

**MASARYK UNIVERSITY**  
**Faculty of Science**

Dissertation

Brno 2021

Stanislav Geidl

**MASARYK UNIVERSITY**  
**Faculty of Science**  
**National Centre for Biomolecular Research**

Chemoinformatical  
Methods for Prediction of  
Physico-chemical Properties  
of Molecules

Dissertation

Stanislav Geidl

Supervisor: prof. RNDr. Jaroslav Koča, DrSc.

Brno 2021

To my grandma.

# Bibliografický záznam

**Autor:** RNDr. Stanislav Geidl  
Přírodovědecká fakulta, Masarykova univerzita  
Národní centrum pro výzkum biomolekul

**Název práce:** Chemoinformatické metody predikce fyzikálně-chemických vlastností molekul

**Studiijní program:** Biomolekulární chemie a bioinformatika

**Vedoucí práce:** prof. RNDr. Jaroslav Koča, DrSc.

**Konzultant práce:** doc. RNDr. Radka Svobodová Vařeková, Ph.D.

**Akademický rok:** 2020/2021

**Počet stran:**

**Klíčová slova:** chemoinformatika, parciální atomové náboje, pKa

# Bibliographic Entry

<b>Author:</b>	RNDr. Stanislav Geidl Faculty of Science, Masaryk University National Centre for Biomolecular Research
<b>Title of Thesis:</b>	Chemoinformatical Methods for Prediction of Physico-Chemical Properties of Molecules
<b>Degree Programme:</b>	Biomolecular chemistry and bioinformatics
<b>Supervisor:</b>	prof. RNDr. Jaroslav Koča, DrSc.
<b>Supervisor Specialist:</b>	doc. RNDr. Radka Svobodová Vařeková, Ph.D.
<b>Academic Year:</b>	2020/2021
<b>Number of Pages:</b>	
<b>Keywords:</b>	chemoinformatics, partial atomic charges, pKa

# Abstrakt

Chemoinformatické přístupy pro výpočet fyzikálních a chemických vlastností organických molekul, speciálně pak molekul dosud nesyntetizovaných, jsou velmi užitečné v rámci procesu vývoje léčiv a v dalších oblastech moderních přírodních věd. Jednou z velmi důležitých vlastností organických molekul je disociační konstanta ( $pK_a$ ).  $pK_a$  lze úspěšně predikovat pomocí chemoinformatických modelů založených na parciálních atomových nábojích. Tyto modely vyžadují molekulární 3D strukturu, kterou však lze připravit mnoha různými způsoby.

Prvním tématem, kterým jsem se v rámci své práce zabýval, je právě vliv zdroje 3D struktury molekul na přesnost predikce  $pK_a$ . Zjistil jsem, že výběr zdroje 3D struktury je klíčový pro úspěšnou predikci  $pK_a$ , přičemž 3D struktury z databází DTP NCI a PubChem je jevily jako nevhodnější. Z této analýzy byla rovněž patrná nutnost kvalitních a rychle vypočítatelných parciálních atomových nábojů, sloužících jako vstupy pro predikci  $pK_a$ . Uvedená oblast se stala druhým tématem mé práce. Konkrétně jsem se zaměřil na parametrizaci metody Electronegativity Equalization Method (EEM), sloužící pro rychlý výpočet parciálních atomových nábojů. Výsledkem mé práce byly EEM parametry, poskytující kvalitní náboje pro léčiva a jím podobné organické molekuly. Výsledkem mé práce byly EEM parametry, poskytující vysoko kvalitní náboje pro léčiva a jím podobné organické molekuly. Při přípravě těchto parametrů jsem si rovněž uvědomil nutnost mít k dispozici softwarový nástroj pro výpočet nábojů a parametrizaci nábojových metod. Tato problematika se stala třetím tématem mé práce – spolupracoval jsem na vývoji software NEEMP, který slouží k parametrizaci EEM, validaci EEM parametrů a výpočtu nábojů pomocí metody EEM.

Celkově má práce poskytuje metodiky a nástroje pro stavbu kompletního workflow, sloužícího pro výpočet i u dosud nesyntetizovaných molekul. Toto workflow zahrnuje získání 3D struktury molekul, výpočet jejich parciálních atomových nábojů metodou EEM a aplikace nábojů pro predikci  $pK_a$ .

# Abstract

Chemoinformatic approaches for predicting physicochemical properties, especially in the case of unsynthesized molecules, are beneficial in the drug design process and other modern life science fields. One of the most important properties of organic molecules is the dissociation constant ( $pK_a$ ).  $pK_a$  is successfully predictable by chemoinformatics models based on partial atomic charges – these models require a molecules' 3D structure that can be obtained using different approaches. The first topic of my work is to analyze the influence of 3D structure sources on the quality of  $pK_a$  prediction. I found out that the correct source of 3D structure is crucial for  $pK_a$  prediction while structures from databases DTP NCI and PubChem appear most suitable. This work shows a need for quality and quickly calculated charges used as input for  $pK_a$  prediction. This field became my second topic. Specifically, I focused on Electronegativity Equalization Method (EEM) for quick partial atomic charges calculation. The result of my work was the EEM parameters, which provide high-quality charges for drug-like molecules. During EEM parameters preparation, I realized a need to have a tool for partial charge calculation and parametrization. This problem became my third topic – I cooperated on the development of NEEMP software that can parametrize EEM, validate EEM parameters and calculate charges via EEM method.

Overall, my work provides a methodology and tools for building the whole workflow used for  $pK_a$  prediction, which can be used for unsynthesized molecules. This workflow contains obtaining 3D structures of molecules, partial atomic charges calculation, and  $pK_a$  prediction.

## **Acknowledgements**

I want to express my deepest gratitude to my supervisor, prof. RNDr. Jaroslav Koča, DrSc. and my supervisor specialist doc. RNDr. Radka Svobodová Vařeková, PhD., for their valuable pieces of advice, support, and help during my whole university study. I am very grateful that I can learn from them. I want to thank the co-authors of all my articles and my former student for their excellent co-operation and dedication.

I am obliged to my father, grandma, and grandpa for their push and unwavering support. Without them, I couldn't go so far. At last but not least, I would like to thank my partner for everything.

## **Declaration**

I hereby declare that this dissertation thesis is my original authorial work, which I have worked on alone. All sources, references and literature used or excerpted during the elaboration of this work are properly cited and listed in a complete reference with regard to the source.

Brno, xxth June 2021

.....

Stanislav Geidl

# Publication list with definition of autor's contribution

This dissertation is based on 3 articles of dissertation author (Stanislav Geidl, SG):

Geidl S, Svobodová Vařeková R, Bendová V, Petrusek L, Ionescu C-M, Jurka Z, Abagyan R, Koča J: **How Does the Methodology of 3D Structure Preparation Influence the Quality of  $pK_a$  Prediction?** *J Chem Inf Model* 2015, **55**:1088–1097.

SG prepared the input dataset (by extension of published datasets), performed all the calculations, participated in the analysis of the results, and wrote a part of the manuscript, including all tables and graphics.

Geidl S, Bouchal T, Raček T, Svobodová Vařeková R, Hejret V, Křenek A, Abagyan R, Koča J: **High-quality and universal empirical atomic charges for chemoinformatics applications.** *J Cheminform* 2015, **7**:59.

SG participated in the study's design, and cooperated in the preparation of the input data (molecules and published EEM parameters) and in QM charge calculation. SG performed the analyses and the interpretation of the data.

Raček T, Pazúriková J, Svobodová Vařekova R, Geidl S, Křenek A, Falginella FL, Horský V, Hejret V, Koča J: **NEEMP: Software for validation, accurate calculation and fast parameterization of EEM charges.** *J Cheminform* 2016, **8**:1.

SG prototyped the DE-Min approach and designed some new NEEMP features, such as the usage of RMSE instead of  $R^2$ . SG also implemented a preparation of validation reports.

---

# **Contents**

# **Part I**

# **Introduction**

# 1

---

## Introduction

In recent years, a vast amount of data about various types of molecules became available. For example, we can obtain the complete human genome of a selected individual in a few days, and about 150 thousand biomacromolecular structures have been determined and published (Protein Data Bank [1]). Furthermore, more than 100 million various small molecules are described in freely accessible databases (e.g., Pubchem [2], ZINC [3], ChEMBL [4]). This richness of data caused the formation of novel modern life-science research fields focused on the utilization of this data. The best-known modern life sciences are bioinformatics, structural bioinformatics, systems biology, genomics, proteomics, and also chemoinformatics. These current research specializations have provided many key results in basic and applied research (e.g. [5–11]).

One fascinating and beneficial field utilizing and processing newly available data about small molecules (i.e., drug-like compounds) is chemoinformatics. This discipline offers methodologies for comparing molecular similarity, molecular database search, virtual screening, and the prediction of molecules' properties and activities. This prediction is based on the idea that molecular structures' similarity has a consequence – a similarity in molecular properties. In chemoinformatics, the structure is first described using mathematical characteristics (so-called descriptors) – numbers containing 3D (or 2D or 1D) structure information and applicable as inputs of mathematical models. Then, these models are constructed based on a relation between descriptors and known values of the property or the activity. Such models are called Quantitative Structure-Property Relationship (QSPR) models or Quantitative Structure-Activity Relationship (QSAR) models.

A property, which is strongly required and is therefore often a target of chemoinformatics prediction models is the acid dissociation constant,  $K_a$ , and its negative logarithm  $pK_a$ . Those  $pK_a$  values are of interest in chemical, biological,

environmental, and pharmaceutical research [12–14].  $pK_a$  values have found applications in many areas, such as evaluating and optimizing drug candidate molecules, pharmacokinetics, ADME profiling, understanding protein-ligand interactions, etc. Moreover, the critical physicochemical properties such as permeability, lipophilicity, solubility, etc., are  $pK_a$  dependent. Unfortunately, experimental  $pK_a$  values are available only for a limited set of molecules. In addition to that, obtaining experimental  $pK_a$  values for newly designed molecules is very time-consuming because they must be synthesized first. Chemoinformatics approaches for  $pK_a$  prediction are therefore currently intensively examined.

For this reason, I also focused on the chemoinformatics way of  $pK_a$  prediction in my work. Very promising descriptors for  $pK_a$  prediction are partial atomic charges [15–20] because they hold information about the distribution of electron density within the molecule. Specifically, electron densities on atoms close to the dissociating hydrogen provide a clue about its dissociation ability. The most common and accurate method for calculating partial atomic charges is an application of quantum mechanics (QM). QM calculation can be performed via various approaches, introducing different approximation levels (i.e., approximating a wave function by different sets of mathematical equations, which are called basis sets). QM outputs electron distribution in orbitals and this distribution can be divided into individual atoms using several charge calculation schemes (e.g., MPA, NPA, AIM, Hirshfeld, MK, etc.). Therefore, the correlation between  $pK_a$  and relevant atomic charges calculated by different QM approaches has been analyzed [19]. I also focused on this file in my bachelor thesis [21], developed a workflow for calculation of  $pK_a$  using QM partial atomic charges and examined, which types of QM are the most suitable.

QM charges are accurate, but their calculation is very time-consuming. A faster Alternative to QM charges is empirical charge calculation approaches. Furthermore, if we would like to apply chemoinformatics  $pK_a$  prediction models practically – for example, in pre-screening large sets of drug candidates – we need a fast approach. Therefore, in my master thesis [22], I developed a  $pK_a$  prediction workflow based on charges (including Electronegativity Equalization Method).

However, several pieces of the puzzle were still missing. For example, the developed  $pK_a$  prediction workflows [20] were strongly dependent on 3D structure source, and also, the quality of available EEM charges was low.

Therefore, my dissertation's goal was to develop a workflow that predicts  $pK_a$  for molecules not synthesized yet and without available experimental 3D structures.

Specifically, the thesis examined how to improve the process of  $pK_a$  prediction via providing suitable inputs. First, the influence of 3D structure source on  $pK_a$  prediction accuracy was analyzed. Afterward, the work focused on obtaining high-quality partial atomic charges, which served as descriptors for  $pK_a$ .

calculation. In the end, the authors also support the development of methodology and software tools for obtaining these high-quality charges.

The thesis structure is the following: First, an overview of key fields is provided (Part II), i.e. – 3D structure and approaches for its prediction, charge calculation methods, and  $pK_a$  prediction approaches. Next, the achieved results, which we published in three research papers, are briefly described (Part III), and full-texts of the respective published papers are attached in Main papers. During the elaboration of this thesis, I was also involved in other projects. Most of them were not related to  $pK_a$  prediction but tightly connected to the field of chemoinformatics or structural bioinformatics. The outcome of these projects consists of several papers and a book I have co-authored. Their title pages are attached in Appendix: Other Publications.

## **Part II**

# **Theory and Methods**

# 2

---

## Structure

### 2.1 Molecular Structure in Computer

The chemical structure is the essential information for chemoinformatics and computational chemistry calculations. We recognize different types of chemical structures according to the complexity of information [23].

The empirical or chemical formula provides information about molecule composition – elements and their count. The structural formula (2D structure) extends this information about topology – bonds between atoms. The three-dimensional structure also provides the conformation of a molecule – the relative placement of atoms in space. We try to provide conformation with the lowest energy representing the most probable conformation of molecule in reality. For some applications, there can even be an assembly of these 3D structures.

In chemoinformatics, two-dimensional structures are often used, but the three-dimensional structure can often bring new information into the *in silico* calculations or models. On the other hand, this 3D structure can be obtained experimentally for a limited number of small molecules. What with other molecules, including those which were not synthesized yet?

### 2.2 3D Structure Calculation

We apply more computationally efficient methods for 3D structure computation because we use them as input for high-throughput methods. For this reason, many resources were devoted to the development of fast and accurate 3D structure prediction methods [24]. These can be classified into the following groups [24]: rule-based and data-based, fragment-based, numerical methods,

and conformational analysis. These rule-based and data-based, fragment-based methods are partially overlying.

### **2.2.1 Rule-Based and Data-Based Methods**

These methods use chemical knowledge of geometric and energetic rules known from experiments and theoretical calculations. In these methods, we use rules explicitly to describe, e.g., bond lengths and angles; we use data implicitly to describe, e.g., ring conformation.

### **2.2.2 Fragment-Based Method**

The fragment-based method is the incremental method using rules in the first step to fragment a structure into parts. According to the following rules, the parts are assembled by linking fragment templates from a library (database). Predicted structures are created from the most similar and largest fragments in a database as possible.

### **2.2.3 Numerical Method**

These numerical methods consist of three methods: molecular mechanics (MM), quantum mechanics, and distance geometry (DG). Distance geometry is a great tool to prepare a reasonable initial structure, which is very close to some low-energy conformation. For this structure, we can use the optimization process from MM or/and QM.

### **2.2.4 Conformational Analysis**

This method generates a set of conformations for one molecule using different approaches - genetic algorithms, systematic methods, random techniques, Monte Carlo or MD simulation. The one or more different structures are selected based on criteria such as the number of conformers, minimum RMSD, only conformations with the lowest MM energy (low-energy conformers).

# 3

---

## Partial Atomic Charges

### 3.1 The Concept of Atomic Charges

Atomic charges are a theoretical concept for the quantitative description of electron density around every atom in a molecule. The first basic concept came from early chemistry, where an integer expressed these charges (e.g. -1, +2). Later, they were a real number (partial charge) in organic chemistry and physical chemistry [25]. It is a great approach to explain the mechanism of a lot of chemical reactions. Recently, partial atomic charges also became popular in chemoinformatics, as they proved to be informative descriptors for QSAR and QSPR modeling [16, 26] and for other applications [17, 27, 28]; they can be utilized in virtual screening [29, 30] and similarity searches [31, 32]. In reality, we are not able to measure these numbers, only calculate or estimate them. For such reasons, many different approaches for the calculation of partial atomic charges were developed.

### 3.2 Overview of Charge Calculation Methods

#### 3.2.1 QM Charge Methods

These methods use a wave function as a starting point and then apply subsequent population analysis, charge calculation scheme, or fit to some physical observation [33].

Mulliken population analysis (MPA) [34, 35] simply calculates a charge of an atom as a sum of an electron density from its molecular orbitals and a half

of an electron density from its bonding orbitals. Natural population analysis (NPA) [36, 37] sophisticatedly improves the MPA method by orthogonalization of specific atoms and after this, NPA performs charge assignment from electron density the same way as in MPA. NPA atomic charges are more stable and independent of the size of basis sets. Other possible population analyses are Löwdin population analysis [38], Hirshfeld population analysis [39].

AIM (atoms-in-molecules) charge calculation scheme is based on the idea that electron density measured by X-ray can help with the calculation of partial charges. Bader and his coworkers [40, 41] defined an atomic volume that is used for charge calculation. Other well-known approaches are electrostatic potential fitting methods (ESP) like CHELPG [42] or MK (Merz-Singh-Kollman) [43] and their extension – RESP methods [44].

Cramer and at [45] also developed semiempirical methods – charge model 5 (CM5), which extends Hirshfeld population analysis by empirical parameters to reproduce charge-dependent observables.

### 3.2.2 Empirical Methods

Empirical approaches use only empirical parameters, and some of these can calculate charges from the 3D structure or only from the topology (2D structure) of a molecule. Therefore, they are distinctly faster than QM approaches.

One of the first empirical methods developed, CHARGE [46], performs a breakdown of the charge transmission by polar atoms into single-bond, double-bond, and triple-bond additive contributions. Other empirical methods have been developed on the electronegativity equalization principle. One group of these empirical approaches are using the Laplacian matrix formalism and the product is a redistribution of electronegativity: Gasteiger-Marsili (PEOE, partial equalization of orbital electronegativity) [47], GDAC (geometry-dependent atomic charge) [48], KCM (Kirchhoff charge model) [49], DENR (dynamic electronegativity relaxation) [50] or TSEF (topologically symmetric energy function) [50].

The second group of approaches applies the full equalization of orbital electronegativity. For example, this group contains EEM (electronegativity equalization method) [51] and its extensions (ABEEM [52], SFKEEM [26]), QEeq (charge equilibration) [53], EQEeq (extended QEeq) [54], or SQE (split charge equilibration) [55].

Group of conformationally independent methods (based on the 2D structure) contains CHARGE, Gasteiger-Marsili, KCM, DENR, and TSEF. Group of conformationally dependent – geometrical charges (based on the 3D structure) also consider an influence of conformation and includes the following methods: GDAC, EEM, ABEEM, SFKEEM, QEeq, EQEeq, and SQE.

A typical representative of the topological method is the Gasteiger-Marsili method, which first assigns charges based on atom types and then iteratively updates atomic charges based on the closest partners. The correction is smaller and smaller in every step until the sixth step when these corrections are too small and atomic charges are final. Empirical parameters for this method were calculated from QM.

On the other hand, the EEM method needs a complete 3D structure and more applicable charges for some of the applications.

### 3.3 EEM Calculation

EEM (electronegativity equalization method) [51] is one of the most popular empirical charge calculation methods and was developed more than twenty years ago. This method's new parameterizations [26, 51, 56–61] and extension [26, 52] are still under development. An advantage of EEM calculation is that it considers the influence of the molecule's conformation on the atomic charges. For this reason, EEM charges are often used in predictive models as chemoinformatics regressors (descriptors) [62].

EEM is based on three principles:

The first principle is Sanderson's electronegativity equalization principle. It assumes that the effective electronegativity of each atom in the molecule is equal to the molecular electronegativity:

$$\chi_1 = \chi_2 = \dots = \chi_x = \bar{\chi} \quad (3.1)$$

where  $\chi_x$  is the effective electronegativity of the atom  $x$  and  $\bar{\chi}$  is the molecular electronegativity.

The second principle is the principle of the charge balance. The sum of all charges is equal to the total charge  $Q$ :

$$\sum_{x=1} q_x = Q \quad (3.2)$$

where  $q_x$  is the charge of the atom  $x$ .

And the last principle is the principle of charge-dependent electronegativity. This principle is the definition of atomic electronegativity, and states that the electronegativity of each atom can be expressed as a function of its charge:

$$\chi_i = A_i + B_i \cdot q_i + \kappa \sum_{j=1, i \neq j}^N \frac{q_j}{R_{i,j}} \quad (3.3)$$

where  $R_{i,j}$  is the distance between atoms  $i$  and  $j$ , and the coefficients  $A_i$ ,  $B_i$  and  $\kappa$  are empirical parameters.

These principles can be summed up to a system of equations with  $N + 1$  unknowns (where  $q_1, q_2, \dots, q_N$  and  $\bar{\chi}$ ):

$$\begin{pmatrix} B_1 & \frac{\kappa}{R_{1,2}} & \cdots & \frac{\kappa}{R_{1,N}} & -1 \\ \frac{\kappa}{R_{2,1}} & B_2 & \cdots & \frac{\kappa}{R_{2,N}} & -1 \\ \vdots & \vdots & \ddots & \vdots & \vdots \\ \frac{\kappa}{R_{N,1}} & \frac{\kappa}{R_{N,2}} & \cdots & B_N & -1 \\ 1 & 1 & 1 & 1 & 0 \end{pmatrix} \cdot \begin{pmatrix} q_1 \\ q_2 \\ \vdots \\ q_N \\ \bar{\chi} \end{pmatrix} = \begin{pmatrix} -A_1 \\ -A_2 \\ \vdots \\ -A_N \\ Q \end{pmatrix} \quad (3.4)$$

The first values of parameters  $A_i$  and  $B_i$  were modifications of experimental hardness and electronegativity [51].  $\kappa$  was equal to 1. Nowadays, these parameters are calculated from the QM charges [26, 56–61]. Therefore, EEM charges were correlated with the QM charge calculated with the same method used for parametrization.

### 3.4 Quality and Usability of EEM parameters

The quality of EEM parameters describes how the empirical charges computed using these EEM parameters correspond with QM charges used for EEM parameterization. Three main characteristics (statistics [63, 64]) can describe the quality of EEM parameters – the coefficient of determination root mean square error (RMSE) and average absolute error ( $\bar{\Delta}$ ).

**The coefficient of determination**  $R^2$  is the squared value of the Pearson coefficient (equation 3.5). This value describes the linear correlation rate. Values close to 1 mean that values correlate very well, and values close to 0 mean no correlation.

$$R = \sqrt{\frac{\sum_{x=1}^N ((q_x^{calc} - \bar{q}_x^{calc}) \cdot (q_x^{ref} - \bar{q}_x^{ref}))}{\sum_{x=1}^N (q_x^{calc} - \bar{q}_x^{calc})^2 \cdot \sum_{x=1}^N (q_x^{ref} - \bar{q}_x^{ref})^2}} \quad (3.5)$$

where  $q^{ref}$  is the reference value of charge calculated by QM and  $q^{calc}$  is charge value calculated by EEM.  $\bar{q}^{ref}$ ,  $\bar{q}^{calc}$  are the average value of  $q^{ref}$ , respectively  $q^{calc}$ .

**Root mean square error** RMSE is the normalized sum of squared error describing the reliability of the model calculated by:

$$RMSE = \frac{\sum_{x=1}^N (q_x^{calc} - q_x^{ref})^2}{N} \quad (3.6)$$

**Average absolute error**  $\bar{\Delta}$  is an averaged difference between corresponding EEM and QM charges in a molecule and is calculated according to an equation:

$$\bar{\Delta} = \frac{\sum_{x=1}^N |q_x^{calc} - q_x^{ref}|}{N} \quad (3.7)$$

Their **coverage** describes the applicability of EEM parameters. Coverage is a percentage value describing EEM parameters' ability to calculate charges for individual molecules in a dedicated dataset. *De facto*, this coverage depends on the representation of atom types in EEM parameters.

$$\text{coverage} = \frac{N_{pos}}{N_{tot}} \quad (3.8)$$

where  $N_{pos}$  is the number of molecules able calculated by EEM parameters and  $N_{tot}$  is the total number of molecules in a dataset.

## 3.5 EEM Parametrization

For the parameterization of EEM charges, a lot of different methods have been introduced. We can summarize it into two groups: one group contains a method that analytically solves equation [?] – linear regression [59, 60] and the second group contains optimization methods [61] such as Accelerated Random Search, Particle Swarm Optimization, and Differential Evolution algorithms. Both of these groups need input – a set of molecules with 3D structures and QM atomic charges. In my work, linear regression and differential evolution were used, and therefore, they are described in more detail below:

**The linear regression (LR)** method is based on these two equations:

$$A_i + B_i \cdot x = y \quad (3.9)$$

$$\begin{aligned} x &= q_i \\ y &= \chi_i - \kappa \sum_{j=1}^N i \neq j \frac{q_j}{R_{i,j}} \end{aligned} \quad (3.10)$$

Equations are derived from equations 3.9 and 3.10, which define the EEM method. In the LR method, the dataset of molecules with QM charges can change in every iteration to improve the quality of resulting charges. Quality criterium can be the Pearson correlation coefficient or the coefficient of determination, and the root mean square error or different types of errors. An advantage of the

LR method is its straightforwardness and the possibility to optimize  $\kappa$  by another iteration. On the other hand, this method is not possible to make parametrization for some extensions of EEM like SFKEEM and ABEEM.

**Differential Evaluation (DE)** [65] is a heuristics method to focus on finding a global minimum of a function. This method works similar to other optimization methods – iteratively optimize parameters to improve the final solution. Parameters of function are set up randomly, mutated, and evaluated until there is no best solution.

# 4

---

## Acid Dissociation Constant Prediction

### 4.1 Motivation

The acid dissociation constant ( $pK_a$ ) is a physicochemical property that characterizes the strength of acids. It is one of the essential properties for pharmaceutical, chemical, biological and environmental research or industry. For example, it can be used in the chemoinformatics pipeline for evaluation and optimization of drug candidate [66–68], ADME profiling [69,70], pharmacokinetics [12], understanding protein-ligand interactions [13,71].

### 4.2 Overview of Approaches

Several different approaches for pKa prediction have been developed [71–74].

#### 4.2.1 LFER (Linear Free Energy Relationships) Methods

This is one of the oldest methods [75,76] for  $pK_a$  prediction. This method uses the linear relation of Gibbs energy and  $pK_a$  or the logarithm of a reaction rate constant – the Hammett and Taft equations. An advantage of this method is a simple, straightforward, and quick calculation, but on the other hand, we need substituent and reaction parameters. This method is still used in the programs ACD/pKa [77], EPIK [78], and SPARC [79].

## 4.2.2 Database Methods

These methods [80, 81] use a library (database) of molecules with known pKa values. The  $pK_a$  value is taken directly from this library, or it is interpolated or triangulated from most similar molecules in this library. Most accurate results are produced only for molecules that are similar to molecules in the database. For this reason, it is essential to have an extensive library.

## 4.2.3 Ab Initio Quantum Mechanical Calculations

These methods [82, 83] use the fact that the dissociation constant can be calculated from the Gibbs energy of the reaction and from the solvation based on equation 4.2. However, there is no general approach, and every specific calculation configuration needs to be calibrated based on experimental values. The significant disadvantage of these methods is that they are time-consuming. On the other hand, these methods can be very accurate if they use correct calibration parameters. It is only one of the few methods that can be used to extend the training dataset with experimental values or validate some of this experimental value. It means that other methods can be improved by this method. This method is implemented as a module of the Jaguar quantum chemical software package [84].

$$pK_a = -\log_{10} K_a \quad (4.1)$$

$$K_a = e^{\frac{-\Delta G^\circ}{RT}} \quad (4.2)$$

## 4.2.4 QSPR Method

The quantitative structure–property relationship method [17, 85, 86] uses mainly a linear model to describe the relationship between molecular structure and a property of a molecule, in our case  $pK_a$ . In those models, structures are presented by descriptors [62] that are numerical expressions of molecular properties. For example, descriptors can be the number of hydrogen atoms, the ratio between carbon atoms and all atoms in the molecule, or solvent accessible surface area.

$pK_a$  correlates well with the polarizability, HOMO energy [87], proton-transfer energy [87], partial atomic charges [15, 16, 18–20], the electrostatic potential of the molecule [88], etc. Partial atomic charges proved as very promising descriptors [15, 16, 18–20] for  $pK_a$  prediction.

# **Part III**

# **Results**

# 5

---

## Synopsis of the Results

We published a series of articles about pKa prediction [19, 20] where we showed that some specific atomic charges correlate with  $pK_a$ . Based on this, we were able to build QSPR models for the prediction of this property. We also compared QM, EEM charges, and their models. In this dissertation, I focused only on the last one:

Geidl S, Svobodová Vařeková R, Bendová V, Petrusek L, Ionescu C-M, Jurka Z, Abagyan R, Koča J: How Does the Methodology of 3D Structure Preparation Influence the Quality of  $pK_a$  Prediction? *J Chem Inf Model* 2015, 55:1088–1097.

In this article, we utilized different approaches to generate the 3D structures of organic molecules. These structures were used for the building of  $pK_a$  prediction models based on charge descriptors. Then we analyzed various influences and relationships and found which methodologies for 3D structure preparation are applicable for  $pK_a$  prediction.

We examined not only pKa prediction models employing QM charges but also the models utilizing EEM charges. An application of EEM charges looked very promising. Moreover, EEM charge calculation is significantly faster than QM charge calculation.

However, in parallel, we found one significant limitation of EEM charges – the parameters. It was available a reasonable number of parameter sets, but they had only a low coverage. For example, Bultinc's parameter set [57] contains parameters only for these elements: C, F, H, N, O. This fact markedly reduced a dataset, which we were able to use for QM charges.

A lack of parameters disallows the usage of EEM charges in many chemoinformatics applications. For this reason, in our follow-up work, we focused on

the development of new and more robust EEM parameters. The first step was to develop a new parameterization tool that was easy to use and extendable. After the prototype, we carefully prepared a new dataset of molecules, and for this dataset, we computed EEM parameters with higher coverage. These new parameters were published in a scientific paper:

Geidl S, Bouchal T, Raček T, Svobodová Vařeková R, Hejret V, Křenek A, Abagyan R, Koča J: **High-quality and universal empirical atomic charges for chemoinformatics applications.** *J Cheminform* 2015, **7**:59.

After some tuning up and extensive research, we released and also published the tool for EEM parametrization – the NEEMP software:

Raček T, Pazúriková J, Svobodová Vařekova R, Geidl S, Křenek A, Falginella FL, Horský V, Hejret V, Koča J: **NEEMP: Software for validation, accurate calculation and fast parameterization of EEM charges.** *J Cheminform* 2016, **8**:1.

Sections 5.1, 5.2, and 5.3 contain a summary and Main papers full text of all these aforementioned articles. I was also involved in other projects, and the outcome of it is a list of additional articles and book chapters in Appendix: Other Publications.

## 5.1 How Does the Methodology of 3D Structure Preparation Influence the Quality of pKa Prediction?

From our previous articles [19, 20], we know that the prediction of  $pK_a$  is possible via QSPR models using partial atomic charges as descriptors. The article's goal was to discover how the methodology of 3D structure preparation influences the quality of pKa prediction. We prepared a dataset containing 60 phenols, 82 carboxylic acids, 48 anilines, and additional testing 53 phenols for these purposes. We took structures from 5 different sources for all these molecules: PubChem [2], DTP NCI database [89]; Balloon [90], Frog2 [91], OpenBabel [92], and RDKit [93] software. We used neutral forms of all the molecules and dissociated forms of phenols and carboxylic acids, and associated forms of anilines. We also optimized these structures with MM (Molecular mechanics) and QM. All combination led to 7220 structures for that we calculated four different QM, one semiempirical QM, four different EEM charges, and Gasteiger-

Marselli charges. We created 516 QSPR models for all these molecules and charges. The robustness of these models was tested by cross-validation and QM charges also by an independent test set of phenols.

