

Dimensionality Reduction and Clustering: single cell RNA-seq Data

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Cancer scRNA data

- Lots of clinical interest in what cells express what genes, especially when it comes to cell pathologies (ex: cancer)
- Since 2006, The Cancer Genome Atlas (TCGA) project has sequenced over 20,000 samples acros 33 cancer types
- Their Pan-Cancer initiative aims to provide data products that let analysts compare gene expression across cancer types
- Our dataset consists of 5 different types of tumors: BRCA, KIRC, COAD, LUAD and PRAD (200 genes, 801 samples)

NATIONAL CANCER INSTITUTE THE CANCER GENOME ATLAS

TCGA BY THE NUMBERS



To put this into perspective, **1 petabyte** of data is equal to

212,000 DVDs

TCGA data describes

DIFFERENT
TUMOR TYPES

...including

RARE
CANCERS

...based on paired tumor and normal tissue sets collected from



...usino





REF: https://www-nature-com.proxy.lib.umich.edu/articles/ng.2764, https://www.cancer.gov/about-nci/organization/cog/research/structural-genomics/tcga, https://computerhistory.org/blog/decoding-cancer-with-the-cancer-genome-atlas-dr-jean-claude-zenkl usen/

Clustering for scRNA data

Challenges:

- Data can be very high dimensional (n ≈ p or n << p)
- Common clustering algorithms' performance can vary wildly depending on the initialization procedure

Our Solutions:

- Use a combination of dimensionality reduction and clustering
- Make dimensionality reduction work well in this high-dimensional setting
- Improve upon existing, widely-used clustering methods

Dimensionality Reduction: Sparse Principal Component Analysis (SPCA)

- SPCA promotes sparsity in the modes; i.e., the sparse modes only have a few active coefficients
- highly localized, more interpretable, avoids overfitting when p >> n
- minimizes the following objective function:

$$\begin{aligned} & \underset{\mathbf{A}, \mathbf{B}}{\text{minimize}} & f(\mathbf{A}, \mathbf{B}) = \frac{1}{2} \| \mathbf{X} - \mathbf{X} \mathbf{B} \mathbf{A}^\top \|_{\mathrm{F}}^2 + \psi(\mathbf{B}) \\ & \text{subject to} & \mathbf{A}^\top \mathbf{A} = \mathbf{I}, \end{aligned}$$

- existing SPCA packages:
 - sparsepca
 - elasticnet
 - EESPCA

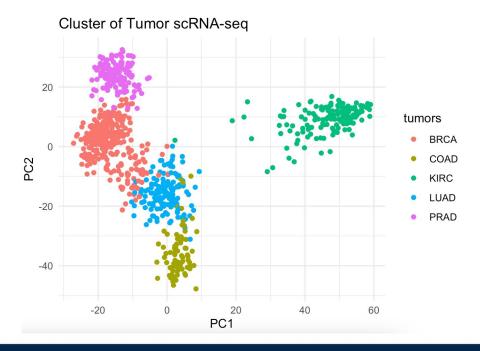
spcaRcpp

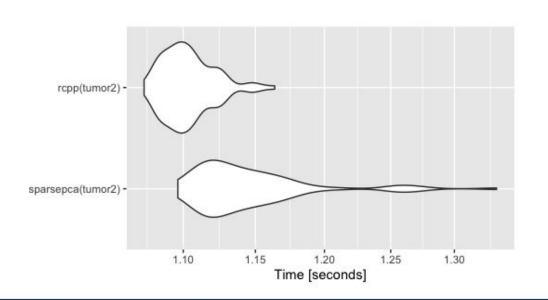
- a RcppArmadillo integration of sparsepca
- https://github.com/BoyaJiang/spcaRcpp

```
devtools::install_github("BoyaJiang/spcaRcpp")
library(spcaRcpp)
spca_out = spcaRcpp(tumor2, k = 21, center = TRUE, alpha = 1e-4, beta = 1e-4)
```

Usage		Argument	s	Value	
	<pre>pcaRcpp(X, k = NULL, alpha = 1e-04, beta = 1e-04, center = TRUE, max_iter = 1000, tol = 1e-05</pre>	Х	a numeric matrix or data.frame which provides the data for the sparse principal components analysis.	spcaRcpp returns a list containing the following six components:	
		k	optional, a number specifying the maximal rank.	loadings	the matrix of variable loadings.
		alpha	Sparsity controlling parameter. Higher values means sparser components.	standard deviations	the approximated standard deviations.
		beta	a Amount of ridge shrinkage to apply in order to improve conditioning.	eigenvalues	the approximated eigenvalues.
		center	a logical value indicating whether the variables should be shifted to be zero centered.	center	the centering used.
)				var	the variance.
		max_iter	maximum number of iterations to perform.	scores	the principal component scores.
		tol	stopping criteria for the convergence.		

