

DNA methylation in *Mytilus chilensis*

Cristian Gallardo¹, Valentina Venezuala¹, Steven Roberts²

¹University of Concepcion,

²University of Washington,

Corresponding author: Cristian Gallardo, cgallardo@udec.cl

Corresponding author: Valentina Venezuala, vvenezuala@udec.cl

Corresponding author: Steven Roberts, sr320@uw.edu

Abstract

We investigate genome-wide DNA methylation patterns in *Mytilus chilensis* from two contrasting seedbeds—Cochamó and Yaldad—with the aim of identifying differentially methylated regions (DMRs) that may underlie local adaptation. By integrating this epigenetic information with known patterns of gene expression and environmental context, we seek to illuminate the molecular basis of ecological divergence and contribute new insights into how environmental and anthropogenic forces shape genomic and epigenomic variation in a key aquaculture species.

Plain Language Summary

This study explores how DNA methylation may help Chilean blue mussels adapt to different environments, providing insights for aquaculture and conservation.

1 Introduction

Understanding how marine species respond to environmental heterogeneity is critical for conserving locally adapted populations and sustaining aquaculture practices in the face of global change. The Chilean blue mussel, *Mytilus chilensis*, a key endemic species in southern Chile, supports one of the world’s largest mussel aquaculture industries. However, the biological foundation of population-level adaptation in this species remains underexplored. Emerging evidence suggests that both genetic and epigenetic mechanisms—particularly DNA methylation—may contribute to phenotypic plasticity and local adaptation in marine invertebrates.

Previous studies have revealed extensive transcriptomic and mitochondrial differences between *M. chilensis* populations from ecologically distinct seedbeds, such as Cochamó (41°S) and Yaldad (43°S), in the inner sea of Chiloé Island. These differences include differential expression of genes involved in immunity, metabolism, and cellular stress responses, as well as tissue-specific shifts in mitochondrial gene expression linked to local environmental pressures, including aquaculture practices and variable oxygenation levels (Yévenes et al., 2021, 2022). Experimental evidence from reciprocal transplants shows population-specific transcriptional responses and reduced growth performance upon translocation, highlighting functional divergence between seedbeds despite high larval connectivity (Yévenes et al., 2025).

In addition to gene expression differences, recent work has uncovered epigenetic signatures that may contribute to adaptive divergence. Long non-coding RNAs (lncRNAs) were found to be differentially expressed between Cochamó and Yaldad populations, with associated changes in immune and metabolic gene networks (Yévenes et al., 2023). These findings underscore the potential for environmentally responsive regulatory elements—including DNA methylation—to mediate local adaptation.

Yet, the role of DNA methylation in shaping population-specific responses in *M. chilensis* remains unexplored. DNA methylation, a heritable and reversible epigenetic modification, can influence gene expression and phenotypic traits without altering the underlying DNA sequence. In bivalves, DNA methylation is known to affect developmental and immune processes and may be responsive to environmental stressors such as hypoxia, toxins, and acidification—conditions that vary across the Chilean inner sea and have been shown to modulate gene expression and microbiota composition in *M. chilensis* (Castillo et al., 2017; Montúfar-Romero et al., 2024, 2025).

In this study, we investigate genome-wide DNA methylation patterns in *Mytilus chilensis* from two contrasting seedbeds—Cochamó and Yaldad—with the aim of identifying differentially methylated regions (DMRs) that may underlie local adaptation. By integrating this epigenetic information with known patterns of gene

expression and environmental context, we seek to illuminate the molecular basis of ecological divergence and contribute new insights into how environmental and anthropogenic forces shape genomic and epigenomic variation in a key aquaculture species.

1.1 Methods

To investigate DNA methylation differences between populations of *Mytilus chilensis* from Cochamó and Yaldad, whole-genome bisulfite sequencing (WGBS) was performed and analyzed using a reproducible, high-throughput bioinformatics workflow. DNA was extracted from gill tissue of adult individuals and subjected to bisulfite treatment and library preparation. Libraries were sequenced on an Illumina NovaSeq 6000 platform, generating 150 bp paired-end reads, with each sample yielding approximately 120–160 million read pairs, providing $>30\times$ average coverage across the 1.93 Gb *M. chilensis* genome.

