

## Programming for Biology Protein Evolution / Similarity Searching

### What BLAST Does / Why BLAST works

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## Protein Evolution/ Similarity Searching

- 9:00 – Homology and Expectation value
- 10:30 – Similarity searching workshop I
- 1:30 – Practical Similarity Searching, improving sensitivity
- 3:00 – Workshop II – Parsing search results

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## Effective Similarity Searching

1. Always search protein databases (possibly with translated DNA)
  2. Use E()-values, not percent identity, to infer homology
    - $E() < 0.001$  is significant in a single search
- 
3. Search smaller (comprehensive) databases
  4. Change the scoring matrix for:
    - short sequences (exons, reads)
    - short evolutionary distances (mammals, vertebrates, a-proteobacteria)
    - high identity (>50% alignments) to reduce over-extension
  5. All methods (pairwise, HMM, PSSM) miss homologs, and find homologs the other methods miss

## Sequence Similarity - Conclusions

- Homologous sequences share a common ancestor, but most sequences are non-homologous
- Always compare Protein Sequences
- Sequence Homology can be reliably inferred from statistically significant similarity (non-homology cannot from non-similarity)
- Homologous proteins share common structures, but not necessarily common functions
- Sequence statistical significance estimates are accurate (verify this yourself)  $10^{-6} < E() < 10^{-3}$  is statistically significant
- Scoring matrices set evolutionary look back horizons - not every discovery is distant
- PSI-BLAST can be more sensitive, but with lower statistical accuracy

*Establishing homology from  
statistically significant similarity*

**Why BLAST works**

- For most proteins, homologs are easily found over long evolutionary distances (500 My – 2 By) using standard approaches (BLAST, FASTA)
- Difficult for distant relationships or very short domains
- Most default search parameters are optimized for distant relationships and work well

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## Protein Evolution and Sequence Similarity

### **Similarity Searching I**

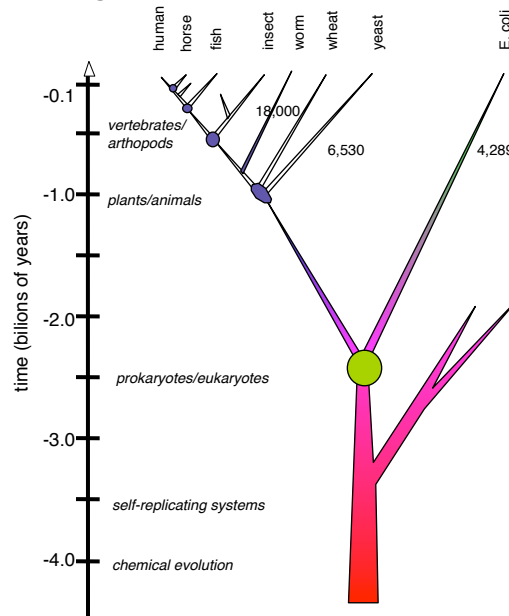
- **What is Homology and how do we recognize it?**
- How do we measure sequence similarity – alignments and scoring matrices?
- DNA vs protein comparison

### **Similarity Searching II**

- More effective similarity searching
  - Smaller databases
  - Appropriate scoring matrices
  - Using annotation/domain information

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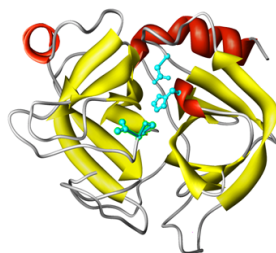
## Homologues share a common ancestor



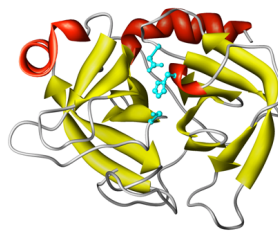
7

## When do we infer homology?

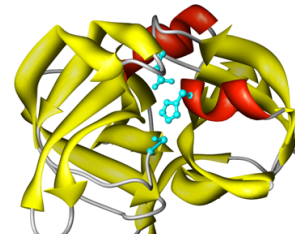
Homology  $\Leftrightarrow$  structural similarity  
? sequence similarity



Bovine trypsin (5ptp)  
Structure:  $E() < 10^{-23}$ ,  
RMSD 0.0 Å  
Sequence:  $E() < 10^{-84}$   
100% 223/223



S. griseus trypsin (1sgt)  
 $E() < 10^{-14}$  RMSD 1.6 Å  
 $E() < 10^{-19}$  36%; 226/223

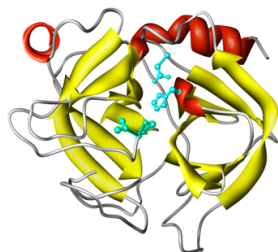


S. griseus protease A (2sga)  
 $E() < 10^{-4}$ ; RMSD 2.6 Å  
 $E() < 2.6$  25%; 199/181

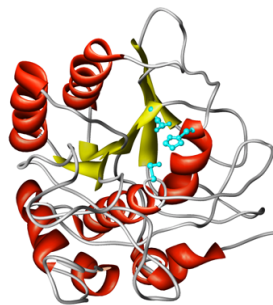
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## When can we infer non-homology?

