

# **FALL 2020: SEMINAR IN GENE REGULATION**

## **COURSE NUMBERS**

Hunter Undergraduate: BIOL 471.93  
Hunter Graduate (M.A.): BIOL 790.52  
CUNY Graduate Center (Ph.D.): BIOL 793.03

**PROFESSOR:** Armin Lahiji

**OFFICE HOURS:** Wed 8:25-9:25 PM

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## **The student learning goals for this seminar course are:**

1. To enhance participant familiarity with current literature on mechanisms of gene regulation
2. To enable students to recognize and formulate testable hypotheses based on existing data
3. To provide opportunities for students to develop skills in oral and written communication of research

**Seminar course overview:** In this course, we will be studying recent review and primary articles on various topics in gene regulation. In the first part of the course we will be going over articles as a group. During this period, students in the course will be independently researching a chosen topic for the purpose of writing a paper on that topic. The type of paper to be written for undergraduate and graduate students are as follows:

**Undergraduates:** A short synthetic review article with the following components:

Brief description of the chosen topic/process and its importance to development and/or disease

The state of knowledge of that field (what is known)

The most important unanswered questions (what is NOT known)

New hypothesis arising from your review that may address one of those unanswered questions

Explanation and synthesis of the key points of prior data that support the new hypothesis

**Graduate students:** A short research proposal to investigate a particular question relevant to mechanisms of gene regulation roughly following the NIH format:

A "Specific aims" page (introducing 1-2 related aims testing a hypothesis arising from your research)

Background, Significance, Experimental Plan, Expected outcomes and interpretations

During the second part of the course, students will make presentations of their plans for their paper/proposal. While students may pool efforts in research and sharing information, the paper must be an individual work.

## **An outline and drafts will be collected at various due dates during the semester as follows:**

**9/22 Paper prospectus due!!!** (approved topic, outline and list of relevant references to be used)

**11/3** First draft due, as complete as possible

**11/24** Second complete draft due

**Final draft due (as e-mail attachment) by 9:00AM on 12/15**

## **COURSE GRADES WILL BE CALCULATED AS FOLLOWS:**

**FINAL PAPER--50%, CLASS PARTICIPATION--25%, PRESENTATION--25%**

**Class participation** means reading and understanding the articles in advance of a given session, contributing regularly and meaningfully to the in-class discussions of them. Included in this component of the course is the submission of short written answers to a short list of general questions about the primary papers we will discuss in sessions 2 through 8. The answers relevant to the primary paper discussed in each session will be due and collected at the beginning of the session. This exercise is designed to help students recognize and understand the hypotheses that were the basis for beginning the work described in the paper.

## **SCHEDULE OF SESSIONS AND ARTICLE LIST (please note: no class on 10/1 and 10/8)**

### **9/1 Session 1, Introduction and Organization**

**Review:** Margueron, et. al. The key to development: interpreting the histone code? *Curr Opin Genet. Dev.* 2005, 15:163-176.

### **9/8 Session 2, Recognition of histone modifications**

**Review:** Chi, et. al. Covalent histone modifications – miswritten, misinterpreted and mis-erased in human cancers. *Nature Rev. Cancer.* 2010, 10:457-469.

Primary: Wysocka, et. al. A PHD finger of NURF couples histone H3 lysine 4 trimethylation with chromatin remodeling. *Nature*. 2006, 442:86-90.

### **9/15 Session 3, Transcription and the “histone code”**

Review: Berger. The complex language of chromatin regulation during transcription. *Nature*. 2007, 447:407-12.

Preview: Lorincz and Schubeler. RNA polymerase II: Just dropping by. *Cell*. 2007, 130:16-18.

Primary: Guenther, et. al. A chromatin landmark and transcription initiation at most promoters in human cells. *Cell*. 2007, 130:77-88.

### **9/22 Session 4, Transcription “pause” and “release”**

Review: Chiba, et. al. Promoter-proximal pausing and its release: Molecular mechanisms and physiological functions. *Exp. Cell Res.* 2010, 316:2723-30.

Preview: Price. Regulation of RNA polymerase II elongation by c-myc. *Cell*. 2010, 141:399-340.

Primary: Rahl, et. al. c-Myc regulates transcriptional pause release. *Cell*. 2010, 141:432-445.

### **10/06 Session 5, DNA methylation: PAPER PROSPECTUS DUE!**

Review: Bogdanovic and Veenstra. DNA methylation and methyl-CpG binding proteins: developmental requirements and function. *Chromosoma* 2009, 118:549-565.

Primary 1: Nan, et. al. Transcriptional repression by the methyl-CpG-binding protein MeCP2 involves a histone deacetylase complex. *Nature*. 1998, 393:386-9.

Primary 2: Fuks F, et. al. The methyl-CpG-binding protein MeCP2 links DNA methylation to histone methylation. *J. Biol Chem.* 2003, 278:4035-40.

### **10/13 Session 6, Small RNAs and gene regulation**

Review: Farazi, et. al. The growing catalog of small RNAs and their association with distinct Argonaute/Piwi family proteins. *Development*. 2008, 135:1201-1214.

Primary: Giles, et. al. Maintenance of a constitutive heterochromatin domain in vertebrates by a dicer-dependent mechanism. *Nature Cell Biol.* 2010, 12:94-99.

### **10/20 Session 7, Boundary/Insulator elements**

Review: Barkess and West. Chromatin insulator elements: establishing barriers to set heterochromatin domain boundaries. *Epigenomics*. 2012, 4:67-80.

Primary: Dickson, et. al. VEZF1 elements mediate protection of DNA methylation. *PLoS Genetics*. 2010, 6(1):e1000804.

### **10/27 Session 8, Chromatin Looping and Gene Regulation**

Review: Deng and Blobel. Do chromatin loops provide epigenetic gene expression states? *Curr. Opin. Genet. Dev.* 2010, 20:548-554.

Primary: Deng, et. al. Controlling long-range genomic interactions at a native locus by targeted tethering of a looping factor. *Cell*. 2012, 149:1233-1244.

### **11/3 Session 9, Student presentations: FIRST DRAFT OF PAPER DUE!**

### **11/10 Session 10, Student presentations**

### **11/17 Session 11, Student presentations**

### **11/24 Session 12, Student presentations: SECOND DRAFT OF PAPER DUE!**

**12/1      Session 13, Student presentations**

**12/8      Session 14, Leftovers and discussion: FINAL PAPER DUE on 12/15!**

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