

Module 1: The Eukaryotic Cell Cycle (Lecture 1-8)

AIMS: a) To derive principles of human cell cycle control from studies in **model systems**. b) To explore the impact of cell cycle controls on **cell and chromosome integrity** and their relevance to **cancer**. c) To dissect **decision-making switches** across cell cycle transitions.

M1 Lecture 1: THE LANGUAGE OF GENETICS

Learning outcomes: understanding the evolving Genetics Viewpoint underpinning experimental design

- . Basic genetic and molecular concepts
- . Relevance of model organisms
- . Forward genetics as a "starting point"
- . Genetics in the whole genome sequencing era

The yeast cAMP pathway, *a case study*. Concept of essential gene, allele, conditional mutant, complementation, genetic interaction (synthetic lethality, redundancy, second site suppressor, dosage suppressor, epistasis). The *relevance* of models: yeast vs human *RAS* genes. Outlook with whole genomes and related tools.

M1 LECTURES 2-8: THE EUKARYOTIC CELL CYCLE.

Learning outcomes:

- . Competent interpretation of source experimental data and problem solving
- . Confidence in experimental design exploiting tools introduced in the lectures and recommended readings
- . With emphasis on evaluation of experimental evidence, develop a broad understanding of cell cycle controls and their significance
- . Gaining molecular insight into human cell cycle control

M1 LECTURE 2: THE EUKARYOTIC CELL CYCLE. PRINCIPLES AND MASTER REGULATORS

Biological importance of cell cycle control. Cell growth and cell division. Embryonic vs. somatic cell cycle. A *universal Master Regulator of the eukaryotic cell cycle*: CDK. Multiple CDK complexes: G1, S-phase and M-phase cyclins. Stage specific Cdk catalytic subunits in higher eukaryotes.

M1 LECTURE 3: THE EUKARYOTIC CELL CYCLE. THE G1/S TRANSITION

Feedback loops sharpen the G1/S transition in yeast. Transcriptional control of G1 cyclins. SBF/MBF and the repressor Whi5. Ubiquitin-dependent proteolysis of the CDK inhibitor (CKI) Sic1. Lessons from yeast: SCF control of cyclin E and CKIs in humans. Control of D-cyclins by *CRL4^{AMBRA1}*. E2F and Rb. Common circuitry themes.

M1 LECTURE 4: THE EUKARYOTIC CELL CYCLE. IN AND OUT OF MITOSIS AND THE PERFECT OSCILLATOR

More post-translational modifications switching CDK on & off. *S. pombe*: *wee1⁺* and *cdc25⁺*. Ubiquitin-dependent proteolysis in M phase. *APC/C targets: mitotic cyclins, mitotic kinases and securin*. Control of sister chromatid cohesion: The players - securin and separase. APC/C and the control of mitotic exit. *Downstream events responding to the oscillator*: cell cycle control of DNA replication. FUCCI.

M1 LECTURE 5: THE EUKARYOTIC CELL CYCLE. CHECKPOINTS

In the beginning - the DNA damage paradigm in yeast. Screening for checkpoint mutants. Brief overview of checkpoints monitoring replication, spindle integrity and orientation. Lessons from yeast: checkpoints in humans.

M1 LECTURE 6: THE EUKARYOTIC CELL CYCLE. TEMPORAL AND SPATIAL CONTROL OF MITOTIC EXIT

Cdc14 and the reversal of CDK phosphorylation. The Fourteen early anaphase release (FEAR) network, the Mitotic Exit Network (MEN) and the Spindle Position Checkpoint (SPOC). Principles for a spatial biosensor at the spindle pole. Conserved core components in higher eukaryotic cells.

M1 LECTURES 7: THE EUKARYOTIC CELL CYCLE. HOW DO CDKS IMPART ORDER —SPECIFICITY OR ACTIVITY THRESHOLDS?

Qualitative (specificity) vs. quantitative (threshold) models, evidence from yeast versus humans. Measuring CDK activity *in vitro* and apparent specificity dictated by cyclins. Studies using analog-sensitive CDK and biosensors. Tug of war between CDK and phosphatases. Time-resolved analysis for in vivo interactors of CDKs in human cells.