Non-homologous proteins have different structures



Bovine trypsin (5ptp)  
 Structure:  $E() < 10^{-23}$   
 RMSD 0.0 Å  
 Sequence:  $E() < 10^{-84}$   
 100% 223/223



Subtilisin (1sbt)  
 $E() > 100$   
 $E() < 280$ ; 25% 159/275



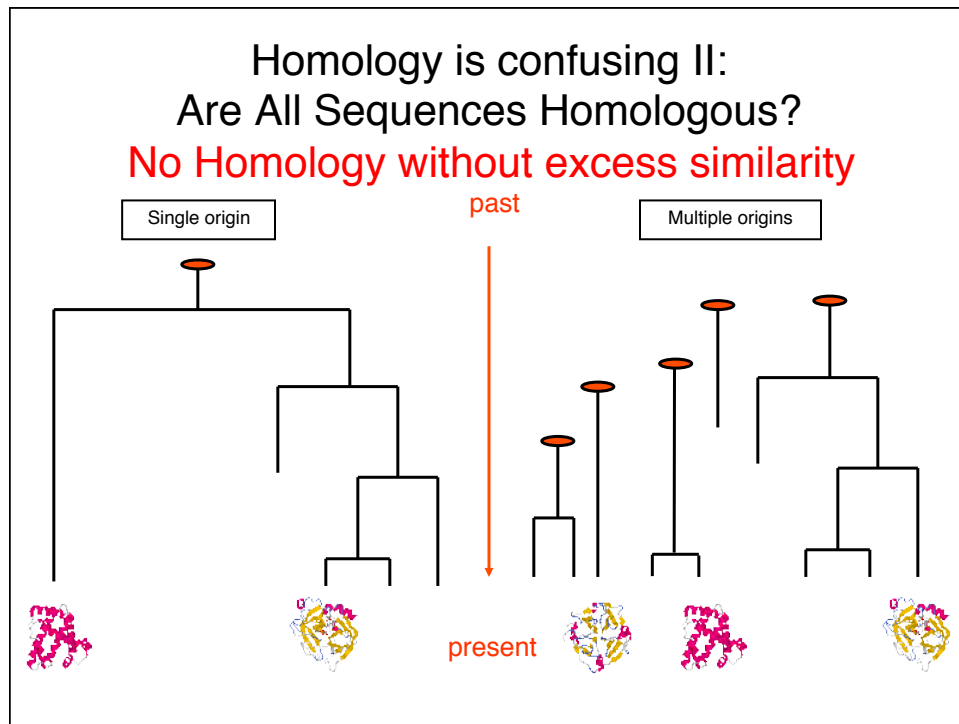
Cytochrome c4 (1etp)  
 $E() > 100$   
 $E() < 5.5$ ; 23% 171/190

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## Homology is confusing I: Homology defined Three(?) Ways

- Proteins/genes/DNA that share a common ancestor
- Specific positions/columns in a multiple sequence alignment that have a 1:1 relationship over evolutionary history
  - sequences are *50% homologous* ???
- Specific (morphological/functional) characters that share a recent divergence (clade)
  - bird/bat/butterfly wings are/are not homologous

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### Homology from sequence similarity

- Sequences are inferred to share a common ancestor based on statistically significant **excess** similarity. Any evidence of **excess** similarity can be used to infer homology
- Lack of sequence evidence **cannot** be used to infer non-homology.
  - Proteins with different structures are non-homologous
- There are always two alternative hypotheses: homology (common ancestry), or independence – one must weigh the evidence for each hypothesis (independence is the *null* hypothesis).

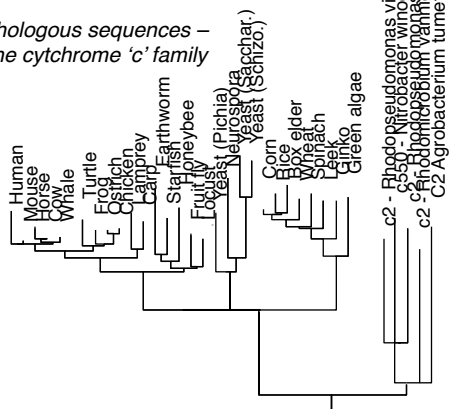
## E. coli proteins vs Human – Ancient Protein Domains