spcaRcpp





Multivariate Normal Mixture EM Clustering

1. Key assumptions:

- a. Independent random variables(features)
- b. Normal distributions
- 2. Likelihood function

$$L(\theta) = p(x, z | \theta) = \prod_{c=1}^{k} \prod_{i=1}^{n} \left\{ \pi_{z_i} \prod_{j=1}^{p} P(x_{i,j} | p_{z_{i,j}}) \right\}^{I_{z_i = c}}$$

3. Log-likelihood function

$$l(\theta) = log L(\theta)$$

$$= \sum_{c=1}^{k} \sum_{i=1}^{n} I_{z_i=c} \left\{ log \pi_{z_i} + \sum_{j=1}^{p} log P(x_{i,j} | \mu_{z_i,j}, \sigma_{z_i,j}) \right\}$$

a. E-step... b. M-step...

normMixEm

- normMixEm(input_dat, num_components):
 (based on Class normMixEm from lecture14)
- normalmix_init(): Initialization function of parameters
 (based on mvnormalmix.init() from mixtools)
- rmvnorm_chol(): Random number generation function of MVNormal based on Cholesky decomposition (based on lecture 9)

Speed

normMixEm V.S. mixtools::mvnormalmixEM

```
set.seed(1000)
  x.1 < - rmvnorm(40, c(0, 0))
  x.2 < - rmvnorm(60, c(3, 4))
  X.1 < - rbind(x.1, x.2)
  class_t <- c(rep(1,40), rep(2,60)) # true labels
                                                                                mixtools Em test() -
  mu \leftarrow list(c(0, 0), c(3, 4))
  normMixEm_test <- function(data = X.1, num_components= 2L ){</pre>
    EM <- normMixEm$new(input_dat = data,num_components = num_components)</pre>
    out.2 <- EM$run.EM(loglik_tol=1e-5)
  mixtools_Em_test <- function(){</pre>
    out.1 <- mixtools::mvnormalmixEM(X.1, arbvar = FALSE, mu = mu,
                                             epsilon = 1e-02
                                                                                 normMixEm test()
  result <- microbenchmark(normMixEm_test(), mixtools_Em_test())</pre>
  autoplot(result)
> result
                                                                                                                           100
                                                                                                        10
                                                                                                                  30
                                                                                                                                     300
Unit: milliseconds
                                                                                                            Time [milliseconds]
                                                            median
                              min
                                           lq
                  expr
                                                   mean
                                                                                      max neval
   normMixEm_test() 5.169736 7.715231 10.18093 8.857127 10.52796 62.17496
                                                                                             100
 mixtools_Em_test() 26.886495 43.181047 62.65075 57.349866 68.84627 200.00337
                                                                                             100
```



normMixEm V.S. mixtools::mvnormalmixEM

Accuracy of clustering in the previous example

Accuracy

Clustering result of normMixEm

```
true est n freq
1 1 1 40 0.4
2 2 60 0.6
```

True data

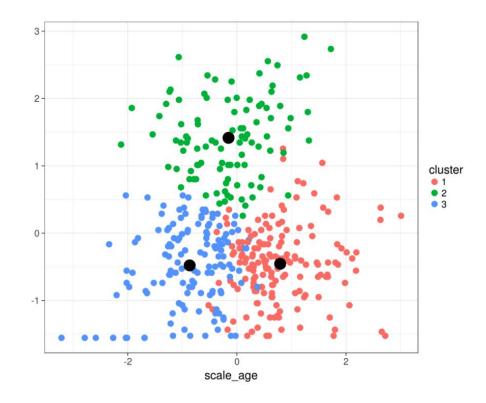
```
set.seed(1000)
x.1 <- rmvnorm(40, c(0, 0))
x.2 <- rmvnorm(60, c(3, 4))
X.1 <- rbind(x.1, x.2)
class_t <- c(rep(1,40),rep(2,60)) # true labels
mu <- list(c(0, 0), c(3, 4))</pre>
```

K-means Clustering

K-means is a very popular and relatively straightforward clustering algorithm. The most common implementation is referred to as **Lloyd's** algorithm.

Steps:

- 1. Randomly assign k datapoints to be the initial centroids (cluster centers)
- 2. Iterate until clusters do not change:
 - a. Assign rest of data to closest centroid (according to squared Euclidean distance)
 - b. Re-calculate centroids



K-means - initialization problem

Quality of clusters assessed by within-cluster sum of squares given by:

For clusters
$$C_1, ..., C_k$$
,

$$WCSS = \sum_{i=1}^{k} \sum_{x \in C_i} ||x - \mu_i||^2$$

where μ_i is the centroid for each cluster

Problem: cluster quality is affected by the random initialization method

How can we improve our initialization method, and thus our cluster quality?

