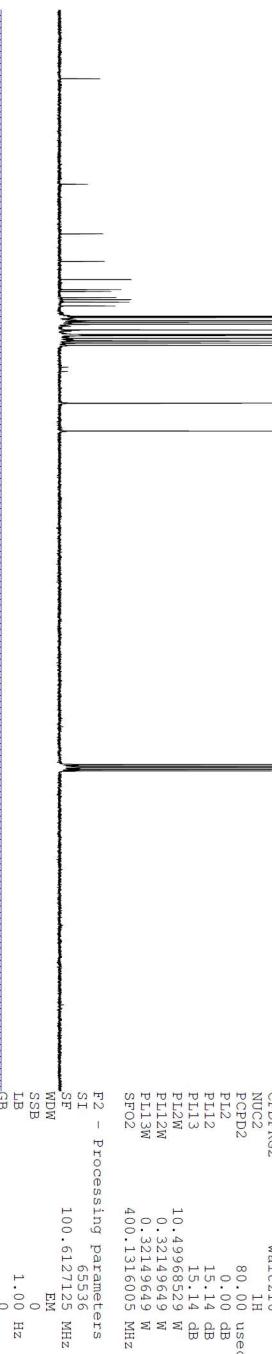
54.54
54.27
54.00
53.73
53.46

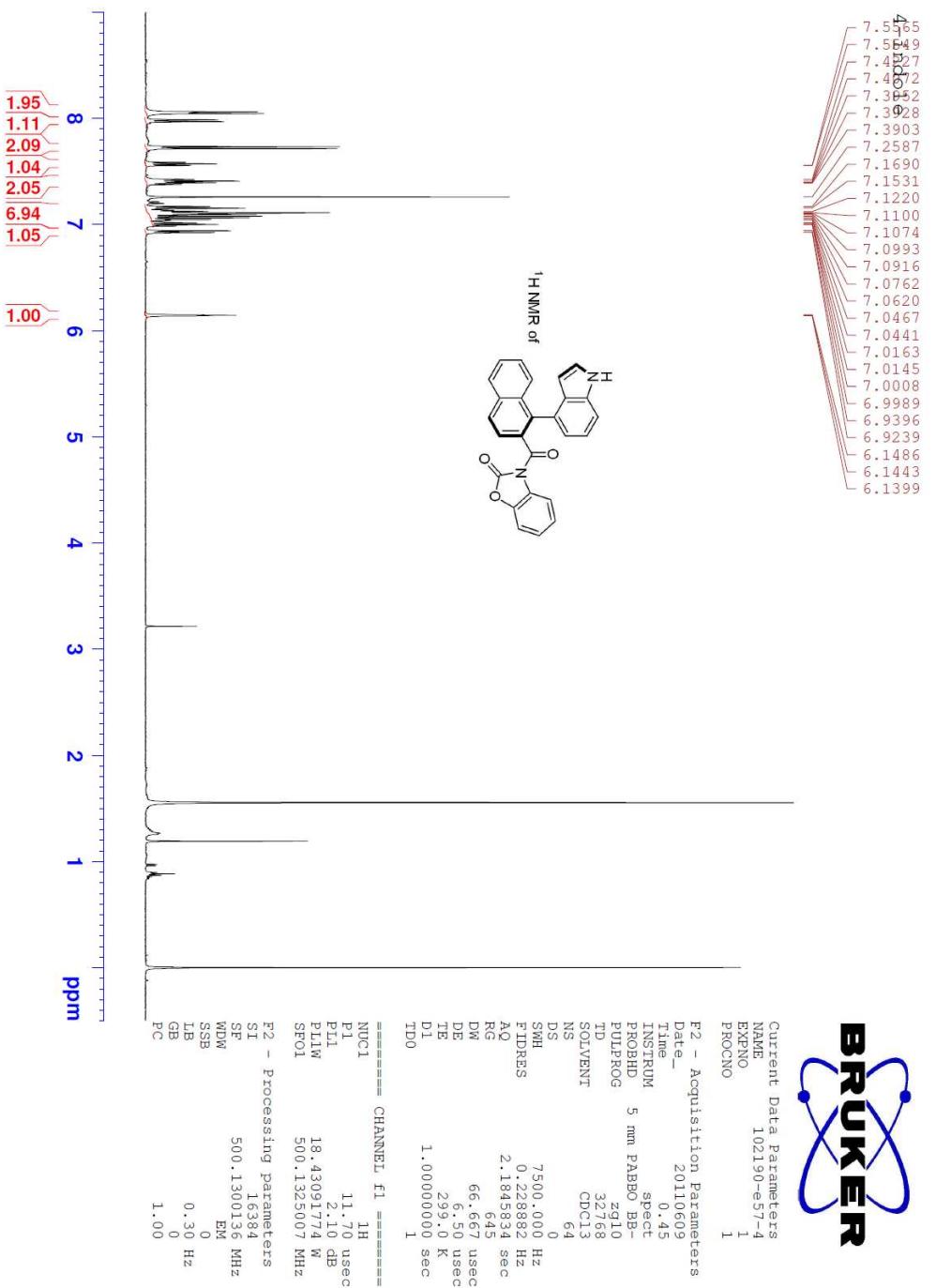


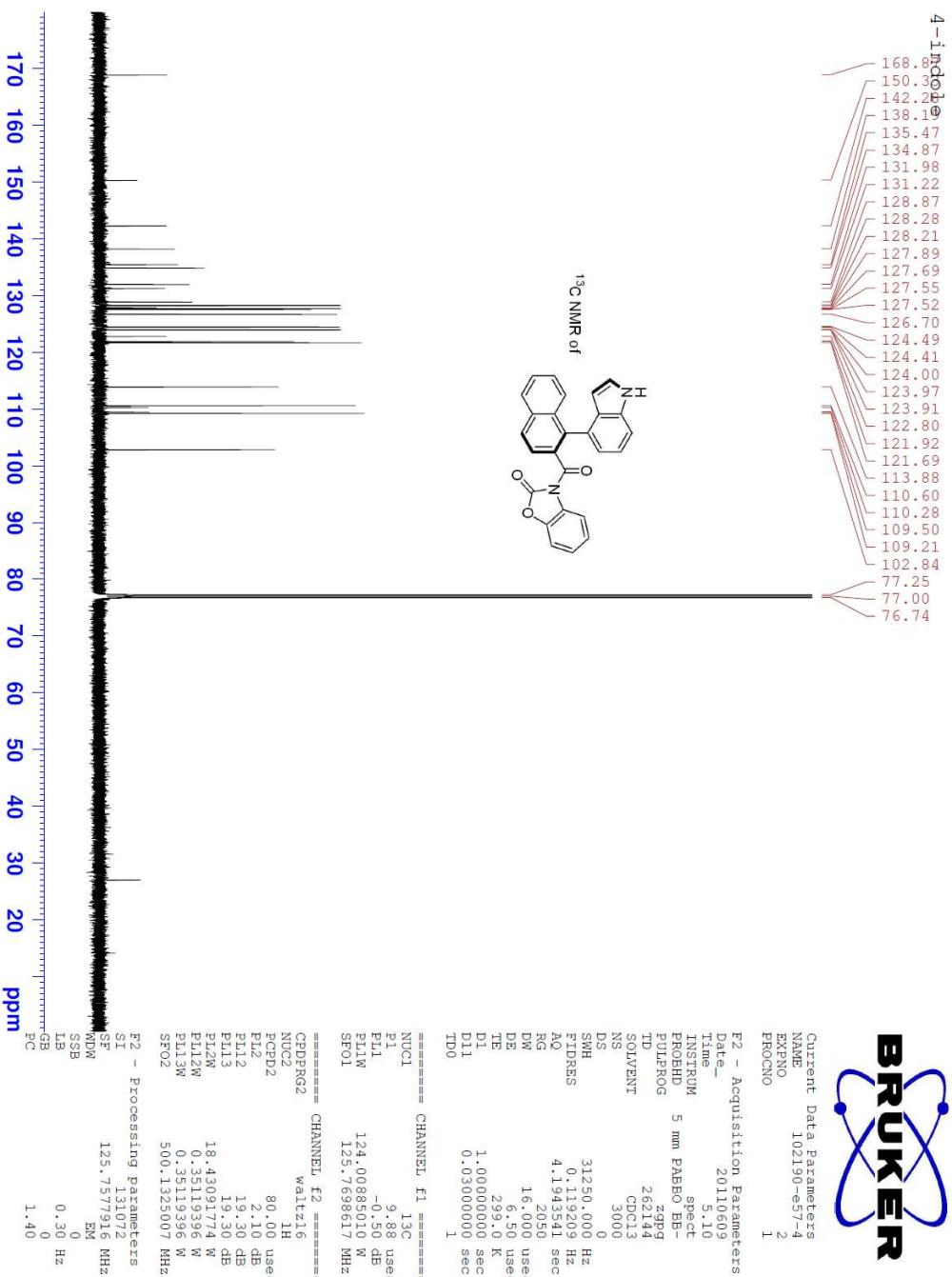
Current Data Parameters
NAME 101990-Pyrene
EXPNO 2
PROCNO 1

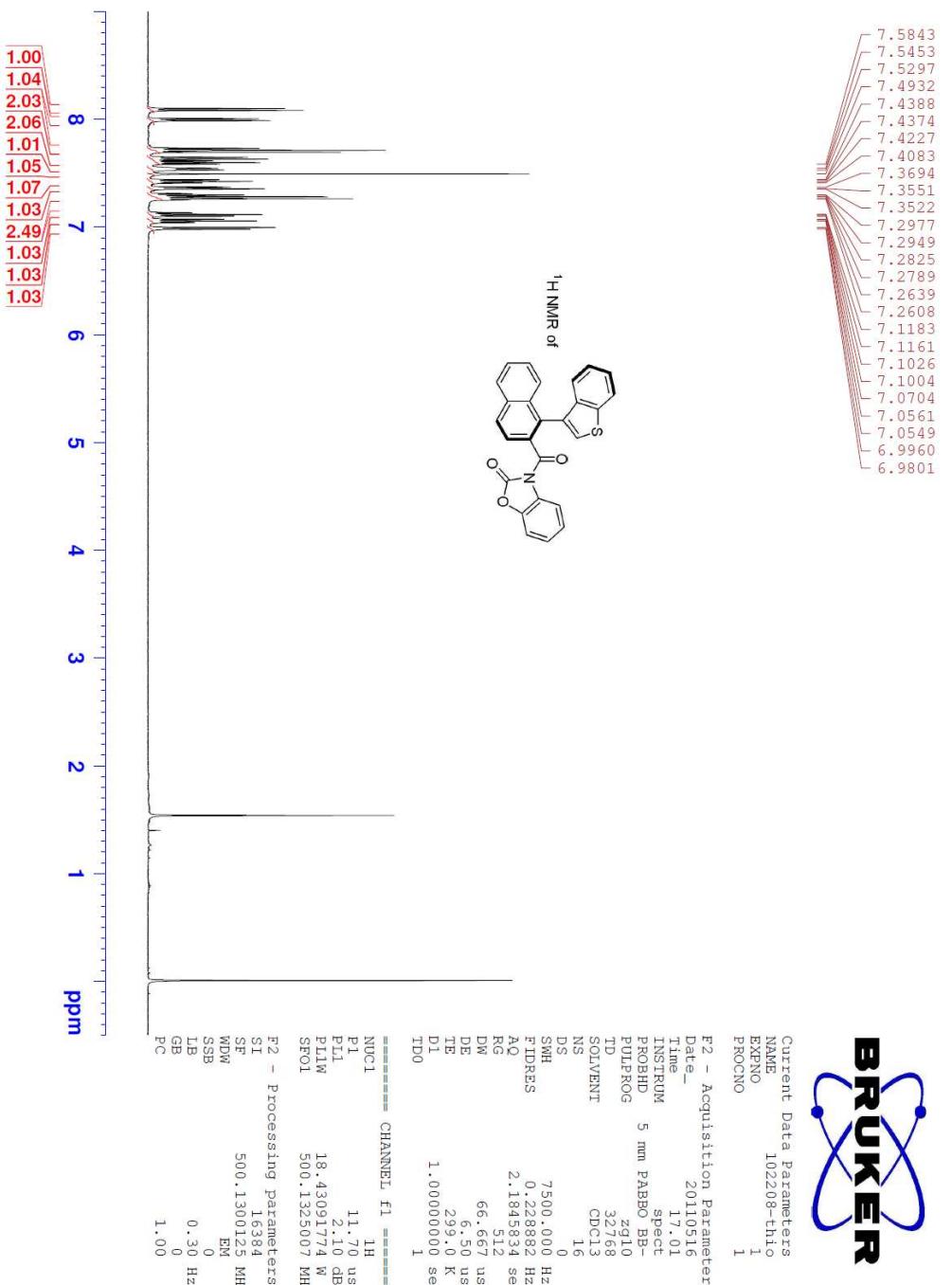
F2 - Acquisition Parameters
Date_ 20110507
Time_ 22.53
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zapg
TD 32768
SOLVENT CDCl₂
NS 5000
DS 4
SWH 24038.461 Hz
FIDRES 0.73396 Hz
AQ 0.6816244 sec
RG 228
DW 20.800 usec
DE 6.50 usec
TE 298.0 K
D1 2.000000 sec
D11 0.0300000 sec
TDO 1