expect	% id	alen	E coli descr	Human descr	sp_name
2.7e-206	53.8	944	glycine decarboxylase, P	Glycine dehydrogenase [de	GCSP_HUMAN
1.2e-176	59.5	706	methylmalonyl-CoA mutase	Methylmalonyl-CoA mutase,	MUTA_HUMAN
3.8e-176	50.6	803	glycogen phosphorylase [E	Glycogen phosphorylase, l	PHS1_HUMAN
9.9e-173	55.6	1222	B12-dependent homocystein	5-methyltetrahydrofolate-	METH_HUMAN
1.8e-165	41.8	1031	carbamoyl-phosphate synth	Carbamoyl-phosphate synth	CPSM_HUMAN
5.6e-159	65.7	542	glucosephosphate isomeras	Glucose-6-phosphate isome	G6PI_HUMAN
8.1e-143	53.7	855	aconitate hydratase 1 [Esch	Iron-responsive element b	IRE1_HUMAN
2.5e-134	73.0	459	membrane-bound ATP syntha	ATP synthase beta chain,	ATPB_HUMAN
3.3e-121	55.8	550	succinate dehydrogenase,	Succinate dehydrogenase [	DHSA_HUMAN
1.5e-113	60.6	401	putative aminotransferase	Cysteine desulfurase, mit	NFS1_HUMAN
4.4e-111	60.9	460	fumarase C= fumarate hydr	Fumarate hydratase, mitoc	FUMH_HUMAN
1.5e-109	56.1	474	succinate-semialdehyde de	Succinate semialdehyde de	SSDH_HUMAN
3.6e-106	44.7	789	maltodextrin phosphorylas	Glycogen phosphorylase, m	PHS2_HUMAN
1.4e-102	53.1	484	NAD+-dependent betaine al	Aldehyde dehydrogenase, E	DHAG_HUMAN
3.8e-98	53.0	449	pyridine nucleotide trans	NAD(P) transhydrogenase,	NNTM_HUMAN
5.8e-96	49.9	489	glycerol kinase [Escheric	Glycerol kinase, testis s	GKP2_HUMAN
2.1e-95	66.8	328	glyceraldehyde-3-phosphat	Glyceraldehyde 3-phosphat	G3P2_HUMAN
5.0e-91	62.5	368	alcohol dehydrogenase cla	Alcohol dehydrogenase cla	ADHX_HUMAN
6.7e-91	56.5	393	protein chain elongation	Elongation factor Tu, mit	EFTU_HUMAN
9.5e-91	56.6	392	protein chain elongation	Elongation factor Tu, mit	EFTU_HUMAN
2.2e-89	59.1	369	methionine adenosyltransf	S-adenosylmethionine synt	METK_HUMAN
6.5e-88	53.3	422	enolase [Escherichia coli	Alpha enolase (2-phospho-	ENOA_HUMAN
9.2e-88	43.3	536	NAD-linked malate dehydro	NADP-dependent malic enzy	MAOX_HUMAN
7.3e-86	55.5	389	2-amino-3-ketobutyrate Co	2-amino-3-ketobutyrate co	KBL_HUMAN
5.2e-83	44.4	543	degrades sigma32, integra	AFG3-like protein 2 (Para	AF32_HUMAN

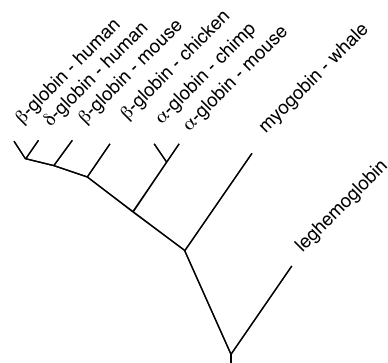
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## Orthologs and Paralogs – Inferring Function

Orthologous sequences –  
the cytochrome 'c' family



Paralogous genes – globins

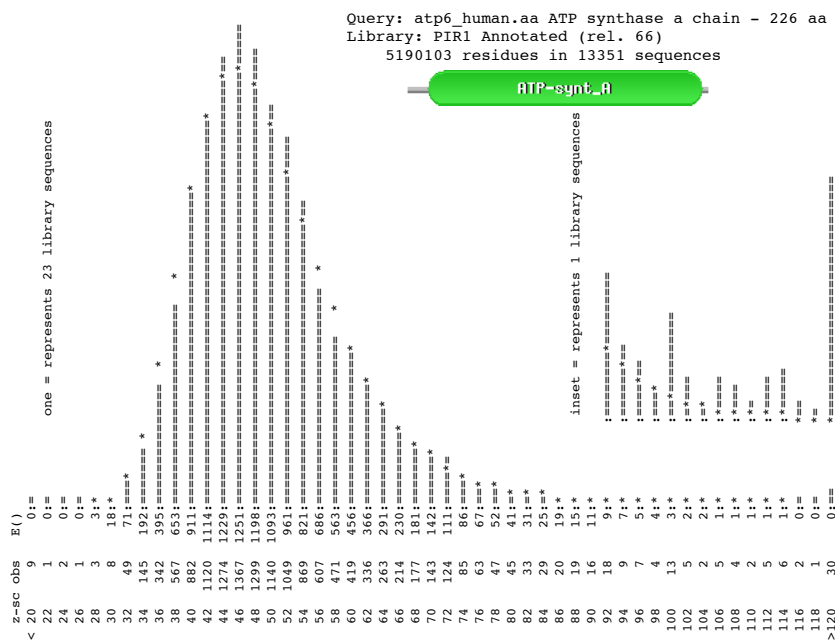


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## Protein Evolution and Sequence Similarity

