

Data sharing and reuse

lennart martens

@compomics

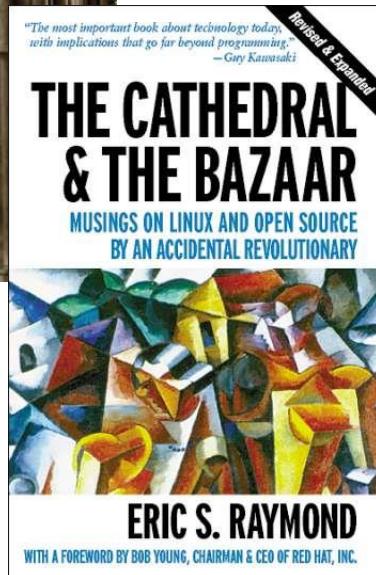
lennart.martens@ugent.be

computational omics and systems biology group

Ghent University and VIB, Ghent, Belgium

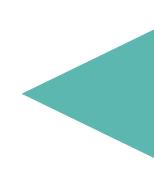


Lincoln Cathedral, Lincolnshire UK



Grand Bazaar, Istanbul, Turkey





Why should we be re-using data?

Four types of (proteomics) data re-use

Unbiased proteome-wide (PT)M discovery as example

A subject of sociological study



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Four types of (proteomics) data re-use

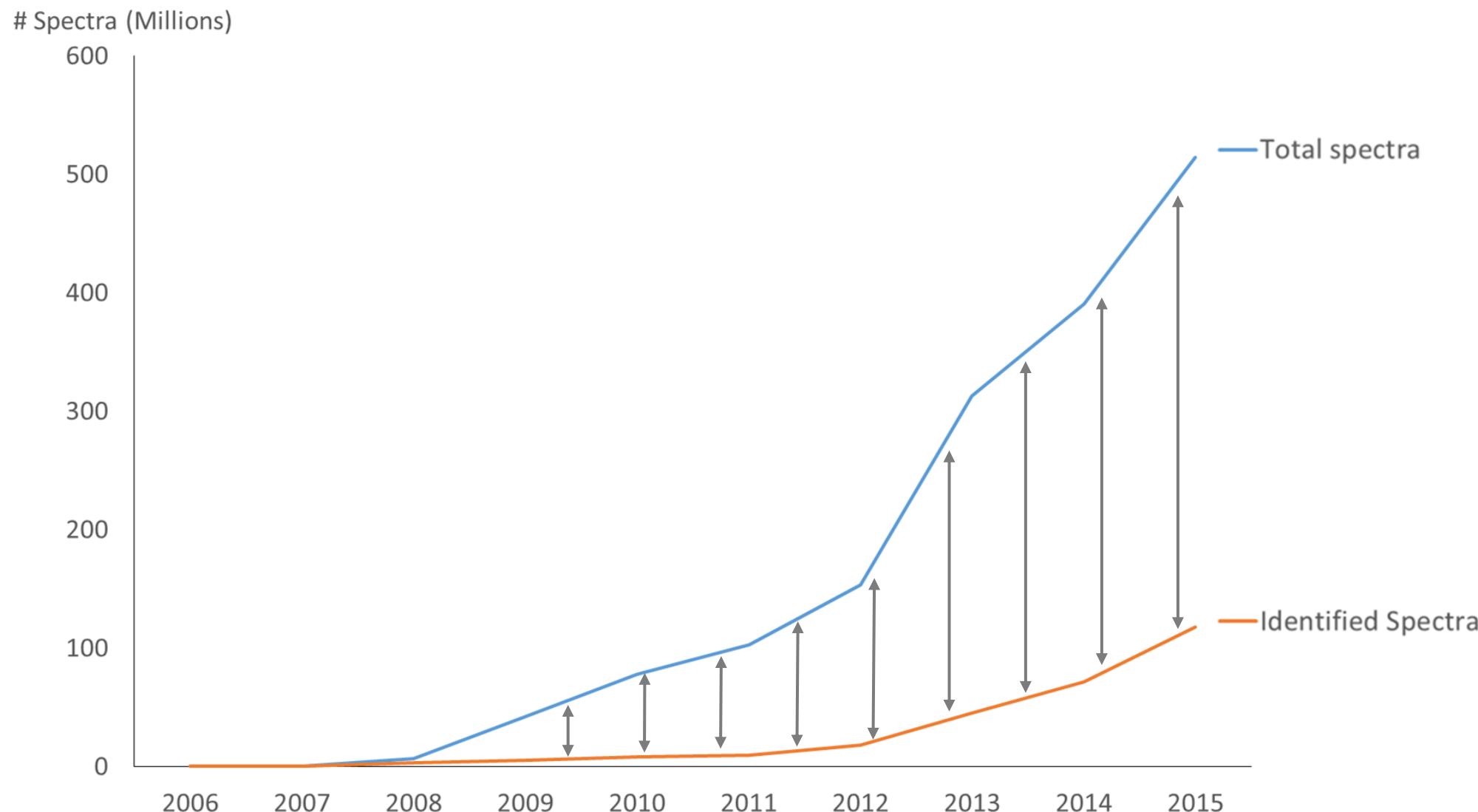
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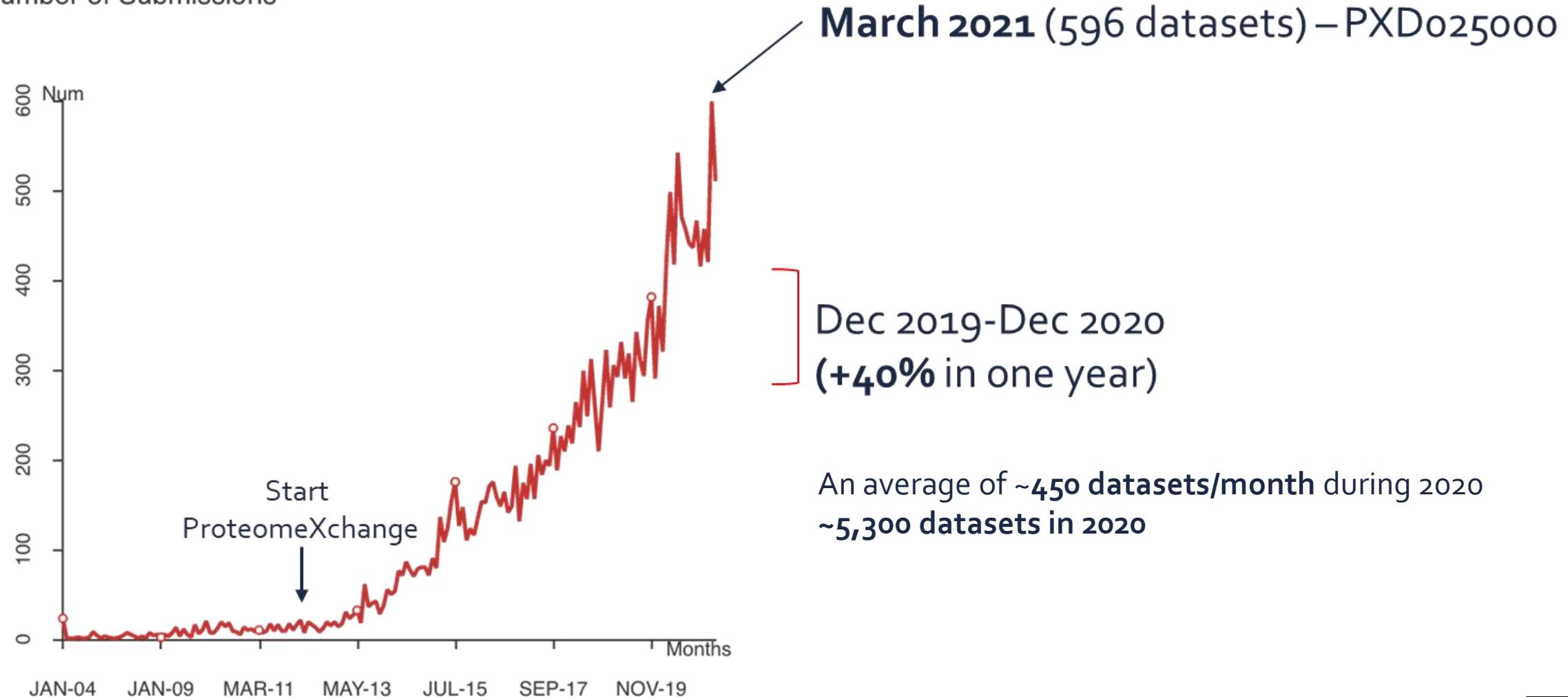
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Mass spectrometry data is high-content, meaning that much more data is acquired than is used in most papers

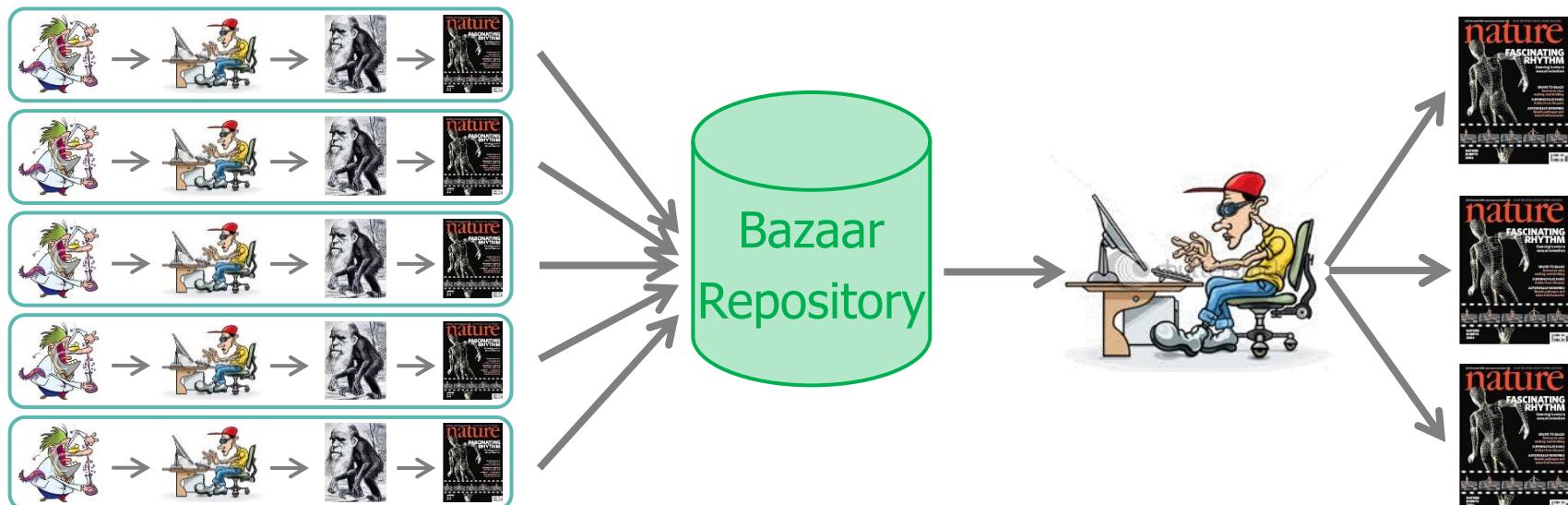
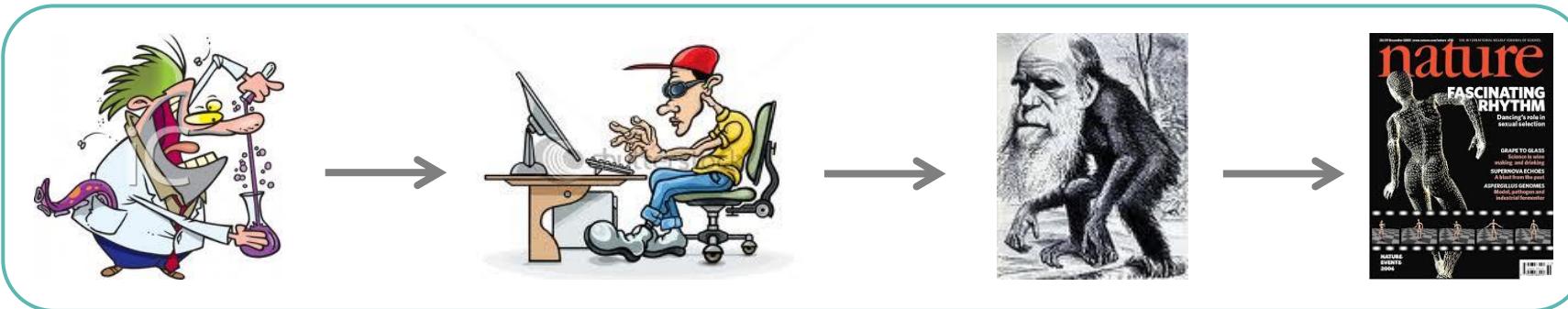