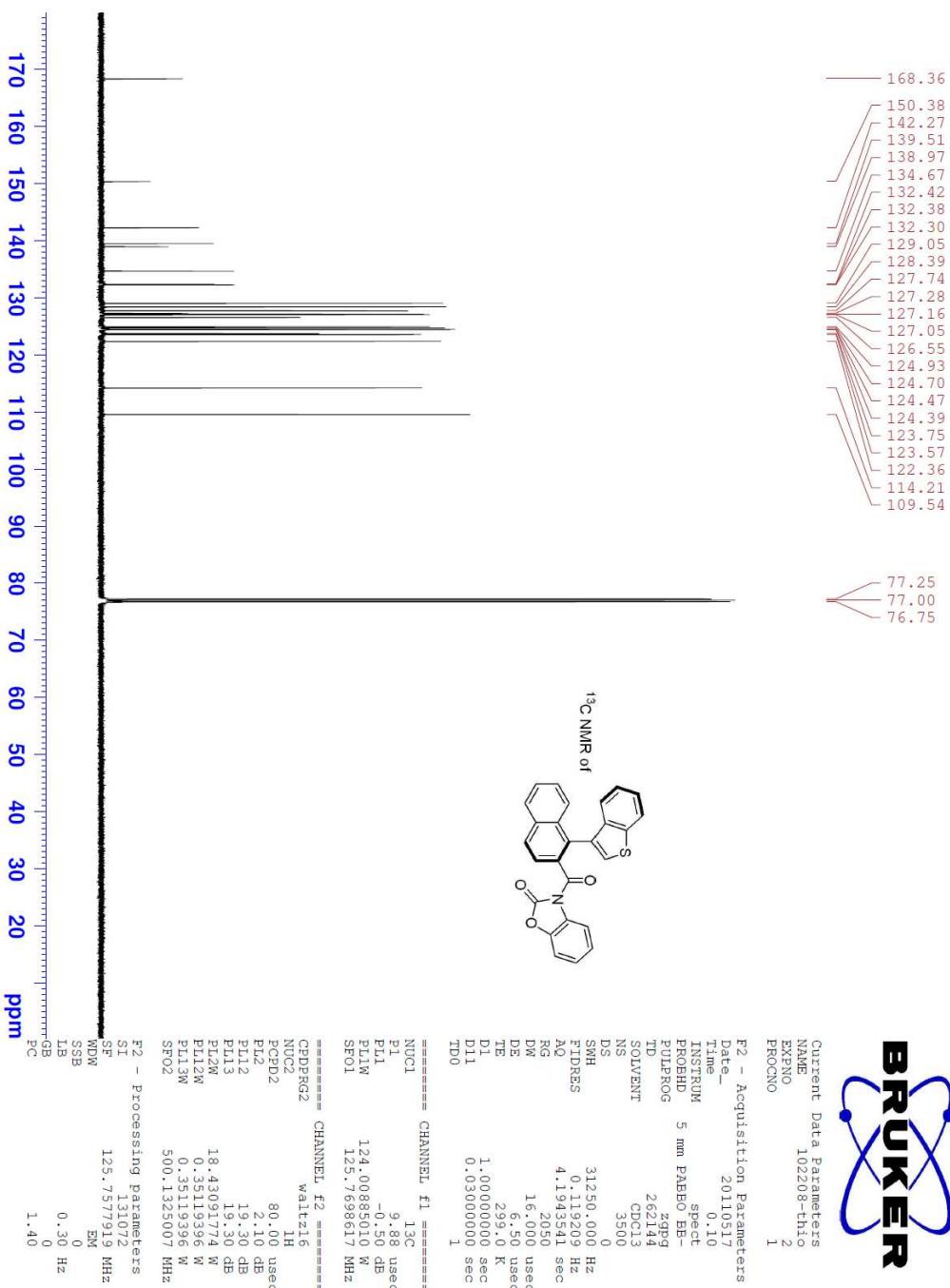
===== CHANNEL f1 =====
NUC1 13C
P1 9.75 usec
PLW -1.00 dB
P1M 43.0969708 MHz
SFOL 100.6243395 MHz
===== CHANNEL f2 =====
CCDPPIR2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL1 0.00 dB
PL12 15.14 dB
PL13 15.14 dB
PL2W 1.0-4.9968929 W
PL12W 0.32149649 W
PL13W 4.00-13.16005 MHz
SF02