- What is Homology and how do we recognize it?
- How do we measure sequence similarity – alignments and scoring matrices?
- DNA vs protein comparison
- More effective similarity searching
  - Smaller databases
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## Inferring Homology from Statistical Significance

- Real **UNRELATED** sequences have similarity scores that are indistinguishable from **RANDOM** sequences
- If a similarity is NOT **RANDOM**, then it must be NOT **UNRELATED**
- Therefore, NOT **RANDOM** (statistically significant) similarity must reflect **RELATED** sequences

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Query: atp6\_human.aa ATP synthase a chain - 226 aa  
Library: 5190103 residues in 13351 sequences

The best scores are:		( len)	s-w	bits	E(13351)	%_id	%_sim	alen
sp P00846	ATP6_HUMAN ATP synthase a chain (AT	( 226)	1400	325.8	5.8e-90	1.000	1.000	226
sp P00847	ATP6_BOVIN ATP synthase a chain (AT	( 226)	1157	270.5	2.5e-73	0.779	0.951	226
sp P00848	ATP6_MOUSE ATP synthase a chain (AT	( 226)	1118	261.7	1.2e-70	0.757	0.916	226
sp P00849	ATP6_XENLA ATP synthase a chain (AT	( 226)	745	176.8	4.0e-45	0.533	0.847	229
sp P00851	ATP6_DROYA ATP synthase a chain (AT	( 224)	473	115.0	1.7e-26	0.378	0.721	222
sp P00854	ATP6_YEAST ATP synthase a chain pre	( 259)	428	104.7	2.3e-23	0.353	0.694	232
sp P00852	ATP6_EMENI ATP synthase a chain pre	( 256)	365	90.4	4.8e-19	0.304	0.691	230
sp P14862	ATP6_COCHE ATP synthase a chain (AT	( 257)	353	87.7	3.2e-18	0.313	0.650	214
sp P68526	ATP6_TRITI ATP synthase a chain (AT	( 386)	309	77.6	5.1e-15	0.289	0.651	235
sp P05499	ATP6_TOBAC ATP synthase a chain (AT	( 395)	309	77.6	5.2e-15	0.283	0.635	233
sp P07925	ATP6_MAIZE ATP synthase a chain (AT	( 291)	283	71.7	2.3e-13	0.311	0.667	180
sp P0AB98	ATP6_ECOLI ATP synthase a chain (AT	( 271)	178	47.9	3.2e-06	0.233	0.585	236
sp P0C2Y5	ATPI_ORYSA Chloroplast ATP synth (A	( 247)	144	40.1	0.00062	0.242	0.580	231
sp P06452	ATPI_PEA Chloroplast ATP synthase a	( 247)	143	39.9	0.00072	0.250	0.586	232
sp P27178	ATP6_SYNY3 ATP synthase a chain (AT	( 276)	142	39.7	0.00095	0.265	0.571	170
sp P06451	ATPI_SPIOL Chloroplast ATP synthase	( 247)	138	38.8	0.0016	0.242	0.580	231
sp P08444	ATP6_SYNPF ATP synthase a chain (AT	( 261)	127	36.3	0.0095	0.263	0.557	167
sp P69371	ATPI_ATRBE Chloroplast ATP synthase	( 247)	126	36.0	0.01	0.221	0.571	231
sp P06289	ATPI_MARPO Chloroplast ATP synthase	( 248)	126	36.0	0.011	0.240	0.575	167
sp P30391	ATPI_EUGGR Chloroplast ATP synthase	( 251)	123	35.4	0.017	0.257	0.579	214
sp P19568	TLCA_RICPR ADP,ATP carrier protein	( 498)	122	35.0	0.043	0.243	0.579	152
sp P24966	CYB_TAYTA Cytochrome b	( 379)	113	33.0	0.13	0.234	0.532	158
sp P03892	NU2M_BOVIN NADH-ubiquinone oxidored	( 347)	107	31.7	0.31	0.261	0.479	211
sp P68092	CYB_STEAT Cytochrome b	( 379)	104	31.0	0.54	0.277	0.547	137
sp P03891	NU2M_HUMAN NADH-ubiquinone oxidored	( 347)	103	30.8	0.58	0.201	0.537	149
sp P00156	CYB_HUMAN Cytochrome b	( 380)	102	30.5	0.74	0.268	0.585	205
sp P15993	AROP_ECOLI Aromatic amino acid tr	( 457)	103	30.7	0.78	0.234	0.622	111
sp P24965	CYB_TRANA Cytochrome b	( 379)	101	30.3	0.87	0.234	0.563	158
sp P29631	CYB_POMTE Cytochrome b	( 308)	99	29.9	0.95	0.274	0.584	113
sp P24953	CYB_CAPHI Cytochrome b	( 379)	99	29.8	1.2	0.236	0.564	140