Mass spectrometry is also high throughput, meaning there is lots of data available!

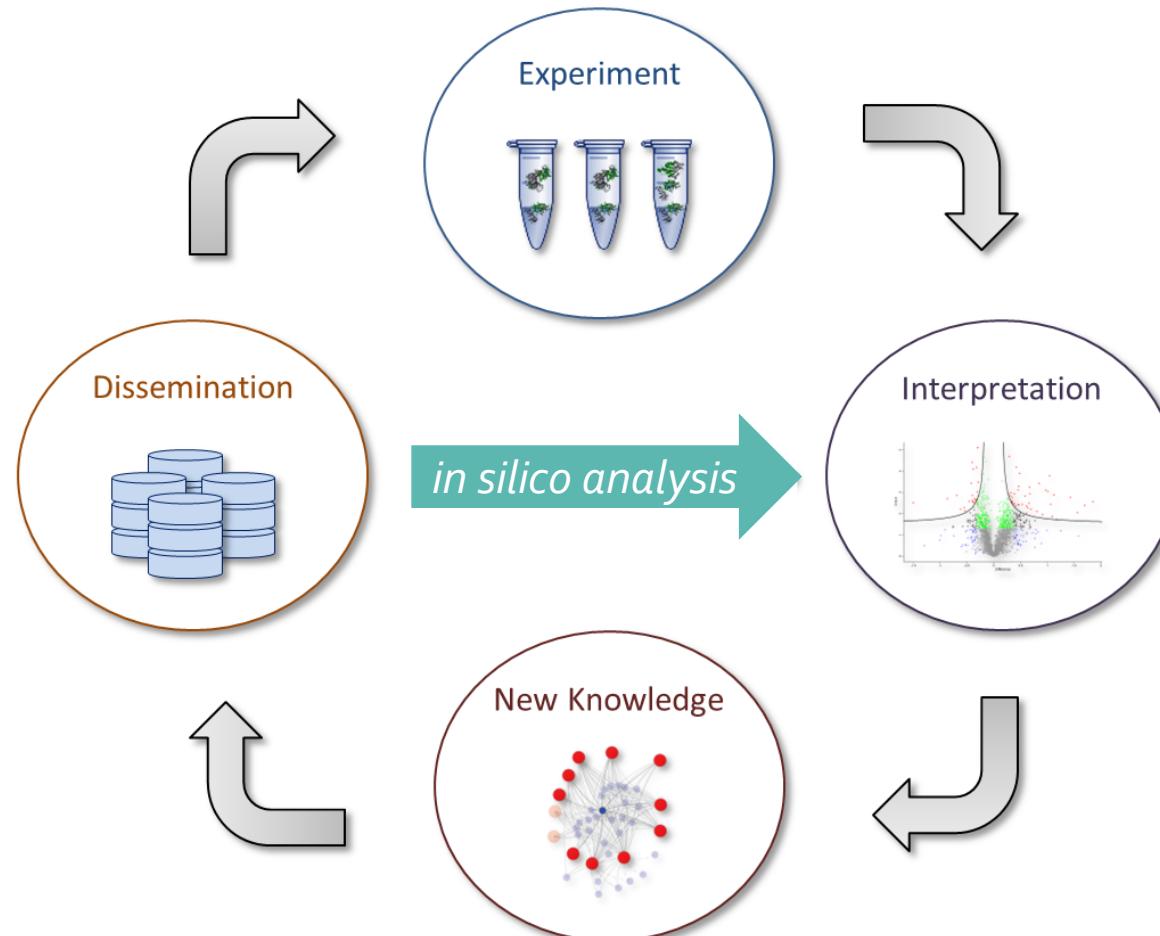
Number of Submissions



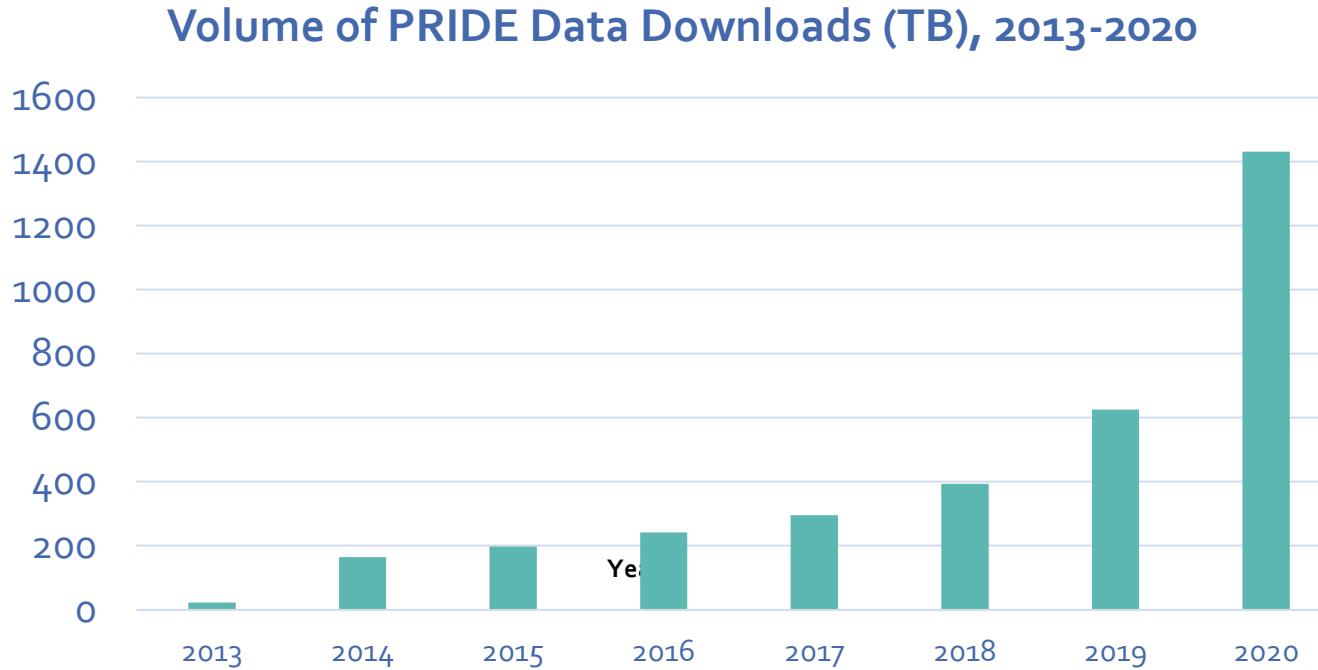
As the volume and content of data increases in a field,
the role of bioinformatics in that field changes as well



The data life cycle shows how *in silico* re-use of data fits in with the overall flow of information

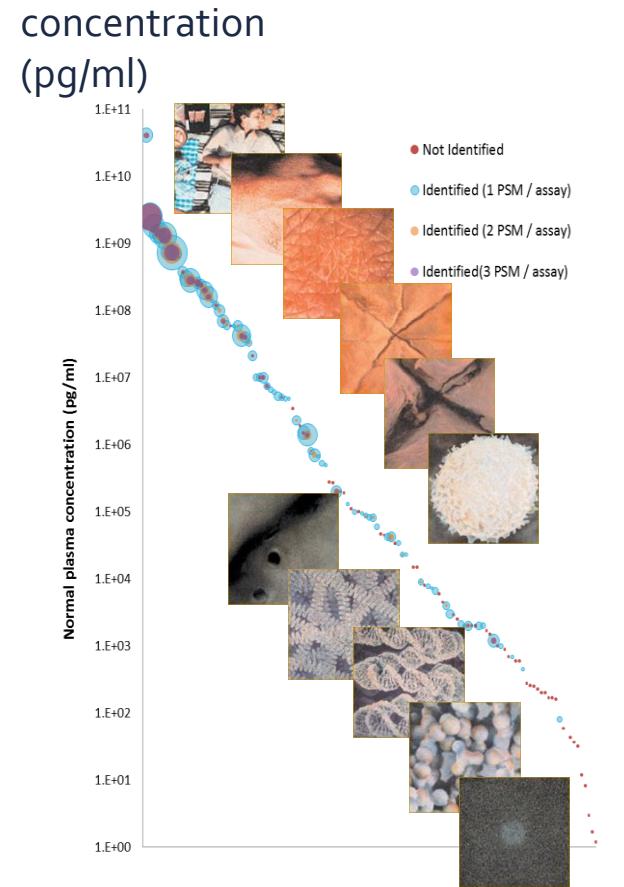
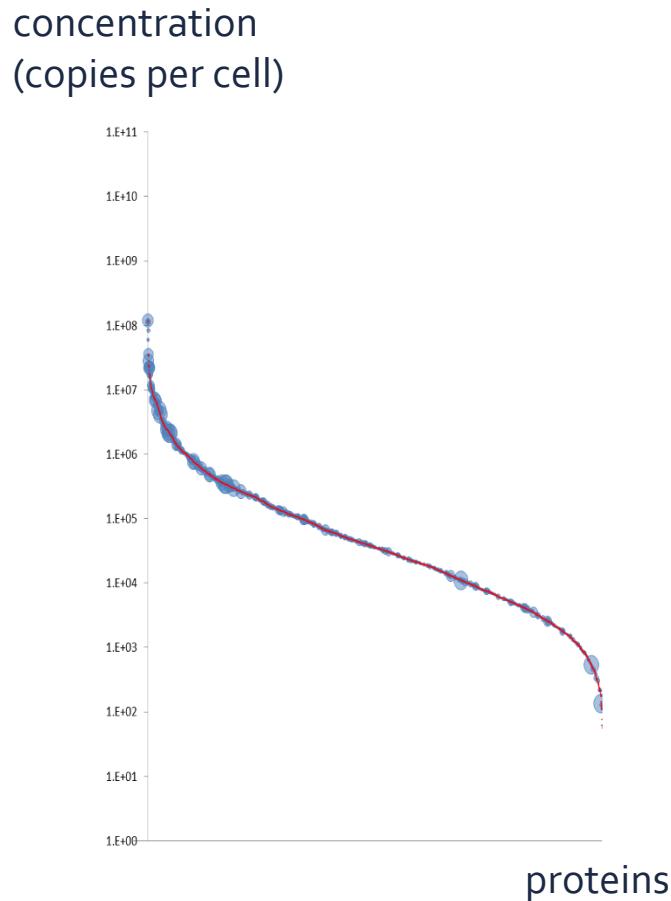
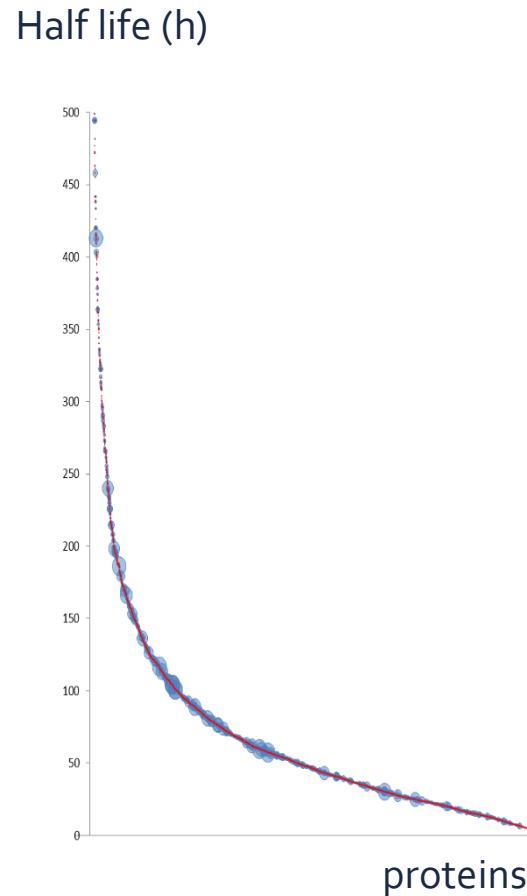