K-means++

Another popular initialization method

Steps:

- 1. Randomly choose one centroid
- 2. Iterate until we have k centroids:
 - a. Compute the squared distance between non-centroids and all current centroids and take the minimum for each non-centroid
 - b. The next centroid is chosen with probability proportional to the distance computed in (a)
- 3. Proceed with Lloyd's k-means algorithm with these centroids

Greedy K-means++

Steps:

- 1. Randomly choose one centroid
- 2. Iterate until we have k centroids:
 - a. Compute the squared distance between non-centroids and all current centroids and take the minimum for each non-centroid
 - b. j new centroids are sampled with probability proportional to the distance computed in (a)
 - c. The next centroid is the one that, if chosen, would result in the lowest total WCSS
- 3. Proceed with Lloyd's k-means algorithm with these centroids

Our K-means implementation

Description

Clusters data according to k-means algorithm

Usage

```
kmeans_clust(X, k, nstart, iter.max, init.method)
```

Arguments

X n x p data matrix k number of clusters

nstart number of times to perform k means on the data

iter.max max iterations per run

init.method method for centroid initialization - choose from random, kmeans++, greedy

kmeans++ (gkmeans++)

Value

A list

- clusters n x p+1 matrix of cluster assignments where the first column is cluster assignments
- iter number of iterations
- centroids k x p+1 matrix of centroids where the first column is cluster assignments
- wcsse min within-cluster SSE over all nstart iterations

Examples

```
kmeans_clust(X, k = 3, nstart = 5, init.method = "kmeans++")
```

Many existing packages:

- stats::kmeans()
- flexclust::kcca()
- pracma::kmeanspp()
- clustR::KMeans_rcpp()



Our K-means - speed

```
Unit: milliseconds

expr min lq mean median uq
kmeans(brain, centers = 5, nstart = 1, algorithm = "Lloyd") 16.56256 17.70935 28.53108 23.02219 36.40693
kmeans_clust(brain, k = 5, nstart = 1, init.method = "random") 2361.56747 2511.42605 2868.15236 2579.80416 3221.93433
kmeans_clust(brain, k = 5, nstart = 1, init.method = "kmeans++") 2561.34022 3136.05081 3449.07094 3169.78173 3470.96459
kmeans_clust(brain, k = 5, nstart = 1, init.method = "gkmeans++") 5260.80046 5659.11857 6439.09512 6329.19468 7150.81366
max neval
49.27628 10
3930.11971 10
4898.88594 10
7898.80726 10
```

- Much slower than stats::kmeans()....
- stats::kmeans is written in C and Fortran, while kmeans_clust() is written in
 R
- As initialization method complexity increases, time also increases, with greedy kmeans++ having the longest runtime

Our K-means - performance

Algorithm	Adjusted Rand Index		
stats::kmeans(X, centers = 5, nstart = 1, algorithm = "Lloyd")	0.7699028		
kmeans_clust(X, k = 5, nstart = 1, init.method = "random")	0.7707366		
kmeans_clust(X, k = 5, nstart = 1, init.method = "kmeans++")	0.7619758		
kmeans_clust(X, k = 5, nstart = 1, init.method = "gkmeans++")	0.8405325		

Results

 EM clustering on principal components generated from spcaRcpp

```
s = Sys.time()
res_EM = normMixEm_test(data = spca_out$scores, num_components= 5L)
Sys.time() - s
class <- apply(res_EM$prob_mat, 1, which.max)
kable(data.frame(est=class, true=TC) %>% count(true,est) %>% mutate(freq=n/sum(n)))
```

- run-time: 0.237 secs
- adjustedRandIndex: 0.7328

true	est	n	freq
BRCA	2	1	0.0012484
BRCA	3	299	0.3732834
COAD	2	3	0.0037453
COAD	4	75	0.0936330
KIRC	2	1	0.0012484
KIRC	5	145	0.1810237
LUAD	2	139	0.1735331
LUAD	3	2	0.0024969
PRAD	1	134	0.1672909
PRAD	3	2	0.0024969

Results

 Kmeans clustering on principal components generated from spcaRcpp

```
s = Sys.time()
res_kmeans = kmeans_clust(spca_out$scores, k =5, nstart = 1, init.method = "gkmeans++")
Sys.time() - s
PC <- res_kmeans$clusters[,1]
kable(data.frame(est=PC, true=TC) %>% count(true,est) %>% mutate(freq=n/sum(n)))
```

- run-time: 1.058733 secs
- adjustedRandIndex: 0.6321

true	est	n	freq
BRCA	2	300	0.3745318
COAD	2	2	0.0024969
COAD	4	76	0.0948814
KIRC	2	1	0.0012484
KIRC	5	145	0.1810237
LUAD	1	136	0.1697878
LUAD	2	5	0.0062422
PRAD	3	136	0.1697878

Discussion

- After dimensionality reduction, EM algorithm outperforms k-means in terms of speed and accuracy
- Speed/accuracy trade-off of k-means (with and without SPCA):
 - o ARI: 0.8405 vs 0.6321
 - Speed: 6.4 secs vs 2 secs
- Limitations:
 - Both k-means and EM algorithm are sensitive to initialization point
 - Speed of clustering algorithms could be improved by writing in compiled language

References:

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- 2. N. B. Erichson, P. Zheng, S. Aravkin, sparsepca, (2018), GitHub repository
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