1.1.1 Read Processing and Alignment

Raw sequencing reads were processed using the nf-core/methylseq pipeline (version 2.5), implemented through Nextflow (version 23.04.1). Reads were aligned to the *M. chilensis* reference genome (*MchilensisGenomeV1.fa*), a chromosome-level assembly comprising 14 scaffolds (Gallardo-Escárate et al. 2023), using Bismark (version 0.24.1) with the Bowtie2 aligner (version 2.5.1). Quality trimming was applied to reverse reads by hard-clipping the final 10 bases (`-clip_r2 10`) to mitigate low-quality base calls at the 3' end. Methylation calls were extracted using the Bismark methylation extractor and aggregated by cytosine context. Pipeline execution was configured via a custom `.config` file to specify HPC resources and run with `-resume`, `-with-trace`, `-with-report`, and `-with-timeline` flags for reproducibility and tracking. Output files were stored in a structured directory and included per-base CpG methylation calls, quality control metrics, and coverage files.

1.1.2 Methylation Data Processing and Filtering

Downstream analysis was performed in R (version 4.3.1) using the methylKit package (version 1.10.0). Due to compatibility issues with newer versions of supporting libraries, `data.table` was pinned to version 1.14.10. Methylation coverage files were imported using `methRead()` with the `pipeline = "bismarkCoverage"` parameter and CpG context specified. Six samples—three each from Cochamó (LCo_BSR1–3) and Yaldad (LYa_BSR1–3)—were analyzed. Samples were assigned to treatment groups with Cochamó designated as the control (`treatment = 0`) and Yaldad as the treatment group (`treatment = 1`).

To ensure data quality, CpG sites were filtered to retain only those with a minimum read coverage of 10 in all samples, excluding those in the top 2% of coverage values to reduce PCR bias. Sites were then unified across samples using `unite()` with `min.per.group = 3` and `destrand = FALSE` to retain strand-specific methylation information.

1.1.3 Exploratory and Differential Methylation Analyses

Global methylation patterns were assessed using hierarchical clustering (Ward's method on correlation distance) and principal component analysis (PCA) to evaluate sample similarity. Population-level differences in DNA methylation were then tested using methylKit's logistic regression model, incorporating a multinomial overdispersion correction (`overdispersion = "MN"`) to account for biological variability. Statistical testing was performed using the Chi-squared test (`test = "Chisq"`), and q-values were adjusted for multiple testing using the SLIM method (`adjust = "SLIM"`).

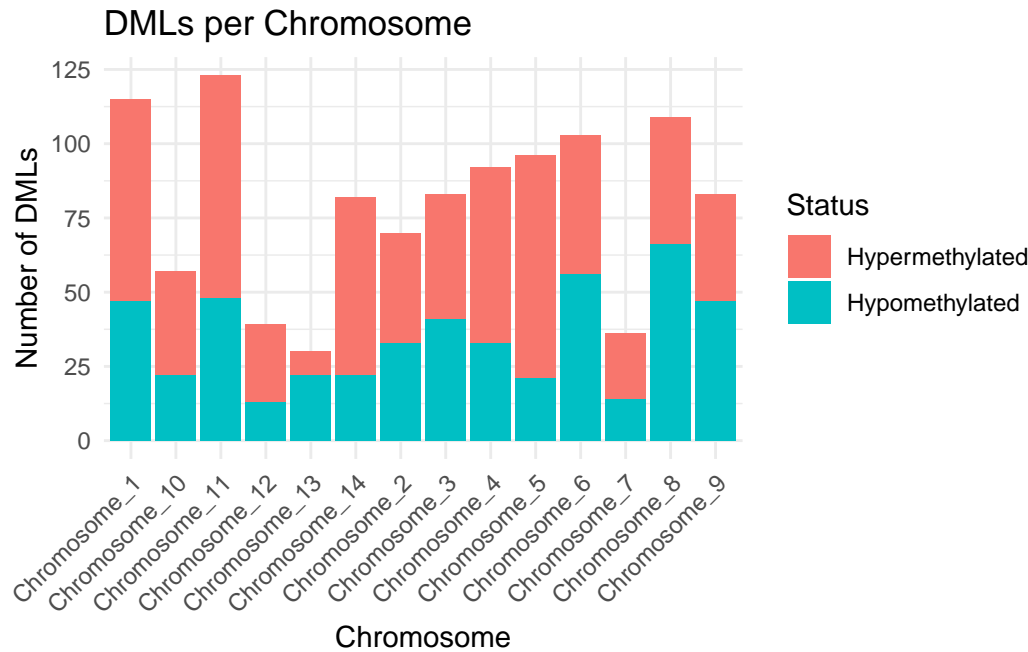
Differentially methylated loci (DMLs) were defined as CpG sites with an absolute methylation difference $\geq 25\%$ and FDR-adjusted q-value ≤ 0.05 . Separate sets of DMLs were extracted to identify hypomethylated (lower in Yaldad) and hyperme-

thylated (higher in Yaldad) loci. Results were saved as tab-delimited text files for downstream annotation and visualization.