And this is seen in practice in proteomics as data is increasingly re-used



benchmarking
spectral-libraries
sORFs
metaanalyses
Proteogenomics
Verification
PTMs
data-integration
isoforms

Large-scale data reprocessing can harness heterogeneity to dig very deep into the proteome





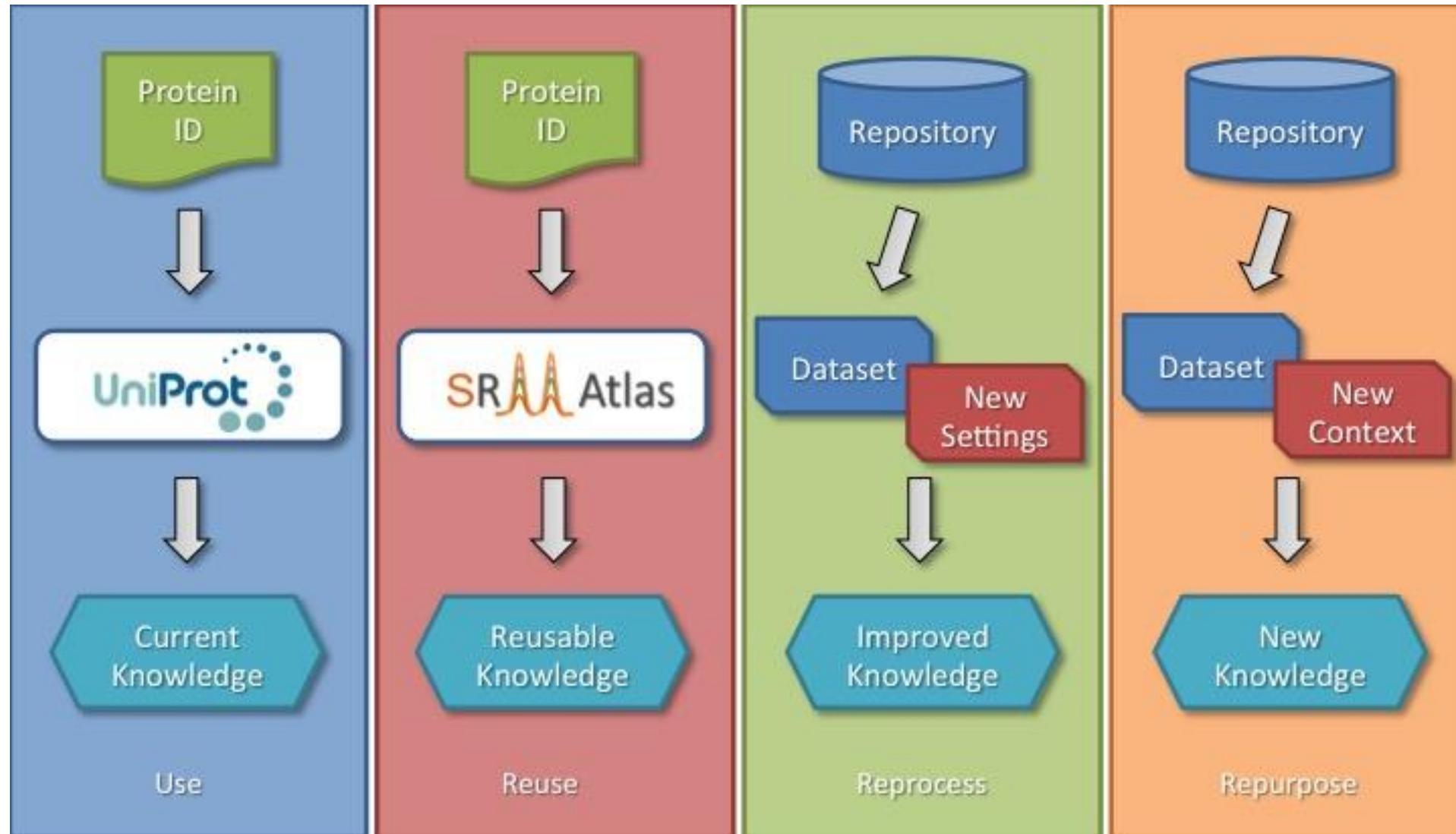
Why should we be re-using data?

Four types of (proteomics) data re-use

Unbiased proteome-wide (PT)M discovery as example

A subject of sociological study

In general, data re-use can take four distinct forms,
all of which are somehow applied in our example





Why should we be re-using data?

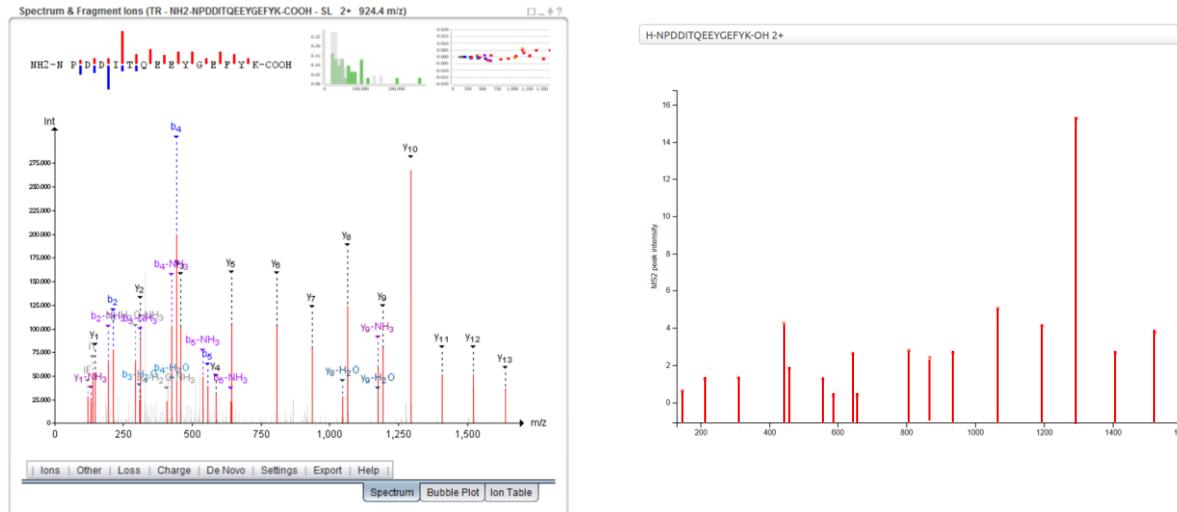
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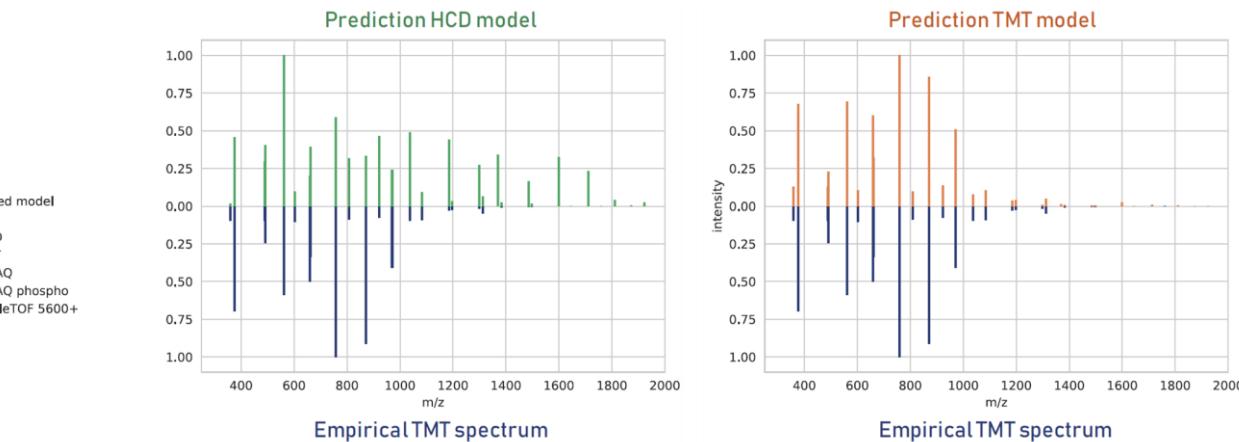
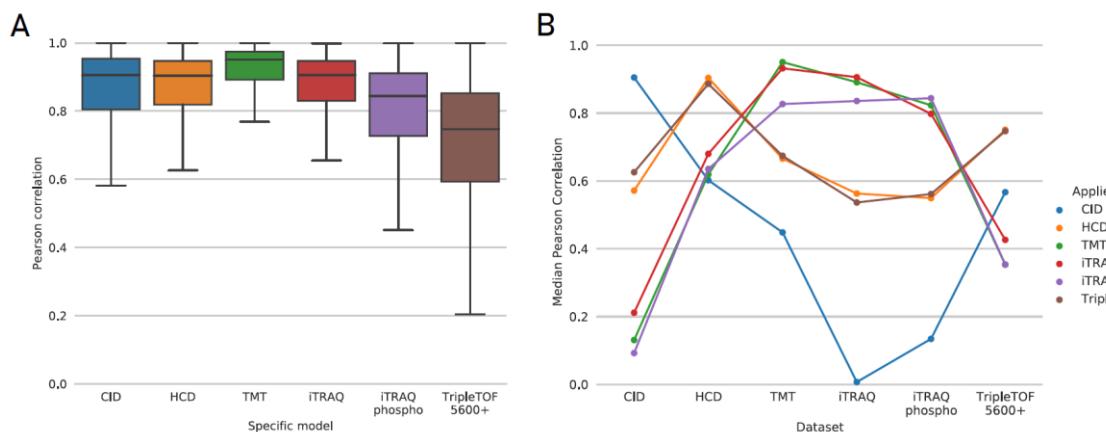
A subject of sociological study