We confirmed that QM and EEM charge descriptors could be used for  $pK_a$  prediction. About half of all models have excellent quality with  $R^2 \geq 0.9$ . We also showed that ab initio and semiempirical charges correlate with  $pK_a$  and their models are very accurate. In EEM, we had models with a little worse quality, but empirical charges are calculated much faster, and an application in chemoinformatics is much more appropriate. In our models, we were not able to use Gasteiger-Marsili charges to get an adequate quality.

We focused on different types of influence. For classes of the benzene derivates (phenols and anilines) was much easier to obtain high-quality models. Nevertheless, for aliphatic hydrocarbon derivates (carboxylic acid), it was more challenging.

The focus of this article was a comparison of the source of the 3D structure. The influence of input structure for models is essential because the result – the quality of QSPR models – depends on input structures and their quality. For example, structures taken from RDKit generated only by distance geometry produced fragile models. On the other hand, the 3D structures from the DTP NCI and PubChem databases, formally structures generated by CORINA and Omega, exhibited the best performance for all the tested molecular classes and charge calculation approaches. Structures generated by Frog2 also performed very well. Other 3D structure sources can also be used, but with caution.

We also tried the influence of structure optimization on the quality of QSPR models. In most cases, differences between original structures and optimized structures were slight.

In this article, we summed up the best workflow for the fast and accurate prediction of  $pK_a$ . This or similar workflow can also be easily applied to other important properties for *in silico* designed molecules. Flow is about preparing 3D structures by CORINA or Omega (with no further optimization), calculation of EEM charges for these structures, and then the EEM QSPR calculation of  $pK_a$ .

### 5.1.1 My contribution

I prepared the input dataset (by extension of published datasets), performed all the calculations, participated in the analysis of the results, and wrote a part of the manuscript, including all tables and graphics.

## 5.2 High-quality and universal empirical atomic charges for chemoinformatics applications

The EEM method for charge calculation was published several decades ago. Before our study, there were done some improvements of EEM [26, 52], parameterizations of empirical parameters [26, 51, 56–61], and developments of new parameterization methods [26, 57, 59]. However, there was a problem with the usability of EEM because the available parameter sets had a low coverage in chemical space.

We prepared a set of 4475 distinct small organic, drug-like molecules containing these elements: H, C, N, O, F, P, S, Cl, Br, and I, in different functional groups. This set was created so that each selected atom type is contained in at least 100 molecules. CORINA calculated the 3D structure for all molecules. The next step was the calculation of reference QM charges. We selected 6 different approaches: B<sub>3</sub>LYP/6-311G/MPA, B<sub>3</sub>LYP/6-311G/NPA, B<sub>3</sub>LYP/6-311G/AIM, HF/6-311G/MPA, HF/6-311G/NPA, and HF/6-311G/AIM. EEM parameterization was performed for six QM charge calculation approaches, and the whole set of prepared molecules was used.

Our new EEM parameters get very high quality – all coefficients of determination had a value greater or equal to 0.9. We also showed that the used QM approach did not prove any difference in the quality of parameters – B<sub>3</sub>LYP and HF produced comparable results. EEM parameters based on NPA and AIM population analysis are slightly better than EEM parameters based on MPA.

We also calculated coverage of our parameters previously published parameters on four big chemoinformatics databases of drug-like molecules — DrugBank, [94] ChEMBL, [4] PubChem, [2] and ZINC [3]. We found out that their coverage is less than 60% for most of the previously published parameters. Our newly produced parameters showed coverage of at least 90% for these databases. Consistency of coverage points out that this problem is not related to a database but concerns a chemical space of drug-like molecules and their atom types.

For evaluation of quality, we selected 657 approved drugs from the DrugBank database. We compared the coefficient of determination, root mean square error and found out that our new parameters outperform the previously published parameters. Coverage of the old parameters on this small evaluation dataset is like coverage on whole databases.

The quality of EEM parameters is also affected by a used QM charge scheme. EEM parameters derived from MPA, NPA, and AIM charges showed high quality. EEM parameters based on Hirshfeld charges were acceptable, and MK and

CHELPG charges cannot be used with EEM. On the other hand, none of the QM theory level and basis set combinations showed any problem in the quality of EEM parameters. This also confirmed that we used an appropriate selection of reference QM charges.

We also evaluated already existing tools for calculating partial atomic charges, and all the tools showed some issues. For example, OpenBabel tool [92] is using Bult2002\_mpa parameter set [57], but developers extended this set about missing atom types with parameters for different atom types. This hack increases coverage paid by decreased EEM charges quality for molecules containing atom type missing in the original parameter set.

### 5.2.1 My contribution

I participated in the study's design, and I cooperated in the preparation of the input data (molecules and published EEM parameters) and in QM charge calculation. I performed the analyses and the interpretation of the data.

## 5.3 NEEMP: software for validation, accurate calculation, and fast parameterization of EEM charges

This article describes NEEMP – a software tool with three main functionalities – parametrization of EEM charges from reference QM charges, validation of EEM parameter sets (including quality and coverage calculation), and EEM charge calculation.

NEEMP provides two different parametrization approaches:

- linear regression (LE),
- differential evolution with the local minimization (DE-MIN) approach.

A combination of a global optimization method with a local optimization method improves EEM parameterization. This combined approach provides a more robust methodology than LR. Therefore it is applicable even for heterogeneous training sets. Specifically, we combined differential evolution (DE) [65] with the local minimization method NEWUOA [95]. Quality criteria for evaluation of each iteration of the parametrization process (both LR and DE-MIN) can be set up to coefficient of determination ( $R^2$ ) or the root mean square error (RMSE).

The validation mode of NEEMP calculates quality metrics, coverage, and a graphical representation of EEM charge correlation with reference QM charges.

The calculation mode of NEEMP provides a calculation of EEM charges using an input parameter set.

The article also presents two case studies to show the functionality and performance of NEEMP – a parametrization and a validation case study.

The parametrization case study targets a comparison of the parameterization method (LR vs. DE-MIN) and metrics for model validation ( $R^2$  vs. RMSE). The case study proved that LR (with both metrics) is suitable for smaller and homogeneous datasets. DE-MIN (with RMSD metric) is a more robust approach that can also handle the parametrization of larger and more heterogeneous datasets. The validation case study provided similar findings to the previous article – low coverage of the older parameter sets. Also, a quality validation agrees with the previous article for smaller datasets with molecules comprised of C, H, N, and O. On the other hand, the case study uncovered an interesting problem: in larger and more heterogeneous datasets – the parameters set from our previous article proved accuracy problems with some atom types. Using NEEMP, we computed parameter sets, which are also applicable for such problematic datasets.

### 5.3.1 My contribution

I prototyped the DE-Min approach and designed some new NEEMP features, such as the usage of RMSE instead of  $R^2$ . I also implemented a preparation of validation reports.

# 6

---

## Follow-up work and future plans

From these results, we were able to create a tool for the calculation of charge descriptors – ChrgDescCalc.py [96], and we also successfully prepared the universal model for  $pK_a$  prediction [97].

The EEM parameters computed in our publications [98, 99] became a part of AtomicChargeCalculator [100] and also its successor AtomicChargeCalculator II [101].

A parameterization approach from an NEEMP article [99], was further extended by my coworker J. Pazúriková in an article [102]. A further extension of the parameterization approach (optGM) was done by my colleague T. Raček and it allowed a parameterization of the majority of empirical charge calculation methods (not only EEM). An article describing optGM is now in a review process.

In the future, the development of an empirical charge calculation approach for proteins and a parameter set fully covering Protein Data Bank is in development.

## **Part IV**

# **Conclusion**

# 7

---

## Conclusion

The acid dissociation constant ( $pK_a$ ) is an important property of organic molecules. Its prediction (especially for unsynthesized molecules) is beneficial in the drug design process and other modern life science fields.

Our previous articles [19, 20] (before my dissertation) proved that  $pK_a$  is successfully predictable by chemoinformatics models based on partial atomic charges.

In my thesis, I focused on the first topic – analysis of 3D structure sources on  $pK_a$  prediction. We proved that the source of the 3D structure had a significant impact on charges and, respectively, on the quality of  $pK_a$  prediction models. The models exhibited the best performance for two databases and two software used by these databases for 3D structure generation – a database DTP NCI (where CORINA generates 3D structures) and a database PubChem (3D structures generated by Omega). Other software tools for 3D structure generation required additional MM optimization to produce acceptable or good  $pK_a$  prediction models. In this work, we also showed that  $pK_a$  prediction models had the best performance when QM or EEM charges (with specific parameter sets) were used. Purely empirical and topological charges (e.g., Gasteiger-Marseli) proved as too approximated for pKa prediction.  $pK_a$  prediction models based on EEM charges seemed very promising because they were fast (no time-demanding QM charge calculation was required), and quality was high (comparable to models based on QM charges). However, we also found that the applicability of EEM parameters for drug-like molecules (e.g., if the parameters cover all atomic types present in the molecule) was significantly limited.

It motivated us to focus on the development of EEM parameters suitable for  $pK_a$  prediction. Specifically, we prepared a new molecule dataset and successfully computed EEM parameter sets with more extensive coverage for drug-like molecules and excellent quality ( $R^2 > 0.9$ ). These newly published parameter

sets can be easily used in chemoinformatics applications such as virtual screening or QSAR/QSPR modeling. We also prepared an OpenBabel patch with these parameter sets.

During EEM parameters preparation, I realized a need to have a tool for partial charge calculation and parametrization. For this reason, I focused on the development of NEEMP software. NEEMP is the only available tool that provides EEM parametrization, validation of EEM parameters, and calculation of EEM charges. In NEEMP, we also included an improved parametrization process, including the DE-MIN method that can markedly increase the quality of final parameters for heterogeneous datasets. We published NEEMP, and in the article, we also provided two case studies demonstrating NEEMP capabilities. The publication also included new EEM parameters tailored for ligand molecules.

These articles together provide a solid base for preparing chemoinformatics workflows for  $pK_a$  prediction, including 3D structures and partial atomic charges. They sum up the current state of the art and distill the best of well-known approaches, tools, and parameters to increase the quality of the final result.

# **Part V**

# **Appendix**

---

# Bibliography

- [1] Berman, H. M., Kleywegt, G. J., Nakamura, H., and Markley, J. L. (2014) The protein data bank archive as an open data resource. *Journal of computer-aided molecular design*, **28**, 1009–1014.
- [2] Bolton, E. E., Wang, Y., Thiessen, P. A., and Bryant, S. H. (2008) Pubchem: integrated platform of small molecules and biological activities. *Annual reports in computational chemistry*, **4**, 217–241.
- [3] Irwin, J. J., Sterling, T., Mysinger, M. M., Bolstad, E. S., and Coleman, R. G. (2012) Zinc: A free tool to discover chemistry for biology. *Journal of chemical information and modeling*, **52**, 1757–1768, PMID: 22587354.
- [4] Gaulton, A., et al. (2012) Chemb: a large-scale bioactivity database for drug discovery. *Nucleic acids research*, **40**, D1100–D1107.
- [5] Paulsen, C. E., Armache, J.-P., Gao, Y., Cheng, Y., and Julius, D. (2015) Structure of the TRPA1 ion channel suggests regulatory mechanisms. *Nature*, **520**, 511–517.
- [6] Cao, E., Liao, M., Cheng, Y., and Julius, D. (2013) Trpv1 structures in distinct conformations reveal activation mechanisms. *Nature*, **504**, 113–118.
- [7] Prota, A. E., Bargsten, K., Zurwerra, D., Field, J. J., Díaz, J. F., Altmann, K.-H., and Steinmetz, M. O. (2013) Molecular mechanism of action of microtubule-stabilizing anticancer agents. *Science*, **339**, 587–590.
- [8] Lu, J., et al. (2005) Microrna expression profiles classify human cancers. *Nature*, **435**, 834–838.
- [9] Puente, X. S., et al. (2011) Whole-genome sequencing identifies recurrent mutations in chronic lymphocytic leukaemia. *Nature*, **475**, 101–105.
- [10] Nayal, M. and Honig, B. (2006) On the nature of cavities on protein surfaces: application to the identification of drug-binding sites. *Proteins*, **63**, 892–906.
- [11] Xie, L., Xie, L., and Bourne, P. (2009) A unified statistical model to support local sequence order independent similarity searching for ligand-binding sites and its application to genome-based drug discovery. *Bioinformatics*, **25**, i305–i312.
- [12] Comer, J. and Tam, K. (2001) *Pharmacokinetic Optimization in Drug Research: Biological, Physicochemical, and Computational Strategies*. Verlag Helvetica Chimica Acta, Postfach, CH-8042 Zürich, Switzerland.
- [13] Klebe, G. (2000) Recent developments in structure-based drug design. *Journal of molecular medicine*, **78**, 269–281.

- [14] Kim, J. H., Gramatica, P., Kim, M. G., Kim, D., and Tratnyek, P. G. (2007) Qsar modelling of water quality indices of alkylphenol pollutants. *SAR and QSAR in environmental research*, **18**, 729–743.
- [15] Citra, M. J. (1999) Estimating the  $pK_a$  of phenols, carboxylic acids and alcohols from semi-empirical quantum chemical methods. *Chemosphere*, **1**, 191–206.
- [16] Gross, K. C., Seybold, P. G., and Hadad, C. M. (2002) Comparison of Different Atomic Charge Schemes for Predicting pKa Variations in Substituted Anilines and Phenols. *International journal of quantum chemistry*, **90**, 445–458.
- [17] Zhang, J., Kleinöder, T., and Gasteiger, J. (2006) Prediction of pKa Values for Aliphatic Carboxylic Acids and Alcohols With Empirical Atomic Charge Descriptors. *Journal of chemical information and modeling*, **46**, 2256–2266.
- [18] Kreye, W. C. and Seybold, P. G. (2009) Correlations between quantum chemical indices and the  $pK_a$ s of a diverse set of organic phenols. *International journal of quantum chemistry*, **109**, 3679–3684.
- [19] Svobodová Vařeková, R., Geidl, S., Ionescu, C.-M., Skrehota, O., Kudera, M., Sehnal, D., Bouchal, T., Abagyan, R., Huber, H., and Koca, J. (2011) Predicting  $pK(a)$  values of substituted phenols from atomic charges: Comparison of different quantum mechanical methods and charge distribution schemes. *Journal of chemical information and modeling*, **51**, 1795–806.
- [20] Svobodová Vařeková, R., Geidl, S., Ionescu, C.-M., Ehota, O., Bouchal, T., Sehnal, D., Abagyan, R., and A, J. (2013) Predicting  $pK_a$  values from eem atomic charges. *Journal of cheminformatics*, **5**, 18.
- [21] Geidl, S. (2011),  $pK_a$  prediction based on atomic charges – *Bachelor's thesis*. URL: <https://is.muni.cz/auth/th/ya74p/?lang=en;setlang=en>.
- [22] Geidl, S. (2013), Predicting  $pK_a$  values from eem atomic charges – *Master's thesis*. URL: <https://is.muni.cz/auth/th/g67zh/?lang=en;setlang=en>.
- [23] Gasteiger, J. and Engel, T. (2006) *Chemoinformatics: a textbook*. John Wiley & Sons.
- [24] Sadowski, J. (2008) *3D Structure Generation*, pp. 231–261.
- [25] Atkins, P. and De Paula, J. (2011) *Physical chemistry for the life sciences*. Oxford University Press.
- [26] Chaves, J., Barroso, J. M., Bultinck, P., and Carbo-Dorca, R. (2006) Toward an alternative hardness kernel matrix structure in the electronegativity equalization method (eem). *Journal of chemical information and modeling*, **46**, 1657–1665.
- [27] Moller, H., Martinez-Yamout, M., Dyson, H., and Wright, P. (2005) Solution structure of the N-terminal zinc fingers of the Xenopus laevis double-stranded RNA-binding protein ZFa. *Journal of molecular biology*, **351**, 718–730.
- [28] Ghafourian, T. and Dearden, J. (2000) The Use of Atomic Charges and Orbital Energies as Hydrogen-bonding-donor Parameters for QSAR Studies: Comparison of MNDO, AM1 and PM<sub>3</sub> Methods. *Journal of pharmacy and pharmacology*, **52**, 603–610.
- [29] Galvez, J., Garcia, R., Salabert, M. T., and Soler, R. (1994) Charge Indexes. New Topological Descriptors. *Journal of chemical information and modeling*, **34**, 520–525.
- [30] Stalke, D. (2011) Meaningful structural descriptors from charge density. *Chemistry*, **17**, 9264–78.
- [31] Lyne, P. D. (2002) Structure-based virtual screening: an overview. *Drug discovery today*, **7**, 1047–1055.

- [32] Bissantz, C., Folkers, G., and Rognan, D. (2000) Protein-Based Virtual Screening of Chemical Databases. I. Evaluation of Different Docking/Scoring Combinations. *Journal of medicinal chemistry*, **43**, 4759–4767.
- [33] Cramer, C. (2003) *Essentials of computational chemistry*.
- [34] Mulliken, R. S. (1955) Electronic Population Analysis on LCAO-MO Molecular Wave Functions. II. Overlap Populations, Bond Orders, and Covalent Bond Energies. *Journal of chemical physics*, **23**, 1841.
- [35] Mulliken, R. S. (1955) Electronic Population Analysis on LCAO-MO Molecular Wave Functions. I. *Journal of chemical physics*, **23**, 1833.
- [36] Reed, A. E. and Weinhold, F. (1983) Natural bond orbital analysis of near-Hartree-Fock water dimer. *Journal of chemical physics*, **78**, 4066–4073.
- [37] Reed, A. E., Weinstock, R. B., and Weinhold, F. (1985) Natural population analysis. *Journal of chemical physics*, **83**, 735.
- [38] Löwdin, P.-O. (1950) On the Non-Orthogonality Problem Connected with the Use of Atomic Wave Functions in the Theory of Molecules and Crystals. *Journal of chemical physics*, **18**, 365.
- [39] Hirshfeld, F. L. (1977) Bonded-atom fragments for describing molecular charge densities. *Theoretica chimica acta*, **44**, 129–138.
- [40] Bader, R. F. W. (1985) Atoms in molecules. *Accounts of chemical research*, **18**, 9–15.
- [41] Bader, R. F. W. (1991) A quantum theory of molecular structure and its applications. *Chemical reviews*, **91**, 893–928.
- [42] Breneman, C. M. and Wiberg, K. B. (1990) Determining atom-centered monopoles from molecular electrostatic potentials. The need for high sampling density in formamide conformational analysis. *J. Comput. Chem.*, **11**, 361–373.
- [43] Besler, B. H., Merz, K. M., and Kollman, P. A. (1990) Atomic charges derived from semiempirical methods. *Journal of computational chemistry*, **11**, 431–439.
- [44] Bayly, C., Cieplak, P., Cornell, W., and Kollman, P. (1992) A well behaved electrostatic potential based method using charge restraints for deriving atomic charges: the resp model. *J. Phys. Chem.*, **97**, 10269–10280.
- [45] Marenich, A. V., Cramer, C. J., and Truhlar, D. G. (2009) Universal solvation model based on solute electron density and on a continuum model of the solvent defined by the bulk dielectric constant and atomic surface tensions. *Journal of physical chemistry B*, **113**, 6378–96.
- [46] Abraham, R., Griffiths, L., and Loftus, P. (2004) Approaches to charge calculations in molecular mechanics. *Journal of Computational Chemistry*, **3**, 407–416.
- [47] Gasteiger, J. and Marsili, M. (1980) Iterative partial equalization of orbital electronegativity—a rapid access to atomic charges. *Tetrahedron*, **36**, 3219–3228.
- [48] Cho, K.-H., Kang, Y. K., No, K. T., and Scheraga, H. A. (2001) A Fast Method for Calculating Geometry-Dependent Net Atomic Charges for Polypeptides. *Journal of physical chemistry B*, **105**, 3624–3634.
- [49] Oliferenko, A. A., Pisarev, S. A., Palyulin, V. A., and Zefirov, N. S. (2006) Atomic Charges via Electronegativity Equalization: Generalizations and Perspectives. *Advances in quantum chemistry*, **51**, 139–156.
- [50] Shulga, D., Oliferenko, A., Pisarev, S., Palyulin, V., and Zefirov, N. (2008) Parameterization of empirical schemes of partial atomic charge calculation for reproducing the molecular electrostatic potential. *Doklady Chemistry - DOKL CHEM*, **419**, 57–61.

- [51] Mortier, W. J., Ghosh, S. K., and Shankar, S. (1986) Electronegativity Equalization Method for the Calculation of Atomic Charges in Molecules. *Journal of the American Chemical Society*, **108**, 4315–4320.
- [52] Yang\*, Z.-Z., , and Wang, C.-S. (1997) Atom-bond electronegativity equalization method. 1. calculation of the charge distribution in large molecules. *The journal of physical chemistry A*, **101**, 6315–6321.
- [53] Rappe, A. K. and Goddard, W. A. (1991) Charge equilibration for molecular dynamics simulations. *Journal of physical chemistry*, **95**, 3358–3363.
- [54] Wilmer, C., Kim, K. C., and Snurr, R. (2012) An extended charge equilibration method. *The Journal of Physical Chemistry Letters*, **3**, 2506–2511.
- [55] Nistor, R. A., Polihirov, J. G., Müser, M. H., and Mosey, N. J. (2006) A generalization of the charge equilibration method for nonmetallic materials. *Journal of chemical physics*, **125**, 094108.
- [56] Baekelandt, B. G., Mortier, W. J., Lievens, J. L., and Schoonheydt, R. A. (1991) Probing the reactivity of different sites within a molecule or solid by direct computation of molecular sensitivities via an extension of the electronegativity equalization method. *Journal of the American Chemical Society*, **113**, 6730–6734.
- [57] Bultinck, P., Langenaeker, W., Lahorte, P., De Proft, F., Geerlings, P., Van Alsenoy, C., and Tollenaere, J. P. (2002) The Electronegativity Equalization Method II: Applicability of Different Atomic Charge Schemes. *Journal of physical chemistry A*, **106**, 7895–7901.
- [58] Bultinck, P., Vanholme, R., Popelier, P. L. A., De Proft, F., and Geerlings, P. (2004) High-speed Calculation of AIM Charges Through the Electronegativity Equalization Method. *Journal of physical chemistry A*, **108**, 10359–10366.
- [59] Svobodová Vařeková, R., Zuzanna, J., Jakub, V., Suchomel, S., and Koca, J. (2007) Electronegativity equalization method: Parameterization and validation for large sets of organic, organohalogen and organometal molecule. *International Journal of Molecular Sciences*, **8**.
- [60] Jiroušková, Z., Vařeková, R. S., Vaněk, J., and Koča, J. (2009) Electronegativity equalization method: parameterization and validation for organic molecules using the Merz-Kollman-Singh charge distribution scheme. *Journal of computational chemistry*, **30**, 1174–8.
- [61] Ouyang, Y., Ye, F., and Liang, Y. (2009) A modified electronegativity equalization method for fast and accurate calculation of atomic charges in large biological molecules. *Physical chemistry chemical physics*, **11**, 6082–9.
- [62] Todeschini, R. and Consonni, V. (2009) *Molecular Descriptors for Chemoinformatics*.
- [63] Urdan, T. (2011) *Statistics in Plain English*.
- [64] Verzani, J. (2018) *Using R for Introductory Statistics*.
- [65] Storn, R. and Price, K. (1997) Differential evolution - a simple and efficient heuristic for global optimization over continuous spaces. *Journal of Global Optimization*, **11**, 341–359.
- [66] Ishihama, Y., Nakamura, M., Miwa, T., Kajima, T., and Asakawa, N. (2002) A rapid method for  $pK_a$  determination of drugs using pressure-assisted capillary electrophoresis with photodiode array detection in drug discovery. *Journal of pharmaceutical sciences*, **91**, 933–942.
- [67] Babić, S., Horvat, A. J., Pavlović, D. M., and Kaštelan-Macan, M. (2007) Determination of  $pK_a$  values of active pharmaceutical ingredients. *TrAC*, **26**, 1043–1061.
- [68] Manallack, D. (2007) The  $pK_a$  distribution of drugs: Application to drug discovery. *Perspectives in medicinal chemistry*, **1**, 25–38.

- [69] Wan, H. and Ulander, J. (2006) High-throughput  $pK_a$  screening and prediction amenable for adme profiling. *Expert opinion on drug metabolism & toxicology*, **2**, 139–155.
- [70] Cruciani, G., Milletti, F., Storchi, L., Sforna, G., and Goracci, L. (2009) *In silico*  $pK_a$  prediction and adme profiling. *Chemistry & biodiversity*, **6**, 1812–1821.
- [71] Lee, A. C. and Crippen, G. M. (2009) Predicting  $pK_a$ . *Journal of chemical information and modeling*, **49**, 2013–2033.
- [72] Rupp, M., Körner, R., and Tetko, I. V. (2010) Predicting the  $pK_a$  of small molecules. *Combinatorial chemistry and high throughput screening*, **14**, 307–327.
- [73] Fraczkiewicz, R. (2006) *In Silico Prediction of Ionization*, vol. 5. Elsevier.
- [74] Ho, J. and Coote, M. (2010) A universal approach for continuum solvent  $pK_a$  calculations: Are we there yet? *Theoretica chimica acta*, **125**, 3–21.
- [75] Clark, J. and Perrin, D. D. (1964) Prediction of the strengths of organic bases. *Quarterly reviews of the Chemical Society*, **18**, 295–320.
- [76] Perrin, D. D., Dempsey, B., and Serjeant, E. P. (1981)  *$pK_a$  prediction for organic acids and bases*. Chapman and Hall: New York.
- [77] Software: acd/pka. URL: <https://www.acdlabs.com/products/percepta/predictors/pKa/>.
- [78] Shelley, J., Cholleti, A., Frye, L., Greenwood, J., Timlin, M., and Uchimaya, M. (2008) Epik: A software program for pka prediction and protonation state generation for drug-like molecules. *Journal of computer-aided molecular design*, **21**, 681–91.
- [79] Hilal, S., Karickhoff, S., and Carreira, L. (1995) A rigorous test for sparc's chemical reactivity models: Estimation of more than 4300 ionization pkas. *Quantitative Structure-Activity Relationships*, **14**, 348 – 355.
- [80] Sayle, R., Physiological ionization and pka prediction. URL: <http://www.daylight.com/meetings/emugoo/Sayle/pkapredict.html>.
- [81] Blower, P. E. and Cross, K. P. (2006) Decision tree methods in pharmaceutical research. *Current topics in medicinal chemistry*, **6**, 31–39.
- [82] Liptak, M. D., Gross, K. C., Seybold, P. G., Feldgus, S., and Shields, G. (2002) Absolute  $pK_a$  determinations for substituted phenols. *Journal of the American Chemical Society*, **124**, 6421–6427.
- [83] Toth, A. M., Liptak, M. D., Phillips, D. L., and Shields, G. C. (2001) Accurate relative  $pK_a$  calculations for carboxylic acids using complete basis set and gaussian-n models combined with continuum solvation methods. *Journal of chemical physics*, **114**, 4595–4606.
- [84] Software: schroinger: Jaguar. URL: <https://www.schrodinger.com/products/jaguar>.
- [85] Dixon, S. L. and Jurs, P. C. (1993) Estimation of pKa for Organic Oxyacids Using Calculated Atomic Charges. *Journal of computational chemistry*, **14**, 1460–1467.
- [86] Jelfs, S., Ertl, P., and Selzer, P. (2007) Estimation of  $pK_a$  for druglike compounds using semiempirical and information-based descriptors. *Journal of chemical information and modeling*, **47**, 450–459.
- [87] Gross, K. and Seybold, P. (2001) Substituent effects on the physical properties and pka of phenol. *International Journal of Quantum Chemistry*, **85**, 569 – 579.
- [88] Liu, S. and Pedersen, L. (2009) Estimation of molecular acidity via electrostatic potential at the nucleus and valence natural atomic orbitals. *The journal of physical chemistry. A*, **113**, 3648–55.
- [89] Nci open database compounds. Retrieved from <http://cactus.nci.nih.gov/> on August 10, 2010.

- [90] Vainio, M. J. and Johnson, M. S. (2007) Generating Conformer Ensembles Using a Multi-objective Genetic Algorithm. *Journal of chemical information and modeling*, **47**, 2462–2474.
- [91] Miteva, M. A., Guyon, F., and Tufféry, P. (2010) Frog2: Efficient 3d conformation ensemble generator for small compounds. *Nucleic acids research*, **38**, W622–W627.
- [92] O’Boyle, N., Banck, M., James, C., Morley, C., Vandermeersch, T., and Hutchison, G. (2011) Open Babel: An Open Chemical Toolbox. *Journal of cheminformatics*, **3**, 33–47.
- [93] Landrum, G., Rdkit: Open-source cheminformatics. Retrieved from <http://www.rdkit.org> on January 10, 2014.
- [94] Law, V., et al. (2014) DrugBank 4.0: shedding new light on drug metabolism. *Nucleic acids research*, **42**, D1091–7.
- [95] Zaslavski, A. and Powell, M. (2006) *The NEWUOA software for unconstrained optimization without derivatives*, pp. 255–297.
- [96] Hejret, V. (2015), Charge descriptors application in chemoinformatics – *Bachelor’s thesis*. URL: <https://is.muni.cz/auth/th/vbxsz/?lang=en;setlang=en>.
- [97] Hejret, V. (2017), Prediction of physico-chemical properties via charge descriptors – *Master’s thesis*. URL: <https://is.muni.cz/auth/th/vbxsz/?lang=en;setlang=en>.
- [98] Geidl, S., Bouchal, T., Raček, T., Svobodová Vařeková, R., Hejret, V., Křenek, A., Abagyan, R., and Koča, J. (2015) High-quality and universal empirical atomic charges for chemoinformatics applications. *Journal of Cheminformatics*, **7**.
- [99] Raček, T., Pazúriková, J., Svobodová Vařeková, R., Geidl, S., Křenek, A., Falginella, F., Horský, V., Hejret, V., and Koča, J. (2016) Neemp: Software for validation, accurate calculation and fast parameterization of eem charges. *Journal of Cheminformatics*, **8**.
- [100] Ionescu, C.-M., Sehnal, D., Falginella, F., Pant, P., Pravda, L., Bouchal, T., Svobodová Vařeková, R., Geidl, S., and Koča, J. (2015) Atomicchargecalculator: Interactive web-based calculation of atomic charges in large biomolecular complexes and drug-like molecules. *Journal of Cheminformatics*, **7**, 50.
- [101] Raček, T., Schindler, O., Toušek, D., Horský, V., Berka, K., Koča, J., and Svobodová, R. (2020) Atomic charge calculator ii: web-based tool for the calculation of partial atomic charges. *Nucleic acids research*, **48**.
- [102] Pazúriková, J., Křenek, A., and Matyska, L. (2016) Guided optimization method for fast and accurate atomic charges computation. Évora Gómez, J. and Hernández-Cabrera, J. J. (eds.), *Proceedings of the 2016 European Simulation and Modelling Conference*, Ghent, Belgicko, pp. 267–274, EUROSIS - ETI.

---

## Main papers

# How Does the Methodology of 3D Structure Preparation Influence the Quality of $pK_a$ Prediction?

Stanislav Geidl<sup>1</sup>, Radka Svobodová Vařeková<sup>1,\*</sup>, Veronika Bendová<sup>1</sup>, Lukáš Petrusek<sup>1</sup>, Crina-Maria Ionescu<sup>1</sup>, Zdeněk Jurka<sup>1</sup>, Ruben Abagyan<sup>2</sup>, Jaroslav Koča<sup>1,\*</sup>

<sup>1</sup> National Centre for Biomolecular Research, Faculty of Science and CEITEC, Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>2</sup> Skaggs School of Pharmacy and Pharmaceutical Sciences, University of California, 9500 Gilman Drive, San Diego, MC 0657, USA.

*Journal of Chemical Information and Modeling* 2015, **55**:1088–1097.

<https://doi.org/10.1021/ci500758w>



# HHS Public Access

Author manuscript

*J Chem Inf Model.* Author manuscript; available in PMC 2016 November 07.