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## Alberts is wrong about sequence similarity (three times in three claims)

"With such a large number of proteins in the database, the search programs find *many nonsignificant matches*, resulting in a background noise level that makes it very difficult to pick out all but the closest relatives. Generally speaking, *one requires a 30% identity* in sequence to consider that two proteins match. However, we know the function of many short signature sequences ("fingerprints"), and *these are widely used to find more distant relationships*."

– Alberts, Molecular Biology of the Cell (5<sup>th</sup> ed) p. 139

- Sequences producing statistically significant alignments ALWAYS share a common structure
- Many significant alignments share < 30% identity (<25% identity is routine, and <20% identity can be significant)
- In the absence of significant similarity, "fingerprints" should never be trusted.

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### ATP-synt\_a

```
>>sp|P0AB98|ATP6_ECOLI ATP synthase a chain (ATPase protein 6) g (271 aa)
s-w opt: 178 Z-score: 218.2 bits: 47.9 E(): 3.2e-06
Smith-Waterman score: 178; 23.3% identity (58.5% similar) in 236 aa overlap (8-222:45-264)
```

	10	20	30	40				
human	M N E N L F A S F I A P T I L G L P A A V L I I L F P P L I P T S K Y L I N N R L I T T Q Q							
E coli	N M T P Q D Y I G H H L N N L Q L D L R T F S L V D P Q N P P A T F W T I N I D S M F F S V V L G L	L F L V L F R S V A K K A T S G	V P G K F Q T A I E					
	10	20	30	40	50	60	70	80
human	50	60	70	80	90	100	110	
human	W L I K L T S K Q M M T M H N T K G R T W S L M L V S L I I F I A T T N L L G L L P	H S F	T P T T Q L S M N L A M A I P L W A G					
E coli	L V I G F V N G S V K D M Y H G K S K L I A P L A L T I F V W V F L M N L M D L L P I D L L P Y I A E H V L G L P A L R V V P S A D V N V T L S M A L G V F							
	90	100	110	120	130	140	150	
human	120	130	140	150	160	170	180	
human	T V I M G F R S K I K N A L A H F L P Q G T P T P L	I P M L V I I E T I S L L I Q P M A L A V R L T A N I T A G H L L M H L I G S A T L A M S T I N L						
E coli	I L I L F Y S I K M K G I G G F T K E L T L Q P F N H W A F I P V N L I E G V S L L S K P V S L G L R L F G N M Y A G E L I F I L I A G L L P W W S Q W I L							
	160	170	180	190	200	210	220	230
human	190	200	210	220				
human	P S T L I I F T I L I L L T I L E I A V A L I Q A Y V F T L L V S L Y L H D N T							
E coli	N V P W A I F H I L I T	L Q A F I F M V L T I V Y L S M A S E E H						
	240	250	260	270				

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Query: atp6\_human.aa ATP synthase a chain - 226 aa  
Library: 5190103 residues in 13351 sequences

The best scores are:

		( len)	s-w	bits	E(13351)	%_id	%_sim	alen
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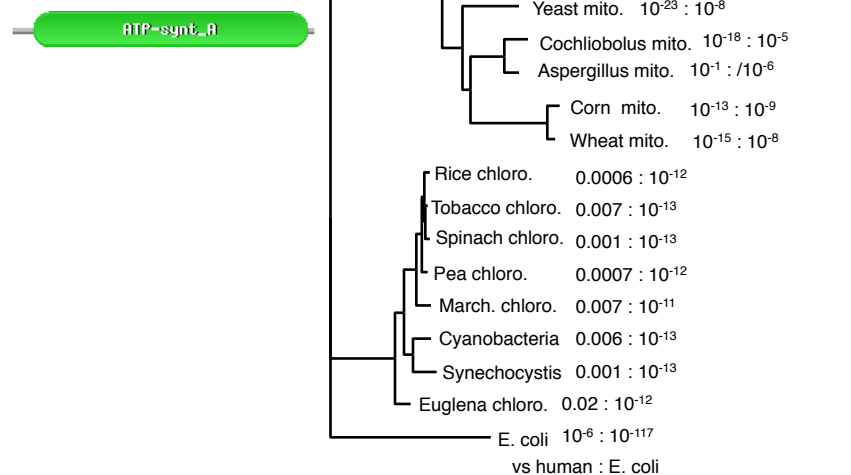
Query: atp6\_ecoli.aa ATP synthase a - 271 aa  
Library: 5190103 residues in 13351 sequences