Our MS2PIP fragmentation model accurately predicts peptide behaviour in varying conditions

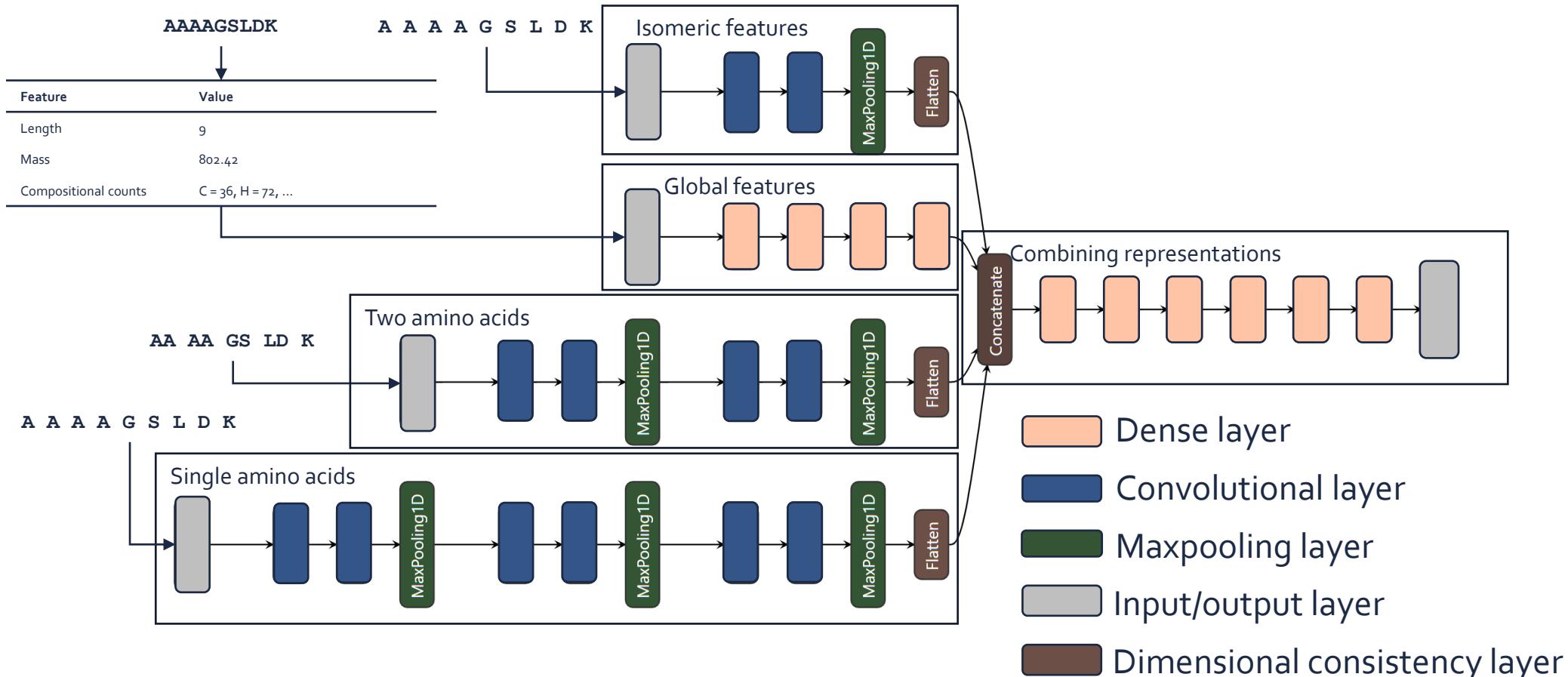
Vaudel, Nat. Biotech., 2015
PeptideShaker



<https://iomics.ugent.be/ms2pip>
Degroeve, Bioinformatics, 2013
Degroeve, Nucleic Acids Research, 2015

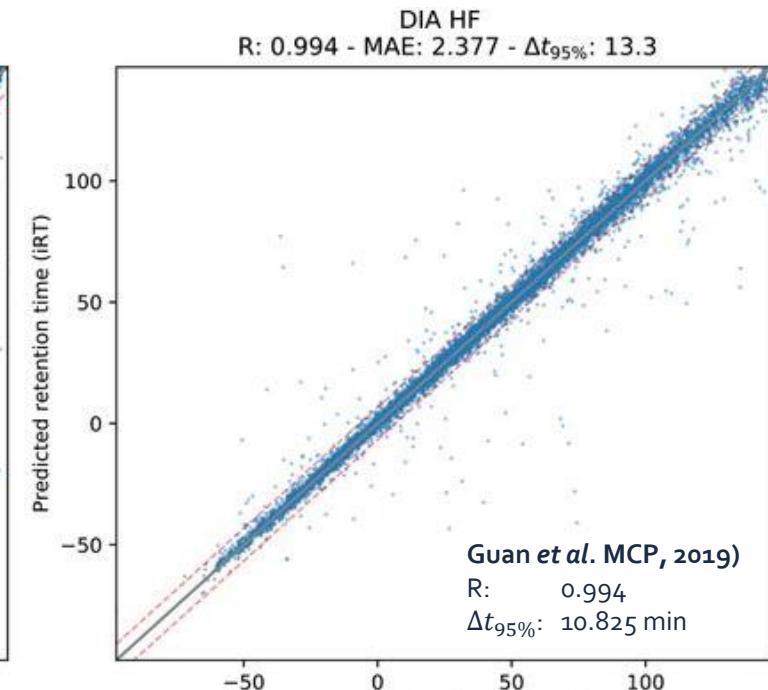
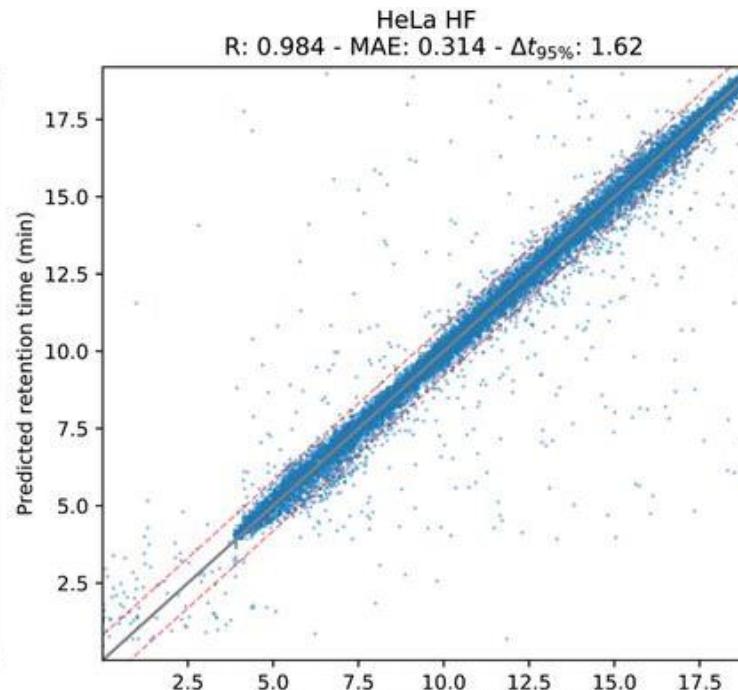
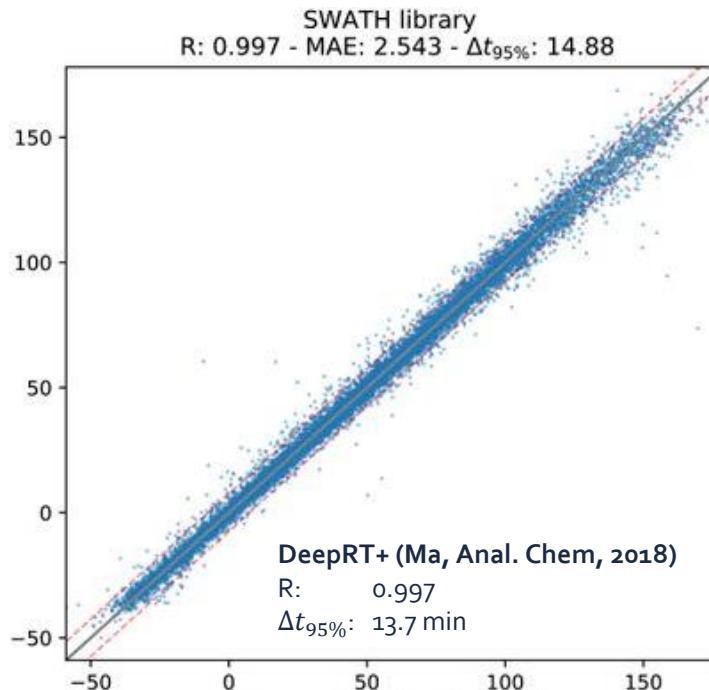


DeepLC is a retention time predictor that can accurately predict retention times of as-yet unseen modifications