Published in final edited form as:

*J Chem Inf Model.* 2015 June 22; 55(6): 1088–1097. doi:10.1021/ci500758w.

## How Does the Methodology of 3D Structure Preparation Influence the Quality of $pK_a$ Prediction?

Stanislav Geidl<sup>†,‡</sup>, Radka Svobodová Vašeková<sup>\*,†,‡</sup>, Veronika Bendová<sup>‡</sup>, Lukáš Petrušek<sup>‡</sup>, Crina-Maria Ionescu<sup>‡</sup>, Zdeněk Jurka<sup>¶</sup>, Ruben Abagyan<sup>§</sup>, and Jaroslav Kocába<sup>\*,‡</sup>

National Centre for Biomolecular Research, Faculty of Science and CEITEC - Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno-Bohunice, Czech Republic, Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic, and Skaggs School of Pharmacy and Pharmaceutical Sciences, University of California, 9500 Gilman Drive, MC 0657, San Diego, USA, svobodova@chemi.muni.cz; jkoca@chemi.muni.cz

### Abstract

The acid dissociation constant is an important molecular property and it can be successfully predicted by Quantitative Structure-Property Relationship (QSPR) models, even for *in silico* designed molecules. We analyzed how the methodology of *in silico* 3D structure preparation influences the quality of QSPR models. Specifically, we evaluated and compared QSPR models based on six different 3D structure sources (DTP NCI, Pubchem, Balloon, Frog2, OpenBabel and RDKit) combined with four different types of optimization. These analyses were performed for three classes of molecules (phenols, carboxylic acids, anilines) and the QSPR model descriptors were quantum mechanical (QM) and empirical partial atomic charges. Specifically, we developed 516 QSPR models and afterwards systematically analyzed the influence of the 3D structure source and other factors on their quality.

Our results confirmed that QSPR models based on partial atomic charges are able to predict  $pK_a$  with high accuracy. We also confirmed that *ab-initio* and semiempirical QM charges provide very accurate QSPR models, and using empirical charges based on electronegativity equalization is also acceptable, as well as advantageous, since their calculation is very fast. On the other hand, Gasteiger-Marsili empirical charges are not applicable for  $pK_a$  prediction. We later found that QSPR models for some classes of molecules (carboxylic acids) are less accurate. In this context, we compared the influence of different 3D structure sources. We found that an appropriate selection of 3D structure source and optimization method is essential for the successful QSPR modeling of  $pK_a$ . Specifically, the 3D structures from the DTP NCI and Pubchem databases performed the best, as they provided very accurate QSPR models for all the tested molecular classes and charge calculation approaches, and they do not require optimization. Also Frog2 performed very well. Other 3D structure sources can also be used, but are not so robust, and an

\*To whom correspondence should be addressed.

†The authors wish it to be known that, in their opinion, the first two authors should be regarded as joint First Authors.

‡NCBR & CEITEC

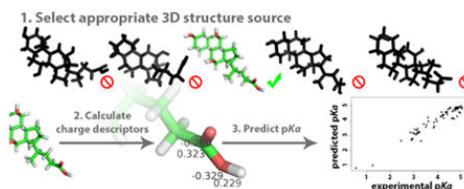
¶Faculty of Informatics

§Skaggs School of Pharmacy and Pharmaceutical Sciences

Supporting Information Available: This material is available free of charge via the Internet at <http://pubs.acs.org/>.

unfortunate combination of molecular class and charge calculation approach can produce weak QSPR models. Additionally, these 3D structures generally need optimization in order to produce good quality QSPR models.

## Graphical abstract



## Introduction

The acid dissociation constant,  $K_a$ , and its logarithmic version  $pK_a$ , is an important molecular property and its values are of interest in chemical, biological, environmental and pharmaceutical research.<sup>1–3</sup> Experimental  $pK_a$  values are usually unavailable for all compounds from the chemical catalogues. Therefore it cannot be used for example in virtual screening, which requires predictions of physico-chemical properties for large sets of *in silico* designed molecules. Several  $pK_a$  prediction methodologies have been published to date and they are summarized in review articles,<sup>4–7</sup> but reliable and accurate  $pK_a$  prediction is still a challenge and a topic of intensive research.<sup>8–10</sup>

A popular and frequently used  $pK_a$  prediction approach is based on the QSPR (Quantitative Structure-Property Relationship) methodology.<sup>11–13</sup> Various types of input values (so-called descriptors) can be used for the calculation of  $pK_a$  via QSPR models. Partial atomic charges are definitely relevant descriptors for  $pK_a$  calculations<sup>12,14–17</sup> and can be calculated directly from the 3D structure of the molecule. The partial atomic charges cannot be determined experimentally or derived from the results of quantum mechanics (QM) in a straightforward manner. For this reason, many different methods have been developed for their calculation. The most common method for charge calculation is using a quantum mechanical approach (a combination of a theory level and a basis set) and the subsequent application of a charge calculation scheme. For example for  $pK_a$  prediction via QSPR models, *ab-initio* QM charges calculated via HF or B3LYP theory levels and STO-3G or 6-31G\* basis sets proved suitable. The most appropriate charge calculation schemes for these purposes seem to be MPA (Mulliken population analysis), NPA (Natural population analysis) and AIM (atoms in molecules).<sup>8,15,17</sup> Semiempirical QM charges have also been employed in QSPR models for  $pK_a$  prediction (e.g., AM1, PM3 or PM6 theory levels in combination with MPA).<sup>11,14,17–19</sup> A major drawback of the QM charges is the computational effort required for the calculation of the wave function. For this reason, the computational complexity of obtaining QM charges is at least  $\Theta(B^4)$ , where  $B$  is the number of basis functions. Therefore, the calculation of *ab-initio* QM charges is very time consuming, while the calculation of semiempirical QM charges is also relatively slow. The Electronegativity Equalization Method<sup>20</sup> is an empirical charge calculation approach which presents a faster alternative to

the QM methods. EEM is able to provide partial atomic charges with comparable accuracy to QM charges, and it is markedly less time consuming than QM charge calculation approaches. EEM is even able to mimic a certain QM charge calculation approach (i.e., the combination of a theory level, a basis set and a charge calculation scheme), because it includes parameters based on the QM charges. EEM charges also proved applicable for  $pK_a$  prediction via QSPR.<sup>8</sup> Last but not least,  $pK_a$  predicting QSPR models based on conformationally independent empirical charges (so called topological charges, e.g., Gasteiger-Marsili charges) have also been evaluated.<sup>13,19</sup>

Therefore, in principle, we can prepare a straightforward and time-efficient workflow for obtaining  $pK_a$  values for molecules designed *in silico*: use the 3D structures of molecules prepared *in silico*, calculate partial atomic charges for them, employ the charges as descriptors in QSPR models and predict the required  $pK_a$  values. Such a workflow can be applied in virtual screening. We can also design similar workflows for other biologically important properties such as  $\log P$ , biodegradability, dioxin-like activity etc.

Nonetheless, before implementing the workflow we need to answer a key question: How does the methodology of *in silico* 3D structure preparation influence the quality of QSPR models for  $pK_a$  prediction? In previous works focused on  $pK_a$  prediction via QSPR,<sup>8,17,19,21,22</sup> 3D structures were mainly obtained from the DTP NCI database<sup>23</sup> (which uses CORINA to generate the 3D structures) or directly designed by CORINA.<sup>24</sup> But there are other tools and databases which are often used as sources of 3D structures. For example, the database Pubchem<sup>25</sup> (employing the software Omega<sup>26</sup>) or software tools such as Balloon,<sup>27</sup> Frog2,<sup>28</sup> OpenBabel<sup>29</sup> or RDKit.<sup>30</sup> These tools create 3D structures via a data or knowledge-based approach (CORINA, OpenBabel, Omega), distance geometry approach (Balloon, RDKit) or other approaches (Frog2). Specifically, Frog2 first generates a graph of rings and acyclic elements, and afterwards performs a Monte Carlo search. Can we use any of these 3D structure sources for the QSPR modeling of  $pK_a$ ? Or is it that only some methodologies for 3D structure preparation provide acceptable QSPR models? In parallel, another important question is whether the 3D structures need to be optimized before they can be used in QSPR models or not. Some articles on this topic use optimization,<sup>14,15,22,31,32</sup> while some provide accurate models even without it.<sup>8,11,17</sup>

In this study, we addressed the above questions. Specifically, we evaluated and compared QSPR models based on six different 3D structure sources combined with four different types of optimization. The 3D structure sources were the databases DTP NCI and Pubchem, and the software tools Balloon, Frog2, OpenBabel and RDKit. The optimization was either skipped or done by molecular mechanics (MMFF94 for all 3D structure sources, MM-UFF for RDKit) or quantum mechanics (B3LYP/6-31G\*). These analyses were performed for three classes of molecules (phenols, carboxylic acids, anilines). We mainly focused on *ab-initio* QM charges, which provide the most accurate  $pK_a$  predicting QSPR models, and on empirical EEM charges, which are a faster and comparably accurate alternative to *ab-initio* QM charges. Specifically, we used four types of QM charges (HF/STO-3G/MPA, B3LYP/6-31G\*/MPA, B3LYP/6-31G\*/NPA, and B3LYP/6-31G\*/AIM) and four corresponding types of EEM charges. To create a complete overview, we provide also QSPR models based on semiempirical charges (i.e., PM6 charges) and on conformationally independent

empirical charges (i.e., Gasteiger-Marsili charges). Thus we developed 516 QSPR models, and afterwards systematically analyzed the influence of the 3D structure source and other factors on their quality.

## Methods

### Data sets

Our training data set is composed of three classes of molecules (i.e., phenols, anilines and carboxylic acids), which represent common classes of organic molecules. These types of molecules are also frequently used for the evaluation of QSPR models.<sup>8,11,14–17,19,22,31</sup> The data set contains 190 molecules: 60 phenols, 82 carboxylic acids and 48 anilines. Additionally, we used a test data set containing 53 phenols which were not included in the training data set. The list of molecules including their figures, NCS numbers and CAS numbers can be found in the Supporting Information (Table S1).

### pK<sub>a</sub> values

The experimental pK<sub>a</sub> values were taken from the Physprop database.<sup>33</sup> The pK<sub>a</sub> values of all molecules can be found in the Supporting Information (Table S1).

### 2D structure of molecules

Information about the 2D structure of individual molecules was obtained from the DTP NCI database. The 2D structures were described in SMILES format. The files with the SMILES of all molecules are in the Supporting Information.

### Sources of 3D structure of molecules

For each molecule, the 3D structure was obtained from six different sources. Specifically, the structure was obtained from two databases (Pubchem, DTP NCI) and in parallel generated by four different freely available software tools (Balloon, Frog2, OpenBabel and RDKit). These sources were selected because they appear to be the most popular, and they also represent the main approaches for 3D structure preparation.

### Optimization

Each molecule was thus associated with six different 3D structures, obtained by the six approaches described above. Afterwards, each 3D structure was processed in two different ways. Specifically, two types of optimization were performed – an optimization via quantum mechanics, and an optimization via molecular mechanics (MM). The QM optimization was performed by Gaussian 09<sup>34</sup> using B3LYP/6-31G\*, and the MM optimization was done with RDKit using MMFF94. These approaches were selected because they are common and frequently used representatives of QM and MM optimization. Additionally, we also performed an optimization via the MM force field UFF (Universal Force Field) for structures prepared with RDKit. The reason is that the RDKit developers recommend applying this particular force field for the structures generated with RDKit.

### 3D structures in the training and test data sets

Each molecule in our training data set was associated with 19 different structures, because there were 6 sources of 3D structure and 3 types of optimization for each (no optimization, QM optimization and MM optimization) plus an additional UFF optimization for RDKit. The test data set contained only phenol molecules. Each molecule was associated with 2 different structures, because we selected 2 sources of 3D structure (i.e., DTP NCI and RDKit) and one type of optimization for each (no optimization).

In our QSPR models, we used neutral forms of all the molecules and also dissociated forms of phenols and carboxylic acids and associated forms of anilines (see Figure 1). The dissociated forms of molecules were created by removing the hydrogen atom of the dissociating group. The associated forms of anilines were created by adding one hydrogen atom to the amino group. The adding of the atom was done via an in-house script which applies the Bioshell library,<sup>35,36</sup> and a detailed description of the procedure is given in the Supporting information.

In this way, our training data set contained 19 ( $6 \times 3 + 1$ ) different structures for each molecule, and 7220 ( $= 19 \times 190 \times 2$ ) structures in total. In parallel, our test data set included 2 different structures for each molecule, therefore 212 ( $= 2 \times 53 \times 2$ ) structures in total.

### QM charges

For each of the 7220 structures from the training set, we calculated *ab-initio* QM partial atomic charges via 4 QM charge calculation approaches (i.e., HF/STO-3G/MPA, B3LYP/6-31G\*/MPA, B3LYP/6-31G\*/NPA, and B3LYP/6-31G\*/AIM) and semiempirical QM charges using PM6. These approaches were selected, because they represent the main types of charge calculation approaches which have been reported as successful for  $pK_a$  prediction via QSPR.<sup>8,15,17</sup> The second reason for selection of the *ab-initio* QM approaches was that corresponding EEM parameters are available for them. For each of the 212 structures from the test set, we calculated *ab-initio* QM charges via B3LYP/6-31G\*/NPA. This charge calculation approach was selected based on the results obtained on the training set. All the *ab-initio* and semiempirical QM charges were calculated by Gaussian 09.<sup>34</sup>

### EEM charges

For each of the 7220 structures in our dataset, the EEM charges were calculated by the program EEM SOLVER<sup>37</sup> using the 4 EEM parameter sets described in Table 1. EEM charges calculated using these parameter sets should mimic QM charges calculated by the relevant QM charge calculation approaches.

### Gasteiger-Marsili charges

We calculated also empirical Gasteiger-Marsili charges for all the molecules from the training set, including their dissociated or associated forms, therefore for 380 ( $= 2 \times 190$ ) molecules. Gasteiger-Marsili charges are based on 2D structure, therefore they do not depend on the source of 3D structure and on the optimization. All these charges were calculated by RD-Kit.<sup>30</sup>

### Descriptors and QSPR models

The descriptors used for QSPR modeling were partial atomic charges from atoms that are close to the dissociation or association site. We employed both charges from neutral and from dissociated (or associated) molecules. The linear model is justified by the linear relationship between  $pK_a$  and the electrostatic potential at the protonation site combined with the linear dependence of the potential on the surrounding charges. The distance dependances are absorbed by the  $p$  coefficients derived from the experimental data.

Thus, the QSPR model employed in this study for phenol molecules has the following equation:

$$pK_a = p_{p(H)} \cdot q_H + p_{p(O)} \cdot q_O + p_{p(C1)} \cdot q_{C1} + p_{p(OD)} \cdot q_{OD} + p_{p(C1D)} \cdot q_{C1D} + p_p \quad (1)$$

where  $q_H$  is the atomic charge of the hydrogen atom from the phenolic OH group of the neutral molecule,  $q_O$  is the charge on the oxygen atom from the phenolic OH group of the neutral molecule,  $q_{C1}$  is the charge on the carbon atom binding the phenolic OH group of the neutral molecule,  $q_{OD}$  is the charge on the phenoxide O<sup>-</sup> from the dissociated molecule, and  $q_{C1D}$  is the charge on the carbon atom binding this oxygen in the dissociated molecule (see Figure 1 a)). The symbols  $p_{p(H)}$ ,  $p_{p(O)}$ ,  $p_{p(C1)}$ ,  $p_{p(OD)}$ ,  $p_{p(C1D)}$  and  $p_p$  are parameters of the QSPR model.

The QSPR model employed in this study for carboxylic acids uses the following equation:

$$pK_a = p_{c(H)} \cdot q_H + p_{c(O1)} \cdot q_{O1} + p_{c(O2)} \cdot q_{O2} + p_{c(C1)} \cdot q_{C1} + p_{c(O1D)} \cdot q_{O1D} + p_{c(O2D)} \cdot q_{O2D} + p_{c(C1D)} \cdot q_{C1D} + p_c \quad (2)$$

where  $q_H$  and  $q_{O1}$  are the atomic charge of the hydrogen and oxygen atoms from the OH group of the neutral molecule, respectively;  $q_{O2}$  is the charge on the oxygen atom from the carbonyl group of the neutral molecule;  $q_{C1}$  is the charge on the carbon atom binding in the COOH group of the neutral molecule;  $q_{O1D}$  is the charge on the O<sup>-</sup> oxygen from the dissociated molecule;  $q_{O2D}$  is the charge on the oxygen atom from the carbonyl group of the dissociated molecule; and  $q_{C1D}$  is the charge on the carbon atom in the carboxyl group of the dissociated molecule (see Figure 1b)). Because the structures of dissociated carboxylic acid molecules were created by removing the H atom with no further correction of the structure, the values  $q_{O1D}$ ,  $q_{O2D}$  and  $q_{C1D}$  describe charge distribution immediately after removing of this hydrogen atom. The symbols  $p_{c(H)}$ ,  $p_{c(O1)}$ ,  $p_{c(O2)}$ ,  $p_{c(C1)}$ ,  $p_{c(O1D)}$ ,  $p_{c(O2D)}$ ,  $p_{c(C1D)}$ , and  $p_c$  are parameters of the QSPR model.

The QSPR model employed in this study for anilines is based on the following equation:

$$\text{p}K_a = p_{a(H)} \cdot q_H + p_{a(N)} \cdot q_N + p_{a(C1)} \cdot q_{C1} + p_{a(HA)} \cdot q_{HA} + p_{a(NA)} \cdot q_{NA} + p_{a(C1A)} \cdot q_{C1A} + p_a \quad (3)$$

where  $q_H$  is the average of charges located on both hydrogens in the amino group of the neutral molecule;  $q_N$  is the charge of the nitrogen from the amino group of the neutral molecule;  $q_{C1}$  is the charge on the carbon atom binding the amino group in the neutral molecule;  $q_{HA}$  is the average of charges located on the three hydrogens in the amino group of the associated molecule;  $q_{NA}$  is the charge on the nitrogen from the amino group of the associated molecule and  $q_{C1A}$  is the charge on the carbon atom binding the amino group in the associated molecule (see Figure 1 c)). The symbols  $p_{a(H)}$ ,  $p_{a(N)}$ ,  $p_{a(C1)}$ ,  $p_{a(HA)}$ ,  $p_{a(NA)}$ ,  $p_{a(C1A)}$ , and  $p_a$  are parameters of the QSPR model.

The QSPR model equations (1) and (2) originate from,<sup>8</sup> and they proved useful for  $\text{p}K_a$  prediction based on QM and EEM charges. Equation (3) was inspired by these two equations.

In this way we created one QSPR model for each of our 3 classes of molecules (phenols, carboxylic acids, anilines), 19 types of structures (6 sources of 3D structures \* 3 methods of optimization + RDKit with MM-UFF) and 9 types of charges (5 types of QM charges and 4 types of EEM charges). For each class of molecules, we additionally created one QSPR model based on Gasteiger-Marsili charges. Thus we created 516 (=3\*19\*9+3) QSPR models. Specifically, 228 QSPR models based on *ab-initio* QM charges (denoted QM QSPR models), 57 models based on semiempirical charges (denoted semiempirical QM QSPR models), 228 models based on EEM charges (denoted EEM QSPR models) and 3 models based on Gasteiger-Marsili charges (GM QSPR models). The parameterization of the QSPR models was done by multiple linear regression (MLR) using the software QSPR Designer.<sup>42</sup>

### Cross-validation

The robustness of all 516 QSPR models was tested by cross-validation. The  $k$ -fold cross-validation procedure was used,<sup>43,44</sup> where  $k = 5$ . Specifically, for each QSPR model, its training data set was divided into five parts (each contained 20% of the molecules). This division was done randomly, and included stratification by  $\text{p}K_a$  value. Afterwards, five cross-validation steps were performed. In the first step, the first part was selected as a test set, and the remaining four parts were taken together as the training set. The test and training sets for the other cross-validation steps were prepared in a similar manner.

### Results and discussion

The quality of the QSPR models, i.e. the correlation between experimental  $\text{p}K_a$  and the  $\text{p}K_a$  calculated by each model, was evaluated using the squared Pearson correlation coefficient ( $R^2$ ), root mean square error (RMSE), and average absolute  $\text{p}K_a$  error ( $\bar{s}$ ), while the statistical criteria were the standard deviation of the estimation ( $s$ ) and Fisher's statistics of the regression ( $F$ ).

Tables 2, 11 and S2 in Supporting Information summarize the squared Pearson correlation coefficients for all QSPR models based on QM charges (QM QSPR models) and for all QM QSPR models, EEM QSPR models and semiempirical QM QSPR models, respectively.

Table S3 in the Supporting Information contains all the quality criteria ( $R^2$ , RMSE,  $\bar{r}$ ) and statistical criteria ( $s$  and  $F$ ) for all the QSPR models analyzed. All these models are statistically significant at  $p = 0.01$ . Since our data sets contained 60 phenols, 82 carboxylic acids and 48 anilines, the appropriate F values to consider were those for 60 samples, 80 samples and 50 samples, respectively. The QSPR models for phenols, carboxylic acids and anilines contained 5, 7 and 6 descriptors, respectively. Thus, the QSPR models for phenols are statistically significant (at  $p = 0.01$ ) when  $F > 3.34$ , the QSPR models for carboxylic acids when  $F > 2.87$  and the QSPR models for anilines when  $F > 3.19$ .

The parameters of the QSPR models are summarized in the Supporting Information (Table S4).

### Quality of QM QSPR models – general summary

The results summarized in Tables 2 and 3 confirmed that the QSPR models based on QM charges are able to predict  $pK_a$  with high accuracy. Specifically, about 24% of the models have excellent quality ( $R^2 \geq 0.95$ ), close to 40% have very good quality ( $R^2 \geq 0.9$ ), 30% have lower quality, but are still applicable ( $R^2 \geq 0.8$ ), and only about 6% have low quality ( $R^2 < 0.8$ ).

### Predictivity of QM QSPR models

In general, the predictivity of QSPR models calculating  $pK_a$  based on charges was shown in the literature (e.g.<sup>11–13</sup>). Additionally, high quality of QM QSPR models based on the same charge descriptors as our models was shown by Svobodová Vašeková et al.<sup>17</sup> To confirm the predictivity, we did a cross-validation for all our QSPR models. Cross-validation results for selected QSPR models are in Table 4 (i.e., based on B3LYP/6-31G\*/NPA charges and non-optimized OpenBabel 3D structures, which show average quality in comparison with other QM QSPR models). All the cross-validation results can be found in the Supporting Information (Table S5). These results showed that the values of  $R^2$  are similar for the test set, the training set and the complete set, therefore the models are stable.

For further confirmation of our QSPR models predictivity, we tested selected QSPR models on an independent test data set prepared only for testing purposes, with a size comparable to that of training data set and which was. Specifically, the test data set includes 53 phenol molecules and we used it for testing two selected QM QSPR models for phenols, namely, one of the best quality models (B3LYP/6-31G\*/NPA charges and non-optimized 3D structures from NCI) and one of the worst quality models (HF/STO-3G/MPA charges and non-optimized 3D structures from RDKit). The quality criteria for the test set and the training set are in Table 5. These results demonstrate that the QSPR models perform comparably for the test set and the training set.

### Influence of *ab-initio* QM charge calculation approach

The results (Tables 2 and 6) show that all four of the *ab-initio* QM charge calculation approaches tested here provide a comparable quality of  $pK_a$  prediction. These results therefore confirmed, that all the selected charge calculation approaches are suitable for the QSPR prediction of  $pK_a$ . Additionally, all the charge calculation approaches are applicable for all three classes of molecules. Specifically, for each class of molecules, any *ab-initio* QM charge calculation approach provides good quality QSPR models ( $R^2$  close to 0.9) at least for some sources of 3D structures. An interesting finding is that the suitability of a certain charge calculation approach strongly depends on the class of molecules. For example, B3LYP/6-31G\*/MPA charges work very well for anilines and markedly poorer for carboxylic acids. The next interesting finding is that the charge calculation approach HF/STO-3G/MPA, which uses the smallest basis set (STO-3G) and the simplest population analysis (MPA), performs very well.

### Influence of the class of molecules

We can see (Table 2 and Table 7), that some classes of molecules are more easily handled by QSPR modeling, while some are more challenging. Specifically, QSPR models work very well for anilines and phenols. These models have high  $R^2$  for all charge calculation approaches and for most of the 3D structure sources. On the other hand, QSPR models provide markedly weaker  $pK_a$  predictions for carboxylic acids. Namely, only a few 3D structure sources are applicable for QSPR modeling for carboxylic acids. One reason for the lower quality of QSPR models for the carboxylic acids is, that the carboxyl group bound some arbitrary chemical scaffold. In contrast, the  $-OH$  group of phenols and  $-NH_2$  group of anilines have the same, conserved neighborhood – the phenolic ring. In parallel, the phenolic ring also allows higher de-localization of electrons, which is better suited for the calculation of QM descriptors than the more rigid electron localization in carboxylic acids.

### Influence of 3D structure preparation methodology on the quality of the QM QSPR model

Tables 2, 8 and 9 show that an appropriate selection of 3D structure source and optimization method is essential for the QSPR modeling of  $pK_a$ .

These results imply that the most appropriate 3D structures were obtained from the DTP NCI and Pubchem databases (i.e., structures prepared with the tools CORINA and Omega, respectively). The QSPR models based on these structures are very accurate, and these 3D structures do not require optimization. A great feature of these 3D structures was that they performed very well for all the tested QM charge calculation approaches and classes of molecules. An interesting finding is that the QM optimization of such 3D structures can markedly decrease the accuracy of the models.

Frog2 also seems to be applicable. QSPR models based on 3D structures from Frog2 are accurate even when the structures were not optimized, and the MM optimization of these structures mainly improves the models. They can be successfully used for all the classes of molecules and all the QM charge calculation approaches tested here.

RDKit, OpenBabel and Balloon are slightly troublesome sources of 3D structures. They can provide accurate QSPR models ( $R^2 > 0.9$ ) for some classes of molecules. In this case, the MM optimization of 3D structures improves the models. But when we process other classes of molecules (carboxylic acids), the QSPR models are weak ( $R^2 \sim 0.85$ ) for most of the charge calculation approaches. And for certain charge calculation approaches the QSPR models can even be unsatisfactory ( $R^2 < 0.7$ ). An interesting fact is that the structures generated by RDKit with no optimization provide the worst performing QSPR models of the whole study. The explanation is clear, these 3D structures are just the raw results of RDKit and, as mentioned in its manual, they need to be optimized by RDKit's internal force field UFF. This case study shows how weak QSPR models can be when based on problematic structures.

Particular geometrical properties, which are incorrectly modelled in certain 3D structure preparation methodologies and which cause worse performance of QSPR models are summarized in Supporting Information.

### Semiempirical QM QSPR models – quality, predictivity and influences

The results summarized in Table 10 and Supplementary Table S2 show that the quality of these models is comparable to the quality of QSPR models based on ab-initio QM charges, just slightly lower for phenols and anilines and slightly better for carboxylic acids. The cross-validation results (see Supplementary Table S5) confirmed the robustness of the semiempirical QM models. When we evaluated the influence of the class of molecules and the 3D structure preparation methodology, we saw the same trends as for the *ab-initio* QM QSPR models (see Table 10 and S2).

### Quality of EEM QSPR models – general summary

The results summarized in Tables 11 and 12 show that the quality of EEM QSPR models is in general lower than for QM QSPR models, but still sufficient. Specifically, about 36% of the models are very good quality ( $R^2 > 0.9$ ), most of the models are acceptable quality ( $R^2$  between 0.9 and 0.8) and only about 2% are low quality ( $R^2 < 0.8$ ). On the other hand, the number of weak models is lower than for QM QSPR models, and there are no models with ( $R^2 < 0.75$ ).

### Predictivity of EEM QSPR models

A high quality of EEM QSPR models based on the same charge descriptors as our models was shown in.<sup>8</sup> We tested the predictivity of our EEM QSPR models the same way as we did for the QM QSPR models – by cross-validation and by testing on a larger set of independent molecules. These results are summarized in Supporting Information (Table S5 and S6, respectively), and confirm that our EEM QSPR models are robust and can handle molecules outside the training set.

### Influence of EEM parameter set

The results (Table 11 and Supplementary Table S7) show that all four EEM parameter sets tested here are applicable for  $pK_a$  prediction. The quality of the QSPR models obtained by

all the EEM parameter sets is comparable. The parameter set Chaves2006 (mimicking B3LYP/6-31G\*/MPA charges) performed slightly better than the remaining sets.

### Influence of the class of molecules

As with QM charges, some classes of molecules are more challenging for the QSPR modeling of  $pK_a$  (carboxylic acids), see Table 11 and Supplementary Table S8. Nonetheless, the differences between the quality of EEM QSPR models for various classes of molecules are markedly smaller than for the QM QSPR models.

### Influence of 3D structure preparation methodology on the quality of the EEM QSPR model

Table 8 and Supplementary Table S6 show that EEM QSPR models are markedly less sensitive to the selection of 3D structure source and optimization method.

As with QM QSPR models, 3D structures from DTP NCI and Pubchem can be successfully used for all of the tested molecular classes and all EEM parameter sets, even without optimization (i.e., more than 90% of EEM QSPR models based on non-optimized NCI 3D structures and all EEM QSPR models based on non-optimized Pubchem 3D structures have  $R^2 > 0.85$ ).

Frog2 also performs very well. More than 80% of EEM QSPR models based on non-optimized Frog2 3D structures have  $R^2 > 0.85$ . Additionally, these models seem to be applicable for all molecular classes and all EEM parameter sets tested here.

For the other four tools, the accuracy of EEM QSPR models depends on the molecular class and EEM parameter set, as certain combinations of these can produce lower accuracy QSPR models.

For all six sources of 3D structures tested in this study, QM optimization produces an improvement in the EEM QSPR models in most cases.

### Quality of GM QSPR models

Gasteiger-Marsili charges does not depend on the 3D structure of molecules, therefore we prepared only one model for each class of molecules. The  $R^2$  values of these models are given in Table 13 and further quality criteria are available in Supplementary Table S3. These results show that GM QSPR models are markedly less accurate than EEM QSPR models and therefore, GM charges are not applicable for  $pK_a$  prediction. These conclusions are in agreement with results published in the past.<sup>15</sup>

## Conclusion

Our results confirmed that QSPR models based on QM and EEM partial atomic charges are able to predict  $pK_a$  with high accuracy. Specifically, more than 60% of *ab-initio* and semiempirical QM QSPR models and nearly 40% of EEM QSPR models are very good quality ( $R^2 > 0.9$ ). We also confirmed that *ab-initio* and semiempirical QM charges provide very accurate QSPR models and using EEM charges is also acceptable, and moreover advantageous because their calculation is very fast. Afterwards, we evaluated the predictivity

of our QM, semiempirical QM and EEM QSPR models via cross-validation and via testing on an independent test data set. This way, we verified that all the types of *ab-initio* and semiempirical and EEM charges used are applicable for QSPR modeling. On the contrary, QSPR models based on empirical Gasteiger-Marsili charges showed low quality, suggesting that Gasteiger-Marsili charges are not suitable descriptors for the prediction of  $pK_a$ .

We then focused on the influence of molecular class. We found that some molecular classes are more amenable to QSPR modeling (phenols and anilines), while some are more challenging (carboxylic acids).

In this context, we compared the influence of the different 3D structure sources. We found that the selection of 3D structure source and optimization method can strongly influence the quality of QSPR models for  $pK_a$  prediction. The 3D structures from the DTP NCI and Pubchem databases, i.e. structures generated by CORINA and Omega, respectively, exhibited the best performance. These 3D structures provided very accurate QSPR models for all the tested molecular classes and charge calculation approaches, and they do not require optimization. Frog2 also performed very well for all of the tested molecular classes and charge calculation approaches. Other 3D structure sources can also be used, but they are not so robust, and an unlucky combination of molecular class and charge calculation approach can lead to weak QSPR models. Additionally, these structures generally need to be optimized in order to produce high quality QSPR models. Specifically, the best approach is to apply MM optimization to 3D structures used with QM QSPR models, and QM optimization to 3D structures used with EEM QSPR models.