M1 LECTURES 8: SINGLE CELL ANALYSIS AND THE ELUSIVE RESTRICTION POINT (R)

Cdk2 biosensor, single cell analysis and R. Multiplex analysis and the structure of the human cell cycle. Revisiting R with START in mind. R biological relevance -- indications from genomic cancer projects.

GENERAL REFERENCES

Forsburg (2001) The art and design of genetic screens: yeast. *Nat Rev Genet* 2, 659-668.

* Morgan (2006) The cell cycle. Principles of control. Oxford University Press. (provided with online lecture material)

Tools for cell cycle analysis

Spiller et al. (2010) Measurement of single-cell dynamics. *Nature* 465, 736–745.

Eastman & Guo (2020) The palette of techniques for cell cycle analysis. *FEBS Letters* 594, 2084–2098.

Core Cell Cycle

**Hartwell et al. (1974) Genetic control of the cell division cycle in yeast. *Science* 183, 46-51

Biggins et al. (2020) Fifty years of cycling. *Mol Biol Cell* 31, 2868-2870.

Nurse (2020) Fission yeast cell cycle mutants and the logic of eukaryotic cell cycle control. *Mol Biol Cell* 31, 2871-2873.

Kirschner (2020) What makes the cell cycle tick? a celebration of the awesome power of biochemistry and the frog egg. *Mol Biol Cell* 31, 2874-2878.

Irniger et al. (1995) Genes involved in sister chromatid separation are needed for B-type cyclin proteolysis in budding yeast. *Cell* 81, 269-278.

Strohmaier et al. (2001) Human F-box protein hCdc4 targets cyclin E for proteolysis and is mutated in a breast cancer cell line. *Nature* 413, 316-322.

**Simoneschi et al. (2021) CRL4^{AMBRA1} is a master regulator of D-type cyclins. *Nature* 592, 789-793.

*Maiani et al. (2021) AMBRA1 regulates cyclin D to guard S-phase entry and genomic integrity. *Nature* 592:799-803.

*Chaikovsky et al. (2021) The AMBRA1 E3 ligase adaptor regulates the stability of cyclin D. *Nature* 592, 794-798.

**Weinert and Hartwell (1988) The *RAD9* gene controls the cell cycle response to DNA damage in *Saccharomyces cerevisiae*. *Science* 241, 317- 322.

Ciccio and Elledge (2010) The DNA damage response: making it safe to play with knives. *Mol Cell* 40, 179-204.

Wieser and Pines (2015) The biochemistry of mitosis. *Cold Spring Harb Perspect Biol.* 7, a015776-17.

Pinsky and Biggins (2005) The spindle checkpoint: tension versus attachment. *Trends Cell Biol* 15, 486-493.

Makrantonis and Marston (2018) Primer - Cohesin and chromosome segregation. *Curr Biol* 28, R679–R694.

Musacchio (2015) The molecular biology of spindle assembly checkpoint signaling dynamics. *Curr Biol* 25, R1002-R1018.

Holder et al.(2019) Getting out of mitosis: spatial and temporal control of mitotic exit and cytokinesis by PP1 and PP2A. *FEBS Lett* 593, 2908-2924.

Lara-Gonzalez, et al. (2021) Spindle assembly checkpoint activation and silencing at kinetochores. *Semin Cell Dev Biol* 117, 86–98.

FEAR, MEN & SPOC

Caydasi and Pereira (2012) SPOC alert--when chromosomes get the wrong direction. *Exp Cell Res* 318, 1421-1427.

Rock and Amon (2009) Primer - The FEAR network. *Curr Biol* 19, R1063-1068.

Scarfone et al. (2015) Coupling spindle position with mitotic exit in budding yeast: The multifaceted role of the small GTPase Tem1. *Small GTPases* 6, 196-201.

**Zhou et al. (2021) Cross-compartment signal propagation in the mitotic exit network. *Elife* 10, e63645.

**Campbell et al. (2019) The Mitotic Exit Network integrates temporal and spatial signals by distributing regulation across multiple components. *Elife* 8, e41139.