The best scores are:

		( len)	s-w	bits	E(13351)	%_id	%_sim	alen
sp P0AB98	ATP6_ECOLI ATP synthase a chain (AT	( 271)	1774	416.8	3.e-117	1.000	1.000	271
sp P06451	ATPI_SPIOL Chloroplast ATP synthase	( 247)	274	70.4	5.8e-13	0.270	0.616	211
sp P69371	ATPI_ATRBE Chloroplast ATP synthase	( 247)	271	69.7	9.3e-13	0.270	0.607	211
sp P08444	ATP6_SYNP6 ATP synthase a chain (AT	( 261)	271	69.7	9.9e-13	0.267	0.600	240
sp P06452	ATPI_PEA Chloroplast ATP synthase a	( 247)	266	68.5	2.1e-12	0.274	0.614	223
sp P30391	ATPI_EUGGR Chloroplast ATP synthase	( 251)	265	68.3	2.5e-12	0.298	0.596	225
sp P0C2Y5	ATPI_ORYSA Chloroplast ATP synthase	( 247)	260	67.2	5.4e-12	0.259	0.603	239
sp P27178	ATP6_SYNY3 ATP synthase a chain (AT	( 276)	260	67.1	6.1e-12	0.264	0.578	258
sp P06289	ATPI_MARPO Chloroplast ATP synthase	( 248)	250	64.8	2.7e-11	0.261	0.621	211
sp P07925	ATP6_MAIZE ATP synthase a chain (AT	( 291)	215	56.7	8.7e-09	0.259	0.578	232
sp P68526	ATP6_TRITI ATP synthase a chain (AT	( 386)	209	55.3	3.1e-08	0.259	0.603	239
sp P00854	ATP6_YEAST ATP synthase a chain pre	( 259)	204	54.2	4.5e-08	0.235	0.578	277
sp P05499	ATP6_TOBAC ATP synthase a chain (AT	( 395)	189	50.7	7.8e-07	0.220	0.582	268
sp P00846	ATP6_HUMAN ATP synthase a chain (AT	( 226)	178	48.2	2.5e-06	0.237	0.589	236
sp P00852	ATP6_EMENI ATP synthase a chain pre	( 256)	178	48.2	2.8e-06	0.209	0.590	244
sp P00849	ATP6_XENLA ATP synthase a chain (AT	( 226)	173	47.1	5.5e-06	0.261	0.630	165
sp P00847	ATP6_BOVIN ATP synthase a chain (AT	( 226)	172	46.8	6.5e-06	0.233	0.581	236
sp P14862	ATP6_COACHE ATP synthase a chain (AT	( 257)	171	46.6	8.7e-06	0.204	0.608	265
sp P00848	ATP6_MOUSE ATP synthase a chain (AT	( 226)	166	45.5	1.7e-05	0.259	0.617	193
sp P00851	ATP6_DROYA ATP synthase a chain (AT	( 224)	139	39.2	0.0013	0.225	0.549	253
sp P24962	CYB_STELO Cytochrome b	( 379)	125	35.9	0.021	0.223	0.575	193
sp P09716	US17_HCMVA Hypothetical protein HVL	( 293)	109	32.3	0.21	0.260	0.565	131
sp P68092	CYB_STEAT Cytochrome b	( 379)	109	32.2	0.27	0.211	0.562	194
sp P24960	CYB_ODOHE Cytochrome b	( 379)	104	31.1	0.61	0.210	0.555	200
sp P03887	NU1M_BOVIN NADH-ubiquinone oxidored	( 318)	98	29.7	1.3	0.287	0.545	167
sp P24992	CYB_ANTAM Cytochrome b	( 379)	99	29.9	1.4	0.192	0.565	193

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## Homology is Transitive (on domains)



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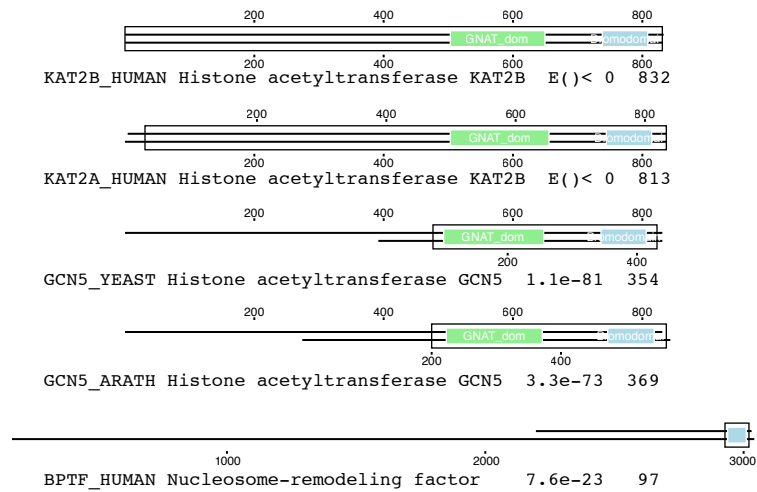
## Homology and Domains – Histone acetyltransferase KAT2B