The main point of this article is that a workflow for the fast and accurate prediction of  $pK_a$  or other important properties for *in silico* designed molecules can be as follows: Preparation of 3D structures by CORINA or Omega (with no further optimization), calculation of EEM charges for these structures and then the EEM QSPR calculation of  $pK_a$ .

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

This work was supported by the Ministry of Education, Youth and Sports of the Czech Republic (LH13055); the European Community's Seventh Framework Programme (CZ.1.05/1.1.00/02.0068) from the European Regional Development Fund and the Capacities specific program (286154); and by the European Social Fund and the state budget of the Czech Republic (CZ.1.07/2.3.00/20.0042, CZ.1.07/2.3.00/30.0009).

This work was also supported in part by NIH grants R01 GM071872, U01 GM094612, and U54 GM094618 to R.A.. The access to MetaCentrum supercomputing facilities provided under the research intent MSM6383917201 is greatly appreciated.

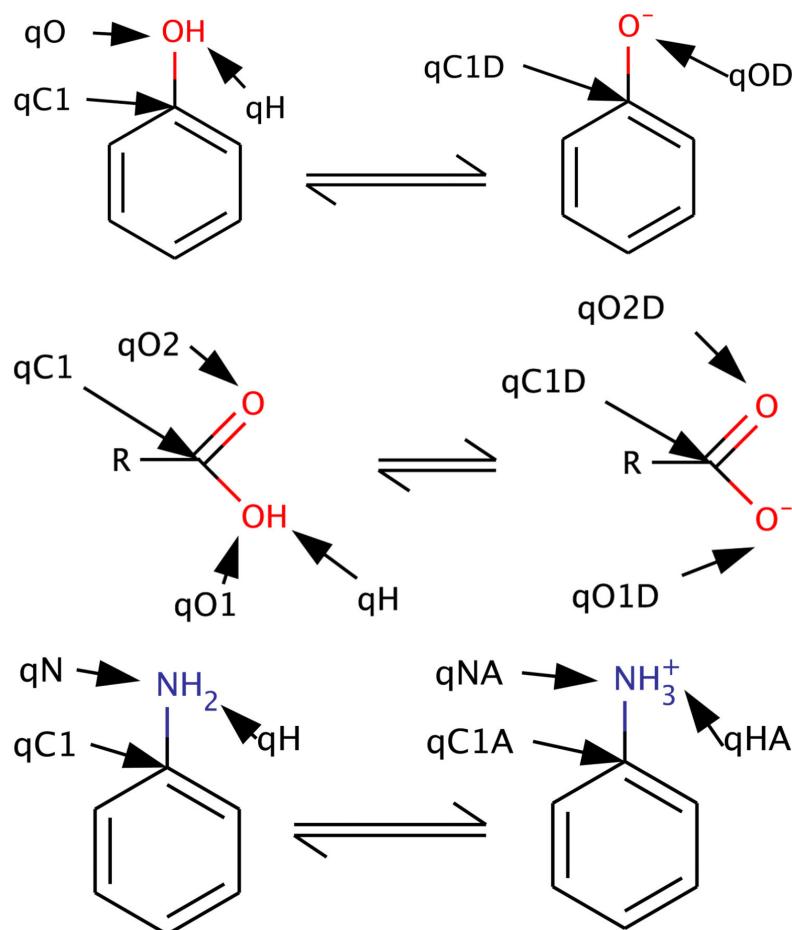
## References

1. Comer, J.; Tam, K. Pharmacokinetic Optimization in Drug Research: Biological, Physicochemical, and Computational Strategies. Verlag Helvetica Chimica Acta, Post-fach; CH-8042 Zürich, Switzerland: 2001.

2. Klebe G. Recent Developments in Structure-Based Drug Design. *J Mol Med.* 2000; 78:269–281. [PubMed: 10954199]
3. Kim JH, Gramatica P, Kim MG, Kim D, Tratnyek PG. QSAR Modelling of Water Quality Indices of Alkylphenol Pollutants. *SAR QSAR Environ Res.* 2007; 18:729–743. [PubMed: 18038370]
4. Lee AC, Crippen GM. Predicting  $pK_a$ . *J Chem Inf Model.* 2009; 49:2013–2033. [PubMed: 19702243]
5. Rupp M, Körner R, Tetko IV. Predicting the  $pK_a$  of Small Molecules. *Comb Chem High Throughput Screen.* 2010; 14:307–327.
6. Ho J. Predicting  $pK_a$  in Implicit Solvents: Current Status and Future Directions. *Aust J Chem.* 2014; 67:1441–1460.
7. Balogh GT, Tarcsey Á, Keser GM. Comparative Evaluation of  $pK_a$  Prediction Tools on a Drug Discovery Dataset. *J Pharm Biomed Anal.* 2012; 6768:63–70.
8. Svobodová Va eková R, Geidl S, Ionescu CM, Sk ehota O, Bouchal T, Sehnal D, Abagyan R, Ko a J. Predicting  $pK_a$  Values From EEM Atomic Charges. *J Cheminf.* 2013; 5:18–34.
9. Fraczkiewicz R, Lobell M, Giller AH, Krenz U, Schoenlein R, Clark RD, Hillisch A. Best of Both Worlds: Combining Pharma Data and State of the Art Modeling Technology to Improve *in silico*  $pK_a$  Prediction. *J Chem Inf Model.* 2015; 55:389–397. [PubMed: 25514239]
10. Settimi L, Bellman K, Knegtel RMA. Comparison of the Accuracy of Experimental and Predicted  $pK_a$  Values of Basic and Acidic Compounds. *Pharmaceut Res.* 2014; 31:1082–1095.
11. Jelfs S, Ertl P, Selzer P. Estimation of  $pK_a$  for Druglike Compounds Using Semiempirical and Information-Based Descriptors. *J Chem Inf Model.* 2007; 47:450–459. [PubMed: 17381168]
12. Dixon SL, Jurs PC. Estimation of  $pK_a$  for Organic Oxyacids Using Calculated Atomic Charges. *J Comput Chem.* 1993; 14:1460–1467.
13. Zhang J, Kleinöder T, Gasteiger J. Prediction of  $pK_a$  Values for Aliphatic Carboxylic Acids and Alcohols With Empirical Atomic Charge Descriptors. *J Chem Inf Model.* 2006; 46:2256–2266. [PubMed: 17125168]
14. Citra MJ. Estimating the  $pK_a$  of Phenols, Carboxylic Acids and Alcohols From Semi-empirical Quantum Chemical Methods. *Chemosphere.* 1999; 1:191–206.
15. Gross KC, Seybold PG, Hadad CM. Comparison of Different Atomic Charge Schemes for Predicting  $pK_a$  Variations in Substituted Anilines and Phenols. *Int J Quantum Chem.* 2002; 90:445–458.
16. Kreye WC, Seybold PG. Correlations Between Quantum Chemical Indices and the  $pK_{as}$  of a Diverse Set of Organic Phenols. *Int J Quantum Chem.* 2009; 109:3679–3684.
17. Svobodová Va eková R, Geidl S, Ionescu CM, Sk ehota O, Kudera M, Sehnal D, Bouchal T, Abagyan R, Huber HJ, Ko a J. Predicting  $pK_a$  Values of Substituted Phenols From Atomic Charges: Comparison of Different Quantum Mechanical Methods and Charge Distribution Schemes. *J Chem Inf Model.* 2011; 51:1795–1806. [PubMed: 21761919]
18. Rayne, S.; Forest, K.; Friesen, K. Examining the PM6 Semiempirical Method for  $pK_a$  Prediction Across a Wide Range of Oxyacids. Available from Nature Precedings. [http://hdl.handle.net/10101/npre.2009\\_2981.1](http://hdl.handle.net/10101/npre.2009_2981.1)
19. Gieleciak R, Polanski J. Modeling Robust QSAR. 2. Iterative Variable Elimination Schemes for CoMSA: Application for Modeling Benzoic Acid  $pK_a$  Values. *J Chem Inf Model.* 2007; 47:547–556. [PubMed: 17381172]
20. Mortier WJ, Ghosh SK, Shankar S. Electronegativity Equalization Method for the Calculation of Atomic Charges in Molecules. *J Am Chem Soc.* 1986; 108:4315–4320.
21. Czodrowski P, Dramburg I, Sottriffer CA, Klebe G. Development, Validation, and Application of Adapted PEOE Charges to Estimate  $pK_a$  Values of Functional Groups in Protein–Ligand Complexes. *Proteins Struct Funct Bioinf.* 2006; 65:424–437.
22. Tehan BG, Lloyd EJ, Wong MG, Pitt WR, Montana JG, Manallack DT, Garcia E. Estimation of  $pK_a$  Using Semiempirical Molecular Orbital Methods. Part 1: Application to Phenols and Carboxylic Acids. *Quant Struct-Act Relat.* 2002; 21:457–472.
23. NCI Open Database Compounds. Retrieved from <http://cactus.nci.nih.gov/> on August 10, 2010

24. Sadowski J, Gasteiger J. From Atoms and Bonds to Three-Dimensional Atomic Coordinates: Automatic Model Builders. *Chem Rev*. 1993; 93:2567–2581.
25. Bolton, EE.; Wang, Y.; Thiessen, PA.; Bryant, SH. In *Annual Reports in Computational Chemistry*. Wheeler, R.; Spellmeyer, D., editors. Vol. 4; Chapter 12. Elsevier; 2008.
26. Hawkins PCD, Skillman AG, Warren GL, Ellingson BA, Stahl MT. Conformer Generation with OMEGA: Algorithm and Validation Using High Quality Structures from the Protein Databank and Cambridge Structural Database. *J Chem Inf Model*. 2010; 50:572–584. [PubMed: 20235588]
27. Vainio MJ, Johnson MS. Generating Conformer Ensembles Using a Multiobjective Genetic Algorithm. *J Chem Inf Model*. 2007; 47:2462–2474. [PubMed: 17892278]
28. Leite TB, Gomes D, Miteva M, Chomilier J, Villoutreix B, Tuffry P. Frog: a FRee Online DruG 3D Conformation Generator. *Nucleic Acids Res*. 2007; 35:W568–W572. [PubMed: 17485475]
29. O’Boyle N, Banck M, James C, Morley C, Vandermeersch T, Hutchison G. Open Babel: An Open Chemical Toolbox. *J Cheminf*. 2011; 3:33–47.
30. Landrum, G. RDKit: Open-Source Cheminformatics. Retrieved from <http://www.rdkit.org> on January 10, 2014
31. Gross KC, Seybold PG. Substituent Effects on the Physical Properties and  $pK_a$  of Phenol. *Int J Quantum Chem*. 2001; 85:569–579.
32. Habibi-Yangjeh A, Danandeh-Jenaghara M, Nooshyar M. Application of Artificial Neural Networks for Predicting the Aqueous Acidity of Various Phenols Using QSAR. *J Mol Model*. 2006; 12:338–347. [PubMed: 16344950]
33. Howard, P.; Meylan, W. *Physical/Chemical Property Database (PHYSPROP)*. Syracuse Research Corporation, Environmental Science Center; North Syracuse NY: 1999.
34. Frisch, MJ.; Trucks, GW.; Schlegel, HB.; Scuseria, GE.; Robb, MA.; Cheeseman, JR.; Montgomery, JA., Jr.; Vreven, T.; Kudin, KN.; Burant, JC.; Millam, JM.; Iyengar, SS.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, GA.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, JE.; Hratchian, HP.; Cross, JB.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, RE.; Yazyev, O.; Austin, AJ.; Cammi, R.; Pomelli, C.; Ochterski, JW.; Ayala, PY.; Morokuma, K.; Voth, GA.; Salvador, P.; Dannenberg, JI.; Zakrzewski, VG.; Dapprich, S.; Daniels, AD.; Strain, MC.; Farkas, O.; Malick, DK.; Rabuck, AD.; Raghavachari, K.; Foresman, JB.; Ortiz, JV.; Cui, Q.; Baboul, AG.; Clifford, S.; Cioslowski, J.; Stefanov, BB.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, RL.; Fox, DJ.; Keith, T.; Al-Laham, MA.; Peng, CY.; Nanayakkara, A.; Challacombe, M.; Gill, PMW.; Johnson, B.; Chen, W.; Wong, MW.; Gonzalez, C.; Pople, JA. *Gaussian 09, Revision E.01*. Gaussian, Inc.; Wallingford, CT: 2004.
35. Gront D, Kolinski A. BioShell – a Package of Tools for Structural Biology Computations. *Bioinformatics*. 2006; 22:621–622. [PubMed: 16407320]
36. Gront D, Kolinski A. Utility Library for Structural. *Bioinformatics*. 2008; 24:584–585. [PubMed: 18227118]
37. Svobodová Va eková R, Ko a J. Optimized and Parallelized Implementation of the Electronegativity Equalization Method and the Atom-Bond Electronegativity Equalization Method. *J Comput Chem*. 2006; 3:396–405.
38. Svobodová Va eková R, Jiroušková Z, Van k J, Suchomel S, Ko a J. Electronegativity Equalization Method: Parameterization and Validation for Large Sets of Organic, Organohalogene and Organometal Molecule. *Int J Mol Sci*. 2007; 8:572–582.
39. Chaves J, Barroso JM, Bultinck P, Carbo-Dorca R. Toward an Alternative Hardness Kernel Matrix Structure in the Electronegativity Equalization Method (EEM). *J Chem Inf Model*. 2006; 46:1657–1665. [PubMed: 16859297]
40. Bultinck P, Langenaeker W, Lahorte P, De Proft F, Geerlings P, Van Alsenoy C, Tollenaere JP. The Electronegativity Equalization Method II: Applicability of Different Atomic Charge Schemes. *J Phys Chem A*. 2002; 106:7895–7901.
41. Bultinck P, Vanholme R, Popelier PLA, De Proft F, Geerlings P. High-speed Calculation of AIM Charges Through the Electronegativity Equalization Method. *J Phys Chem A*. 2004; 108:10359–10366.

42. Skáehota O, Svobodová Vaňková R, Geidl S, Kudera M, Sehnal D, Ionescu CM, Kočáka J. QSPPR Designer – a Program to Design and Evaluate QSPPR models. Case Study on  $pK_a$  Prediction. *J. Cheminf.* 2011; 3(Suppl 1):16.
43. Lemm S, Blankertz B, Dickhaus T, Müller KR. Introduction to Machine Learning for Brain Imaging. *NeuroImage*. 2011; 56:387–399. [PubMed: 21172442]
44. Katritzky AR, Lobanov VS, Karelson M. QSPPR: The Correlation and Quantitative Prediction of Chemical and Physical Properties From Structure. *Chem Soc Rev.* 1995; 24:279–287.

**Figure 1.**

a) dissociation of phenols, b) dissociation of carboxylic acids and c) association of anilines.  
The particular atomic charges used in our QSPR models are marked by their denotations.

**Table 1**

Summary information about the EEM parameter sets used in this study.

Parameter set name	QM charge calculation approach	Published by
Svob2007_chal2	HF/STO-3G/MPA	Svobodova et al. <sup>38</sup>
Chaves2006	B3LYP/6-31G*/MPA	Chaves et al. <sup>39</sup>
Bult2002_npa	B3LYP/6-31G*/NPA	Bultinck et al. <sup>40</sup>
Bult2004_aim	B3LYP/6-31G*/AIM	Bultinck et al. <sup>41</sup>

**Table 2**

$R^2$  describing the correlation between calculated and experimental  $pK_a$  for QM-QSPR models.

$R^2$	Class of molecules	Phenols				Carboxylic acids				Amines				Average
		HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	B3LYP, 6-31G*, MPA	HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	
Balloon	none	0.896	0.939	0.908	0.904	0.823	0.720	0.819	0.846	0.836	0.903	0.912	0.805	0.859
	MM	0.917	0.881	0.933	0.891	0.867	0.887	0.865	0.843	0.874	0.953	0.927	0.921	0.867
	QM	0.915	0.871	0.901	0.856	0.890	0.618	0.824	0.807	0.948	0.967	0.933	0.921	0.871
	none	0.894	0.912	0.906	0.891	0.896	0.876	0.876	0.884	0.934	0.911	0.924	0.916	0.902
	MM	0.967	0.931	0.907	0.938	0.907	0.830	0.903	0.922	0.938	0.973	0.965	0.926	0.927
	QM	0.969	0.963	0.953	0.939	0.917	0.853	0.906	0.917	0.875	0.973	0.911	0.853	0.919
Frog2	none	0.947	0.971	0.960	0.973	0.931	0.891	0.911	0.910	0.951	0.970	0.966	0.903	0.940
	MM	0.958	0.963	0.959	0.956	0.938	0.889	0.929	0.922	0.954	0.955	0.967	0.914	0.940
	QM	0.891	0.935	0.861	0.902	0.925	0.854	0.903	0.921	0.942	0.959	0.937	0.892	0.910
	none	0.955	0.961	0.957	0.963	0.869	0.658	0.845	0.876	0.952	0.973	0.966	0.930	0.909
	MM	0.961	0.965	0.959	0.961	0.863	0.665	0.841	0.875	0.958	0.975	0.967	0.927	0.910
	QM	0.955	0.957	0.956	0.936	0.845	0.674	0.804	0.827	0.874	0.974	0.928	0.880	0.884
NCI	none	0.960	0.950	0.935	0.900	0.909	0.873	0.891	0.907	0.938	0.939	0.921	0.937	0.922
	MM	0.963	0.911	0.927	0.864	0.916	0.885	0.892	0.916	0.942	0.979	0.966	0.916	0.923
	QM	0.943	0.926	0.922	0.886	0.901	0.871	0.896	0.908	0.934	0.974	0.885	0.928	0.907
	none	0.782	0.895	0.796	0.882	0.780	0.723	0.804	0.817	0.853	0.816	0.851	0.796	0.816
	MM-UFF	0.947	0.961	0.941	0.924	0.894	0.821	0.842	0.860	0.965	0.979	0.973	0.980	0.925
	MM	0.931	0.969	0.934	0.950	0.902	0.750	0.797	0.862	0.959	0.976	0.967	0.927	0.905
PubChem	QM	0.935	0.944	0.933	0.922	0.861	0.696	0.814	0.855	0.940	0.964	0.927	0.908	0.892
	Average	0.931	0.934	0.924	0.917	0.886	0.776	0.858	0.878	0.926	0.953	0.936	0.899	
	Legend	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	$R^2 < 0.7$

**Table 3**

Number and percentage of QM QSPR models with  $R^2$  higher than a defined limit.

$R^2$	0.95	(0.95, 0.9>	(0.9, 0.8>	< 0.8
Number of models	55	90	69	14
Percentage of models	24%	39%	30%	6%

$R^2$  values for cross-validation of selected QM QSPPR models.

Table 4

QSPPR model description: phenols, charges: B3LYP/6-31G <sup>g</sup> /NPA, 3D structure: OpenBabel with no optimization					
Cross-validation step	1	2	3	4	5
$R^2$ for training set	0.955	0.956	0.964	0.959	0.957
$R^2$ for test set	0.956	0.967	0.939	0.952	0.957
$R^2$ for complete set				0.957	

QSPPR model description: carboxylic acids, charges: B3LYP/6-31G <sup>g</sup> /NPA, 3D structure: OpenBabel with no optimization					
Cross-validation step	1	2	3	4	5
$R^2$ for training set	0.818	0.825	0.889	0.863	0.852
$R^2$ for test set	0.928	0.785	0.609	0.850	0.816
$R^2$ for complete set			0.845		

QSPPR model description: anilines, charges: B3LYP/6-31G <sup>g</sup> /NPA, 3D structure: OpenBabel with no optimization					
Cross-validation step	1	2	3	4	5
$R^2$ for training set	0.966	0.965	0.973	0.963	0.970
$R^2$ for test set	0.937	0.925	0.910	0.988	0.932
$R^2$ for complete set			0.966		

**Table 5**

Quality criteria for testing of selected QM QSPR models.

QSPR model description: phenols, charges: B3LYP/6-31G*/NPA, 3D structure: NCI with no optimization			
Quality criteria	$R^2$	RMSE	-
Training set	0.960	0.415	0.333
Test set	0.948	0.532	0.437

QSPR model description: phenols, charges: HF/STO-3G/MPA, 3D structure: RDKit with no optimization			
Quality criteria	$R^2$	RMSE	-
Training set	0.782	1.067	0.896
Test set	0.715	0.421	0.328

**Table 6**

Number and percentage of QM QSPR models with  $R^2$  higher than a defined limit for individual charge calculation approaches.

QM charge calculation approach	$R^2$			$\bar{R}_{\text{chrg}}^2$
	0.9	(0.9, 0.8>	< 0.8	
HF/STO-3G/MPA	67%	30%	4%	0.914
B3LYP/6-31G*/MPA	60%	25%	16%	0.888
B3LYP/6-31G*/NPA	68%	28%	4%	0.906
B3LYP/6-31G*/AIM	60%	39%	2%	0.898

Note:  $\bar{R}_{\text{chrg}}^2$  is the average value of  $R^2$  for all QSPR models, which use charges calculated by a given QM charge calculation approach.

**Table 7**

Number and percentage of QM QSPR models with  $R^2$  higher than a defined limit for individual classes of molecules.

Class of molecules	$R^2$			$\bar{R}_{\text{mol}}^2$
	0.9	(0.9, 0.8>	< 0.8	
Phenols	32%	49%	17%	0.927
Carboxylic acids	0%	29%	57%	0.849
Anilines	41%	41%	17%	0.929

Note:  $\bar{R}_{\text{mol}}^2$  is the average value of  $R^2$  for all QSPR models, which were built for a given class of molecules.

Percentage of QM QSPR models with given  $R^2$  for individual 3D structure sources.**Table 8**

Source	Optimization	$R^2$			
		0.95	(0.95, 0.9>	(0.9, 0.85>	(0.85, 0.8>
Balloon	none	0%	42%	8%	42%
	MM	8%	33%	33%	17%
QM	8%	42%	25%	17%	8%
	none	0%	50%	50%	0%
Frog2	MM	33%	58%	0%	8%
	QM	33%	42%	25%	0%
NCI	none	50%	42%	8%	0%
	MM	50%	42%	8%	0%
OpenBabel	QM	8%	58%	33%	0%
	none	58%	8%	17%	8%
PubChem	MM	58%	8%	17%	8%
	QM	33%	17%	17%	25%
RDKit	none	8%	75%	17%	0%
	MM	25%	50%	25%	0%
	QM	8%	50%	33%	8%
	none	0%	0%	33%	25%
	UFF	42%	25%	17%	17%
	MM	25%	50%	8%	0%
	QM	8%	58%	17%	8%

Note: The optimization procedures which produce the best QSPR models for each source of 3D structures are marked in bold font.

Sensitivity of a 3D structure source to a change of molecular class.

**Table 9**

Percent of insensitive QSPR models	Source				
	Balloon	Frog2	NCI	OpenBabel	PubChem
Optimization					RDKit
None	50%	100%	25%	0%	75%
MM	50%	25%	75%	0%	0%
QM	25%	50%	75%	0%	75%
UFF	-	-	-	-	25%
Total	42%	58%	58%	0%	67%
					31%

Note: The sensitivity of a particular QSPR model to a change coefficient of molecular class was analyzed via a statistical test, which compared correlation coefficient of three independent populations (i.e. molecular classes), employed Fisher's *z*-transformation and used the significance level 0.05. Detailed information about this statistical test are in Supporting Information.

**Table 10**

Number and percentage of semiempirical QM QSPR models with  $R^2$  higher than a defined limit.

$R^2$	0.95	(0.95, 0.9 >	(0.9, 0.8 >	< 0.8
Number of models	15	25	17	0
Percentage of models	26%	44%	30%	0%

**Table 11** $R^2$  describing the correlation between calculated and experimental  $pK_a$  for EEM QSPR models.

$R^2$	Class of molecules	Phenols				Carboxylic acids				Amines				Average
		HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	HF, STO-4C, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	HF, STO-3G, MPA	B3LYP, 6-31G*, NPA	B3LYP, 6-31G*, AIM	
<b>Kelton</b>	none	0.873	0.904	0.903	0.888	0.832	0.924	0.888	0.853	0.837	0.847	0.826	0.870	0.868
	MM	0.952	0.906	0.907	0.895	0.890	0.917	0.883	0.917	0.889	0.851	0.930	0.908	0.945
<b>Frog2</b>	QM	0.869	0.908	0.906	0.890	0.772	0.889	0.858	0.832	0.875	0.831	0.870	0.894	0.879
	MM	0.918	0.906	0.917	0.868	0.859	0.888	0.860	0.848	0.863	0.857	0.852	0.902	0.878
<b>NCI</b>	QM	0.921	0.907	0.918	0.869	0.841	0.898	0.866	0.874	0.939	0.926	0.907	0.939	0.900
	MM	0.906	0.906	0.899	0.890	0.875	0.926	0.891	0.879	0.870	0.852	0.839	0.882	0.884
<b>Source + Optimization</b>	QM	0.896	0.926	0.926	0.916	0.860	0.920	0.888	0.820	0.844	0.834	0.848	0.889	0.881
	MM	0.900	0.920	0.912	0.908	0.830	0.898	0.848	0.826	0.860	0.849	0.851	0.899	0.875
<b>OpenBabel</b>	MM	0.900	0.919	0.911	0.907	0.827	0.903	0.849	0.835	0.858	0.851	0.857	0.897	0.876
	QM	0.896	0.917	0.911	0.904	0.807	0.911	0.856	0.851	0.946	0.935	0.939	0.934	0.901
<b>PubChem</b>	none	0.896	0.918	0.913	0.902	0.888	0.891	0.866	0.873	0.874	0.881	0.874	0.907	0.890
	MM	0.887	0.917	0.915	0.899	0.874	0.902	0.876	0.871	0.886	0.852	0.872	0.900	0.888
<b>RDKit</b>	QM	0.898	0.921	0.925	0.899	0.825	0.923	0.894	0.892	0.890	0.905	0.867	0.927	0.897
	MM-UFF	0.923	0.917	0.912	0.895	0.801	0.919	0.866	0.844	0.838	0.845	0.843	0.875	0.873
<b>Average</b>	QM	0.909	0.919	0.916	0.895	0.753	0.915	0.881	0.851	0.933	0.892	0.869	0.923	0.888
	Legend	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>	R <sup>2</sup>

**Table 12**

Number and percentage of EEM QSPR models with  $R^2$  higher than a defined limit.

$R^2$	0.95	(0.95, 0.9>	(0.9, 0.8>	< 0.8
Number of models	82	106	38	2
Percentage of models	36%	46%	17%	1%

**Table 13**

$R^2$  describing the correlation between calculated and experimental  $pK_a$  for GM QSPR models.

Class of molecules	Phenols	Carboxylic acids	Anilines
$R^2$	0.747	0.737	0.870

# **High-quality and universal empirical atomic charges for chemoinformatics applications**

Stanislav Geidl<sup>1</sup>, Tomáš Bouchal<sup>1</sup>, Tomáš Raček<sup>1,2</sup>, Radka Svobodová  
Vařeková<sup>1,\*</sup>, Václav Hejret<sup>1</sup>, Aleš Křenek<sup>3</sup>, Ruben Abagyan<sup>4</sup>, Jaroslav Koča<sup>1,\*</sup>

<sup>1</sup> National Centre for Biomolecular Research, Faculty of Science and CEITEC,  
Central European Institute of Technology, Masaryk University Brno, Kamenice  
5, 625 00 Brno, Czech Republic.

<sup>2</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00  
Brno, Czech Republic.

<sup>3</sup> Institute of Computer Science, Masaryk University Brno, Botanická 68a, 602  
00 Brno, Czech Republic.

<sup>4</sup> Skaggs School of Pharmacy and Pharmaceutical Sciences, University of  
California, 9500 Gilman Drive, San Diego, MC 0657, USA.

*Journal of Cheminformatics* 2015, **7**:59.

<https://doi.org/10.1186/s13321-015-0107-1>

RESEARCH ARTICLE

Open Access



# High-quality and universal empirical atomic charges for chemoinformatics applications

Stanislav Geidl<sup>1†</sup>, Tomáš Bouchal<sup>1†</sup>, Tomáš Raček<sup>1,2†</sup>, Radka Svobodová Vařeková<sup>1\*</sup>, Václav Hejret<sup>1</sup>, Aleš Křenek<sup>3</sup>, Ruben Abagyan<sup>4</sup> and Jaroslav Koča<sup>1\*</sup>

## Abstract

**Background:** Partial atomic charges describe the distribution of electron density in a molecule and therefore provide clues to the chemical behaviour of molecules. Recently, these charges have become popular in chemoinformatics, as they are informative descriptors that can be utilised in pharmacophore design, virtual screening, similarity searches etc. Especially conformationally-dependent charges perform very successfully. In particular, their fast and accurate calculation via the Electronegativity Equalization Method (EEM) seems very promising for chemoinformatics applications. Unfortunately, published EEM parameter sets include only parameters for basic atom types and they often miss parameters for halogens, phosphorus, sulphur, triple bonded carbon etc. Therefore their applicability for drug-like molecules is limited.

**Results:** We have prepared six EEM parameter sets which enable the user to calculate EEM charges in a quality comparable to quantum mechanics (QM) charges based on the most common charge calculation schemes (i.e., MPA, NPA and AIM) and a robust QM approach (HF/6-311G, B3LYP/6-311G). The calculated EEM parameters exhibited very good quality on a training set ( $R^2 > 0.9$ ) and also on a test set ( $R^2 > 0.93$ ). They are applicable for at least 95 % of molecules in key drug databases (DrugBank, ChEMBL, Pubchem and ZINC) compared to less than 60 % of the molecules from these databases for which currently used EEM parameters are applicable.

**Conclusions:** We developed EEM parameters enabling the fast calculation of high-quality partial atomic charges for almost all drug-like molecules. In parallel, we provide a software solution for their easy computation ([http://ncbr.muni.cz/eem\\_parameters](http://ncbr.muni.cz/eem_parameters)). It enables the direct application of EEM in chemoinformatics.

**Keywords:** Partial atomic charges, Electronegativity Equalization Method, EEM, Quantum mechanics, QM, Drug-like molecules

## Background

Partial atomic charges are real numbers describing the distribution of electron density in a molecule, thus providing clues as to the chemical behaviour of molecules. The concept of charges began to be used in physical

chemistry and organic chemistry. Afterwards, partial atomic charges were adopted by computational chemistry and molecular modelling, where they serve for calculating electrostatic interactions, describe the reactivity of the molecule etc. Specifically, they are applied in molecular dynamics, docking, conformational searches, binding site predictions etc. Recently, partial atomic charges also became popular in chemoinformatics, as they proved to be informative descriptors for QSAR and QSPR modelling [1–9] and for other applications [10–12]; they can be utilised in pharmacophore design [13–15], virtual

\*Correspondence: radka.svobodova@ceitec.muni.cz;  
jkoca@chemi.muni.cz

†Stanislav Geidl, Tomáš Bouchal and Tomáš Raček are joint first authors

<sup>1</sup>National Centre for Biomolecular Research, Faculty of Science and CEITEC, Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic

Full list of author information is available at the end of the article

screening [16–18], similarity searches [19–21], molecular structure comparison [22–24] etc.