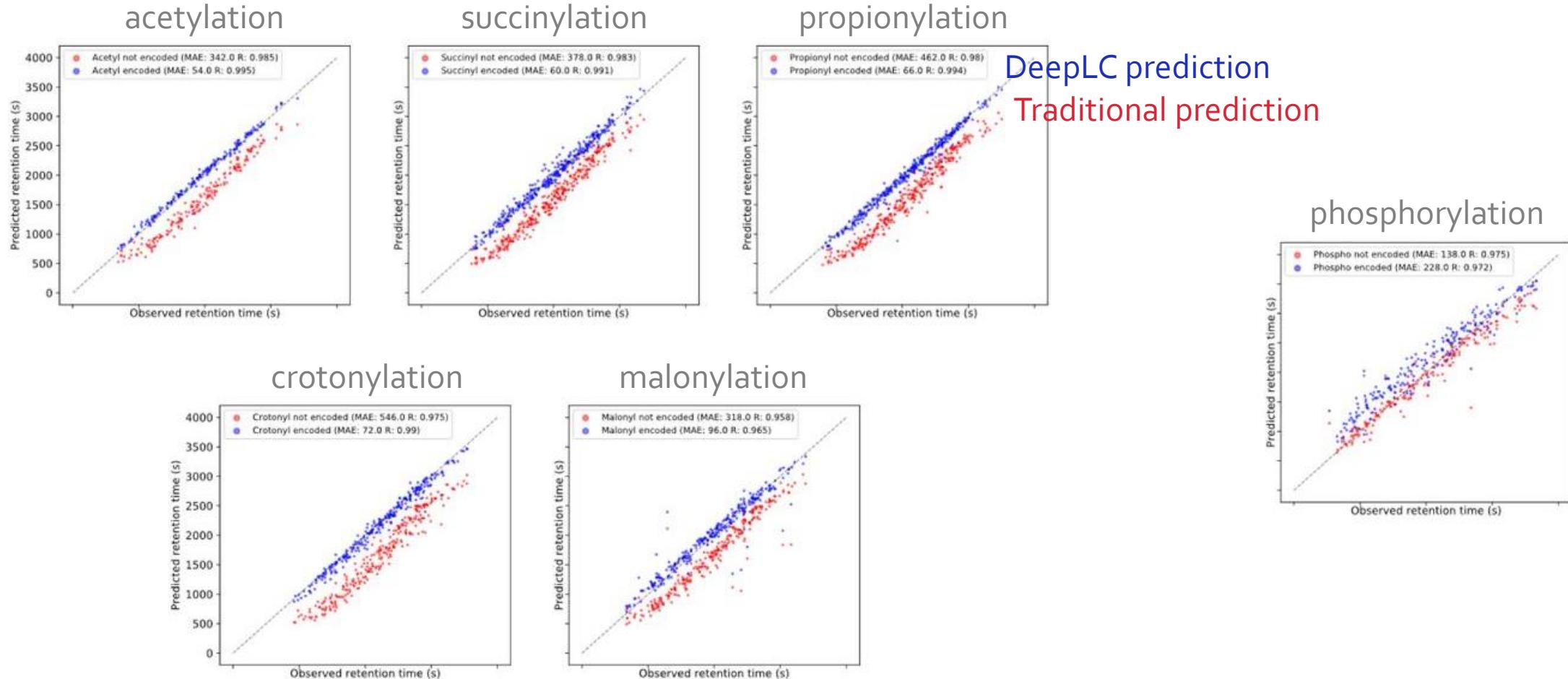


The performance of DeepLC is on par with the state-of-the-art

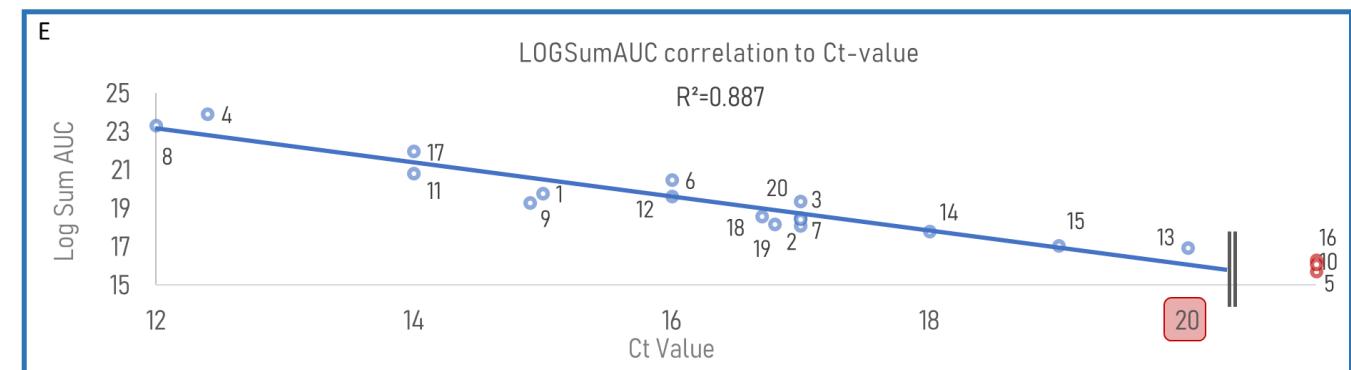
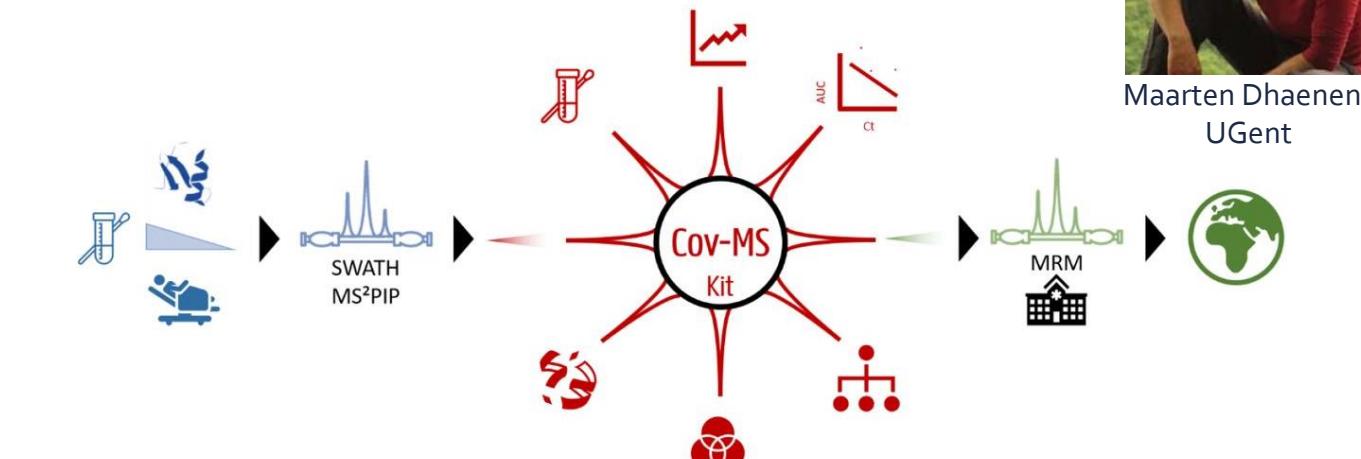
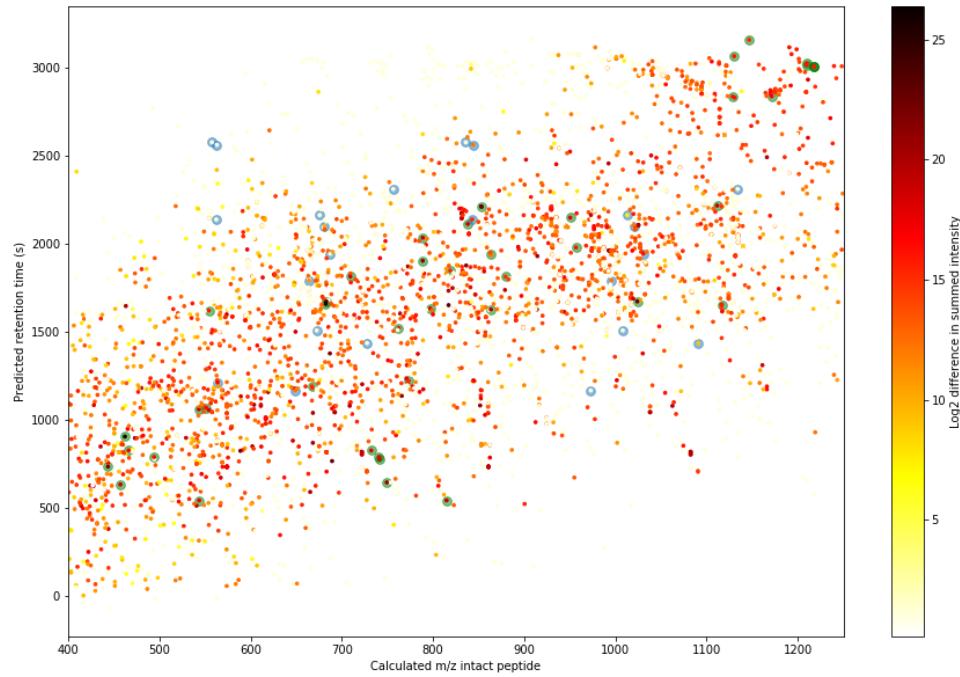
Predicted retention time (min)



DeepLC can accurately predict t_R for many modifications, despite never having seen these before

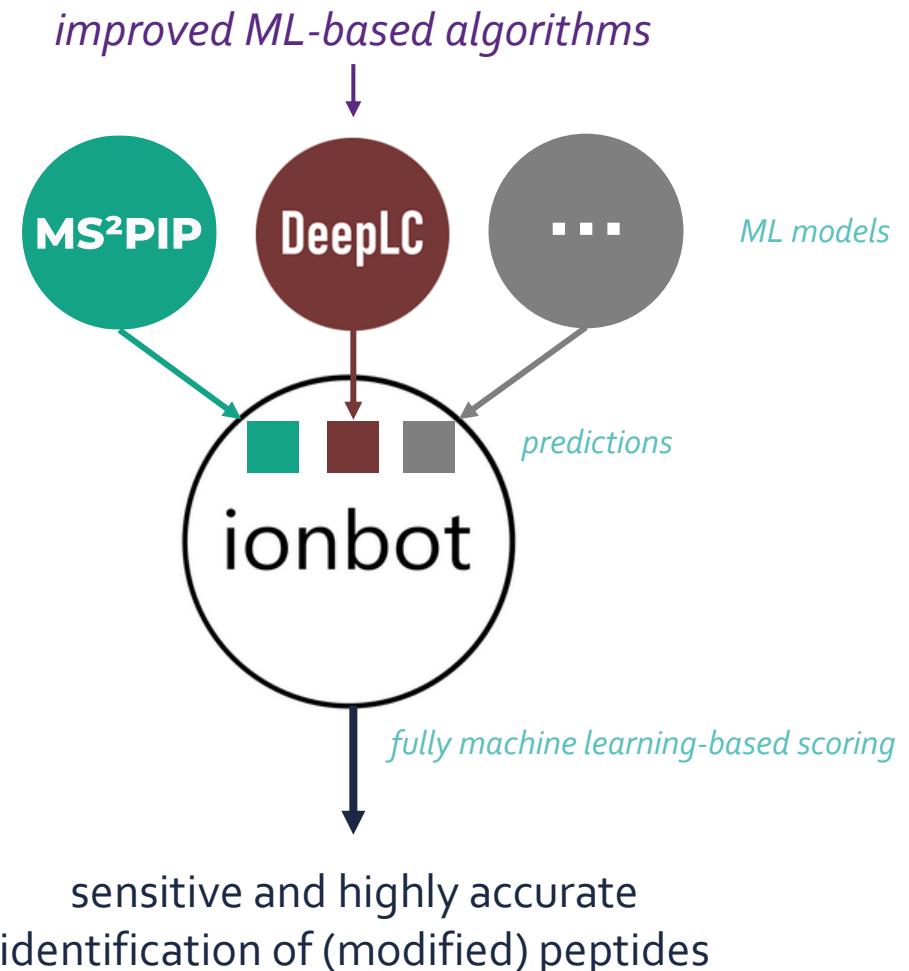


MS2PIP and DeepLC were crucial in the development of a targeted MS-based COVID-19 test that runs in 38 mins