Order in the cell cycle, switches, feedback, thresholds and more...

Bloom and Cross (2007) Multiple levels of cyclin specificity in cell cycle control. *Nat Rev Mol Cell Biol* 8, 149-160.

Loog & Morgan (2005) Cyclin specificity in the phosphorylation of cyclin-dependent kinase substrates. *Nature* 434, 104-108.

* Ferrel (2013) Feedback loops and reciprocal regulation: recurring motifs in the systems biology of the cell cycle. *Curr Op Cell Biol* 25, 676–686

**Skotheim et al. (2008) Positive feedback of G₁ cyclins ensures coherent cell cycle entry. *Nature* 454, 291-297.

****Yang et al. (2013)** Design Principles of the yeast G1/S switch. *PLoS Biol* 11, e1001673.

Fisher et al. (2012) Phosphorylation network dynamics in the control of cell cycle transitions. *J Cell Sci* 125, 4703-4711.

Coudreuse and Nurse (2010) Driving the cell cycle with a minimal CDK control network. *Nature* 468, 1074-1079.

Kamenz and Ferrell (2017) The temporal ordering of cell-cycle phosphorylation. *Mol. Cell* 65, 371-373.

*Swaffer et al. (2016) CDK Substrate phosphorylation and ordering the cell cycle. *Cell* 167, 1750-1761.

****Basu et al. (2022)** Core control principles of the eukaryotic cell cycle. *Nature* 607, 381-386.

Kõivomägi et al. (2011) Cascades of multisite phosphorylation control Sic1 destruction at the onset of S phase. *Nature* 480, 128-131.

Kõivomägi et al. (2021) G₁ cyclin-Cdk promotes cell cycle entry through localized phosphorylation of RNA polymerase II. *Science* 374, 347-351.

Gelens et al. (2018) The importance of kinase-phosphatase integration: lessons from mitosis. *Trends Cell Biol* 28, 6-21.

Pagliuca et al. Quantitative proteomics reveals the basis for the biochemical specificity of the cell-cycle machinery (2011) *Mol Cell* 43, 406-417.

****Merrick et al. (2011)** Switching Cdk2 on or off with small molecules to reveal requirements in human cell proliferation. *Mol Cell* 42, 624-636.

Faustova et al (2022) A synthetic biology approach reveals diverse and dynamic CDK response profiles via multisite phosphorylation of NLS-NES modules. *Sci Adv* 8, eabp8992.

To cycle or not to cycle?

Hanahan and Weinberg (2000) The hallmarks of cancer. *Cell* 100, 57-70.

Hanahan and Weinberg (2011) Hallmarks of cancer: the next generation. *Cell* 144, 646-674.

Malumbres and Barbacid (2001) To cycle or not to cycle: a critical decision in cancer. *Nat Rev Cancer* 1, 222-231.

*Spencer et al. (2013) The proliferation-quiescence decision is controlled by a bifurcation in CDK2 activity at mitotic exit. *Cell* 155, 369 – 83.

Yang et al. (2017) Competing memories of mitogen and p53 signalling control cell-cycle entry. *Nature* 549, 404-408.

****Arora et al., (2017)** Endogenous Replication Stress in Mother Cells Leads to Quiescence of Daughter Cells. *Cell Rep* 19, 1351–1364.

****Schwarz et al. (2018)** A Precise Cdk activity threshold determines passage through the restriction point. *Mol Cell* 69, 253–264.

Orr et al. (2015) Primer - Aneuploidy. *Curr Biol* 25, R538-R542.

Sanchez-Vega et al. (2018) Oncogenic signaling pathways in The Cancer Genome Atlas. *Cell* 173, 321-337.

Rubin et al (2020) Integrating old and new paradigms of G1/S control. *Mol Cell* 80,183-192.

*Liu et al. (2020) Altered G1 signaling order and commitment point in cells proliferating without CDK4/6 activity. *Nat Commun* 11, 5305.

Stallaert et al. (2022) The structure of the human cell cycle. *Cell Syst* 13, 230-240.e3.