The partial atomic charges cannot be determined experimentally or derived straightforwardly from the results of quantum mechanics (QM), and many different methods have been developed for their calculation. The most common method for charge calculation is an application of the QM approach and afterwards the utilisation of a charge calculation scheme. Charge calculation schemes can be based on orbital-based population analysis, on wave-function-dependent physical observables or on reproducing charge-dependent observables. Examples of orbital-based population analyses are Mulliken population analysis (MPA) [25, 26], Löwdin population analysis [27] and Natural population analysis (NPA) [28, 29]. Wave-function-dependent physical observables are used in the atoms-in-molecules (AIM) approach [30, 31], Hirshfeld population analysis [32–34], CHELPG [35] and Merz-Singh-Kollman (MK) [36, 37] method. The reproduction of charge-dependent observables is applied in the CM1, CM2, CM3, CM4, and CM5 approaches [38, 39].

Unfortunately, QM charge calculation approaches are very time-consuming. A markedly faster alternative is to employ empirical charge calculation approaches, which can also provide high-quality charges. These approaches can be divided into conformationally-independent, which are based on 2D structure (e.g., Gasteiger's and Marsili's PEOE [40, 41], GDAC [42], KCM [43], DENR [44]) and conformationally-dependent, calculated from 3D structure (e.g., EEM [45], QEeq [46] or SQE [47, 48]). We would like to highlight that conformationally-dependent charges are considered to be more suitable for chemoinformatics applications [1–3, 7, 12, 20]. The reason is that these charges contain extensive information not only about chemical surrounding of atoms, i.e., its topology (2D structure based charges) but also geometry and “chemical quality” of the surrounding. Such information is missing, for example, in force field charges which use averaged atomic charges from large sets of structures. Therefore we only focus on conformationally-dependent atomic charges.

Electronegativity equalization method (EEM) is the most frequently used conformationally-dependent empirical charge calculation approach. It calculates charges using the following system of linear equations:

$$\begin{pmatrix} B_1 & \frac{\kappa}{R_{1,2}} & \cdots & \frac{\kappa}{R_{1,N}} & -1 \\ \frac{\kappa}{R_{2,1}} & B_2 & \cdots & \frac{\kappa}{R_{2,N}} & -1 \\ \vdots & \vdots & \ddots & \vdots & \vdots \\ \frac{\kappa}{R_{N,1}} & \frac{\kappa}{R_{N,2}} & \cdots & B_N & -1 \\ 1 & 1 & \cdots & 1 & 0 \end{pmatrix} \cdot \begin{pmatrix} q_1 \\ q_2 \\ \vdots \\ q_N \\ \bar{\chi} \end{pmatrix} = \begin{pmatrix} -A_1 \\ -A_2 \\ \vdots \\ -A_N \\ Q \end{pmatrix} \quad (1)$$

where  $q_i$  is the charge of an atom  $i$ ;  $R_{i,j}$  is the distance between atoms  $i$  and  $j$ ;  $Q$  is the total charge of the molecule;  $N$  is the number of atoms in the molecule;  $\kappa$  is the molecular electronegativity, and  $A_i$ ,  $B_i$  and  $\kappa$  are empirical parameters. The parameters  $A_i$  and  $B_i$  vary for individual atom types, where atom type is a combination of element type and maximal bond order of the atom  $i$ . For example, atom type C2 means that the atom is carbon and it creates at least one double bond with its neighbors. An atom X in the aromatic ring is therefore also included into X2 atom type. The parameters  $A_i$ ,  $B_i$  and  $\kappa$  are molecule independent and they are calculated from QM atomic charges by a process of EEM parameterization [49]. EEM is not only a fast charge calculation approach, but it can also provide highly accurate charges, i.e., they can mimic the QM charges for which EEM has been parameterized. On the other hand, EEM charges can be outperformed in certain situations. Specifically, QEeq showed better agreement with experimental dipole moments [46] and SQE is presented as an extension of the EEM to obtain the correct size-dependence of the molecular polarizability [47]. But this drawback is compensated by a fact that the quality of EEM charges was documented by many successful applications [2, 3, 50–55] and they are clearly the most cited empirical conformationally-dependent charges.

Therefore, many EEM parameter sets for various QM charge calculation approaches were published later or recently (see Table 1). In parallel, a few freely available software tools also include an EEM charge calculation method (see Table 2).

EEM recently began to be also used in chemoinformatics, giving very promising results [1–3, 64, 65]. Because of their rapid calculation, they can be easily computed for large sets of molecules (e.g., drug-like compounds). Unfortunately, a broader utilisation of EEM charges in chemoinformatics is now limited by the fact that available EEM parameter sets can only cover part of common organic molecules, as they only contain the parameters for some elements and certain bond orders (Table 1). For the above reasons, our aim with this work is to provide EEM parameter sets that cover most of the drug-like molecules and with accuracy comparable to QM charges. Specifically, we have parameterized EEM for frequently used charge calculation schemes, high enough QM theory levels and a large basis set. Afterwards, we compared the coverage and quality of our EEM parameter sets with previously published EEM parameter sets (see Table 1) and with EEM parameter sets embedded in software tools (see Table 2). Additionally, we have prepared a software solution, enabling the user to easily calculate EEM charges via our EEM parameters.

**Table 1** Summary information about published EEM parameters evaluated in this study

QM theory Level + basis set	Charge calc. scheme	EEM parameter set name	Published by	Elements and bond orders included <sup>†</sup>
HF/STO-3G	MPA	Baek1991	Baekelandt et al. [56]	C, O, N, H, P, Al, Si
		Svob2007_cbeg2	Svobodova et al. [49]	C1, C2, O, N1, N2, H, S1
		Svob2007_cmet2	Svobodova et al. [49]	C1, C2, O, N1, N2, H, S1, Fe, Zn
		Svob2007_chal2	Svobodova et al. [49]	C1, C2, O, N1, N2, H, S1, Br, Cl, F, I
		Svob2007_hm2	Svobodova et al. [49]	C1, C2, O, N1, N2, H, S1, F, Cl, Br, I, Fe, Zn
HF/6-31G*	MK	Jir2008_hf	Jirouskova et al. [57]	C1, C2, O, N1, N2, H, S1, F, Cl, Br, Zn
	MPA	Bult2002_mpa	Bultinck et al. [58]	C, O, N, H, F
B3LYP/6-31G*		Bult2002_npa	Bultinck et al. [58]	C, O, N, H, F
NPA	Ouy2009 <sup>‡</sup>	Ouyang et al. [59]	C, O, N, H	
	Ouy2009_elem	Ouyang et al. [59]	C, O, N, H	
	Hir.	Bultinck et al. [58]	C, O, N, H, F	
MK	Bult2002_mk	Bultinck et al. [58]	C, O, N, H, F	
CHELPG	CHELPG	Jir2008_mk	Jirouskova et al. [57]	C1, C2, O, N1, N2, H, S1, F, Cl, Br, Zn
		Bult2002_che	Bultinck et al. [58]	C, O, N, H, F
		AIM	Bultinck et al. [60]	C, O, N, H, F

<sup>†</sup> An element symbol with no further information (e.g., C) means that the EEM parameters are available for this element bound by all possible bond orders. The element symbol followed by a number (e.g., C1) means that the EEM parameters are only available for this element bound by a bond with an order described using this number

<sup>‡</sup> For this parameter set, C1 represents  $sp^3$  hybridization, C2  $sp^2$  hybridization, C3 sp hybridization, etc.

**Table 2** Information about freely available software tools enabling EEM charge calculation

Software	EEM parameters used by a software
OpenBabel [61]	It contains the embedded EEM parameter set Bult2002_mpa, which was parameterized for B3LYP/6-31G*/MPA charges. It does not allow any other EEM parameter set to be used
Balloon [23]	It contains an embedded EEM parameter set published by Puranen et al. [62], which was calculated by fitting to the MEP field. Balloon's developers claim that the EEM charges calculated via Balloon should be comparable to B3LYP/cc-pVTZ/MPA. It does not allow any other EEM parameter set to be used
EEM SOLVER [63]	It allows the use of any input EEM parameter sets provided by the user. It does not contain any embedded EEM parameter sets

## Methods

### EEM parameterization (step 1)

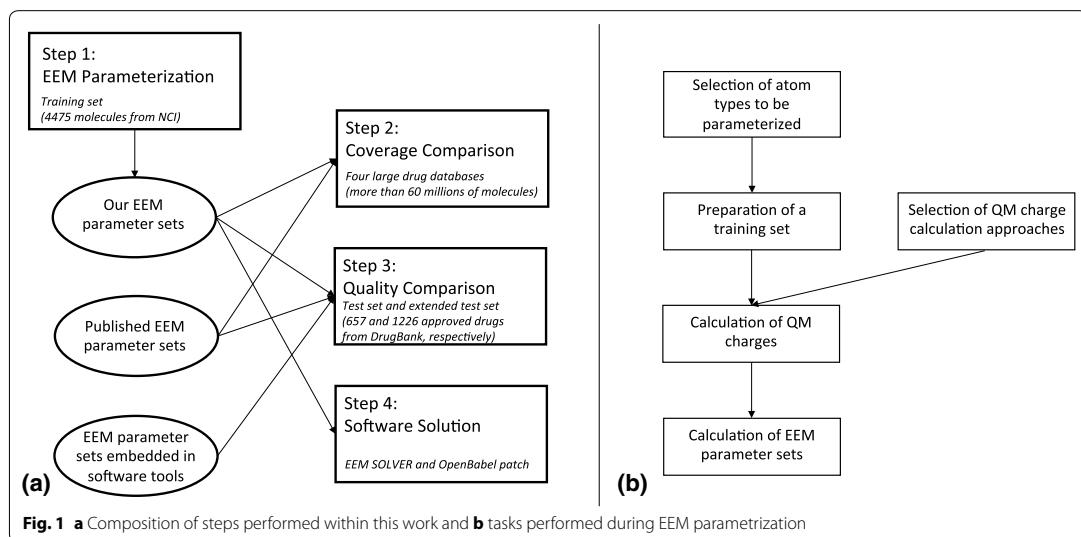
All the steps performed during our work are depicted in Fig. 1a. The most challenging part of our work was the EEM parameterization. This step required several tasks (see Fig. 1b) and the quality of the calculated EEM parameters sets depends on the proper accomplishment of all these tasks.

### EEM parameterization: selection of atom types to be parameterized

Our goal is to provide EEM parameter sets applicable for most common drug-like molecules. Therefore, we provide EEM parameters for the majority of atom types occurring in these molecules. These atom types are summarized in Table 3 (columns 1–3).

### EEM parameterization: preparation of the training set

Our training set contains the 3D structures of 4475 distinct small organic molecules. The molecules were obtained from the DTP NCI database [66] and their 3D structures were generated with CORINA 3.60 [67], without any further geometry optimization. The DTP NCI database collects compounds tested as anticancer drugs (with positive or negative results), therefore it is a database of common drug-like molecules. The training set was created in such a way that each selected atom type is contained in at least 100 molecules. The occurrences of individual atom types in the training set are summarized in Table 3. The list of training set molecules, including their NSC numbers and summary formulas, can be found in (Additional file 1: Table S1).

**Table 3 Occurrence of atom types in the training set**

Denotation of atom type	Element symbol	Maximal bond order	Number of atoms with this atom type in the training set	Number of molecules containing this atom type in the training set
H1	H	1	57,119	4442
C1	C	1	15,220	3447
C2		2	38,097	4149
C3		3	345	266
N1	N	1	4151	2483
N2		2	3383	1879
N3		3	345	266
O1	O	1	5016	2525
O2		2	5793	3069
F1	F	1	938	395
P1	P	1	153	143
P2		2	251	213
S1	S	1	1034	770
S2		2	1391	1211
Cl1	Cl	1	1084	676
Br1	Br	1	336	261
I1	I	1	1734	1365
Total	-	-	136,390	4475

#### EEM parameterization: selection of QM charge calculation approach

We performed the EEM parameterization for two QM theory levels (B3LYP and HF), one basis set (6-311G) and three charge calculation schemes (MPA, NPA and AIM). We provide the EEM parameters for all combinations of these theory levels, the basis sets and the charge

calculation schemes (see Table 4). Theory levels HF and B3LYP were selected, because they are very often used for QM charge calculation and were also successfully used for EEM parameterization several times [49, 56–60]. The basis set 6-311G was used, because it is robust, also covers iodine and moreover, Pople basis sets are very suitable for EEM parameterization. MPA and NPA

**Table 4 Quality criteria of our EEM parameter sets**

EEM parameter set name	Relevant QM charges	$R^2$	RMSD	$\bar{\Delta}$
Cheminf_b3lyp_mpa	B3LYP/6-311G/MPA	0.9007	0.1038	0.0727
Cheminf_b3lyp_npa	B3LYP/6-311G/NPA	0.9651	0.0746	0.0540
Cheminf_b3lyp_aim	B3LYP/6-311G/AIM	0.9499	0.0785	0.0558
Cheminf_hf_mpa	HF/6-311G/MPA	0.9178	0.1125	0.0776
Cheminf_hf_npa	HF/6-311G/NPA	0.9633	0.0805	0.0574
Cheminf_hf_aim	HF/6-311G/AIM	0.9441	0.0919	0.0651

**Table 5 Size of database, used for comparison of EEM parameter set coverages**

Database	Number of compounds
DrugBank	6874
ChEMBL	1,456,020
PubChem	63,676,639
ZINC	21,957,378

population analyses were employed, because they are the most known charge calculation schemes and additionally, EEM is able to mimic MPA and NPA charges very successfully [49, 58, 59]. AIM was selected, because it is based on a different principle from the other two, and EEM can also mimic AIM charges very efficiently [60]. Note that we do not provide EEM parameters for ESP and RESP charges, because it is known that EEM does not mimic these charges well [2, 58].

#### EEM parameterization: calculation of QM charges

For each molecule from the training set, six sets of QM charges were calculated via the above-mentioned six QM charge calculation approaches. The calculations of QM charges were carried out using Gaussian09 [68]. With the AIM population analysis, the output from Gaussian03 was further processed with the software package AIMAll [69].

#### EEM parameterization: calculation of EEM parameter sets

For each set of QM charges, the EEM parameterization was performed and the values of the parameters are provided in (Additional file 2: EEM parameters). The software NEEMP [70] was used for the parameterization. This software implements the parameterization methodology described by [49] and introduces several marked improvements into it. NEEMP provides EEM parameter sets together with their quality criteria, i.e., squared Pearson correlation coefficient ( $R^2$ ), root mean square deviation (RMSD), and average absolute error ( $\bar{\Delta}$ ), calculated via Eqs. (2), (3) and (4), respectively

$$R^2 = \frac{\left( \sum_{i=1}^N (q_i^{EEM} - \bar{q}^{EEM})(q_i^{QM} - \bar{q}^{QM}) \right)^2}{\sum_{i=1}^N (q_i^{EEM} - \bar{q}^{EEM})^2 \sum_{i=1}^N (q_i^{QM} - \bar{q}^{QM})^2} \quad (2)$$

$$\text{RMSD} = \sqrt{\frac{\sum_{i=1}^N (q_i^{EEM} - q_i^{QM})^2}{N}} \quad (3)$$

$$\bar{\Delta} = \frac{\sum_{i=1}^N |q_i^{EEM} - q_i^{QM}|}{N} \quad (4)$$

where  $q_i^{EEM}$  is the EEM charge of an atom  $i$ ;  $q_i^{QM}$  is the QM charge of an atom  $i$ ;  $\bar{q}^{EEM}$  is an average of all EEM charges;  $\bar{q}^{QM}$  is an average of all QM charges,  $N$  is the number of atoms in the molecule.

#### Coverage comparison (step 2)

For comparison, we used our six EEM parameter sets and 15 published EEM parameter sets, described in Table 1 (all 21 of these EEM parameter sets will be below referred to as the tested EEM parameter sets). The coverage comparison was done on four very well-known databases of drug-like chemical compounds: DrugBank [71, 72], ChEMBL [73], PubChem [74], and ZINC [75]. The number of compounds in all these databases (from 10<sup>th</sup> February 2015) are summarized in Table 5. For each tested EEM parameter set, we analysed how many compounds from the four databases can be covered by them (i.e., contains only atom types present in the tested EEM parameter sets). This coverage analysis was done using NEEMP.

#### Quality comparison (step 3)

This evaluation was done for the 21 above-mentioned tested EEM parameter sets and was performed on two data sets—a test set (657 molecules) and an extended test set (1226 molecules). The extended test set contained all approved drugs (i.e., drugs which have received approval in at least one country) from the DrugBank database (downloaded 10<sup>th</sup> February 2015), for which it was possible to calculate all QM charges necessary for testing. The test set was a subset of the extended test set, which contained only molecules covered by all the tested EEM parameter sets. The 2D structures of all molecules were obtained from DrugBank. The lists of molecules from the test set and the extended test set, including their DrugBank IDs and summary formulas, can be found in (Additional file 3: Table S2a; Additional file 4: Table S2b, respectively). The 3D structures of all the molecules were

generated with CORINA 2.6 [67], without any further geometry optimization. For all the molecules, we calculated all the types of QM charges which corresponded to the tested EEM parameters. This means we used the 8 QM charge calculation approaches mentioned in Table 1 and the six QM charge calculation approaches employed for calculating our EEM parameter sets. The calculations of QM charges were done with Gaussian09 and the AIMAll software package was used for AIM charges. We compared the quality of the tested EEM parameter set on both the test set and the extended test set. The comparison was done using NEEMP, which provided quality criteria for all the tested EEM parameter sets. In the extended test set, some molecules were not covered by certain EEM parameter set(s). Therefore, we calculated quality criteria based purely on the covered molecules and in parallel, we also computed the coverage.

#### **Quality comparison: EEM parameter sets embedded in software tools**

The calculation of EEM charges can be done with a few software tools, e.g., EEM SOLVER, OpenBabel or Balloon. The software tools OpenBabel and Balloon contain embedded EEM parameter sets (see Table 2). Therefore, we also evaluated the quality of these embedded EEM parameter sets. This evaluation was done for the same data sets and via the same procedure as with the tested EEM parameter sets. The only difference was that the EEM charges were not calculated with NEEMP, but with OpenBabel and Balloon. Afterwards, these EEM charges were compared with the relevant QM charges using R statistical software [76], which provided their quality criteria.

#### **Software solution (step 4)**

We provide the user two such solutions, the first based on EEM SOLVER and the second on OpenBabel.

### **Results and discussion**

#### **EEM parameterization (step 1)**

EEM parameterization was performed for six QM charge calculation approaches, and a training set containing 4475 drug-like molecules was used. Squared Pearson correlation coefficient ( $R^2$ ), root mean square deviation (RMSD) and average absolute error ( $\bar{\Delta}$ ) of the obtained EEM parameter sets, calculated for the training set, are summarized in Table 4. These quality criteria describe the correlation between QM charges and the corresponding EEM charges and they were calculated using NEEMP software.

These results show that the quality of our EEM parameter sets is very high, i.e., all the  $R^2$  values are higher or equal to 0.9. Table 4 also illustrates that QM theory levels

B3LYP and HF are both applicable for EEM parameterization, and EEM charges based on them have similar accuracy. From this table, we can also see that the quality of EEM parameters based on NPA and AIM population analysis is slightly better than for MPA.

#### **Coverage comparison (step 2)**

Information about the coverages of published EEM parameter sets and our EEM parameter sets are summarized in Table 6. The coverages were computed on four well-known databases of drug-like molecules—DrugBank, ChEMBL, PubChem and ZINC. Table 6 shows that the coverages of the published EEM parameter sets are low (<60%). The only exception are the EEM parameter sets published by Svobodova et al. and Jirouskova et al., which have coverage between 70 and 80%. In contrast, our EEM parameter sets have very high coverage—about 95% or more for all the databases. The not covered molecules include atom types rare for drug-like molecules, e.g., metals or boron. An interesting fact is that the coverages are very similar for all four analyzed databases. Therefore, low EEM parameter set coverage is not merely an isolated issue related to one database, but a general problem.

#### **Quality comparison (step 3)**

Table 6 summarizes the main quality criteria (i.e.,  $R^2$  values) of all tested EEM parameter sets for the test set, which contained 657 approved drugs from DrugBank. Other quality criteria (RMSD and  $\bar{\Delta}$ ) can be found in (Additional file 5: Table S3) and all values of partial atomic charges (represented as tables and as graphs) are in (Additional file 6). The table shows that our EEM parameter sets are among the best performing EEM parameter sets to have been published so far. The table also illustrates that the quality of EEM parameters is strongly influenced by the selection of QM charge calculation scheme. Specifically, EEM parameters based on MPA, NPA and AIM charges are very high quality, and EEM parameters based on Hirshfeld charges are still acceptable. EEM parameters based on MK and CHELPG charges are very low quality, which is in agreement with published data [2, 58]. Both theory levels (HF and B3LYP) and all three basis sets used (STO-3G, 6-31G\* and 6-311G) are applicable for EEM parameterization. These results also confirm that our selection of QM theory level, basis set and charge calculation schemes is appropriate.

For the extended test set, the quality criteria exhibit similar trends (see Additional file 7: Table S4). In parallel, the coverages for this data set are slightly higher than for the complete DrugBank database. An interesting fact is that even for such common compounds as approved drugs, the

**Table 6** Summary information about coverage and quality of all tested EEM parameters (see below for meaning of colours)

Relevant QM charges		EEM parameter set name	Coverage comparison				Quality comparison		
QM theory level + basis set	Charge calc. scheme		Coverage [%]						
			DrugBank	ChEMBL	PubChem	ZINC			
HF/STO-3G	MPA	Back1991	58.1	42.3	40.5	40.1	0.8981		
		Svob2007_cbeg2	55.0	49.5	47.3	51.9	0.9758		
		Svob2007_chal2	71.7	75.2	77.2	80.2	0.9668		
		Svob2007_chm2	72.2	75.2	77.3	80.2	0.9623		
		Svob2007_cmet2	55.5	49.5	47.3	51.9	0.9676		
HF/6-31G*	MK	Jir2008_hf	70.8	74.7	76.5	79.8	0.6872		
B3LYP/6-31G*	MPA	Bult2002_mpa	55.4	49.4	48.2	49.6	0.9658		
		Bult2002_npa	55.4	49.4	48.2	49.6	0.8131		
	NPA	Ouy2009	49.0	41.1	39.1	40.0	0.9655		
		Ouy2009_elem	50.0	41.2	39.1	40.0	0.9633		
	Hirshfeld	Bult2002_hir	55.4	49.4	48.2	49.6	0.9061		
	MK	Bult2002_mk	55.4	49.4	48.2	49.6	0.7844		
		Jir2008_mk	70.8	74.7	76.5	79.8	0.7022		
	CHELPG	Bult2002_che	55.4	49.4	48.2	49.6	0.7803		
	AIM	Bult2004_aim	55.4	49.4	48.2	49.6	0.9739		
HF/6-311G	MPA	Cheminf_hf_mpa	94.6	95.7	96.9	100.0	0.9606		
		Cheminf_hf_npa					0.9713		
		Cheminf_hf_aim					0.9791		
B3LYP/6-311G	MPA	Cheminf_b3lyp_mpa					0.9552		
		Cheminf_b3lyp_npa					0.9695		
		Cheminf_b3lyp_aim					0.9800		

Coverage	> 90%	> 80%	> 70%	> 60%	< 60%
R <sup>2</sup>	> 0.95	> 0.9	> 0.85	> 0.8	< 0.8

coverages of published EEM parameter sets are low. Specifically, most published EEM parameter sets have coverages between 55 and 65 %. Further remarkable fact is that quality criteria of our EEM parameters are better for the test set than for the training set. The reason is that the training set is much larger and heterogeneous than the test set.

#### Quality comparison: EEM parameter sets embedded in software tools

EEM charges produced with OpenBabel were compared with QM charges calculated with B3LYP/6-31G\*/MPA. The quality criteria for the test set were the same as for the EEM parameters Bult2002\_mpa (i.e.,  $R^2$  about 0.97). This was expected, because OpenBabel uses Bult2002\_mpa as its embedded EEM parameters. Very surprising was the behavior of OpenBabel on the extended set. The coverage was 100 %, but the quality criteria were markedly lower (e.g.,  $R^2$  about 0.82). The reason for this is that

OpenBabel replaces the EEM parameters for atom types which are not provided in Bult2002\_mpa with the EEM parameters for some other atom types. Unfortunately, this approach is not very reliable, i.e., the quality criteria for molecules which are in the extended test set but are not in the test set are very low ( $R^2 = 0.66$ ). Additionally, this approach is relatively tricky. The user does not know whether the correct or the estimated EEM parameters are used and, therefore, whether the resulting EEM charges will be of a good quality.

The EEM charges produced by Balloon were compared with the QM charges calculated by the B3LYP/cc-pVTZ/MPA approach. The coverage was close to 100 %, but the correlation was also low ( $R^2 < 0.8$ ). On the other hand, the Balloon developers mentioned that the EEM charges provided by Balloon do not correspond directly to some particular QM charges, and they should only be close to B3LYP/cc-pVTZ/MPA charges.

All the quality criteria and coverages for EEM parameter sets embedded in OpenBabel and Balloon are summarized in (Additional file 8: Table S5).

#### Coverage comparison and quality comparison combined

To date, there have been no EEM parameter sets available which would provide both high coverage and high-quality EEM charges (see Table 6). On the other hand, the EEM parameter sets calculated in this paper solve this problem, because they exhibit coverage close to 100 % and excellent quality criteria. Therefore, they can be used for chemoinformatics applications.

#### Software solution (step 4)

For the actual applicability of EEM in chemoinformatics, the user doesn't just need EEM parameter sets that are high quality and cover almost all molecules. They also need a software package that embeds these EEM parameter sets and calculates EEM charges based on them. We provide the user with two such solutions. First, we provide our EEM parameter sets in a format that can be directly used in EEM SOLVER (Additional file 2: EEM parameter sets). Second, we provide an OpenBabel patch which allows our EEM parameter sets to be used directly in OpenBabel (Additional file 9: OpenBabel patch). All the information including documentation is also accessible on the web: [http://ncbr.muni.cz/eem\\_parameters](http://ncbr.muni.cz/eem_parameters). The parameters are also accessible via ACC web application [77].

#### Conclusion

We provide here six EEM parameter sets which enable the user to calculate EEM charges with quality comparable to frequently used QM charges computed by well-known charge calculation schemes (i.e., MPA, NPA and AIM) and based on a robust QM approach (HF/6-311G, B3LYP/6-311G). The training set for EEM parameterization contained more than 4000 molecules from the DTP NCI drug database, and all six calculated EEM parameter sets exhibited a very good quality on this training set ( $R^2 > 0.9$ ).

The coverage of these computed EEM parameter sets was then compared with the coverages of 15 EEM parameter sets published in the past. This comparison was done on four key databases of drug-like molecules—DrugBank, ChEMBL, Pubchem and ZINC. The comparison showed that our EEM parameter sets enable us to calculate EEM charges for almost all molecules in these databases.

We then compared the quality of computed and published EEM parameter sets on two test data sets composed of approved drugs from DrugBank. This comparison also included EEM parameter sets embedded in

the software tools OpenBabel and Balloon. The comparison showed that our EEM parameter sets are among the best performing EEM parameter sets published to date ( $R^2 > 0.93$ ).

To summarize, charge calculation methodology suitable for chemoinformatics applications like virtual screening or QSAR should be fast, conformationally-dependent and accurate. EEM fulfils all these requirements. However, EEM parameter sets that would exhibit high coverage of drug-like molecule databases and provide high quality charges have not been available to date. The EEM parameters calculated in this paper solve this problem. They exhibit coverage close to 100 % and excellent quality criteria, therefore they are applicable in chemoinformatics.

Last but not least, we provide a software solution for the easy computing of EEM charges based on these EEM parameter sets—input files for EEM SOLVER and OpenBabel patch.

#### Additional files

**Additional file 1: Table S1.** List of training set molecules, including their NSC numbers and summary formulas.

**Additional file 2:** EEM parameters. Values of EEM parameter sets for these six charge calculation approaches (i.e. B3LYP/6-311G/MPA, B3LYP/6-311G/NPA, B3LYP/6-311G/AIM, HF/6-311G/MPA, HF/6-311G/NPA, and HF/6-311G/AIM). These EEM parameter sets are in a format which can be used as an input file for EEM SOLVER.

**Additional file 3: Table S2a.** A list of molecules from the test set including their DrugBank IDs and summary formulas.

**Additional file 4: Table S2b.** A list of molecules from the extended test set including their DrugBank IDs and summary formulas.

**Additional file 5: Table S3.** RMSD and  $\bar{\Delta}$  values of all tested EEM parameter sets on the test set.

**Additional file 6:** Charge details. Values of partial atomic charges (represented as tables and as graphs) for all tested EEM parameter sets on the testset.

**Additional file 7: Table S4.**  $R^2$ , RMSD,  $\bar{\Delta}$  and coverage values of all tested EEM parameter sets on the extended test set.

**Additional file 8: Table S5.** RMSD and  $\bar{\Delta}$  values for OpenBabel and Balloon on the test set and extended test set.

**Additional file 9:** OpenBabel patch. A patch for OpenBabel, which enables it to use the EEM parameter sets calculated in this paper.

#### Authors' contributions

The concept of the study originated from JK and was reviewed and extended by RA, while the design was put together by RSV and SG and reviewed by JK and RA. TB and SG prepared the input data (molecules and published EEM parameters). TB, SG and VH performed QM charge calculation. TR updated and extended NEEMP software. TB and TR performed EEM parameterizations, EEM charges validation and calculation of statistical data. VH prepared an automatic workflow, which is able to reproduce all steps preformed in the article. AK reviewed, corrected and improved this workflow. TR wrote the OpenBabel patch. The data were analyzed and interpreted by RSV, SG and JK. The manuscript was written by RSV in cooperation with JK, and reviewed by all authors. All authors read and approved the final manuscript.

**Author details**

<sup>1</sup> National Centre for Biomolecular Research, Faculty of Science and CEITEC, Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic. <sup>2</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic. <sup>3</sup> Institute of Computer Science, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic. <sup>4</sup> Skaggs School of Pharmacy and Pharmaceutical Sciences, University of California, 9500 Gilman Drive, San Diego, MC 0657, USA.

**Acknowledgements**

This work was supported by the Grant Agency of the Czech Republic [13-25401S]; the European Community's Seventh Framework Programme (CZ.1.05/1.1.00/02.0068) from the European Regional Development Fund; and by the European Social Fund and the state budget of the Czech Republic (CZ.1.07/2.3.00/20.0042, CZ.1.07/2.3.00/30.0009).

This work was also supported in part by NIH Grants R01 GM071872, U01 GM094612, and U54 GM094618 to R.A. The access to MetaCentrum supercomputing facilities provided under research intent MSM6383917201 is greatly appreciated.

**Authors' information**

Stanislav Geidl, Tomáš Bouchal and Tomáš Raček wish it to be known that, in their opinion, the first three authors should be regarded as joint First Authors. Radka Svobodová Vářeková and Jaroslav Kočá wish it to be known that, in their opinion, they should be regarded as joint Corresponding Authors.

**Competing interests**

The authors declare that they have no competing interests.