Maarten Dhaenens
UGent

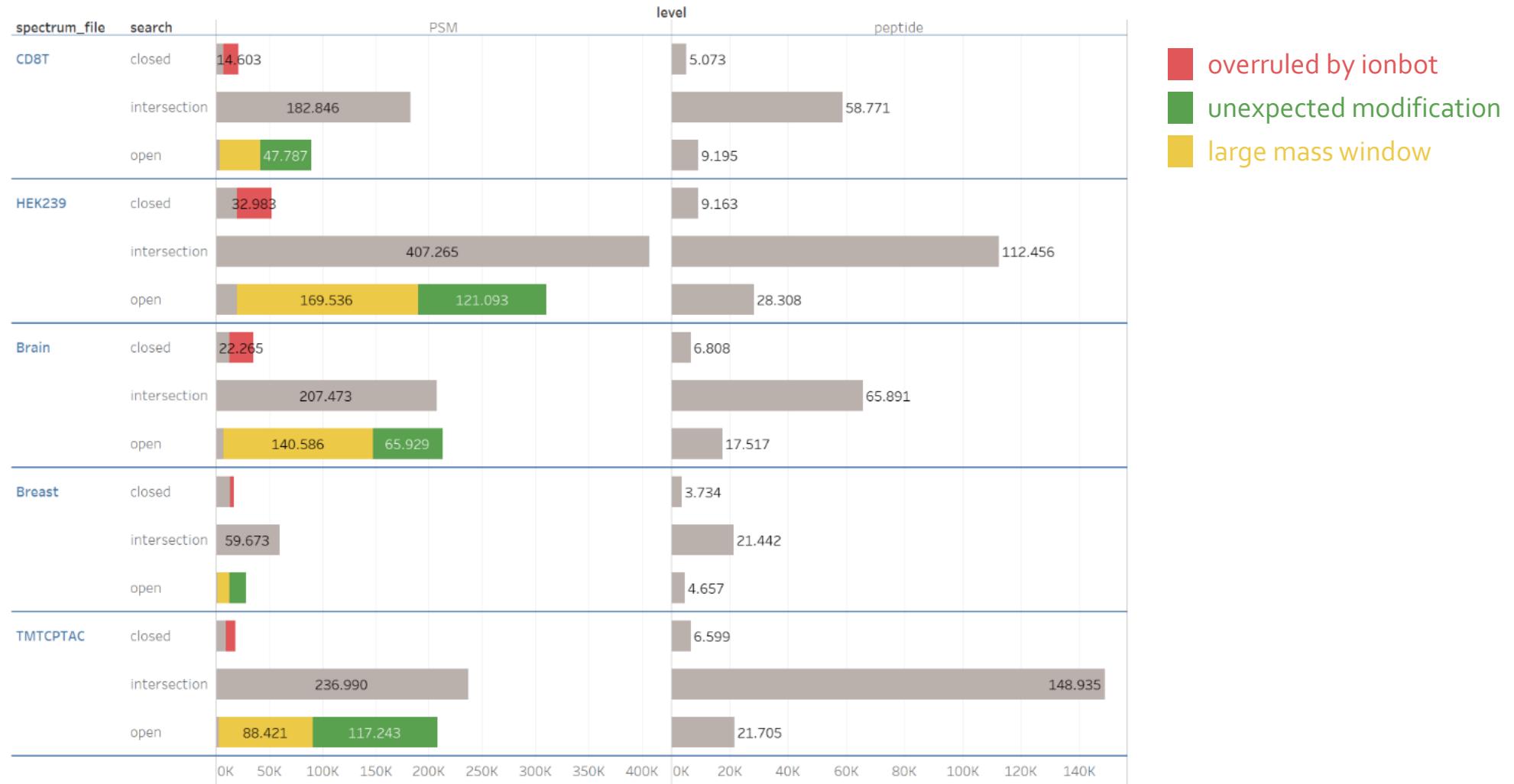
MS²PIP and DeepLC power ionbot, a novel and extensible open modification search engine with high reliability



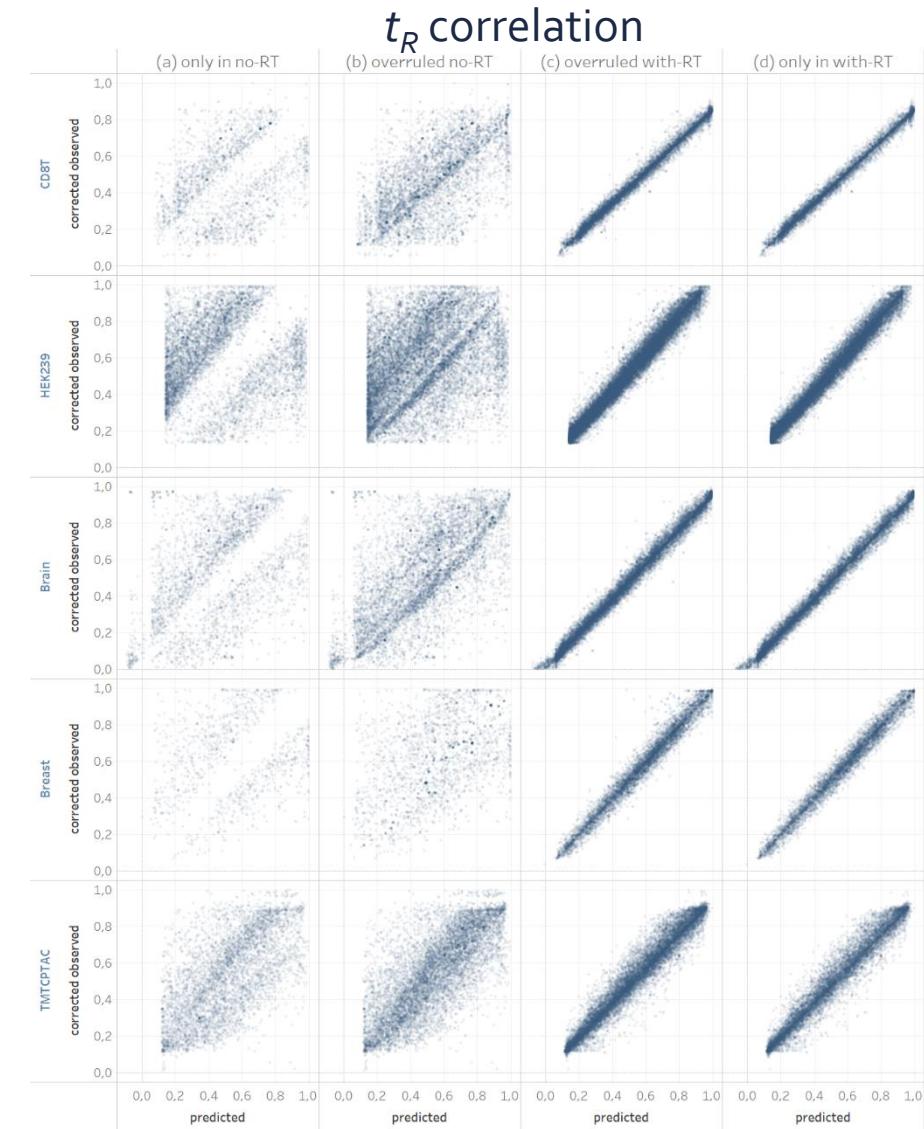
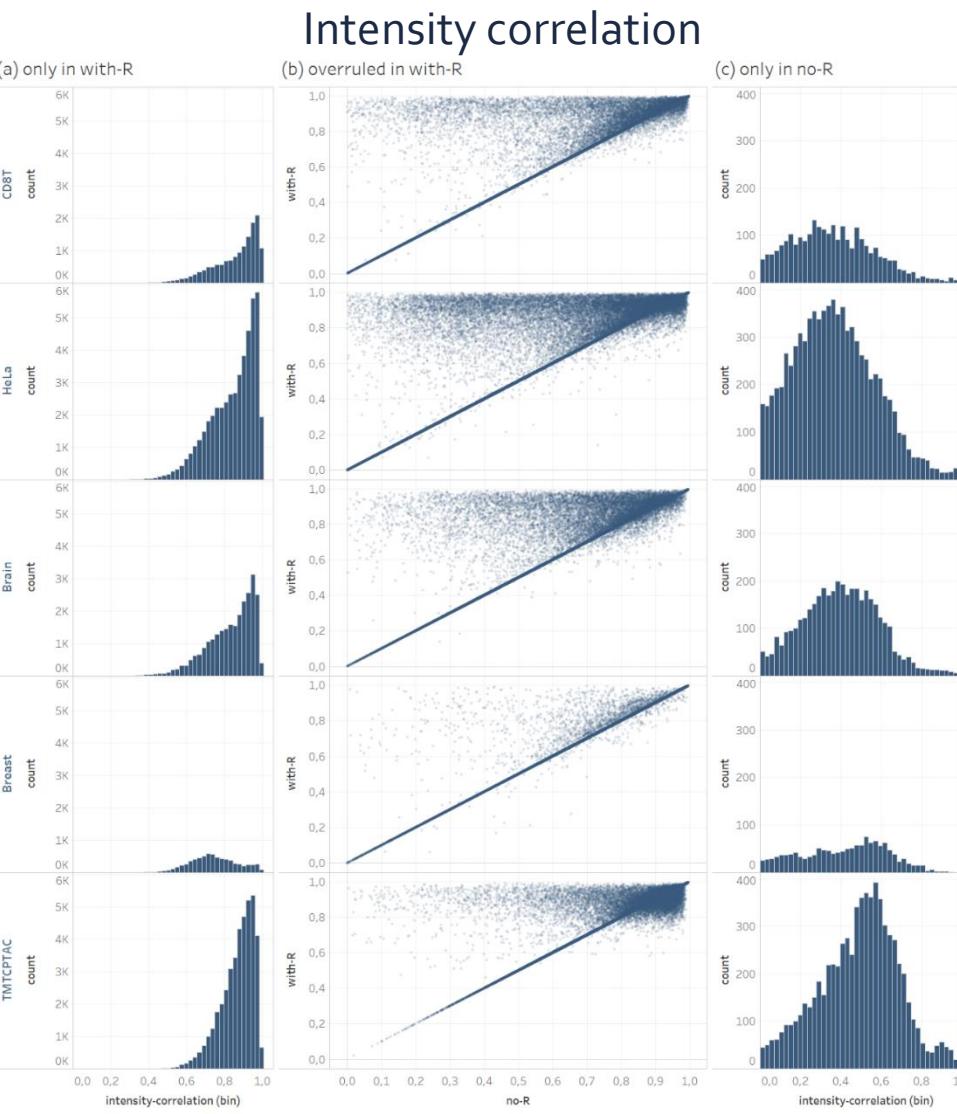
Ionbot shows the value of open modification searches, as well as the value of accurate prediction models