The best scores are:

	s-w	bits	E(454402)	%_id	%_sim	alen
KAT2B_HUMAN Histone acetyltransferase KAT2B ( 832)	3820	1456.	0	1.000	1.000	832
KAT2A_HUMAN Histone acetyltransferase KAT2A ( 837)	2747	1049.	0	0.721	0.870	813
GCN5_SCHPO Histone acetyltransferase gcn5 ( 454)	867	334.7	3e-90	0.483	0.768	354
GCN5_YEAST Histone acetyltransferase GCN5 ( 439)	792	306.2	1.1e-81	0.469	0.760	354
GCN5_ORYSJ Histone acetyltransferase GCN5 ( 511)	760	294.0	5.9e-78	0.436	0.755	376
GCN5_ARATH Histone acetyltransferase GCN5; ( 568)	719	278.4	3.3e-73	0.434	0.740	369
BPTF_HUMAN Nucleosome-remodeling factor sub (3046)	286	113.6	7.6e-23	0.495	0.804	97
NU301_DROME Nucleosome-remodeling factor su (2669)	276	109.8	9.1e-22	0.511	0.819	94
CECR2_HUMAN Cat eye syndrome critical regio (1484)	232	93.2	5e-17	0.371	0.790	105
BRD4_HUMAN Bromodomain-containing protein 4 (1362)	214	86.4	5.2e-15	0.379	0.698	116
BRD4_MOUSE Bromodomain-containing protein 4 (1400)	214	86.4	5.3e-15	0.379	0.698	116
BAZ2A_HUMAN Bromodomain adjacent to zinc fi (1905)	211	85.2	1.7e-14	0.382	0.683	123
BAZ2A_XENLA Bromodomain adjacent to zinc fi (1698)	206	83.3	5.5e-14	0.350	0.684	117
FSH_DROME Homeotic protein female sterile; (2038)	205	82.9	8.8e-14	0.341	0.667	129
BAZ2A_MOUSE Bromodomain adjacent to zinc fi (1889)	204	82.5	1e-13	0.368	0.680	125
BRDT_MACFA Bromodomain testis-specific prot ( 947)	197	80.0	3e-13	0.367	0.697	109
BRD3_HUMAN Bromodomain-containing protein 3 ( 726)	194	78.9	4.9e-13	0.362	0.664	116

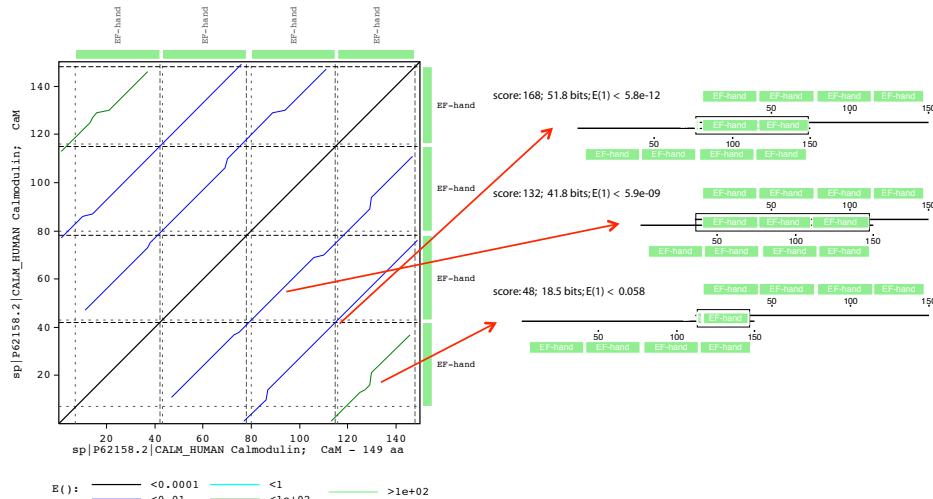
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## Homology and Domains – Histone deacetylase KAT2B



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## LALIGN – Identifying mobile domains: mobile (duplicated) domains in local alignments



## Protein Evolution and Sequence Similarity

- What is Homology and how do we recognize it?
- How do we measure sequence similarity – alignments and scoring matrices?
- **DNA vs protein comparison**
- More effective similarity searching
  - Smaller databases
  - Appropriate scoring matrices
  - Using annotation/domain information