Received: 7 July 2015 Accepted: 16 November 2015

Published online: 02 December 2015

**References**

1. Svobodová Vářeková R, Geidl S, Ionescu C-M, Skřehota O, Kudera M, Sehnal D, Bouchal T, Abagyan R, Huber HJ, Kočá J (2011) Predicting pKa values of substituted phenols from atomic charges: comparison of different quantum mechanical methods and charge distribution schemes. *J Chem Inf Model* 51(8):1795–1806
2. Svobodová Vářeková R, Geidl S, Ionescu C-M, Skřehota O, Bouchal T, Sehnal D, Abagyan R, Kočá J (2013) Predicting pKa values from EEM atomic charges. *J Chem Inf Model* 53(1):18–28
3. Geidl S, Svobodová Vářeková R, Bendová V, Petrusk L, Ionescu C-M, Jurka Z, Abagyan R, Kočá J (2015) How does the methodology of 3D structure preparation influence the quality of pKa prediction? *J Chem Inf Model* 55(6):1088–1097
4. Dixon SL, Jurs PC (1993) Estimation of pKa for organic oxyacids using calculated atomic charges. *J Comput Chem* 14:1460–1467
5. Zhang J, Kleinöder T, Gasteiger J (2006) Prediction of pKa values for aliphatic carboxylic acids and alcohols with empirical atomic charge descriptors. *J Chem Inf Model* 46:2256–2256
6. Gross KC, Seybold PG, Hadad CM (2002) Comparison of different atomic charge schemes for predicting pKa variations in substituted anilines and phenols. *Int J Quantum Chem* 90:445–58
7. Ghafourian T, Dearden JC (2000) The use of atomic charges and orbital energies as hydrogen-bonding-donor parameters for QSAR studies: comparison of MNDO, AM1 and PM3 methods. *J Pharm Pharmacol* 52(6):603–610
8. Dudek AZ, Arodz T, Gálvez J (2006) Computational methods in developing quantitative structure-activity relationships (QSAR): a review. *Comb Chem High Throughput Screen* 9(3):213–228
9. Karelson M, Lobanov VS, Katritzky AR (1996) Quantum-chemical descriptors in QSAR/QSPR studies. *Chem Rev* 96(3):1027–1044
10. Todeschini R, Consonni V (2008) Handbook of molecular descriptors. Wiley-VCH Verlag GmbH, Weinheim
11. Galvez J, Garcia R, Salabert MT, Soler R (1994) Charge indexes. New topological descriptors. *J Chem Inf Model* 34(3):520–525
12. Stalke D (2011) Meaningful structural descriptors from charge density. *Chemistry* 17(34):9264–9278
13. Wermuth CG (2006) Pharmacophores: historical perspective and viewpoint from a medicinal chemist. In: Langer T, Hoffmann RD (eds) *Pharmacophores and pharmacophore searches*, vol 32. Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim
14. MacDougall PJ, Henze CE (2007) Fleshing-out pharmacophores with volume rendering of the Laplacian of the charge density and hyperwall visualization technology. In: Matta CF, Boyd RJ (eds) *The quantum theory of atoms in molecules: from solid state to DNA and drug design*. Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, pp 499–514
15. Clement OO, Mehl AT (2000) HipHop: pharmacophores based on multiple common-feature alignments. In: Güner OF (ed) *Pharmacophore perception, development, and use in drug design*. International University Line, La Jolla, pp 69–84
16. Lyne PD (2002) Structure-based virtual screening: an overview. *Drug Discov Today* 7(20):1047–1055
17. Bissantz C, Folkers G, Rogman D (2000) Protein-based virtual screening of chemical databases. 1. Evaluation of different docking/scoring combinations. *J Med Chem* 43(25):4759–4767
18. Park H, Lee J, Lee S (2006) Critical assessment of the automated AutoDock as a new docking tool for virtual screening. *Proteins* 65(3):549–554
19. Kearsley SK, Sallamack S, Fluder EM, Andose JD, Mosley RT, Sheridan RP (1996) Chemical similarity using physicochemical property descriptors. *J Chem Inf Model* 36(1):118–127
20. Nikolova N, Jaworska J (2003) Approaches to measure chemical similarity—a review. *QSAR Comb Sci* 22(910):1006–1006
21. Holliday JD, Jelfs SP, Willett P, Gedeck P (2003) Calculation of intersubstituent similarity using R-group descriptors. *J Chem Inf Comput Sci* 43(2):406–411
22. Tervo AJ, Rönkkö T, Nyronen TH, Poso A (2005) BRUTUS: optimization of a grid-based similarity function for rigid-body molecular superposition. 1. Alignment and virtual screening applications. *J Med Chem* 48(12):4076–4086
23. Vainio MJ, Johnson MS (2007) Generating conformer ensembles using a multiobjective genetic algorithm. *J Chem Inf Model* 47(6):2462–2474
24. Lemmen C, Lengauer T, Klebe G (1998) FLEXS: a method for fast flexible ligand superposition. *J Med Chem* 41(23):4502–4520
25. Mulliken RS (1955) Electronic Population Analysis on LCAO-MO Molecular Wave Functions. I. *J Chem Phys* 23(10):1833
26. Mulliken RS (1955) Electronic population analysis on LCAO-MO molecular wave functions. II. Overlap populations, bond orders, and covalent bond energies. *J Chem Phys* 23(10):1841
27. Löwdin P-O (1950) On the non-orthogonality problem connected with the use of atomic wave functions in the theory of molecules and crystals. *J Chem Phys* 18(3):365
28. Reed AE, Weinhold F (1983) Natural bond orbital analysis of near-Hartree-Fock water dimer. *J Chem Phys* 78(6):4066–4073
29. Reed AE, Weinstock RB, Weinhold F (1985) Natural population analysis. *J Chem Phys* 83(2):735
30. Bader RFW (1985) Atoms in molecules. *Acc Chem Res* 18(1):9–15
31. Bader RFW (1991) A quantum theory of molecular structure and its applications. *Chem Rev* 91(5):893–928
32. Hirshfeld FL (1977) Bonded-atom fragments for describing molecular charge densities. *Theor Chim Acta* 44(2):129–138
33. Ritchie JP (1985) Electron density distribution analysis for nitromethane, nitromethide, and nitramide. *J Am Chem Soc* 107(7):1829–1837
34. Ritchie JP, Bachrach SM (1987) Some methods and applications of electron density distribution analysis. *J Comput Chem* 8(4):499–509
35. Breneman CM, Wiberg KB (1990) Determining atom-centered monopoles from molecular electrostatic potentials. The need for high sampling density in formamide conformational analysis. *J Comput Chem* 11(3):361–373
36. Singh UC, Kollman PA (1984) An approach to computing electrostatic charges for molecules. *J Comput Chem* 5(2):129–145
37. Besler BH, Merz KM, Kollman PA (1990) Atomic charges derived from semiempirical methods. *J Comput Chem* 11(4):431–439
38. Kelly CP, Cramer CJ, Truhlar DG (2005) Accurate partial atomic charges for high-energy molecules using class IV charge models with the MID! basis set. *Theor Chem Acc* 113(3):133–151
39. Marenich AV, Cramer CJ, Truhlar DG (2009) Universal solvation model based on solute electron density and on a continuum model of the solvent defined by the bulk dielectric constant and atomic surface tensions. *J Phys Chem B* 113(18):6378–6396

40. Gasteiger J, Marsili M (1978) A new model for calculating atomic charges in molecules. *Tetrahedron Lett* 19(34):3181–3184
41. Gasteiger J, Marsili M (1980) Iterative partial equalization of orbital electronegativity—a rapid access to atomic charges. *Tetrahedron* 36(22):3219–3228
42. Cho K-H, Kang YK, No KT, Scheraga HA (2001) A fast method for calculating geometry-dependent net atomic charges for polypeptides. *J Phys Chem B* 105(17):3624–3624
43. Olierenko AA, Pisarev SA, Palyulin VA, Zefirov NS (2006) Atomic charges via electronegativity equalization: generalizations and perspectives. *Adv Quantum Chem* 51:139–156
44. Shulga DA, Olierenko AA, Pisarev SA, Palyulin VA, Zefirov NS (2010) Fast tools for calculation of atomic charges well suited for drug design. *SAR QSAR Environ Res* 19(1–2):153–165
45. Mortier WJ, Ghosh SK, Shankar S (1986) Electronegativity equalization method for the calculation of atomic charges in molecules. *J Am Chem Soc* 108:4315–4320
46. Rappe AK, Goddard WA (1991) Charge equilibration for molecular dynamics simulations. *J Phys Chem* 95(8):3358–3363
47. Nistor RA, Poliakov JG, Müser MH, Mosey NJ (2006) A generalization of the charge equilibration method for nonmetallic materials. *J Chem Phys* 125(9):094108
48. Mathieu D (2007) Split charge equilibration method with correct dissociation limits. *J Chem Phys* 127(22):224103
49. Svobodová Vařeková R, Jiroušková Z, Vaněk J, Suchomel S, Koča J (2007) Electronegativity equalization method: parameterization and validation for large sets of organic, organohalogen and organometal molecule. *Int J Mol Sci* 8:572–572
50. Janssens GOA, Baekelandt BG, Toufar H, Mortier WJ, Schoonheydt RA (1995) Comparison of cluster and infinite crystal calculations on zeolites with the electronegativity equalization method (EEM). *J Phys Chem* 99(10):3251–3258
51. Heidler R, Janssens GOA, Mortier WJ, Schoonheydt RA (1996) Charge sensitivity analysis of intrinsic basicity of Faujasite-type zeolites using the electronegativity equalization method (EEM). *J Phys Chem* 100(50):19728–19734
52. Sorich MJ, McKinnon RA, Miners JO, Winkler DA, Smith PA (2004) Rapid prediction of chemical metabolism by human UDP-glucuronosyltransferase isoforms using quantum chemical descriptors derived with the electronegativity equalization method. *J Med Chem* 47(21):5311–5317
53. Bultinck P, Langenaeker W, Carbó-Dorca R, Tollenen JP (2003) Fast calculation of quantum chemical molecular descriptors from the electronegativity equalization method. *J Chem Inf Comput Sci* 43(2):422–428
54. Smirnov KS, van de Graaf B (1996) Consistent implementation of the electronegativity equalization method in molecular mechanics and molecular dynamics. *J Chem Soc Faraday Trans* 92(13):2469
55. Ionescu C-M, Geidl S, Svobodová Vařeková R, Koča J (2013) Rapid calculation of accurate atomic charges for proteins via the electronegativity equalization method. *J Chem Inf Model* 53(10):2548–2548
56. Baekelandt BG, Mortier WJ, Lievens JL, Schoonheydt RA (1991) Probing the reactivity of different sites within a molecule or solid by direct computation of molecular sensitivities via an extension of the electronegativity equalization method. *J Am Chem Soc* 113(18):6730–6734
57. Jiroušková Z, Vařeková RS, Vaněk J, Koča J (2009) Electronegativity equalization method: parameterization and validation for organic molecules using the Merz-Kollman-Singh charge distribution scheme. *J Comput Chem* 30(7):1174–1178
58. Bultinck P, Langenaeker W, Lahorte P, De Proft F, Geerlings P, Van Alsenoy C, Tollenen JP (2002) The electronegativity equalization method II: applicability of different atomic charge schemes. *J Phys Chem A* 106(34):7895–7901
59. Ouyang Y, Ye F, Liang Y (2009) A modified electronegativity equalization method for fast and accurate calculation of atomic charges in large biological molecules. *Phys Chem Chem Phys* 11(29):6082–6089
60. Bultinck P, Vanholme R, Popelier PLA, De Proft F, Geerlings P (2004) High-speed calculation of AIM charges through the electronegativity equalization method. *J Phys Chem A* 108(46):10359–10366
61. O'Boyle N, Banck M, James C, Morley C, Vandermeersch T, Hutchison G (2011) Open Babel: an open chemical toolbox. *J Chem Inf Model* 51(3):33–47
62. Puranen JS, Vainio MJ, Johnson MS (2010) Accurate conformation-dependent molecular electrostatic potentials for high-throughput in silico drug discovery. *J Comput Chem* 31(8):1722–1732
63. Svobodová Vařeková R, Koča J (2006) Optimized and parallelized implementation of the electronegativity equalization method and the atom-bond electronegativity equalization method. *J Comput Chem* 3:396–405
64. Bultinck P, Carbó-Dorca R, Langenaeker W (2003) Negative Fukui functions: new insights based on electronegativity equalization. *J Chem Phys* 118(10):4349
65. Burden FR, Polley MJ, Winkler DA (2009) Toward novel universal descriptors: charge fingerprints. *J Chem Inf Model* 49(3):710–715
66. Open NCI Database (2012) Release 4. <http://cactus.nci.nih.gov/download/nci/>
67. Sadowski J, Gasteiger J (1993) From atoms and bonds to three-dimensional atomic coordinates: automatic model builders. *Chem Rev* 93:2567–2581
68. Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Montgomery JA Jr, Vreven T, Kudin KN, Burant JC, Millam JM, Iyengar SS, Tomasi J, Barone V, Mennucci B, Cossi M, Scalmani G, Rega N, Petersson GA, Nakatsuji H, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Klene M, Li X, Knox JE, Hratchian PH, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Ayala PY, Morokuma K, Voth GA, Salvador P, Dannenberg JJ, Zakrzewski VG, Dapprich S, Daniels AD, Strain MC, Farkas O, Malick DK, Rabuck AD, Raghavachari K, Foresman JB, Ortiz JV, Cui Q, Baboul AG, Clifford S, Cioslowski J, Stefanov BB, Liu G, Liashenko A, Piskorz P, Komaromi I, Martin RL, Fox DJ, Keith T, Al-Laham MA, Peng CY, Nanayakkara A, Challacombe M, Gill PMW, Johnson B, Chen W, Wong MW, Gonzalez C, Pople JA. Gaussian 09, Revision E.01. <http://www.gaussian.com>
69. Todd A Keith (2015) AIMAll 15.05.18. <http://aim.tkristmill.com>
70. Raček T, Svobodová Vařeková R, Krének A, Koča J NEEMP—tool for parameterization of empirical charge calculation method EEM. <http://ncbr.muni.cz/neemp/>
71. Wishart DS, Knox C, Guo AC, Cheng D, Shrivastava S, Tzur D, Gautam B, Hassanali M (2008) DrugBank: a knowledgebase for drugs, drug actions and drug targets. *Nucleic Acids Res* 36(Database issue):901–906
72. Law Y, Knox C, Djoumbou Y, Jewison T, Guo AC, Liu Y, Maciejewski A, Arndt D, Wilson M, Neveu V, Tang A, Gabriel G, Ly C, Adamjee S, Dame ZT, Han B, Zhou Y, Wishart DS (2004) DrugBank 4.0: shedding new light on drug metabolism. *Nucleic Acids Res* 32(Database issue):1091–1097
73. Bento AP, Gaulton A, Hersey A, Bellis LJ, Chambers J, Davies M, Krüger FA, Light Y, Mak L, McGlinchey S, Nowotka M, Papadatos G, Santos R, Overington JP (2014) The ChEMBL bioactivity database: an update. *Nucleic Acids Res* 42(Database issue):1083–1090
74. Bolton EE, Wang Y, Thiessen PA, Bryant SH (2008) PubChem: integrated platform of small molecules and biological activities. In: Wheeler R, Spellmeyer D (eds) Annual Reports in Computational Chemistry, vol. 4, Chap 12. Elsevier, Oxford
75. Irwin JJ, Sterling T, Mysinger MM, Bolstad ES, Coleman RG (2012) ZINC: a free tool to discover chemistry for biology. *J Chem Inf Model* 52(7):1757–1768
76. R Core Team R: A Language and Environment for Statistical Computing. <http://www.r-project.org/>
77. Ionescu CM, Sehnal D, Falginella FL, Pant P, Pravda L, Bouchal T, Svobodová Vařeková R, Geidl S, Koča J (2015) AtomicChargeCalculator: interactive web-based calculation of atomic charges in large biomolecular complexes and drug-like molecules. *J Cheminf* 7(1):50

# **NEEMP: Software for validation, accurate calculation and fast parameterization of EEM charges**

Tomáš Raček<sup>1,2,3</sup>, Jana Pazúriková<sup>1,3</sup>, Radka Svobodová Vařeková<sup>1,2,\*</sup>,  
Stanislav Geidl<sup>1, 2</sup>, Aleš Křenek<sup>1,2</sup>, Francesco Luca Falginella<sup>1</sup>, Vladimír  
Horský<sup>1, 3</sup>, Václav Hejret<sup>1</sup>, Jaroslav Koča<sup>1, 2</sup>

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University  
Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk  
University Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>3</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00  
Brno, Czech Republic.

<sup>4</sup> Institute of Computer Science, Masaryk University Brno, Botanická 68a, 602  
00 Brno, Czech Republic.

*Journal of Cheminformatics* 2016, **8**:1

<https://doi.org/10.1186/s13321-016-0171-1>

SOFTWARE

Open Access



# NEEMP: software for validation, accurate calculation and fast parameterization of EEM charges

Tomáš Raček<sup>1,2,3</sup>, Jana Pazúriková<sup>1,3</sup>, Radka Svobodová Vařeková<sup>1,2\*</sup>, Stanislav Geidl<sup>1,2</sup>, Aleš Křenek<sup>1,4</sup>, Francesco Luca Falginella<sup>1</sup>, Vladimír Horský<sup>1,3</sup>, Václav Hejret<sup>1</sup> and Jaroslav Koča<sup>1,2</sup>

## Abstract

**Background:** The concept of partial atomic charges was first applied in physical and organic chemistry and was later also adopted in computational chemistry, bioinformatics and chemoinformatics. The electronegativity equalization method (EEM) is the most frequently used approach for calculating partial atomic charges. EEM is fast and its accuracy is comparable to the quantum mechanical charge calculation method for which it was parameterized. Several EEM parameter sets for various types of molecules and QM charge calculation approaches have been published and new ones are still needed and produced. Methodologies for EEM parameterization have been described in a few articles, but a software tool for EEM parameterization and EEM parameter sets validation has not been available until now.

**Results:** We provide the software tool NEEMP (<http://ncbr.muni.cz/NEEMP>), which offers three main functionalities: EEM parameterization [via linear regression (LR) and differential evolution with local minimization (DE-MIN)]; EEM parameter set validation (i.e., validation of coverage and quality) and EEM charge calculation. NEEMP functionality is shown using a parameterization and a validation case study. The parameterization case study demonstrated that LR is an appropriate approach for smaller and homogeneous datasets and DE-MIN is a suitable solution for larger and heterogeneous datasets. The validation case study showed that EEM parameter set coverage and quality can still be problematic. Therefore, it makes sense to verify the coverage and quality of EEM parameter sets before their use, and NEEMP is an appropriate tool for such verification. Moreover, it seems from both case studies that new EEM parameterizations need to be performed and new EEM parameter sets obtained with high quality and coverage for key structural databases.

**Conclusion:** We provide the software tool NEEMP, which is to the best of our knowledge the only available software package that enables EEM parameterization and EEM parameter set validation. Additionally, its DE-MIN parameterization method is an innovative approach, developed by ourselves and first published in this work. In addition, we also prepared four high-quality EEM parameter sets tailored to ligand molecules.

**Keywords:** Partial atomic charges, Electronegativity equalization method, EEM, EEM parameterization, wwPDB CCD database

\*Correspondence: radka.svobodova@ceitec.muni.cz

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic

Full list of author information is available at the end of the article

## Background

Information about electron density distribution in a molecule is very useful, because it gives us an insight into the chemical behavior of the molecule and helps us to understand its reactivity. We can express this information via the electron populations of orbitals. But this approach is highly complex, resource-demanding and inconvenient for applications. A markedly more efficient solution is to summarize the electron density “belonging” to each atom into one overall number—partial atomic charge. The concept of partial atomic charges was first applied in physical and organic chemistry, and because of its usefulness and intuitiveness it was also adopted in computational chemistry (e.g., docking [1], conformers generation [2] or molecular dynamics [3, 4]), bioinformatics (e.g., similarity searches [5, 6]), molecular structure comparison [7, 8]) and chemoinformatics (e.g., QSAR and QSPR modelling [9–14], pharmacophore design [15], virtual screening [16]).

The most common and also the most accurate charge calculation method is the application of quantum mechanics (QM). Specifically, QM is employed for calculating electron orbital populations, and the populations are divided among the individual atoms using a charge calculation scheme. Unfortunately, there is no one universal and best method for QM charge calculation. We can use various combinations of QM theory level and basis set to obtain information about electron distribution in the orbitals. In addition, we can also apply different charge calculation schemes to process this information and obtain a sum of electron density for each individual atom. Well-known charge calculation schemes are for example Mulliken population analysis (MPA) [17, 18], Natural population analysis (NPA) [19, 20], the atoms-in-molecules (AIM) approach [21, 22], CHELPG [23] and Merz-Singh-Kollman (MK) [24, 25] method. Therefore, many various combinations of QM theory level, basis set and charge calculation schemes can be used for QM charge calculation. Different combinations are suitable for different types of applications.

Although a wide spectrum of QM charge calculation methods are available, all the methods have a major limitation—they are very time-demanding. For this reason, empirical charge calculation approaches have been developed [3, 26–33]. One of the most frequently used empirical approaches is the electronegativity equalization method (EEM). It is based on DFT and it calculates the charges via the following equation set:

$$\begin{bmatrix} B_1 & \frac{\kappa}{R_{1,2}} & \cdots & \frac{\kappa}{R_{1,N}} & -1 \\ \frac{\kappa}{R_{2,1}} & B_2 & \cdots & \frac{\kappa}{R_{2,N}} & -1 \\ \vdots & \vdots & \ddots & \vdots & \vdots \\ \frac{\kappa}{R_{N,1}} & \frac{\kappa}{R_{N,2}} & \cdots & B_N & -1 \\ 1 & 1 & \cdots & 1 & 0 \end{bmatrix} \cdot \begin{bmatrix} q_1 \\ q_2 \\ \vdots \\ q_N \\ \bar{\chi} \end{bmatrix} = \begin{bmatrix} -A_1 \\ -A_2 \\ \vdots \\ -A_N \\ Q \end{bmatrix} \quad (1)$$

where  $q_i$  is the charge of an atom  $i$ ;  $R_{i,j}$  is the distance between atoms  $i$  and  $j$ ;  $Q$  is the total charge of the molecule;  $N$  is the number of atoms in the molecule;  $\bar{\chi}$  is the molecular electronegativity, and  $A_i$ ,  $B_i$  and  $\kappa$  are empirical parameters. The parameters  $A_i$  and  $B_i$  vary for individual atom types, where atom type is a combination of element type and maximal bond order of the atom  $i$ . For example, the atom type N3 means that the atom is nitrogen and it creates at least one triple bond with its neighbors.

The main advantages of EEM are the following: It provides conformationally dependent charges (i.e., charges sensitive to conformational change), it has low time complexity (i.e.,  $\theta(N^3)$ ) and its accuracy is comparable to QM approaches. A limitation of EEM is that it requires a set of empirical parameters (i.e.,  $A_i$  and  $B_i$  and  $\kappa$ ). These empirical parameters are calculated from QM charges using a process of EEM parameterization. Consequently, EEM can mimic the QM charge calculation approach for which it was parameterized. In addition, because the EEM parameter set is calculated for a specific dataset of molecules, it provides the highest quality of charges on molecules similar to this dataset. Therefore, the EEM parameterizations are often performed for different QM charge calculation approaches and also for various types of molecules (small organic molecules, peptides, proteins, ligands, organometals etc.) to achieve the best accuracy of EEM charges. A lot of EEM parameter sets were published in the past [34–39] and new EEM parameter sets are still in development [40]. Unfortunately, the EEM parameter sets published in the past often only contain parameters for a few atom types and therefore cannot be used for molecules including other atoms.

Because of the strong demand for EEM parameterization, several EEM parameterization approaches were developed. The most widely known is an application of linear regression (LR), described by [31] and [35] and utilized for the preparation of many EEM parameter sets, e.g., in [34–37, 39, 40]. An alternative approach is differential evolution, described and used in [38]. Also other approaches (e.g., accelerated random search [38], particle swarm optimization algorithm [38]) were tested for EEM parameterization, but they were not applicable. Unfortunately, no software is currently available for EEM parameterization or for the validation of EEM parameters. All the software tools related to EEM (e.g., OpenBabel [41], Balloon [42], EEM Solver [43]) are focused purely on EEM charge calculation.

This motivated us to create such a tool and to provide it to the research community. Specifically, we developed NEEMP—a software for fast EEM parameterization, EEM parameters validation and also EEM charge calculation. NEEMP offers two approaches for EEM parameterization—the standard LR method and differential evolution with local minimization (DE-MIN) approach,

recently developed by ourselves. NEEMP also provides two validation modes—a validation of EEM charge quality and coverage. The quality validation compares EEM charges with relevant QM charges and reports common correlation coefficients. The coverage validation analyzes how large a proportion of the molecules from the input database can be processed using the validated EEM parameter set (therefore the validated EEM parameter set covers these molecules).

NEEMP is available here: <http://ncbr.muni.cz/NEEMP>, source codes are also in (Additional file 1). NEEMP is also documented in Bio.Tools [57]—a portal of bioinformatics resources world-wide.

NEEMP performance was demonstrated via two case studies—the first was focused on EEM parameterization and the second on EEM parameter validation. In both case studies, we worked with molecules from the databases which are very interesting and important for the life science community. Specifically, the wwPDB CCD database [44] of all ligands present in biomacromolecular structures, the DrugBank database [45] of drug compounds and the PubChem database [46], containing a huge amount of organic molecules.

### Description of the tool

NEEMP offers the user three modes—calculation, parameterization and validation mode.

#### Calculation mode

In this mode, NEEMP calculates EEM charges for the input molecule(s) using a user-defined EEM parameter set. Therefore, this mode requires 3D structure(s) of the input molecule(s), information about their total charge (0 for neutral molecules, nonzero real number for charged molecules) and the input EEM parameter set. The charge calculation is performed using Eq. (1) and the values of EEM charges are returned.

#### Parameterization mode

This mode is for calculating EEM parameters. An input for this calculation is a training set of molecules (i.e., their 3D structures) and QM charges for each molecule. NEEMP can calculate EEM parameters for neutral molecules and also for ions. The parameterization can be performed via two approaches: LR and DE-MIN.

The LR approach is implemented according to its description in [35]. The only extension is that the previous implementation only enables the best performing EEM parameter set to be selected via searching for the highest squared Pearson coefficient ( $R^2$ ). NEEMP also offers a selection based on the lowest average atom type root mean square difference ( $\text{avg}(RMSD_a)$ ). The  $\text{avg}(RMSD_a)$  is calculated as an average of root mean square difference

values for individual atom types ( $RMSD_a$ ). For simplification, the  $\text{avg}(RMSD_a)$  metrics will be abbreviated below as “RMSD metrics”. Optionally, the program can also attempt to discard some of the molecules in the training set, which may yield better results in some situations.

The DE-MIN algorithm is one that we recently developed ourselves. Advanced EEM parameterization approaches [38] usually combine global optimization methods (evolution algorithms, genetic algorithms, simulated annealing) with local optimization methods (simplex method, conjugated gradients or other). These advanced approaches search for the set of EEM parameters that fit QM charges from the training set in the best possible way. They offer a more robust approach than LR, therefore they are applicable even for the heterogeneous training set. We combined differential evolution (DE) with local minimization, which has not been done before. DE starts with generating a random population of vectors, each vector consisting of  $\kappa$ ,  $A_i$  and  $B_i$  for all atom types. Afterwards, all vectors (i.e., EEM parameter sets) are evaluated: EEM charges are computed using the parameter set and compared to QM charges via the chosen metrics ( $R^2$ ,  $RMSD$ ). Vectors with at least slightly promising results (e.g.,  $R^2 > 0.2$  and  $R > 0$ ) are minimized by the local minimization method NEWUOA [47]. This step significantly increases the quality of population vectors. Then evolution is mimicked over many iterations: a new vector is created as a combination of two vectors randomly selected from the population. Again, if the vector is promising, we apply local minimization. The best vector found during the evolution iterations is polished again via a few more iterations of NEWUOA and presented as the result. Because of the random generation of the vectors, the DE-MIN approach works stochastically, i.e., even for identical inputs, the results will slightly differ.

#### Validation mode

This mode enables us to perform two types of EEM parameter set validation—coverage validation and quality validation.

The coverage validation analyzes, how large a proportion of the molecules from the input database are composed only of the atom types included in the input EEM parameter set. This means they are “covered” by this EEM parameter set. Therefore, the coverage validation requires an input database (containing the 3D structures of molecules) and the validated EEM parameters. This validation returns a count and a percentage of molecules from the database which are covered by the parameters. Additionally, it identifies which particular molecules are covered and which are not.

The quality validation tests the accuracy of the EEM charges produced by the input EEM parameter set on the

validated dataset. Therefore, the inputs are the validated EEM parameters, the dataset (3D structures of molecules) and relevant QM charges for each molecule. This validation of quality provides three types of quality criteria—summary criteria (calculated for the whole dataset), atom type criteria (calculated for all the atom types available in the validated EEM parameter set) and criteria for individual molecules. The summary validation criteria are the Pearson coefficient ( $R$ ), the squared Pearson coefficient ( $R^2$ ), the Spearman coefficient, the squared Spearman coefficient, root mean square deviation ( $RMSD$ ), absolute average difference ( $\Delta$ ) and maximal absolute difference ( $\Delta_{max}$ ). The atom type criteria and the criteria for individual molecules are the same, but they are calculated for all the relevant atom types or molecules, respectively. In addition, NEEMP also generates graphs depicting the correlation between reference charges and EEM charges. Specifically, it creates graphs showing the dependency for all the atoms (see Fig. 1a) and also graphs for individual atomic types (see Fig. 1b).

## Implementation

NEEMP is implemented as a single C program which switches among its three modes (calculation, parameterization, and validation) according to a command line option. Therefore, its distribution is trivial—only a single binary and a few libraries for a particular platform are downloaded. In total, the program size is approximately 5000 lines of code.

The most compute-intensive part (common to all program modes) is the solution of the linear equation

system (1). We use LAPACK DSPSV/DSPSVX calls [48] (Cholesky matrix factorization followed by backward substitution and optionally iterative refinement). The LR parameterization method solves another system of linear equations to do the least squares fitting; in this case we use a LAPACK DGELS call (QR factorization) which can handle nearly singular matrices more accurately.

Both open-source and Intel MKL LAPACK implementations are supported.

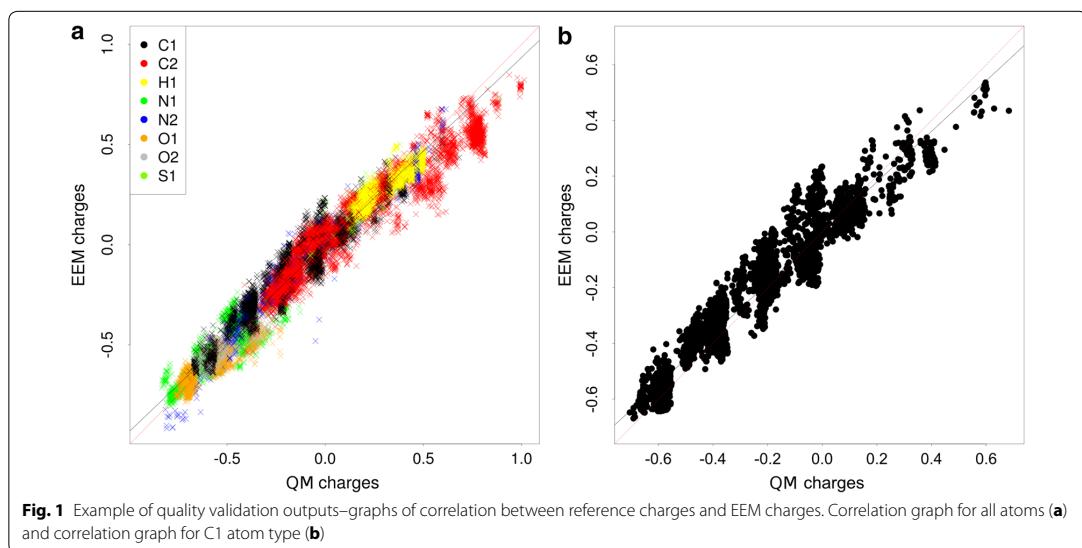
The DE-MIN parameterization method uses NEWUOA local minimization, the program links to Powell's implementation in its references [47].

The program utilizes simple, coarse grain parallelism. Using the Open MP programming paradigm, several loops—charge calculation for multiple molecules, evaluation over different  $\kappa$  values in the LR method, and minimization of multiple parameter sets in DE-MIN—run in parallel on available CPU cores. Of these, the first provides the best speedup.