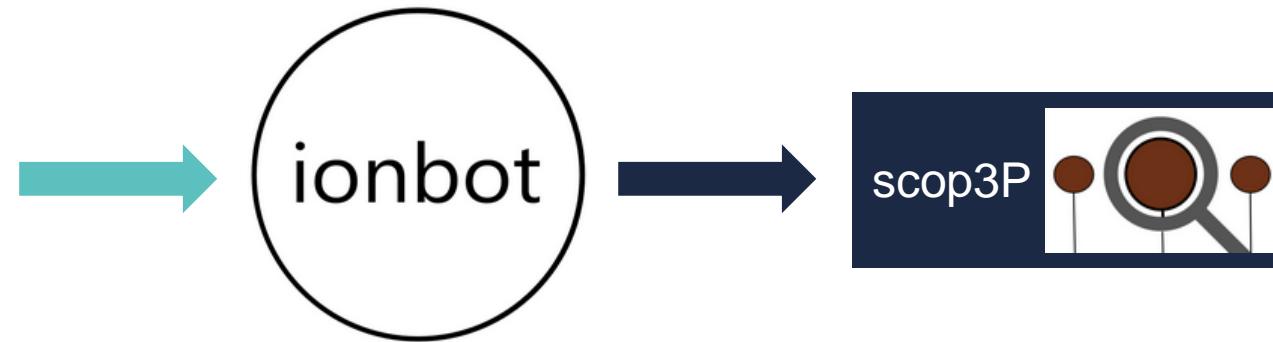
Interestingly, many identifications from the closed search are overruled by ionbot in the open search



Overruled identifications are better, as shown for results obtained with, and without predictions provided to ionbot



We reprocessed a large amount of phosphoproteomics data using ionbot, and made it available through Scop3P



www.ebi.ac.uk/pride

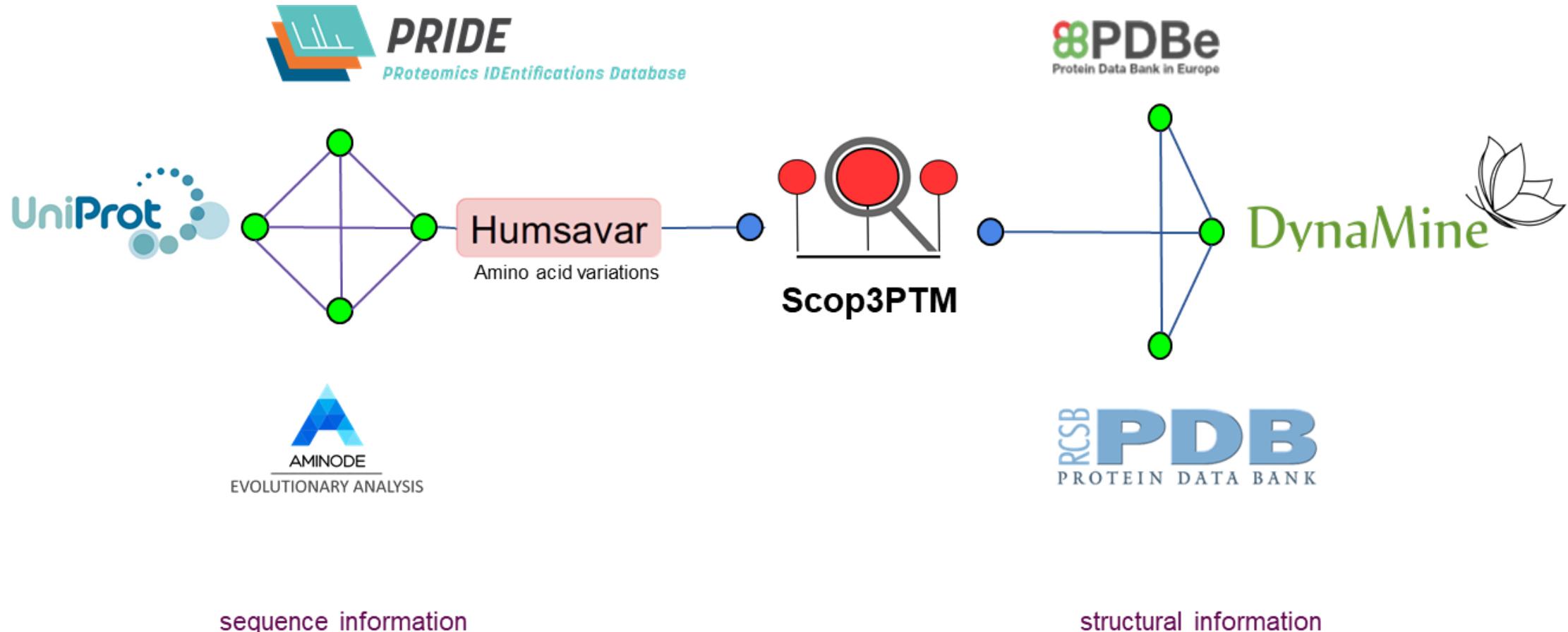
2016 RAW files
60.2 million spectra

1490 UniMod modifications
all possible AA mutations

omics.ugent.be/scop3p

19.2 million PSMs
139 048 P-sites
94 111 sites, PhosphoRS > 0.5
14 261 phosphoproteins

Scop3PTM integrates protein information at the residue level from a variety of resources

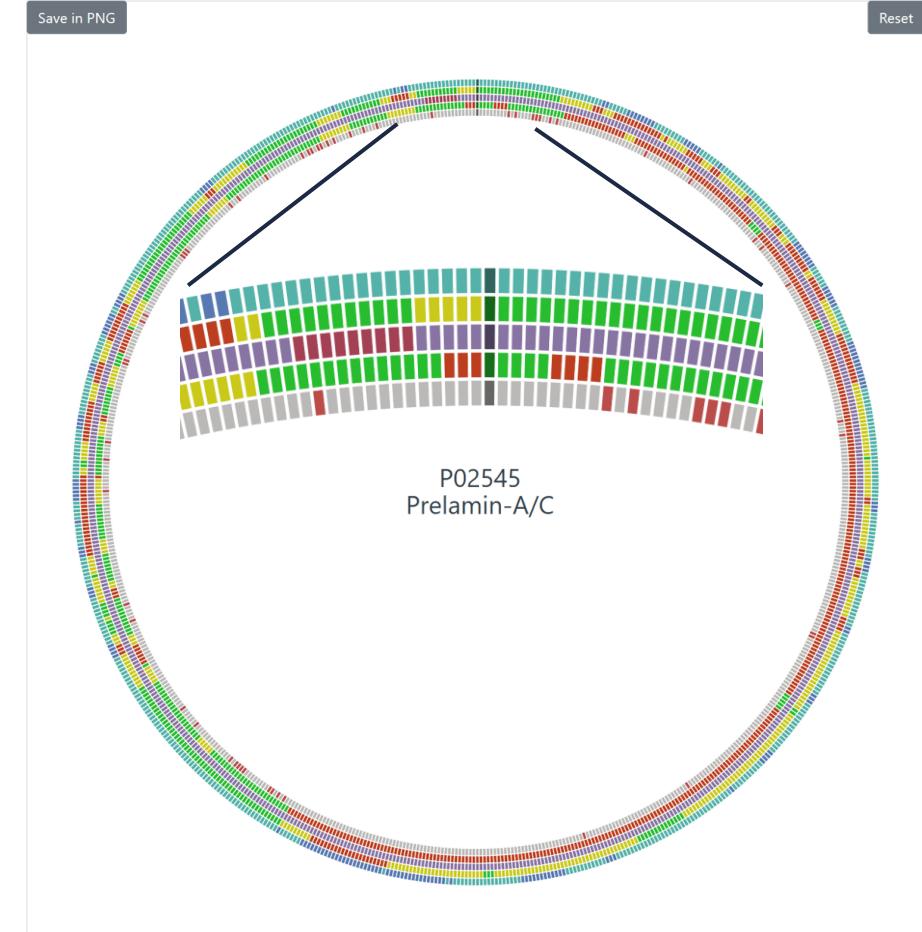
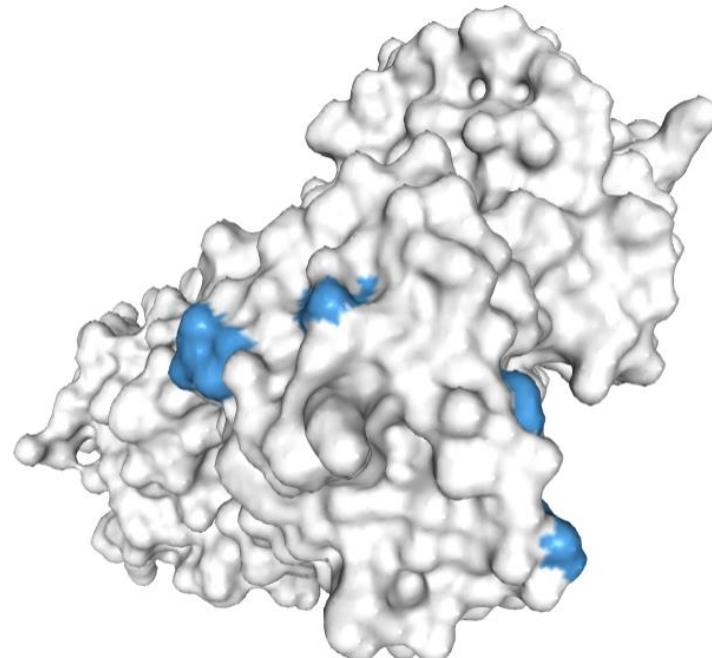


Scop3P shows these results interactively on the web, and presents REST APIs for 3rd party re-use