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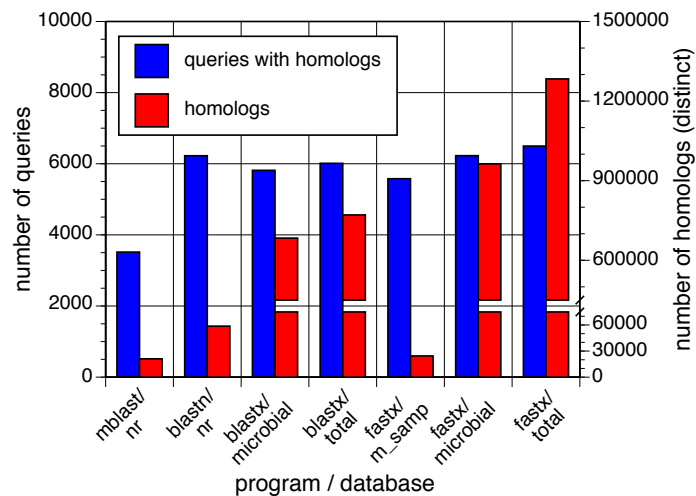
### *DNA vs protein sequence comparison*

The best scores are:

		DNA E(188,018)	tfastx3 E(187,524)	prot. E(331,956)
<del>BMGST</del>	<del>D.melanogaster GST1-1</del>	<del>1.3e-164</del>	<del>4.1e-109</del>	<del>1.0e-109</del>
MDGST1	M.domestica GST-1 gene	2e-77	3.0e-95	1.9e-76
LUCGLTR	Lucilia cuprina GST	1.5e-72	5.2e-91	3.3e-73
MDGST2A	M.domesticus GST-2 mRNA	9.3e-53	1.4e-77	1.6e-62
MDNF1	M.domestica nf1 gene. 10	4.6e-51	2.8e-77	2.2e-62
MDNF6	M.domestica nf6 gene. 10	2.8e-51	4.2e-77	3.1e-62
MDNF7	M.domestica nf7 gene. 10	6.1e-47	9.2e-77	6.7e-62
AGGST15	A.gambiae GST mRNA	3.1e-58	4.2e-76	4.3e-61
CVU87958	Culicoides GST	1.8e-41	4.0e-73	3.6e-58
AGG3GST11	A.gambiae GST1-1 mRNA	1.5e-46	2.8e-55	1.1e-43
BMO6502	Bombyx mori GST mRNA	1.1e-23	8.8e-50	5.7e-40
AGSUGST12	A.gambiae GST1-1 gene	2.3e-16	4.5e-46	5.1e-37
MOTGLUSTRA	Manduca sexta GST	5.7e-07	2.5e-30	8.0e-25
RLGSTARGN	R.leguminosarum gsta	0.0029	3.2e-13	1.4e-10
HUMGSTT2A	H. sapiens GSTT2	0.32	3.3e-10	2.0e-09
HSGSTT1	H.sapiens GSTT1 mRNA	7.2	8.4e-13	3.6e-10
ECAE000319	E. coli hypothet. prot.	—	4.7e-10	1.1e-09
MYMDCMA	Methyl. dichlorometh. DH	—	1.1e-09	6.9e-07
BCU19883	Burkholderia maleylacetate red.	—	1.2e-09	1.1e-08
NFU43126	Naegleria fowleri GST	—	3.2e-07	0.0056
SP505GST	Sphingomonas paucim	—	1.8e-06	0.0002
EN1838	H. sapiens maleylaceto. iso.	—	2.1e-06	5.9e-06
HSU86529	Human GSTZ1	—	3.0e-06	8.0e-06
SYCCPNC	Synechocystis GST	—	1.2e-05	9.5e-06
HSEF1GMR	H.sapiens EF1g mRNA	—	9.0e-05	0.00065

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### Improving search strategies (windshield splatter metagenomics)



- always use protein/translated DNA comparisons
- smaller databases are more sensitive

### Effective Similarity Searching

1. Always search protein databases (possibly with translated DNA)
2. Use E()-values, not percent identity, to infer homology
  - $E() < 0.001$  is significant in a single search (proteins)
3. Search smaller (comprehensive) databases
4. Change the scoring matrix for:
  - short sequences (exons, reads)
  - short evolutionary distances (mammals, vertebrates, a-proteobacteria)
  - high identity (>50% alignments) to reduce over-extension
5. All methods (pairwise, HMM, PSSM) miss homologs, and find homologs the other methods miss



Computer lab:  
[fasta.bioch.virginia.edu/mol\\_evol](http://fasta.bioch.virginia.edu/mol_evol)

- Significant hits are homologous
- Non-significant hits? Homologous or not?
- Are *all* aligned residues homologous
- Are *unaligned* residues non-homologous
- Are domains really missing?

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