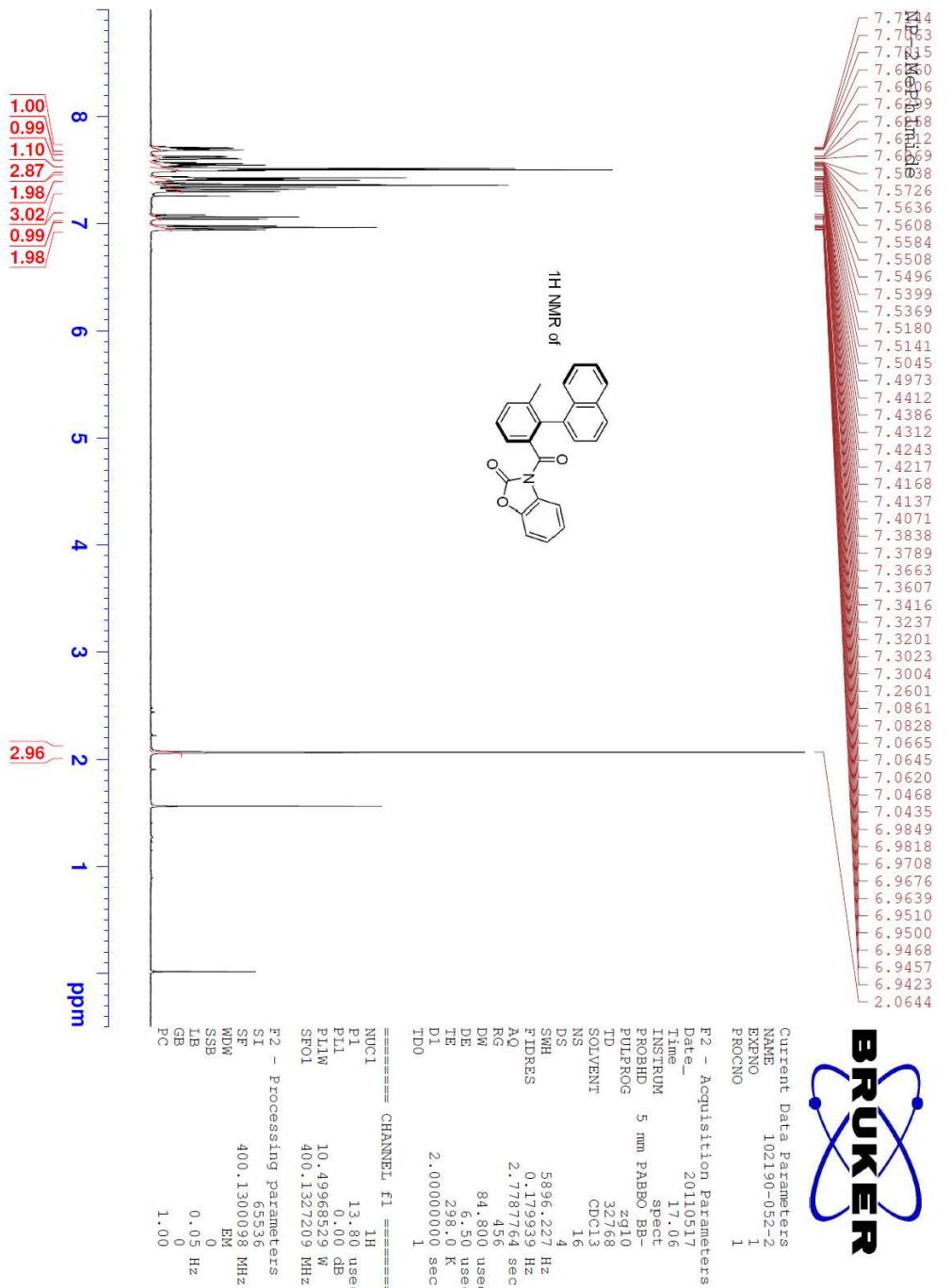


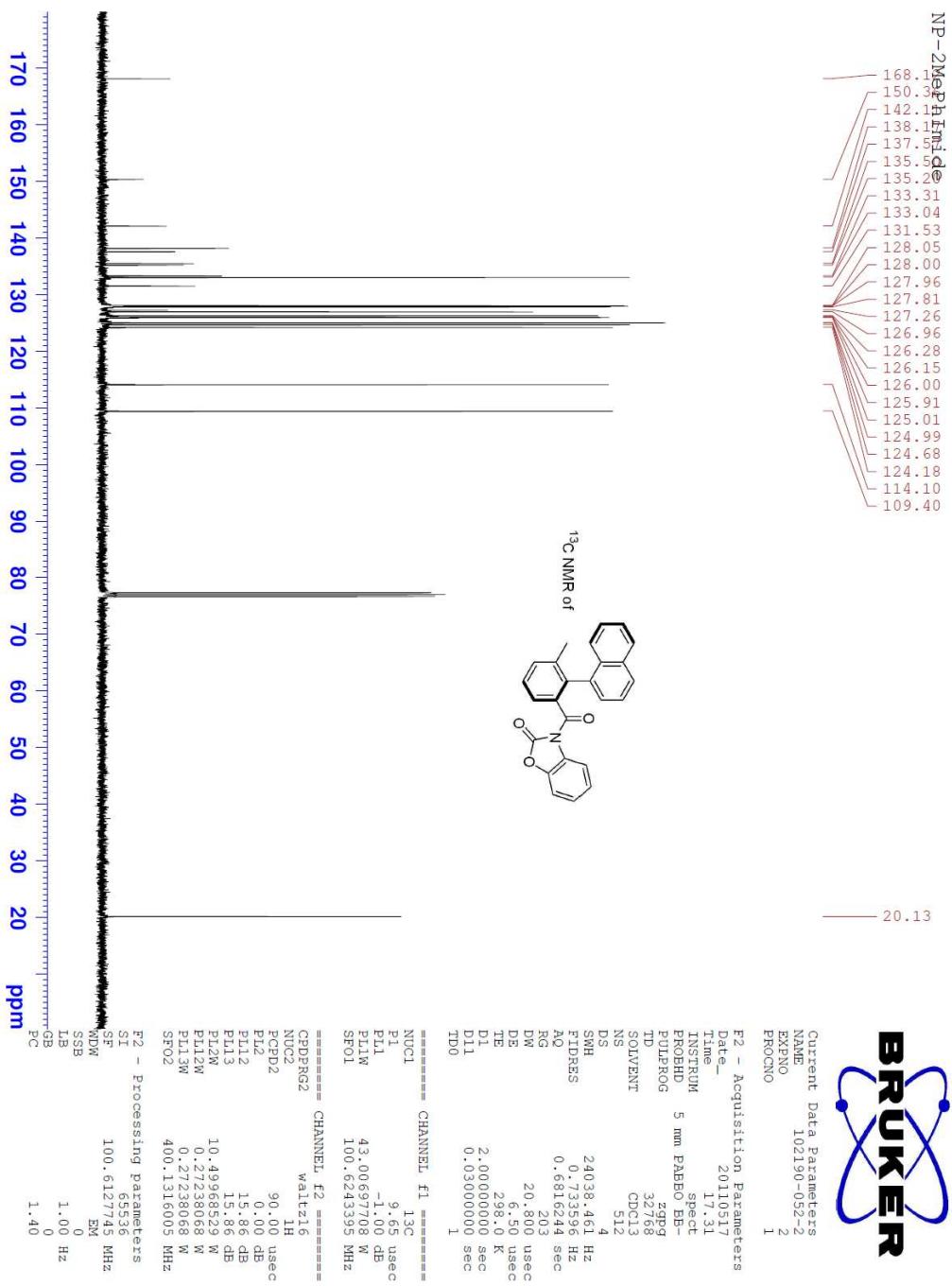


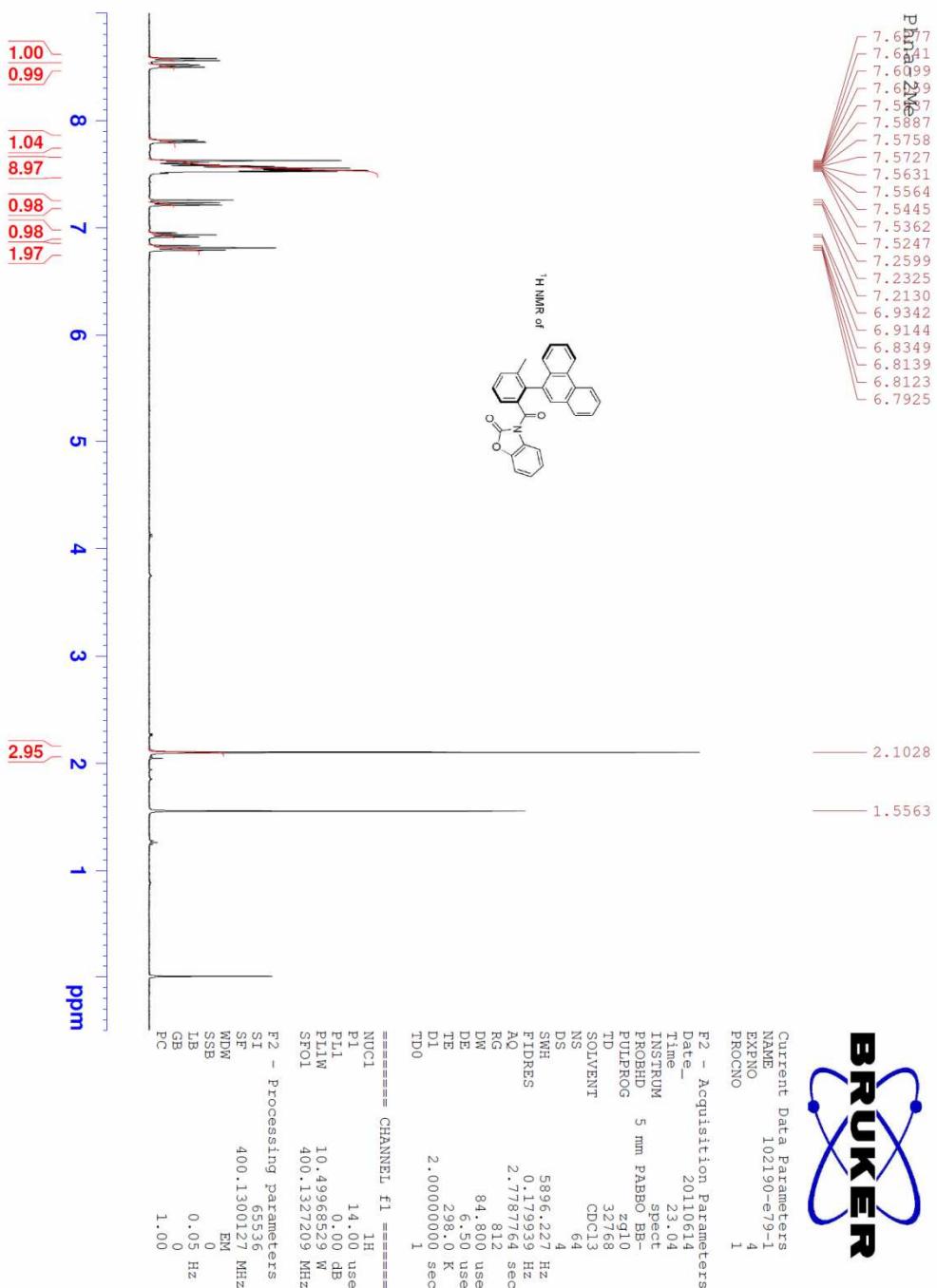


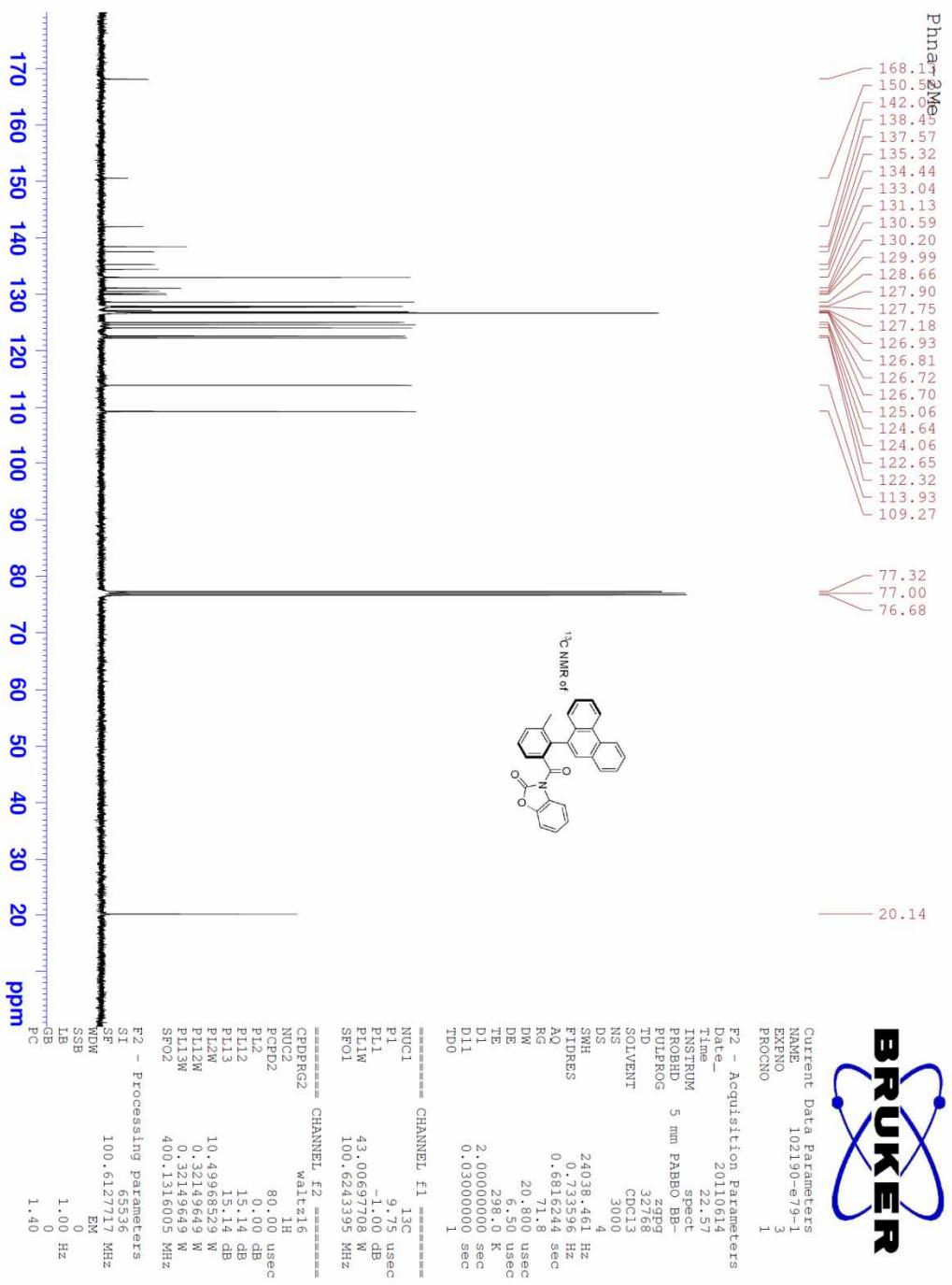