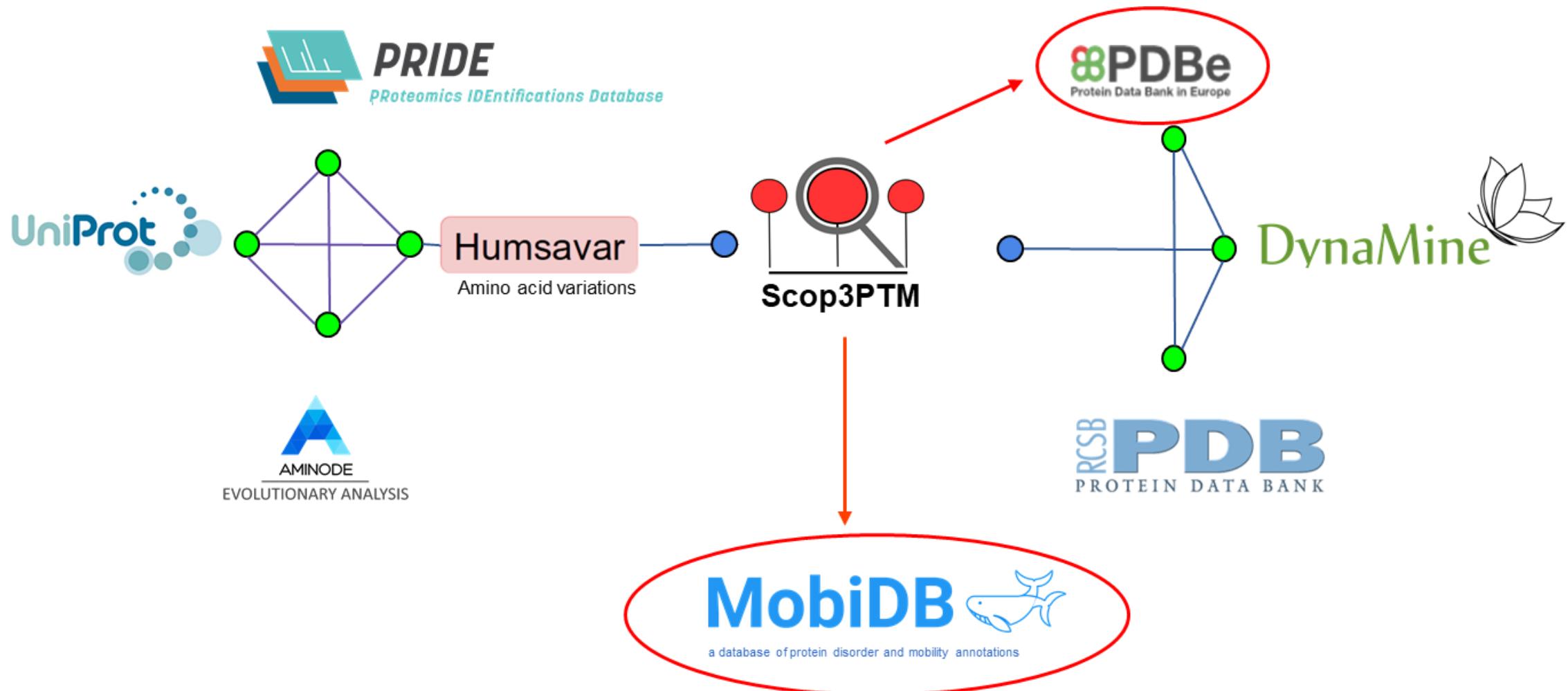
Phospho peptides

Showing 69 results

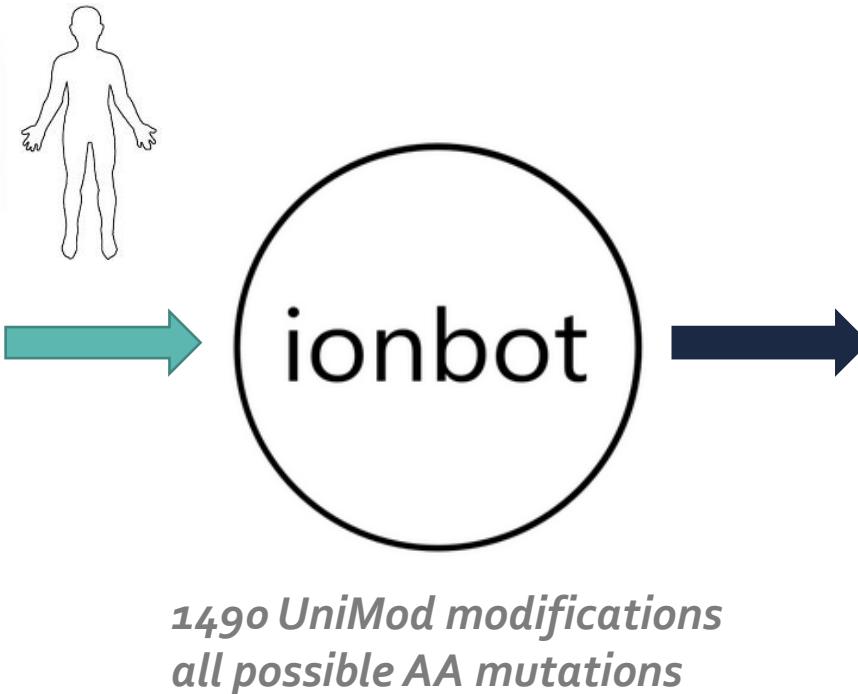
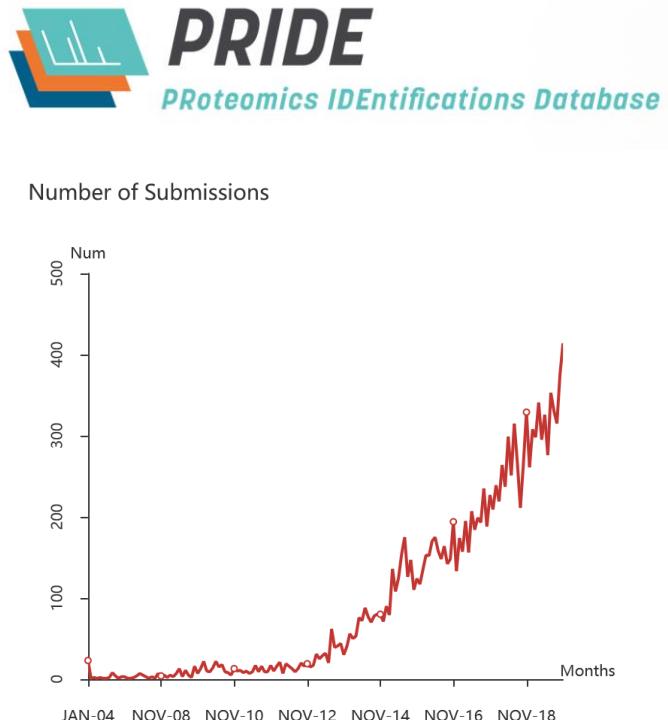
Sequence	Modified position (Swiss-Prot)	Peptide start	Peptide end	Modified position (Peptide)	Number of projects		
RATRSGAQASSTPLSPTR		10	8	25	3	1	
RATRSGAQASSTPLSPTR		12	8	25	5	1	
ProteomeXchange accession		Peptide frequency	Project Title	Species	Submission Type	Publication Date	Tissues
PXD002436	3		Monitoring cellular phosphorylation signaling pathways into chromatin and down to the gene level	Homo sapiens (Human)	COMPLETE	2015-11-09	HeLa cell
SGAQASSTPLSPTR		12	12	25	1		3
SGAQASSTPLSPTR		17	12	25	6		4
SGAQASSTPLSPTR		18	12	25	7		3



Closing the circle, Scop3P itself becomes a resource for use and re-use by others!



And we are now running ionbot on all human spectra contained in the PRIDE database



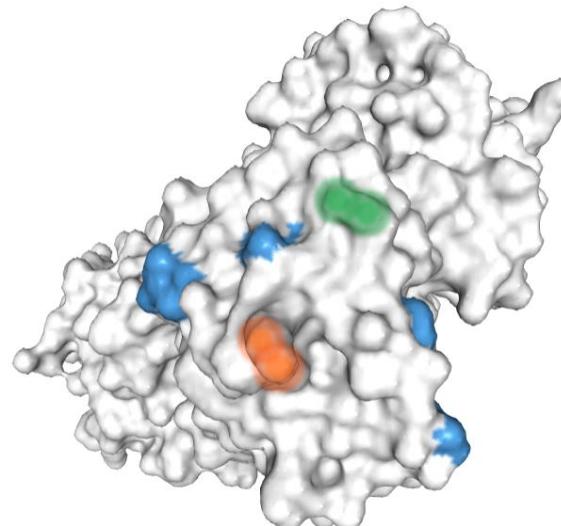
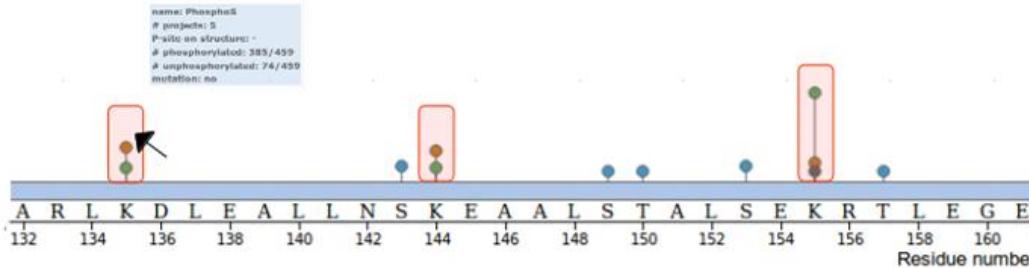
215,046,444 PSMs
742,422 unique peptides
20,246 proteins
444 modifications
4,869,660 modified residues

25 314 RAW files
1 billion spectra

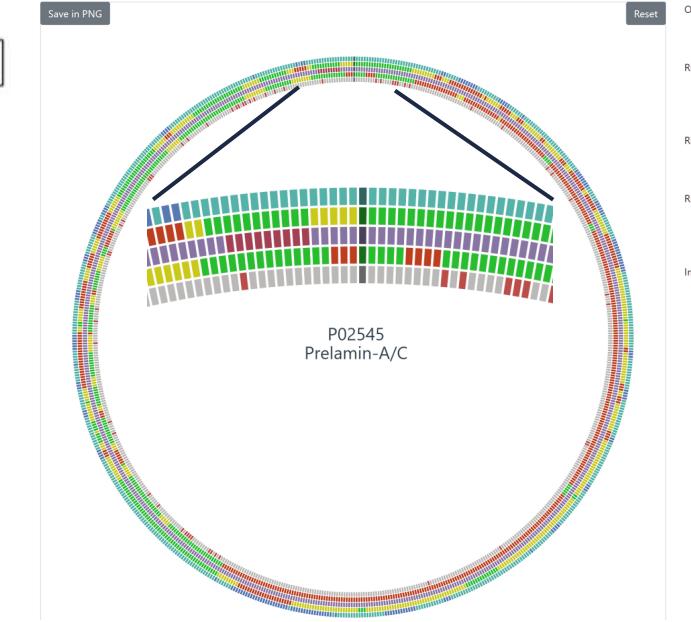


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Scop3PTM will become a proteome-wide PTM detection knowledgebase



- Radio buttons with select/deselect option
- Phosphorylation
 - Ubiquitylation
 - Acetylation
 -
 -
 -
 -
 - Crotonylation



PTM peptides

Sequence	Modified position (Swiss-Prot)	Peptide start	Peptide end	Modified position (Peptide)	Number of projects	
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\$GAQASSTPLSPTR		12	12	25	1	3
SGAQAS\$STPLSPTR		17	12	25	6	4
SGAQAS\$TPLSPTR		18	12	25	7	3



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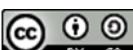


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A sociological take on work of my group highlights the key benefits of open science

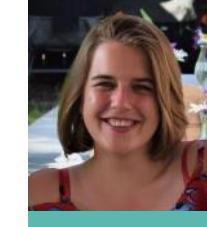
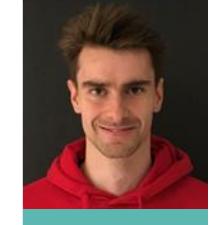
“This desire to reactivate data is widespread, and Klie et al. are not alone in wanting to show that ‘far from being places where data goes to die’ (Klie et al., 2007: 190), **such data collections can be mined for valuable information that could not be obtained in any other way.**”

“In attempting to **reactivate sedimented data** in order to enable its re-use, their first step was ...”

“... they are experiments in seeing, in furnishing ways of seeing how data on proteins could become re-usable, could be reactivated as **collective property rather than the by-product of publication.**”



Gift shop at Chester Cathedral, UK



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omics



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