The program only supports a single file format (SDF), using its internal file parsing routine, hence does not introduce dependencies on other libraries. Other file formats can be easily converted using 3rd-party tools (e.g., Open Babel).

## Results and discussion

We prepared two case studies to show the functionality and performance of NEEMP. The first case study is focused on EEM parameterization and the second on the validation of EEM parameters.



### Parameterization case study

This case study first compares EEM parameterization approaches (Parameterization comparison case study), then shows the parameterization running times (Parameterization running time case study) and afterwards focuses on EEM parameterization for wwPDB CCD (Parameterization calculation case study).

### Parameterization comparison case study

**Goal** Comparison of EEM parameterization approaches (LR vs. DE-MIN,  $R^2$  metric versus  $RMSD$  metric) and evaluation of which are the most suitable for which types of data.

**Datasets preparation** In this case study, we used four datasets, which are described in Table 1.

We wanted to demonstrate that NEEMP is able to produce results comparable with previously published data. Therefore, we focused first on datasets for which EEM parameterization has been performed in the past. Specifically, our first two datasets—DTP\_small and DTP\_large (see Table 1) originate from the DTP NCI database [49] and were used in publications [35] and [40], respectively. In addition, we also wanted to provide new interesting and useful results for the research community. For this reason, we then focused on datasets of interest to bioinformatics and chemoinformatics, and which have never been subjected to EEM parameterization. Specifically, the next two datasets (CCD\_gen and CCD\_exp, see Table 1) were obtained from the wwPDB Chemical component dictionary (wwPDB CCD) database.

This database contains molecules which are parts of biomacromolecular structures deposited in Protein Data Bank [50]. Therefore, these molecules are highly biologically important and include drug molecules, metabolites, compounds from biochemical pathways, etc. For each molecule, the wwPDB CCD contains two types of coordinates, i.e., ideal coordinates generated by CORINA software [51] (included in our dataset CCD\_gen) and model coordinates originating from experimental data (included in our dataset CCD\_exp). wwPDB CCD is a database of “raw” structural data, therefore we had to perform several preprocessing steps to create our datasets. In this way we obtained the datasets CCD\_gen\_all and CCD\_exp\_all, which we used in the validation case study. But for our EEM parameterization goals, these datasets were too large (about four times larger than the dataset DTP\_large). Therefore we reduced the size of datasets by a factor of four. Details about wwPDB CCD preprocessing and a summary of its results can be found in (Additional file 2) and (Additional file 3), respectively. Lists of the molecules in all datasets are in (Additional file 4).

The four datasets enable us to increase how demanding our EEM parameterization was in a stepwise manner and therefore show the strong and weak points of the LR and DE-MIN EEM parameterization approaches. The first dataset (DTP\_small) is the easiest—small, with low variability of atomic types, molecules and structure sources. The second dataset (DTP\_large) is more

**Table 1 Description of datasets used in parameterization case study**

Dataset				
Denotation	DTP_small	DTP_large	CCD_gen	CCD_exp
Source database	DTP NCI			wwPDB CCD
Number of molecules	1956	4475	4443	
Atomic types (elements and bond orders)	C1, C2, O1,O2, N1, N2,H, S1	H1, C1, C2,C3, N1, N2,N3, O1, O2,F1, P1, P2,S1, S2, Cl1,Br1,I1	H1, C1, C2, C3, N1,N2, N3, O1, O2, F1,P2, S1, S2, Cl1, Br1	
Size of molecules	6-176 atoms	5-124 atoms	3-305 atoms	
Type of molecules	Small organic molecules	Small organic molecules	Small organic and inorganic molecules, organometals, peptides	
Source of 3D structures	Generated by CORINA			Experimental structures
Characterization of a dataset	Variability of atomic types Variability of molecules Variability of structure sources	Low Low Low	High High	High
Reference to publication	[35] (set beg2)	[40]	–	–

ambitious, because it is large and contains a large number of atomic types. The third dataset (CCD\_gen) brings further complexity, since it contains heterogeneous types of molecules. The fourth dataset (CCD\_exp) is the most challenging, because it has all the demands of CCD\_gen and in addition, its structures originate from different experiments performed under highly varied conditions by various scientific teams.

**Selection and calculation of QM charges** The QM charge calculation approach B3LYP/6-311G/NPA was selected for calculating the QM charges used as inputs for the EEM parameterization. These charges were selected, because the B3LYP theory level, 6-311G basis set and NPA proved to be very suitable for EEM parameterization [37, 38, 40]. In addition, the same combination of B3LYP, 6-311G and NPA was used in publication [40], from which we took the dataset DTP\_large. The QM charges were calculated by Gaussian [52] for all molecules from datasets DTP\_small, DTP\_large, CCD\_gen and CCD\_exp.

**EEM parameterization** The EEM parameterization was performed using NEEMP on all four prepared datasets and four different parameterization methodologies were used (LR with  $R^2$  metrics, LR with RMSD metrics, DE-MIN with  $R^2$  metrics and DE-MIN with RMSD metrics). Thus we obtained 16 EEM parameter sets, including their quality criteria. The molecules in all the datasets were not optimized before performing the EEM parameterization. This strategy was motivated by a fact, that the resulting EEM parameters should be utilized also for non optimized molecules, to keep the procedure of EEM charge calculation quick. The same strategy was successfully used in the past (e.g., in articles [9, 35, 40, 53]).

**Comparison of EEM parameterization methods LR and DE-MIN using metrics  $R^2$  and RMSD.** The main quality criteria of the calculated EEM parameter sets are summarized in Table 2. Complete validation reports for all the EEM parameter sets are in (Additional file 5) and the particular EEM parameter sets are stored in (Additional file 6).

For the simple dataset DTP\_small, both LR and DE-MIN provide excellent results and both  $R^2$  and RMSD metrics are applicable. Only the combination of DE-MIN with  $R^2$  metrics performs slightly weaker.

For the bigger dataset DTP\_large, which contains more atom types, differences between the tested approaches started to appear. Summary quality criteria are still very good for all of the approaches, but only the combinations LR+RMSD and DE-MIN+RMSD also have acceptable atom types criteria. Interestingly, the performance of LR+RMSD and DE-MIN+RMSD is still almost the same.

For the dataset CCD\_gen, which brings a heterogeneity of molecules, the differences between the approaches markedly increase. LR still has good summary quality

criteria, but the atom types quality criteria significantly worsen, even with LR+RMSD. Therefore, only the combination DE-MIN+RMSD seems to be applicable for this dataset and provides very good quality criteria.

In the last and the most challenging dataset CCD\_exp, the tested approaches demonstrate similar trends as for CCD\_gen, but even more pronounced. LR also has weak summary quality criteria and the atom types quality criteria are highly problematic. Fortunately, the DE-MIN+RMSD approach is still applicable and provides quality criteria only slightly worse than for CCD\_gen. A graph of the QM and EEM charges correlation for CCD\_exp and the approaches LR+RMSD and DE-MIN+RMSD are shown in Fig. 2, and demonstrate that with such a large and heterogeneous dataset, the proper choice of EEM parameterization approach is crucial.

**Summary of comparison results** To conclude, we found that LR (with both metrics) is an appropriate approach for smaller and homogeneous datasets. On the other hand, DE-MIN (with RMSD metric) is a markedly more suitable solution for larger and more heterogeneous datasets.

#### Parameterization running time case study

The performance of NEEMP, measured on a standard personal computer is showed in Table 3.

All measurements were repeated 3 times, and we always considered the minimum running time of all the repetitions (in this way random interference of background activity of the operating system is masked out). Running time varies from a few minutes to several hours. As expected, there is no observable difference between CCD\_gen and CCD\_exp—the complexity depends on the number of molecules and atoms but not the specific values of atom coordinates or charges. In general, DE-MIN performs significantly better for all datasets. The difference becomes more apparent with a larger number of molecules, being caused by the discard algorithm, which has to examine more options (this step is not necessary for DE-MIN).

As described in the Implementation section, the code can run on multiple CPU cores in the heaviest computations, therefore the computation time can be markedly shortened. Figure 3 shows speedup of the parallel version on different number of CPU cores, i.e., how many times faster the parallel program runs compared to the single-core version. The experiments were run with the DE-MIN method and RMSD metric, on the CCD\_exp dataset and using a machine with 4 Intel Xeon E7-4860 @ 2.27 GHz CPUs. Again, all measurements were repeated 3 times, and we always considered the minimum running time. The particular values of minimum running time are summarized in (Additional file 7: Table S2).

In the ideal case, if the workload was uniformly distributed among all the cores, the speedup would be the

**Table 2** Quality criteria of EEM parameter sets calculated in parameterization comparison case study

Dataset	EEM parameterization		Quality criteria				
			Summary criteria			Atom types criteria	
	Method	Metric	R <sup>2</sup>	RMSD	Δ	max(RMSD <sub>a</sub> )	max(Δ <sub>a</sub> )
DTP_small	LR	R <sup>2</sup>	0.9718	0.0555	0.0416	0.1061	0.0882
		RMSD	0.9686	0.0592	0.0418	0.0917	0.0709
	DE-MIN	R <sup>2</sup>	0.9728	0.1023	0.0809	0.2427	0.2392
		RMSD	0.9700	0.0584	0.0408	0.0926	0.0714
DTP_large	LR	R <sup>2</sup>	0.9629	0.0780	0.0554	1.5287	1.3864
		RMSD	0.9531	0.0729	0.0545	0.1767	0.1402
	DE-MIN	R <sup>2</sup>	0.9674	0.1875	0.1439	2.0937	2.0671
		RMSD	0.9599	0.0693	0.0515	0.1774	0.1436
CCD_gen	LR	R <sup>2</sup>	0.9662	0.0732	0.0497	1.7118	0.6637
		RMSD	0.9484	0.0881	0.0609	0.7908	0.5174
	DE-MIN	R <sup>2</sup>	0.9764	1.2229	0.9972	0.9267	6.9413
		RMSD	0.9696	0.0648	0.0449	0.1595	0.1128
CCD_exp	LR	R <sup>2</sup>	0.9620	0.0803	0.0526	2.2310	0.8494
		RMSD	0.8848	0.1376	0.0830	2.1176	1.9218
	DE-MIN	R <sup>2</sup>	0.9738	0.2764	0.2230	1.4040	1.3970
		RMSD	0.9687	0.0665	0.0466	0.1933	0.1383

**Legend:**

R <sup>2</sup>	> 0.95	> 0.925	> 0.9	> 0.85	> 0.8	< 0.8	
RMSD, Δ, max(RMSD <sub>a</sub> ), max(Δ <sub>a</sub> )	< 0.05	< 0.1	< 0.15	< 0.2	< 0.3	< 0.4	≥ 0.4

same as the number of cores. However, the measurement shows a decrease in efficiency of the parallel execution, which is a consequence of the non-uniform distribution of the workload (existence of non-parallel sections in fact). We can conclude that it is worth running NEEMP with **up to 20 CPU cores**, where we still get an approximately 6x speedup, but using more cores becomes a waste of resources. In general, with larger training sets, when there is more work to evaluate a single parameter vector, the efficiency will improve.

**Parameterization calculation case study**

**Goal** In this case study, we would like to obtain high-quality EEM parameters for the wwPDB CCD database and based on several frequently used QM charge calculation approaches. For this purpose, we will apply the knowledge obtained during our comparison of EEM parameterization approaches.

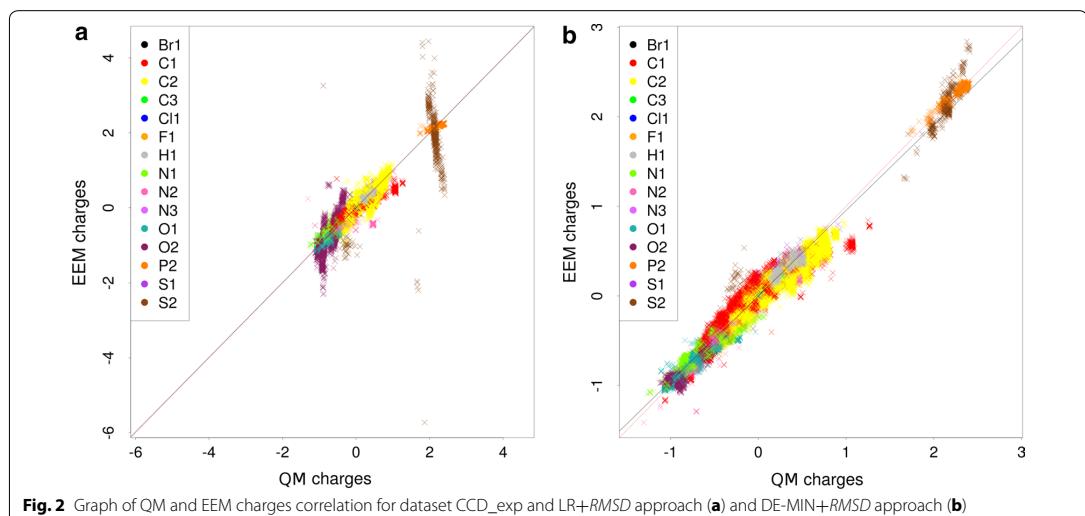
**Dataset preparation** During this comparison, we prepared two datasets for wwPDB CCD: CCD\_gen and CCD\_exp. CCD\_gen provided EEM parameter sets with better quality criteria, therefore we will use this dataset.

**Selection and calculation of QM charges** The QM charge calculation approach B3LYP/6-311G/NPA was

again selected—for the same reasons as in the parameterization comparison case study. Furthermore, B3LYP/6-311G/MPA was selected, because MPA is often used for EEM parameterization [31, 34–37] as well. Moreover, it was also used in combination with B3LYP/6-311G [40]. Then, the approaches B3LYP/6-311G\*/NPA and B3LYP/6-311G\*/MPA were selected. The reason for this was that the 6-311G\* basis set had never been used for EEM parameterization, and EEM parameters for these approaches can be interesting and useful for the research community. The QM charges were calculated by Gaussian [52] for all molecules from the CCD\_gen dataset, except for the B3LYP/6-311G/NPA charges, which were taken from the parameterization comparison case study.

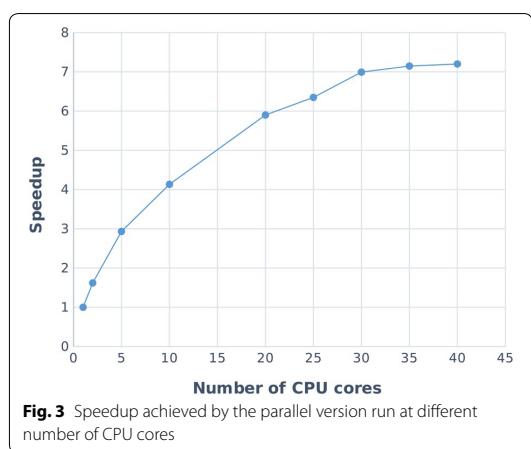
**EEM parameterization** The EEM parameterization was performed by NEEMP on the CCD\_gen dataset for the four above-mentioned QM charges. The DE-MIN+RMSD approach was used, because it provides the best results for CCD\_gen in the parameterization comparison case study. Thus we obtained 4 EEM parameter sets, including their quality criteria.

**Summary of EEM parameterization results** The denotations and the quality criteria of the obtained EEM



**Table 3** NEEMP performance on a standard personal computer (Intel i7-4790K CPU @ 4.00GHz)

Dataset	DTP_small	DTP_large	CCD_gen and CCD_exp
EEM parameterization method	LR	DE-MIN	LR
Running time	54 m	14 m	16 m
		4 h 25 m	9 h 24m
			25 m



parameter sets are summarized in Table 4. These results show that NEEMP provided us with four high-quality EEM parameter sets for the wwPDB CCD database. These EEM parameter sets are in (Additional file 6) and validation reports for them are in (Additional file 5).

### Validation case study

This case study first analyses the coverage of selected EEM parameter sets (Coverage validation case study) and then also the quality of these sets (Quality validation case study).

#### Coverage validation case study

*Goal* In this case study, we would like to compare the coverage of selected EEM parameter sets on key databases of small molecules. In this way, we introduce NEEMP functionality focused on the validation of coverage.

*EEM parameter sets* Several sets of published EEM parameter sets [34, 35, 37, 38, 40], i.e., the sets which proved to be of good quality in the past [10, 11, 40], and also four EEM parameter sets calculated in the parameterization calculation case study were selected for the coverage comparison. A list of the compared EEM parameter sets, including basic information about them, can be seen in the first three columns of (Additional file 8: Table S3).

*Databases* The coverage comparison was done on three well-known databases of biologically important small molecules: wwPDB CCD, DrugBank, and PubChem. The number of compounds in all these databases (from March 2016) are summarized in Table 5. wwPDB CCD,

**Table 4 Denotations and main quality criteria of EEM parameter sets calculated in parameterization calculation case study**

EEM parameter set denotation	Relevant QM charges	Quality criteria				
		Summary criteria			Atom types criteria	
		R <sup>2</sup>	RMSD	Δ	max(RMSD <sub>a</sub> )	max(Δ <sub>a</sub> )
Ccd2016_npa	B3LYP/6-311G/NPA	0.9696	0.0648	0.0449	0.1595	0.1128
Ccd2016_mpa	B3LYP/6-311G/MPA	0.9502	0.0711	0.0491	0.2138	0.1276
Ccd2016_npa2	B3LYP/6-311G*/NPA	0.9747	0.0595	0.0438	0.1904	0.1588
Ccd2016_mpa2	B3LYP /6-311G*/MPA	0.9676	0.0582	0.0417	0.1319	0.1019

**Legend:**

R <sup>2</sup>	> 0.95	> 0.925	> 0.9	>0.85	> 0.8	< 0.8
RMSD, Δ, max(RMSD <sub>a</sub> ), max(Δ <sub>a</sub> )	< 0.05	< 0.1	< 0.15	< 0.2	< 0.3	< 0.4 &gt;= 0.4

**Table 5 Size of database, used for comparison of EEM parameter set coverages**

Database	Number of compounds
DrugBank	7097
wwPDB CCD	21,741
PubChem	71,632,601

which was also used in the parameterization case study, is a medium-sized database including ligands incorporated in biomacromolecules. DrugBank is a relatively small database containing chemical compounds with medical applications. The PubChem database intends to include all common chemical substances, therefore it is very large and heterogeneous.

*Coverage comparison procedure* The coverage of all the tested EEM parameter sets was calculated via NEEMP for all three databases of interest. The results are summarized in (Additional file 8: Table S3).

*Summary of results* Interestingly, even though the databases are very different, the coverage values are very similar for all of them. Only the EEM parameter sets calculated recently (i.e., Cheminf2015 and Ccd2016 sets) exhibit sufficient coverage (> 93 % for all the databases). The other parameter sets have low coverage, specifically, they are only applicable for 40–80% of molecules from the tested databases. The coverage values for DrugBank and PubChem agree with information published in [40]. In general, this confirms that coverage is a weakness of the majority of currently published EEM parameter sets. We also showed that NEEMP enables us to easily obtain information about the EEM parameter set coverage for each database of interest.

**Quality validation case study**

*Goal* This case study compares the quality of selected EEM parameter sets on two datasets, which contain wwPDB CCD structures. It also shows NEEMP functionality focused on the validation of EEM parameter set quality.

*Preparation of datasets* Two datasets containing molecules from wwPDB CCD were used for quality comparison—a simple dataset for basic testing and a challenging dataset for deep analysis of EEM parameter set quality. The challenging dataset is specifically the dataset CCD\_gen\_all, which was prepared in the parameterization comparison case study and which includes structures generated by CORINA. This dataset contains all the wwPDB CCD molecules composed of atoms of C, H, N, O, S, P, F, Cl and Br and that have no structural errors. Therefore, it includes about 82 % of the whole of wwPDB CCD, it is highly chemically heterogeneous and demanding for calculating high-quality EEM charges. The simple dataset (denoted CCD\_gen\_CHNO) is a subset of CCD\_gen\_all. The list of molecules in this dataset can be found in (Additional file 4). This dataset was designed for a basic quality test of all the EEM parameter sets used in the coverage validation case study, and so it had to be completely covered by all these EEM parameter sets. For this reason, its molecules contain only the atoms C, H, N and O and do not include triple bonds. This fact implies its low chemical variability. Information about both datasets are summarized in Table 6.

*EEM parameter sets* The quality comparison was analyzed on the same EEM parameter sets as the coverage comparison. Specifically, when the quality comparison was performed on the dataset CCD\_gen\_CHNO, all these EEM parameter sets were used. The quality comparison on CCD\_gen\_all was only performed for the

**Table 6** Description of datasets used in quality validation case study

Datasets		
Designation	CCD_gen_CHNO*	CCD_gen_all*
Source database	wwwPDB CCD	wwwPDB CCD
Number of molecules	8144	17,769
Atomic types (elements and bond orders)	H1, C1, C2, N1, N2, O1, O2	H1, C1, C2, C3, N1, N2, N3, O1, O2, F1, P2, S1, S2, Cl1, Br1

\*All other information about the dataset is the same as for the dataset CCD\_gen, described in Table 1

Cheminf2015 and Ccd2016 EEM parameter sets, because only these sets can be applied on all the molecules from this dataset.

*Calculation of QM charges* For molecules from the dataset CCD\_gen\_CHNO, we calculated the same charges as in the coverage validation case study, because EEM charges calculated using the tested EEM parameter sets had to be compared with corresponding QM charges. Therefore, the following QM charges were calculated: HF/STO-3G/MPA and NPA, B3LYP/6-31G\*/MPA and NPA, B3LYP/6-311G/MPA and NPA, and B3LYP/6-311G\*/MPA and NPA. For molecules from the dataset CCD\_gen\_all, we only calculated QM charges corresponding to the EEM parameter sets Cheminf2015 and Ccd2016. Therefore we calculated the QM charges B3LYP/6-311G/MPA and NPA, and B3LYP/6-311G\*/MPA. The QM charges were calculated by Gaussian [52] or (where possible) taken from the parameterization case study.

*Quality comparison procedure* For the dataset CCD\_gen\_CHNO, the EEM charges were calculated using the same EEM parameter sets as in the coverage validation case study. Afterwards, these EEM charges were compared with the corresponding QM charges via NEEMP and the validation reports were created. A summary of the most important quality criteria and the validation reports are in (Additional file 9: Table S4) and (Additional file 5), respectively. For the dataset CCD\_gen\_all, the EEM charges were calculated using only the EEM parameter sets Cheminf2015 and Ccd2016. We then employed NEEMP to compare EEM charges with relevant QM charges and produce validation reports. The most important quality criteria are summarized in Table 7, selected correlation graphs are shown in Fig. 4 and all the validation reports are in (Additional file 5).

*Summary of results* All the EEM parameter sets proved to be of very high quality on the dataset CCD\_gen\_CHNO. Both the summary quality criteria and the atom type quality criteria were excellent. Specifically,  $R^2$  was mostly higher than 0.95,  $RMSD < 0.08$  and  $\max(RMSD_a) < 0.12$ . This documents the fact that all these EEM parameter sets are very well adjusted for EEM charge calculation on datasets with low atom type variability. The results for the dataset CCD\_gen\_all are more heterogeneous (see Table 7). The summary criteria are

excellent ( $R^2 > 0.95$ ) or at least acceptable ( $R^2 \sim 0.9$ ) for all the EEM parameter sets. But the atom type quality criteria are sometimes problematic. The Cheminf2015\_mpa parameter set in particular produced very high  $\max(RMSD_a)$  and  $\max(\Delta_a)$  values. Figure 4a and the validation report shows that there is a problem with the correlation of charges on carbon atoms with triple bonds (C3 atoms). Further EEM parameter sets have sufficiently low atom type quality criteria. Two of the EEM parameter sets (Ccd2016\_mpa and Cheminf2015\_npa) contain slide correlation issues for S2 or C3—see an example in Fig. 4b. The remaining EEM parameter sets exhibited no problems or issues. Furthermore, the quality criteria of all Ccd2016 parameter sets are comparable to the quality criteria obtained during the calculation process (see Table 4). This fact confirmed the robustness of the EEM parameterization performed via NEEMP. In general, these results show that the datasets with high atom type variability can still represent a challenge for the available EEM parameter sets. Therefore, the EEM parameter set quality validation implemented in NEEMP is a very important step in EEM usage and application.

## Conclusion

We provide the software tool NEEMP, which offers three main functionalities: EEM parameterization (via the LR and DE-MIN method, with  $R^2$  and  $RMSD$  metrics); EEM parameter set validation (i.e., validation of coverage and quality) and EEM charge calculation. NEEMP was implemented in C, contains parallelization and provides a fast and accurate solution for work with EEM. To the best of our knowledge, NEEMP is the only available software tool enabling EEM parameterization and EEM parameter set validation. In addition, the DE-MIN parameterization method is an innovative approach, developed by ourselves and first published in this work.

NEEMP functionality is demonstrated on two case studies—a parameterization and a validation case study.

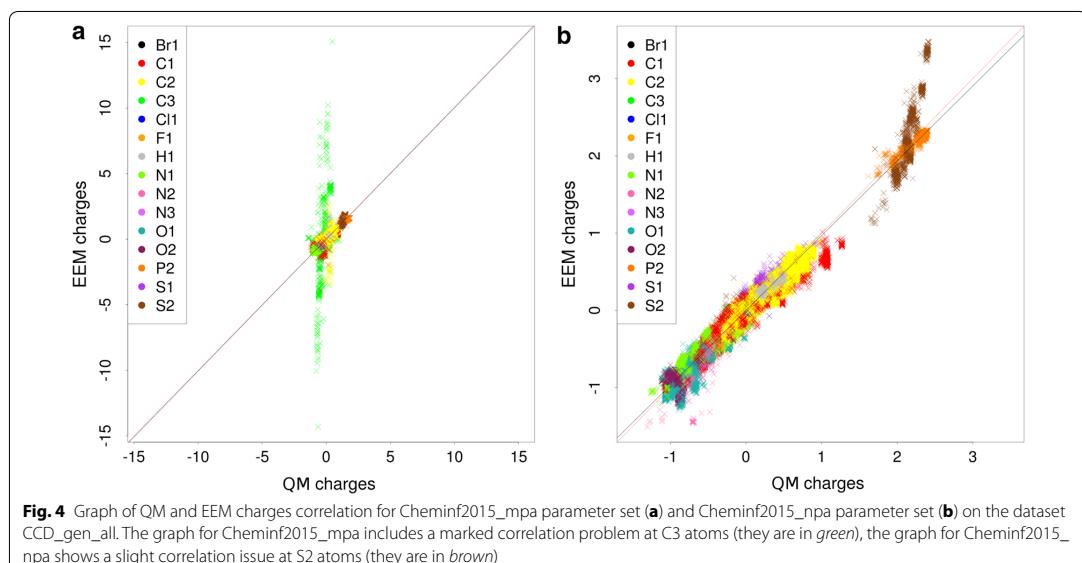
The parameterization case study analyses the performance of both parameterization methods (LR and DE-MIN) and metrics ( $R^2$  and  $RMSD$ ) using four different datasets which increase the demands of EEM parameterization in a stepwise manner. We found that LR (with both metrics) is an appropriate approach for

**Table 7** Quality criteria of the EEM parameter sets on the dataset CCD\_gen\_all

Relevant QM charges		EEM parameter set name	Quality criteria					
QM theory level + basis set	Charge calc. scheme		Summary criteria			Atom types criteria		
			R <sup>2</sup>	RMSD	Δ	max(RMSD <sub>a</sub> )	max(Δ <sub>a</sub> )	
B3LYP/6-311G	MPA	Cheminf2015_mpa	0.8986	0.1189	0.0597	2.2752	1.1929	
		Ccd2016_mpa	0.9568	0.0732	0.0473	0.2884	0.0911	
	NPA	Cheminf2015_npa	0.9763	0.0635	0.0446	0.281	0.1916	
		Ccd2016_npa	0.9872	0.9254	0.0662	0.1682	0.1219	
B3LYP/6-311G*	MPA	Ccd2016_mpa2	0.97	0.0595	0.0406	0.1365	0.102	
	NPA	Ccd2016_npa2	0.9796	0.0603	0.0428	0.2002	0.1671	

**Legend:**

R <sup>2</sup>	> 0.95	> 0.925	> 0.9	> 0.85	> 0.8	< 0.8	
RMSD, Δ, max(RMSD <sub>a</sub> ), max(Δ <sub>a</sub> )	< 0.05	< 0.1	< 0.15	< 0.2	< 0.3	< 0.4	>= 0.4

**Fig. 4** Graph of QM and EEM charges correlation for Cheminf2015\_mpa parameter set (a) and Cheminf2015\_npa parameter set (b) on the dataset CCD\_gen\_all. The graph for Cheminf2015\_mpa includes a marked correlation problem at C3 atoms (they are in green), the graph for Cheminf2015\_npa shows a slight correlation issue at S2 atoms (they are in brown)

smaller and homogeneous datasets. On the other hand, DE-MIN (with RMSD metric) is a markedly more suitable solution for larger and more heterogeneous datasets. We also showed that NEEMP is able to perform EEM parameterizations in a reasonable time, and its execution on multiple processors produces a marked speedup. We then performed EEM parameterization via the DE-MIN method with RMSD metrics on wwPDB CCD—a database of ligands found in biomacromolecular structures. This database is frequently used by the life science

community and it has never been subjected to EEM parameterization. Despite the high heterogeneity of the database, we produced 4 high-quality EEM parameter sets. This demonstrated, that NEEMP is highly applicable for the computation of new EEM parameter sets.

The validation case study focused first on coverage validation. Specifically, we validated the coverage of selected EEM parameter sets (i.e., several published EEM parameter sets and EEM parameter sets provided in this article) on three well-known databases of small molecules

(wwPDB CCD, PubChem and DrugBank). It was shown that the coverage of older EEM parameter sets is problematic. Specifically, they are only applicable for 40–80 % of molecules from the tested databases. Only the recently published Cheminf2015 EEM parameter sets and the EEM parameter sets provided in this article had sufficient coverage (>90–95 %). The case study then also focused on quality validation of the selected EEM parameter sets. All the sets performed very well on a small dataset with molecules comprised of C, H, N and O. On the other hand, the larger and more heterogeneous dataset (17,769 molecules; 15 atom types) was a challenge for most of the tested EEM parameter sets. The older parameter sets could not cover the dataset and the newer ones (i.e., Cheminf 2015) had accuracy problems with some atom types. The only applicable EEM parameter sets were the Ccd2016 sets provided in this article. From these results it can be seen that EEM parameter set coverage and quality can still be problematic. Therefore it makes sense to verify the coverage and quality of EEM parameter sets before their use, and NEEMP is an appropriate tool for such verification.

Moreover, from both case studies it seems that it is still necessary to perform new EEM parameterizations and obtain EEM parameter sets with high quality and coverage on key structural databases.

Last but not least, NEEMP can potentially help the community to perform EEM parameterizations which are challenging. For example, EEM parameterization based on HF/6-31G\*/MK QM charges. Mimicking these QM charges via EEM is very important because they are used for AMBER partial-charge parameterization routine focused on biomolecular ligands [54–56]. On the other hand, EEM is documented as an approach which performs very weakly for MK charge calculation scheme [10, 37, 40]. Employing NEEMP can help us to override the problems with MK based EEM parameterizations, or it can confirm limitations of EEM in this domain. Further challenging EEM parameterizations, which can be potentially solved via NEEMP, are parameterizations focused on proteins or metalloproteins—large macromolecules containing long-range interactions.