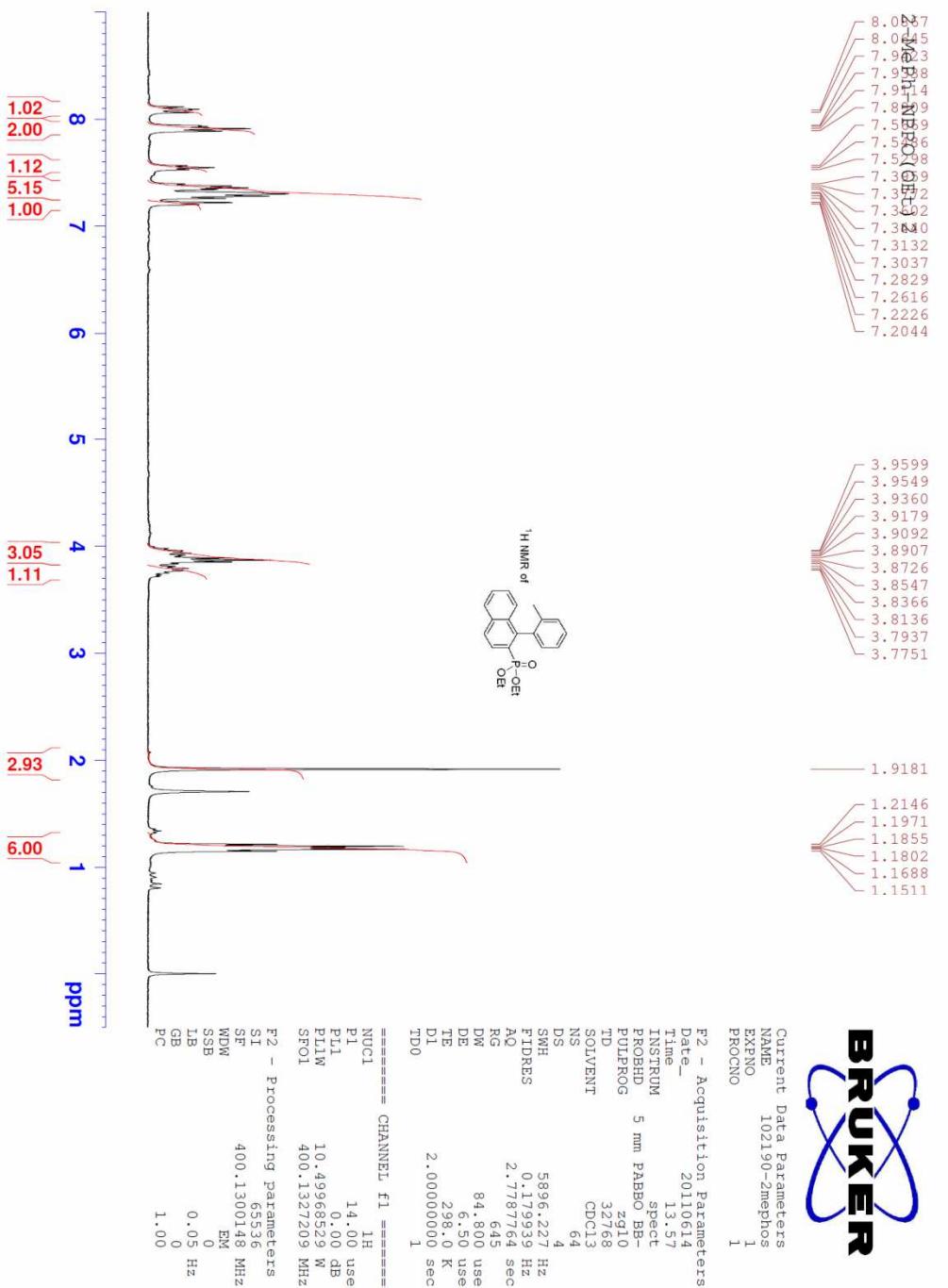


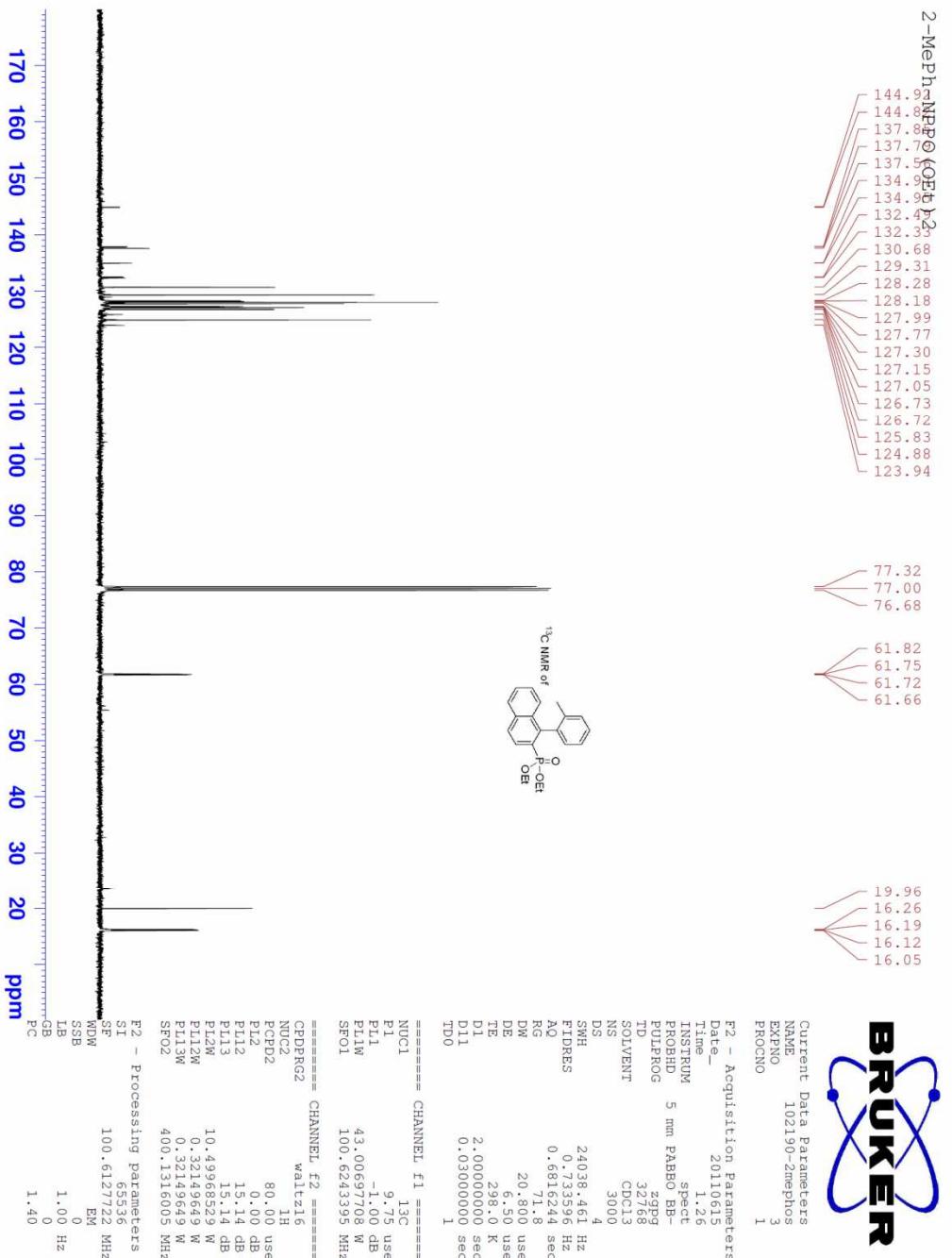


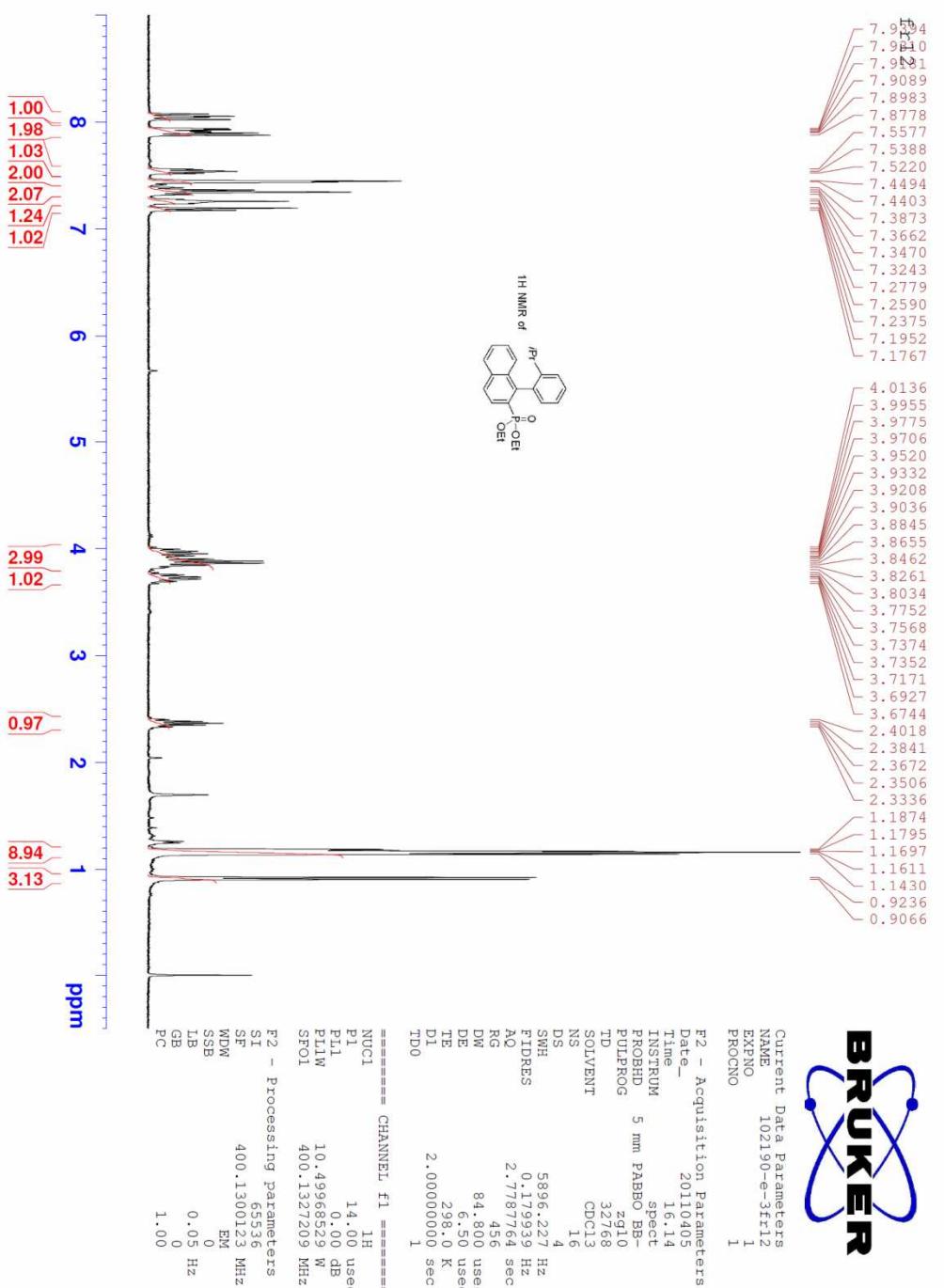












fr12

147.96
144.76
144.65
136.87
136.82
134.80
134.77
133.22
133.06
130.44
128.36
128.03
127.94
127.86
127.80
127.39
127.24
126.34
126.33
126.29
125.32
124.99
124.75
124.39

61.77
61.72
61.66

30.54
24.56
23.51
16.32
16.25
16.19

BRUKER

Current Data Parameters
NAME 102190-e3fr12
EXNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110405