1.2 Results

Bisulfite sequencing across six libraries yielded approximately 99.8 million reads per sample. Alignment to the reference genome was successful for 60.5–61.4% of reads, with 22–24% unaligned and 15–16% aligning ambiguously. After removal of duplicate reads (2.2–3.5 million per sample), 57.0–58.8 million uniquely aligned reads remained for methylation analysis.

Genome-wide cytosine methylation was assessed in CpG, CHG, and CHH contexts. Across samples, ~2.1–2.2 billion cytosines were examined. CpG methylation was the most prominent, with an average of ~36 million methylated CpGs and ~205 million unmethylated CpGs per sample, corresponding to a methylation rate of approximately 15%. In contrast, CHG and CHH methylation was much lower, with methylation levels below 1% in both contexts. These patterns are consistent with expectations for invertebrate methylomes, where methylation is primarily restricted to CpG sites.



Source: Article Notebook

1.2.1 Location

Source: Article Notebook

Castillo, N., Saavedra, L. M., Vargas, C. A., Gallardo-Escárate, C., & Détrée, C. (2017). Ocean acidification and pathogen exposure modulate the immune response of the edible mussel *Mytilus chilensis*. *Fish & Shellfish Immunology*, 70, 149–155. <https://doi.org/10.1016/j.fsi.2017.08.047>

Montúfar-Romero, M., Valenzuela-Muñoz, V., Valenzuela-Miranda, D., & Gallardo-Escárate, C. (2024). Hypoxia in the blue mussel *Mytilus chilensis* induces a transcriptome shift associated with endoplasmic reticulum stress, metabolism, and immune response. *Genes*, 15(6), 658. <https://doi.org/10.3390/genes15060658>

Montúfar-Romero, M., Valenzuela-Miranda, D., Valenzuela-Muñoz, V., Morales-Rivera, M. F., & Gallardo-Escárate, C. (2025). Microbiota dysbiosis in *Mytilus chilensis* is induced by hypoxia, leading to molecular and functional consequences. *Microorganisms*, 13(4), 825. <https://doi.org/10.3390/microorganisms13040825>

- 139 Yévenes, M., Núñez-Acuña, G., Gallardo-Escárate, C., & Gajardo, G. (2021). Adap-
 140 tive differences in gene expression in farm-impacted seedbeds of the native blue
 141 mussel *mytilus chilensis*. *Frontiers in Genetics*, 12, 666539. [https://doi.org/](https://doi.org/10.3389/fgene.2021.666539)
 142 [10.3389/fgene.2021.666539](https://doi.org/10.3389/fgene.2021.666539)
- 143 Yévenes, M., Núñez-Acuña, G., Gallardo-Escárate, C., & Gajardo, G. (2022). Adap-
 144 tive mitochondrial genome functioning in ecologically different farm-impacted
 145 natural seedbeds of the endemic blue mussel *mytilus chilensis*. *Comparative*
 146 *Biochemistry and Physiology Part D: Genomics and Proteomics*, 42, 100955.
 147 <https://doi.org/10.1016/j.cbd.2021.100955>
- 148 Yévenes, M., Gallardo-Escárate, C., & Gajardo, G. (2023). Epigenetic variation me-
 149 diated by lncRNAs accounts for adaptive genomic differentiation of the endemic
 150 blue mussel *mytilus chilensis*. *Heliyon*, 10(1), e23695. [https://doi.org/](https://doi.org/10.1016/j.heliyon.2023.e23695)
 151 [10.1016/j.heliyon.2023.e23695](https://doi.org/10.1016/j.heliyon.2023.e23695)
- 152 Yévenes, M., Gajardo, G., & Gallardo-Escárate, C. (2025). Decoding local adapta-
 153 tion in the exploited native marine mussel *mytilus chilensis*: Genomic evidence
 154 from a reciprocal transplant experiment. *International Journal of Molecular*
 155 *Sciences*, 26(3), 931. <https://doi.org/10.3390/ijms26030931>