## Availability and requirements

**Project name:** NEEMP

**Project home page:** <http://ncbr.muni.cz/neemp>

**Operating system(s):** Linux (recommended), Windows, Mac OS X

**Programming language:** C, external library in Fortran

**Other requirements:** GNU Fortran, libxml2, LAPACK, zlib, OpenMP

**License:** GNU GPLv3

**Any restrictions on use by non-academics:** no restrictions

## Additional files

**Additional file 1.** Source codes. Archive with source codes of NEEMP.

**Additional file 2.** Preprocessing information. Detailed description of wwPDB CCD preprocessing.

**Additional file 3.** Summarization of preprocessing results for individual molecules from wwPDB CCD.

**Additional file 4.** List of molecules. Lists of molecules in all datasets.

**Additional file 5.** Validation reports. Validation reports for EEM parameter sets, calculated in parameterization case study and for EEM parameter sets, tested in validation case study.

**Additional file 6.** EEM parameter sets. EEM parameter sets, calculated in the parameterization case study.

**Additional file 7: Table S2.** NEEMP running times on more CPUs.

**Additional file 8: Table S3.** Summary information about coverage of tested EEM parameter sets, performed on databases wwPDB CCD, DrugBank and PubChem.

**Additional file 9: Table S4.** Quality criteria of the EEM parameter sets on the dataset CCD\_gen\_CHNO.

## Authors' contributions

TR: Design and development of NEEMP, work coordination; JP: DE-MIN module design and development, performing of analyses; RSV: Case study experiments design, results evaluation, writing publication; SG: Prototyping of DE-MIN and suggesting of RMSD instead R2, validation reports implementation; AK: NEEMP development, measuring NEEMP running times; FLF: Charge calculation, NEEMP testing, writing NEEMP documentation; VHo: wwPDB validation; VHe: Charge calculation; JK: Review of experimental design, writing publication. All authors read and approved the manuscript.

## Author details

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic. <sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic. <sup>3</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic. <sup>4</sup> Institute of Computer Science, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic.

## Acknowledgements

This research has been financially supported by the Ministry of Education, Youth and Sports of the Czech Republic under the project CEITEC 2020 (LQ1601).

Access to computing and storage facilities owned by parties and projects contributing to the National Grid Infrastructure MetaCentrum provided under the programme “Projects of Projects of Large Research, Development, and Innovations Infrastructures” (CESNET LM2015042), is greatly appreciated. In addition, access to the CERTI-SC computing and storage facilities provided by the CERTI-SC Center under the programme “Projects of Projects of Large Research, Development, and Innovations Infrastructures” (CERTI Scientific Cloud LM2015085), is greatly appreciated

## Competing interests

The authors declare that they have no competing interests.

Received: 8 July 2016 Accepted: 5 October 2016

Published online: 17 October 2016

## References

- Park H, Lee J, Lee S (2006) Critical assessment of the automated AutoDock as a new docking tool for virtual screening. Proteins 65(3):549–554

2. Vainio MJ, Johnson MS (2007) Generating conformer ensembles using a multiobjective genetic algorithm. *J Chem Inf Model* 47(6):2462–2474
3. Rappe AK, Goddard WA (1991) Charge equilibration for molecular dynamics simulations. *J Phys Chem* 95(8):3358–3363
4. Chenoweth K, Van Duin AC, Goddard WA (2008) Reaxff reactive force field for molecular dynamics simulations of hydrocarbon oxidation. *J Phys Chem A* 112(5):1040–1053
5. Kearsley SK, Sallamack S, Fluder EM, Andose JD, Mosley RT, Sheridan RP (1996) Chemical similarity using physicochemical property descriptors. *J Chem Inf Model* 36(1):118–127
6. Holliday JD, Jelfs SP, Willett P, Gedeck P (2003) Calculation of intersubstituent similarity using R-group descriptors. *J Chem Inf Comput Sci* 43(2):406–411
7. Tervo AJ, Rönkkö T, Nyroën TH, Poso A (2005) BRUTUS: optimization of a grid-based similarity function for rigid-body molecular superposition. 1. Alignment and virtual screening applications. *J Med Chem* 48(12):4076–4086
8. Lemmen C, Lengauer T, Klebe G (1998) FLEXS: a method for fast flexible ligand superposition. *J Med Chem* 41(23):4502–4520
9. Svobodová Vařeková R, Geidl S, Ionescu C-M, Skřehota O, Kudera M, Sehnal D, Bouchal T, Abagyan R, Huber HJ, Koča J (2011) Predicting pKa values of substituted phenols from atomic charges: comparison of different quantum mechanical methods and charge distribution schemes. *J Chem Inf Model* 51(8):1795–1806
10. Svobodová Vařeková R, Geidl S, Ionescu C-M, Skřehota O, Bouchal T, Sehnal D, Abagyan R, Koča J (2013) Predicting pKa values from EEM atomic charges. *J Cheminf* 5(1):18
11. Geidl S, Svobodová Vařeková R, Bendová V, Petrusk L, Ionescu C-M, Jurka Z, Abagyan R, Koča J (2015) How does the methodology of 3D structure preparation influence the quality of pKa prediction? *J Chem Inf Model* 55(6):1088–1097
12. Gross KC, Seybold PG, Hadad CM (2002) Comparison of different atomic charge schemes for predicting pKa variations in substituted anilines and phenols. *Int J Quantum Chem* 90:445–458
13. Galvez J, García R, Salabert MT, Soler R (1994) Charge indexes: new topological descriptors. *J Chem Inf Model* 34(3):520–525
14. Stalke D (2011) Meaningful structural descriptors from charge density. *Chemistry* 17(34):9264–9278
15. MacDougall PJ, Henze CE (2007) Fleshing-out pharmacophores with volume rendering of the laplacian of the charge density and hyperwall visualization technology. In: Matta CF, Boyd RJ (eds) The quantum theory of atoms in molecules: from solid state to DNA and drug design. Wiley, Weinheim, pp 499–514
16. Bissantz C, Folkers G, Rognan D (2000) Protein-based virtual screening of chemical databases. 1. Evaluation of different docking/scoring combinations. *J Med Chem* 43(25):4759–4767
17. Mulliken RS (1955) Electronic population analysis on LCAO-MO molecular wave functions. II. Overlap populations, bond orders, and covalent bond energies. *J Chem Phys* 23(10):1841
18. Mulliken RS (1955) Electronic population analysis on LCAO-MO molecular wave functions. I. *J Chem Phys* 23(10):1833
19. Reed AE, Weinhold F (1983) Natural bond orbital analysis of near-Hartree-Fock water dimer. *J Chem Phys* 78(6):4066–4073
20. Reed AE, Weinstock RB, Weinhold F (1985) Natural population analysis. *J Chem Phys* 83(2):735
21. Bader RFW (1985) Atoms in molecules. *Acc Chem Res* 18(1):9–15
22. Bader RFW (1991) A quantum theory of molecular structure and its applications. *Chem Rev* 91(5):893–928
23. Breneman CM, Wiberg KB (1990) Determining atom-centered monopoles from molecular electrostatic potentials. The need for high sampling density in formamide conformational analysis. *J Comput Chem* 11(3):361–373
24. Singh UC, Kollman PA (1984) An approach to computing electrostatic charges for molecules. *J Comput Chem* 5(2):129–145
25. Besler BH, Merz KM, Kollman PA (1990) Atomic charges derived from semiempirical methods. *J Comput Chem* 11(4):431–439
26. Gasteiger J, Marsili M (1978) A new model for calculating atomic charges in molecules. *Tetrahedron Lett* 19(34):3181–3184
27. Gasteiger J, Marsili M (1980) Iterative partial equalization of orbital electronegativity—a rapid access to atomic charges. *Tetrahedron* 36(22):3219–3228
28. Cho K-H, Kang YK, No KT, Scheraga HA (2001) A fast method for calculating geometry-dependent net atomic charges for polypeptides. *J Phys Chem B* 105(17):3624–3634
29. Olfirerenko AA, Pisarev SA, Palyulin VA, Zefirov NS (2006) Atomic charges via electronegativity equalization: generalizations and perspectives. *Adv Quantum Chem* 51:139–156
30. Shulga DA, Olfirerenko AA, Pisarev SA, Palyulin VA, Zefirov NS (2010) Fast tools for calculation of atomic charges well suited for drug design. *SAR QSAR Environ Res* 19(1–2):153–165
31. Mortier WJ, Ghosh SK, Shankar S (1986) Electronegativity equalization method for the calculation of atomic charges in molecules. *J Am Chem Soc* 108:4315–4320
32. Nistor RA, Polihrivn JG, Müser MH, Mosey NJ (2006) A generalization of the charge equilibration method for nonmetallic materials. *J Chem Phys* 125(9):094108
33. Mathieu D (2007) Split charge equilibration method with correct dissociation limits. *J Chem Phys* 127(22):224103
34. Baekelandt BG, Mortier WJ, Lievens JL, Schoonheydt RA (1991) Probing the reactivity of different sites within a molecule or solid by direct computation of molecular sensitivities via an extension of the electronegativity equalization method. *J Am Chem Soc* 113(18):6730–6734
35. Svobodová Vařeková R, Jiroušková Z, Vaněk J, Suchomel S, Koča J (2007) Electronegativity equalization method: parameterization and validation for large sets of organic, organohalogen and organometal molecule. *Int J Mol Sci* 8:572–582
36. Jiroušková Z, Vařeková RS, Vaněk J, Koča J (2009) Electronegativity equalization method: parameterization and validation for organic molecules using the Merz-Kollman-Singh charge distribution scheme. *J Comput Chem* 30(7):1174–1178
37. Bultinck P, Langenaeker W, Lahorte P, De Proft F, Geerlings P, Van Alsenoy C, Tollenaere JP (2002) The electronegativity equalization method II: applicability of different atomic charge schemes. *J Phys Chem A* 106(34):7895–7901
38. Ouyang Y, Ye F, Liang Y (2009) A modified electronegativity equalization method for fast and accurate calculation of atomic charges in large biological molecules. *Phys Chem Chem Phys* 11(29):6082–6089
39. Bultinck P, Vanholme R, Popelier PLA, De Proft F, Geerlings P (2004) High-speed calculation of AIM charges through the electronegativity equalization method. *J Phys Chem A* 108(46):10359–10366
40. Geidl S, Bouchal T, Raček T, Svobodová Vařeková R, Hejret V, Křenek A, Abagyan R, Koča J (2015) High-quality and universal empirical atomic charges for chemoinformatics applications. *J Cheminf* 7(1):59
41. O'Boyle N, Banck M, James C, Morley C, Vandermeersch T, Hutchison G (2011) Open babel: an open chemical toolbox. *J Cheminf* 3(1):33–47
42. Vainio MJ, Johnson MS (2007) Generating conformer ensembles using a multiobjective genetic algorithm. *J Chem Inf Model* 47(6):2462–2474
43. Svobodová Vařeková R, Koča J (2006) Optimized and parallelized implementation of the electronegativity equalization method and the atom-bond electronegativity equalization method. *J Comput Chem* 3:396–405
44. Feng Z, Chen L, Maddula H, Akcan O, Oughtred R, Berman HM, Westbrook J (2004) Ligand depot: a data warehouse for ligands bound to macromolecules. *Bioinformatics* 20(13):2153–2155
45. Wishart DS, Knox C, Guo AC, Cheng D, Shrivastava S, Tzur D, Gautam B, Hassanali M (2008) Drugbank: a knowledgebase for drugs, drug actions and drug targets. *Nucleic Acids Res* 36(suppl 1):901–906
46. Bolton EE, Wang Y, Thiessen PA, Bryant SH (2008) Pubchem: integrated platform of small molecules and biological activities. *Ann Rep Comput Chem* 4:217–241
47. MJD Powell (2006) The NEWUOA software for unconstrained optimization without derivatives. In: Large-scale nonlinear optimization, pp. 255–297. Springer, Oxford
48. Anderson E, Bai Z, Bischof C, Blackford S, Demmel J, Dongarra J, Du Croz J, Greenbaum A, Hammarling S, McKenney A, Sorensen D (1999) LAPACK users' guide, 3rd edn. Society for Industrial and Applied Mathematics, Philadelphia
49. Open NCI Database, Release 4. <http://cactus.nci.nih.gov/download/nci/>
50. Berman H, Henrick K, Nakamura H (2003) Announcing the worldwide protein data bank. *Nat Struct Mol Biol* 10(12):980–980
51. Sadowski J, Gasteiger J (1993) From atoms and bonds to three-dimensional atomic coordinates: automatic model builders. *Chem Rev* 93:2567–2581

52. MJ Frisch, GW Trucks, HB Schlegel, GE Scuseria, MA Robb, JR Cheeseman, JA Montgomery Jr, T Vreven, KN Kudin, JC Burant, JMMillam, SS Iyengar, J Tomasi, V Barone, B Mennucci, M Cossi, GS Calman, N Rega, GA Petersson, H Nakatsuji, M Hada, M Ehara, K Toyota, R Fukuda, J Hasegawa, M Ishida, T Nakajima, Y Honda, OKitao, H Nakai, M Klene, X Li, JE Knox, HP Hratchian, JB Cross, VBakken, C Adamo, J Jaramillo, R Gomperts, RE Stratmann, O Yazyev, AJ Austin, R Cammi, C Pomelli, JW Ochterski, PY Ayala, K Morokuma, GA Voth, P Salvador, JJ Dannenberg, VG Zakrzewski, S Dapprich, ADDaniels, MC Strain, O Farkas, DK Malick, AD Rabuck, K Raghavachari, JB Foresman, JV Ortiz, Q Cui, AG Baboul, S Clifford, J Cioslowski, BB Stefanov, G Liu, A Liashenko, P Piskorz, I Komaromi, RL Martin, DJ Fox, T Keith, MA Al-Laham, CY Peng, A Nanayakkara, M Challacombe, PMW Gill, B Johnson, W Chen, MW Wong, C Gonzalez, JA Pople, Gaussian09, Revision E.01. <http://www.gaussian.com>
53. Jelfs S, Ertl P, Selzer P (2007) Estimation of pka for druglike compounds using semiempirical and information-based descriptors. *J Chem Inf Model* 47(2):450–459
54. Wang J, Wolf RM, Caldwell JW, Kollman PA, Case DA (2004) Development and testing of a general amber force field. *J Comput Chem* 25(9):1157–1174
55. Bren U, Hodošek M, Koller J (2005) Development and validation of empirical force field parameters for netropsin. *J Chem Inf Model* 45(6):1546–1552
56. Udornmaneethanakit T, Rungrotmongkol T, Bren U, Frecer V, Stanislav M (2009) Dynamic behavior of Avian Influenza A Virus Neuraminidase Subtype H5N1 in Complex with Oseltamivir, Zanamivir, Peramivir, and Their Phosphonate Analogues. *J Chem Inf Model* 49(10):2323–2332
57. Ison J, Rapacki K, Ménager H, Kalař M, Rydza E, Chmura P, Anthon C, Beard N, Berka K, Bolser D, Booth T, Bretaudéau A, Brezovsky J, Casadio R, Cesareni G, Coppens F, Cornell M, Cuccuru G, Davidsen K, Vedova GD, Dogan T, Doppelt-Azeroual O, Emery L, Gasteiger E, Gatter T, Goldberg T, Grosjean M, Grüning B, Helmer-Citterich M, Ienasescu H, Ioannidis V, Jespersen MC, Jimenez R, Juty N, Juvan P, Koch M, Laibe C, Li J-W, Licata L, Mareuil F, Mičetić I, Friberg RM, Moretti S, Morris C, Möller S, Nenadic A, Peterson H, Profitti G, Rice P, Romano P, Roncaglia P, Saidi R, Schafferhans A, Schwämmle V, Smith C, Sperotto MM, Stockinger H, Váfková RS, Tosatto SCE, de la Torre V, Uva P, Via A, Yachdav G, Zambelli F, Vriend G, Rost B, Parkinson H, Longreen P, Brunak S (2016) Tools and data services registry: a community effort to document bioinformatics resources. *Nucleic Acids Res* 44(D1):D38–D47

Submit your manuscript to a SpringerOpen® journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► [springeropen.com](http://springeropen.com)

---

# Appendix: Other Publications

This part contains the publication outcome of the projects I was collaborating on during my PhD studies. In total there are 4 publications sorted as the list by a year in a descending order. Further, each manuscript is represented by its title page.

Koča J, Svobodová Vařeková R, Pravda L, Berka K, Geidl S, Otyepka M: **Structural Bioinformatics Tools for Drug Design** Springer International Publishing, Cham 2016.

Ionescu CM, Sehnal D, Falginella F L, Pant P, Pravda L, Bouchal T, Svobodová Vařeková R, Geidl S, Koča J: **AtomicChargeCalculator: Interactive Web-based calculation of atomic charges in large biomolecular complexes and drug like molecules** *J Cheminform* 2015, **7**:50.

Sehnal D, Svobodová Vařeková R, Pravda L, Ionescu CM, Geidl S, Horský V, Jaiswal D, Wimmerová M, Koča J: **ValidatorDB: database of up-to-date validation results for ligands and non-standard residues from the Protein Data Bank** *Nucleic Acids Res* 2015, **43**:D368–D375.

Svobodová Vařeková R, Jaiswal D, Sehnal D, Ionescu CM, Geidl S, Pravda L, Horský V, Wimmerová M, Koča J: **MotiveValidator: interactive web-based validation of ligand and residue structure in biomolecular complexes** *Nucleic Acids Res* 2014, **42**:W227–W233.

# **Structural Bioinformatics Tools for Drug Design**

## **Extraction of Biologically Relevant Information from Structural Databases**

Jaroslav Koča<sup>1</sup>, Radka Svobodová Vařeková<sup>1</sup>, Lukáš Pravda<sup>1</sup>, Karel Berka<sup>2</sup>,  
Stanislav Geidl<sup>1</sup>, David Sehnal<sup>1</sup>, Michal Otyepka<sup>2</sup>

<sup>1</sup> National Centre for Biomolecular Research, Masaryk University Brno,  
Faculty of Science National Centre Biomolecular Research, Brno-Bohunice,  
Czech Republic

<sup>2</sup> Regional Centre of Advanced Technologies and Materials, Department of  
Physical Chemistry, Palacký University Olomouc, Faculty of Science Olomouc,  
Czech Republic

Book in series *SpringerBriefs in Biochemistry and Molecular Biology*, published by  
*Springer, Cham, 2016.*

<http://doi.org/10.1007/978-3-319-47388-8>

SPINGER BRIEFS IN BIOCHEMISTRY AND  
MOLECULAR BIOLOGY

Jaroslav Koča

Radka Svobodová Vařeková

Lukáš Pravda

Karel Berka

Stanislav Geidl

David Sehnal

Michal Otyepka

# Structural Bioinformatics Tools for Drug Design

## Extraction of Biologically Relevant Information from Structural Databases



Springer

# **AtomicChargeCalculator: Interactive Web-based calculation of atomic charges in large biomolecular complexes and drug like molecules**

Crina-Maria Ionescu<sup>1</sup>, David Sehnal<sup>1,2,3</sup>, Francesco L. Falginella<sup>1</sup>, Purbaj Pant<sup>2</sup>, Lukáš Pravda<sup>1,2</sup>, Tomáš Bouchal<sup>1,2</sup>, Radka Svobodová Vařeková<sup>1,2</sup>,  
Stanislav Geidl<sup>1,2</sup>, Jaroslav Koča<sup>1,2</sup>

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University  
Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk  
University Brno, Kotlářská 2, 611 37, Brno, Czech Republic.

<sup>3</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00  
Brno, Czech Republic.

*Journal of Cheminformatics 2015, 7:50.*

<https://doi.org/10.1186/s13321-015-0099-x>

**SOFTWARE**

**Open Access**



# AtomicChargeCalculator: interactive web-based calculation of atomic charges in large biomolecular complexes and drug-like molecules

Crina-Maria Ionescu<sup>1†</sup>, David Sehnal<sup>1,2,3†</sup>, Francesco L. Falginella<sup>1</sup>, Purbaj Pant<sup>2</sup>, Lukáš Pravda<sup>1,2</sup>, Tomáš Bouchal<sup>1,2</sup>, Radka Svobodová Vařeková<sup>1,2</sup>, Stanislav Geidl<sup>1,2</sup> and Jaroslav Koča<sup>1,2\*</sup>

## Abstract

**Background:** Partial atomic charges are a well-established concept, useful in understanding and modeling the chemical behavior of molecules, from simple compounds, to large biomolecular complexes with many reactive sites.

**Results:** This paper introduces AtomicChargeCalculator (ACC), a web-based application for the calculation and analysis of atomic charges which respond to changes in molecular conformation and chemical environment. ACC relies on an empirical method to rapidly compute atomic charges with accuracy comparable to quantum mechanical approaches. Due to its efficient implementation, ACC can handle any type of molecular system, regardless of size and chemical complexity, from drug-like molecules to biomacromolecular complexes with hundreds of thousands of atoms. ACC writes out atomic charges into common molecular structure files, and offers interactive facilities for statistical analysis and comparison of the results, in both tabular and graphical form.

**Conclusions:** Due to high customizability and speed, easy streamlining and the unified platform for calculation and analysis, ACC caters to all fields of life sciences, from drug design to nanocarriers. ACC is freely available via the Internet at <http://ncbr.muni.cz/ACC>.

**Keywords:** Conformationally dependent atomic charges, Biomacromolecules , Drug-like molecules, Paracetamol, Benzoic acids, Protegrin, Proteasome, Allostery, Chemical reactivity

## Background

Partial atomic charges are real numbers meant to quantify the uneven distribution of electron density in the molecule, and have been used for decades in theoretical and applied chemistry in order to understand the chemical behavior of molecules. Atomic charges are extensively used in many molecular modeling and chemoinformatics applications. With respect to biomacromolecules, charges can elucidate electrostatic effects critical for long range molecular recognition phenomena, protein folding,

dynamics and allostery, directed adduction of substrates and egression of products in enzymes, ligand binding and complex formation for proteins and nucleic acids, etc. [1–3]. With respect to drug-like molecules, atomic charges provide information related to reactivity and can be used in the prediction of various pharmacological, toxicological or environmental properties [4, 5].

Although, in principle, it is possible to estimate atomic charges based on experimental measurements (e.g., [6, 7]), such calculations are impractical. Most commonly, atomic charges are estimated based on theoretical approaches. Quantum mechanical (QM) approaches first solve the Schrödinger equation [8] and calculate the electron density using a combination of theory level and basis set. They then partition the obtained molecular

\*Correspondence: jaroslav.koca@ceitec.muni.cz

†Crina-Maria Ionescu and David Sehnal contributed equally

<sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk University, Kotlářská 2, 611 37 Brno, Czech Republic

Full list of author information is available at the end of the article

# **ValidatorDB: database of up-to-date validation results for ligands and non-standard residues from the Protein Data Bank**

David Sehnal<sup>1,2,3</sup>, Radka Svobodová Vařeková<sup>1,2</sup>, Lukáš Pravda<sup>1,2</sup>,  
Crina-Maria Ionescu<sup>1</sup>, Stanislav Geidl<sup>1,2</sup>, Vladimír Horský<sup>3</sup>, Deepti Jaiswal<sup>1</sup>,  
Michaela Wimmerová<sup>1,2</sup>, Jaroslav Koča<sup>1,2</sup>

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University  
Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk  
University Brno, Kotlářská 2, 611 37, Brno, Czech Republic.

<sup>3</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00  
Brno, Czech Republic.

*Nucleic Acids Research* 2015, **43**:D368–D375.

<https://doi.org/10.1093/nar/gku1118>

# ValidatorDB: database of up-to-date validation results for ligands and non-standard residues from the Protein Data Bank

David Sehnal<sup>1,2,3,†</sup>, Radka Svobodová Vařeková<sup>1,2,†</sup>, Lukáš Pravda<sup>1,2</sup>, Crina-Maria Ionescu<sup>1</sup>, Stanislav Geidl<sup>1,2</sup>, Vladimír Horský<sup>3</sup>, Deepti Jaiswal<sup>1</sup>, Michaela Wimmerová<sup>1,2</sup> and Jaroslav Koča<sup>1,2,\*</sup>

<sup>1</sup>CEITEC—Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic, <sup>2</sup>National Centre for Biomolecular Research, Faculty of Science, Masaryk University, Kotlářská 2, 611 37 Brno, Czech Republic and <sup>3</sup>Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic

Received August 29, 2014; Revised October 24, 2014; Accepted October 24, 2014

## ABSTRACT

Following the discovery of serious errors in the structure of biomacromolecules, structure validation has become a key topic of research, especially for ligands and non-standard residues. ValidatorDB (freely available at <http://ncbr.muni.cz/ValidatorDB>) offers a new step in this direction, in the form of a database of validation results for all ligands and non-standard residues from the Protein Data Bank (all molecules with seven or more heavy atoms). Model molecules from the wwPDB Chemical Component Dictionary are used as reference during validation. ValidatorDB covers the main aspects of validation of annotation, and additionally introduces several useful validation analyses. The most significant is the classification of chirality errors, allowing the user to distinguish between serious issues and minor inconsistencies. Other such analyses are able to report, for example, completely erroneous ligands, alternate conformations or complete identity with the model molecules. All results are systematically classified into categories, and statistical evaluations are performed. In addition to detailed validation reports for each molecule, ValidatorDB provides summaries of the validation results for the entire PDB, for sets of molecules sharing the same annotation (three-letter code) or the same PDB entry, and for user-defined selections of annotations or PDB entries.

## INTRODUCTION

Validation of biomacromolecular structures has become a very important topic, because some published structures have been found to contain serious errors (1–4). The first step in the validation of biomacromolecules and their complexes is checking the standard building blocks, namely, standard amino acids and nucleotides. The usual procedure is to evaluate specific properties of each residue (e.g. electron density, atom clashes, bond lengths, bond angles, torsion angles, etc.). Various software tools have been developed to perform such analyses, e.g. WHAT-CHECK (5), PROCHECK (6), MolProbity (7) and OOPS (8).

The next key step is the validation of ligands and non-standard residues in biomacromolecular structures, which can be performed in a similar manner as for standard residues (focus on electron density, atom clashes, etc.). An example of software specialized on this type of validation is ValLigURL (9). This approach was also added to several software tools focused on the validation of standard residues (Mogul (10), Coot (11), PHENIX (12)).

A different ligand validation approach, which can be denoted as validation of annotation, was developed later. The goal of this approach is to evaluate if the ligand or non-standard residue is annotated correctly (i.e. if its structure corresponds to the three-letter code it was assigned in the Protein Data Bank (PDB) file format). Specifically, the topology and stereochemistry of the validated molecule are compared to those of a reference molecule (model), and any differences found are reported. The first software tool implementing this methodology has been pdb-care (13), a tool specialized on carbohydrates. The next step has been MotiveValidator (14), which allows validation of all ligands and residues, performs basic validation analyses and reports

\*To whom correspondence should be addressed. Tel: +420 54949 4947; Fax: +420 54949 2556; Email: Jaroslav.Koca@ceitec.muni.cz  
†The authors wish it to be known that, in their opinion, the first two authors should be regarded as Joint First Authors.

Svobodová Vařeková R, Jaiswal D, Sehnal D, Ionescu CM, Geidl S, Pravda L, Horský V, Wimmerová M, Koča J: *Nucleic Acids Res* 2014, **42**:W227–W233.

## MotiveValidator: interactive web-based validation of ligand and residue structure in biomolecular complexes

Radka Svobodová Vařeková<sup>1,2</sup>, Deepti Jaiswal<sup>1</sup>, David Sehnal<sup>1,2,3</sup>, Crina-Maria Ionescu<sup>1</sup>, Stanislav Geidl<sup>1,2</sup>, Lukáš Pravda<sup>1,2</sup>, Vladimír Horský<sup>3</sup>, Michaela Wimmerová<sup>1,2</sup>, Jaroslav Koča<sup>1,2</sup>

<sup>1</sup> CEITEC – Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic.

<sup>2</sup> National Centre for Biomolecular Research, Faculty of Science, Masaryk University Brno, Kotlářská 2, 611 37, Brno, Czech Republic.

<sup>3</sup> Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic.

*Nucleic Acids Research* 2015, **43**:D368–D375.

<https://doi.org/10.1093/nar/gku1118>

# MotiveValidator: interactive web-based validation of ligand and residue structure in biomolecular complexes

Radka Svobodová Vařeková<sup>1,2,†</sup>, Deepti Jaiswal<sup>1,†</sup>, David Sehnal<sup>1,2,3</sup>, Crina-Maria Ionescu<sup>1</sup>, Stanislav Geidl<sup>1,2</sup>, Lukáš Pravda<sup>1,2</sup>, Vladimír Horský<sup>3</sup>, Michaela Wimmerová<sup>1,2,\*</sup> and Jaroslav Koča<sup>1,2,\*</sup>

<sup>1</sup>CEITEC - Central European Institute of Technology, Masaryk University Brno, Kamenice 5, 625 00 Brno, Czech Republic, <sup>2</sup>National Centre for Biomolecular Research, Faculty of Science, Masaryk University, Kotlářská 2, 611 37 Brno, Czech Republic and <sup>3</sup>Faculty of Informatics, Masaryk University Brno, Botanická 68a, 602 00 Brno, Czech Republic

Received January 30, 2014; Revised May 02, 2014; Accepted May 2, 2014

## ABSTRACT

Structure validation has become a major issue in the structural biology community, and an essential step is checking the ligand structure. This paper introduces MotiveValidator, a web-based application for the validation of ligands and residues in PDB or PDBx/mmCIF format files provided by the user. Specifically, MotiveValidator is able to evaluate in a straightforward manner whether the ligand or residue being studied has a correct annotation (3-letter code), i.e. if it has the same topology and stereochemistry as the model ligand or residue with this annotation. If not, MotiveValidator explicitly describes the differences. MotiveValidator offers a user-friendly, interactive and platform-independent environment for validating structures obtained by any type of experiment. The results of the validation are presented in both tabular and graphical form, facilitating their interpretation. MotiveValidator can process thousands of ligands or residues in a single validation run that takes no more than a few minutes. MotiveValidator can be used for testing single structures, or the analysis of large sets of ligands or fragments prepared for binding site analysis, docking or virtual screening. MotiveValidator is freely available via the Internet at <http://ncbr.muni.cz/MotiveValidator>.

## INTRODUCTION

Validation arose as a major issue in the structural biology community when it became apparent that some published structures contained serious errors (1–6). Various tools for the validation of the protein and nucleic acid 3D structures are well established, such as WHAT\_CHECK (7), PROCHECK (8), MolProbity (9) and OOPS (10).

An essential step in the validation process is checking the ligand structure. Ligands are chemical compounds which form a complex with a biomacromolecule (e.g. sugar, drug, heme) and play a key role in its function. The ligands are also the main source of errors in structures (11,12). Nonetheless, ligand validation is a very challenging task (13), because of the high diversity and nontriviality of their structure and the general lack of information about correct structures. Therefore, early validation tools focused on selected types of ligands (PDB-care (14) focused on carbohydrates) and their scope only widened later (ValLigURL (15)). Ligand validation features were recently added to existing software (e.g. Mogul (16), Coot (17)). New tools such as PHENIX (18) were developed to include ligand validation functionality. However, the functionality of some available tools (i.e. ValLigURL, Mogul, Coot, PHENIX) is aimed at the validation of selected properties (atom clashes, bond lengths, bond angles, etc.) or is limited to a selected type of molecules (e.g. PDB-care validates only carbohydrates).

This article presents the web-based application MotiveValidator, which offers a user-friendly, interactive and platform-independent environment for the validation of ligands and residues in PDB (<http://www.wwpdb.org/docs.html>) or PDBx/mmCIF (19) files provided by the user.

\*To whom correspondence should be addressed. Tel: +420 54949 4947; Fax: +420 54949 2556; Email: Jaroslav.Koca@ceitec.muni.cz  
Correspondence may also be addressed to Michaela Wimmerová. Tel: +420 54949 3805; Fax: +420 54949 2690; Email: michaw@chemi.muni.cz

†The authors wish it to be known that, in their opinion, the first two authors should be regarded as Joint First Authors.

---

# **Curriculum Vitae**