Time_ 20:38

INSTRUM spect

PROBHD 5 mm PABBO BB-

PULPROG zspgr

TD 32768

SOLVENT CDCl3

NS 2048

DS

SWH 24038.461 Hz

FDRES 0.73396 Hz

AQ 0.61624 sec

RG 256

DW 20.800 usec

DE 6.50 usec

TE 298.0 K

D1 2.0000000 sec

D11 0.0300000 sec

TDO 1 sec

===== CHANNEL f1 =====

NUCL 13C

PL1 9.75 usec

PLL -1.00 dB

PLW 43.000597708 W

SFO1 100.6243395 MHz

===== CHANNEL f2 =====

NUCL 1H

PCPD2 80.00 usec

PCPD2 0.00 dB

PL2 15.14 dB

PL12 15.14 dB

PL13 10.49968529 W

PL2W 0.32149649 W

PL12W 400.1316005 MHz

SFO2

PL2 15.14 dB

PL12 15.14 dB

PL13 10.49968529 W

PL2W 0.32149649 W

PL12W 400.1316005 MHz

SFO2

PL2 15.14 dB

PL12 15.14 dB

PL13 10.49968529 W

PL2W 0.32149649 W

PL12W 400.1316005 MHz

SFO2

PL2 15.14 dB

PL12 15.14 dB

PL13 10.49968529 W

PL2W 0.32149649 W

PL12W 400.1316005 MHz

SFO2

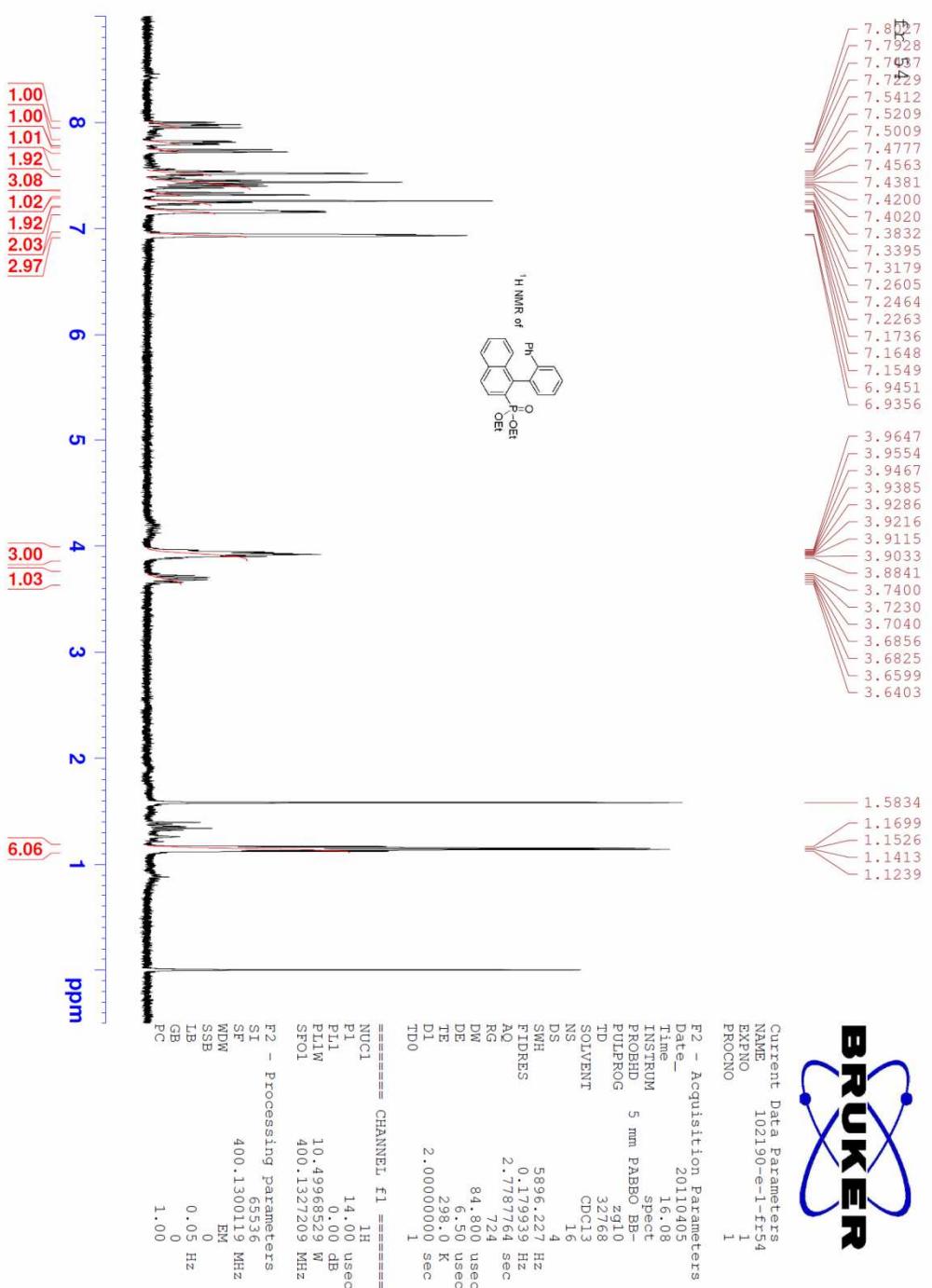
PL2 15.14 dB

PL12 15.14 dB

PL13 10.49968529 W

PL2W 0.32149649 W

PL12W 400.1316005 MHz



fr 54

142.01
141.25
136.89
134.57
131.68
129.66
129.10
128.33
127.88
127.78
127.73
127.69
127.51
127.40
127.33
127.26
126.50
126.33
126.29

61.87
61.84
61.82
61.78

16.30
16.24
16.19
16.13



Current Data Parameters
NAME 102100-e-1-fr54
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110406
Time_ 0.46
INSTRUM spect
PROBID 5 mm PABBO BB-
PULPROG 29PG
TD 3768
SOLVENT CICL3
NS 2048
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.681624 sec
RG 256
DW 20.800 usec
DE 6.50 usec
TE 238.0 K
D1 2.0000000 sec
D11 0.0300000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
PL 9.75 usec
P1L -1.00 dB
PL1W 43.00697708 W
SP01 100.6243395 MHz

===== CHANNEL f2 =====
NUC2 1H
PCPD2 80.00 usec
CPDPG2 Wallz16

PL2 0.00 dB
PL12 15.14 dB
PL13 15.14 dB
PL2W 10.4996529 W
PL12W 0.3214964 W
PL13W 0.32149649 W
SP02 400.1316005 MHz

F2 - Processing Parameters
SI 6536
SF 100.6127690 MHz
RMW EM
SSB 0
LB 1.00 Hz
GB 0
FC 1.40

