# Package 'sp.gwas'

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Title Selection probabilities usin	g glmnet for GWAS		
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<b>Description</b> Selection probabilities using generalized linear model with regularization for a SNP data in the hapmap format.			
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#### **Details**

The penalty function of elastic-net is defined as

$$\alpha ||\beta||_1 + (1 - \alpha)||\beta||_2/2,$$

where  $\alpha$  is a mixing proportion of ridge and the lasso, and  $\beta$  is regression coefficients. This penalty is equivalent to the Lasso penalty if alpha=1.

#### Value

A list of data files(genotype, phenotype, etc.), results for selection probabilities, and manhattan plot for multiple traits.

#### References

Zou, H., & Hastie, T. (2005). Regularization and variable selection via the elastic net. Journal of the royal statistical society: series B (statistical methodology), 67(2), 301-320.

qqman\_manhattan

Creates a manhattan plot

### **Description**

Creates a manhattan plot from PLINK assoc output (or any data frame with chromosome, position, and p-value).

#### Usage

```
qqman_manhattan(x, chr = "CHR", bp = "BP", p = "P", snp = "SNP",
  col = c("gray10", "gray60"), col.highlight = "green",
  chrlabs = NULL, suggestiveline = -log10(1e-05),
  genomewideline = -log10(5e-08), highlight = NULL, logp = TRUE,
  annotatePval = NULL, annotateTop = TRUE, ...)
```

# **Arguments**

X	A data.frame with columns "BP," "CHR," "P," and optionally, "SNP."
chr	A string denoting the column name for the chromosome. Defaults to PLINK's "CHR." Said column must be numeric. If you have X, Y, or MT chromosomes, be sure to renumber these 23, 24, 25, etc.
bp	A string denoting the column name for the chromosomal position. Defaults to PLINK's "BP." Said column must be numeric.
p	A string denoting the column name for the p-value. Defaults to PLINK's "P." Said column must be numeric.
snp	A string denoting the column name for the SNP name (rs number). Defaults to PLINK's "SNP." Said column should be a character.
col	A character vector indicating which colors to alternate.
col.highlight	A character vector of colors corresponding to the list of "highlight".

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chrlabs A character vector equal to the number of chromosomes specifying the chromo-

some labels (e.g., c(1:22, "X", "Y", "MT")).

suggestiveline Where to draw a "suggestive" line. Default -log10(1e-5). Set to FALSE to

disable.

genomewideline Where to draw a "genome-wide sigificant" line. Default -log10(5e-8). Set to

FALSE to disable.

highlight A list of character vector of SNPs in your dataset to highlight. These SNPs

should all be in your dataset.

logp If TRUE, the -log10 of the p-value is plotted. It isn't very useful to plot raw p-

values, but plotting the raw value could be useful for other genome-wide plots,

for example, peak heights, bayes factors, test statistics, other "scores," etc.

annotatePval If set, SNPs below this p-value will be annotated on the plot.

annotateTop If TRUE, only annotates the top hit on each chromosome that is below the an-

notatePval threshold.

... Arguments passed on to other plot/points functions

#### Value

A manhattan plot.

#### **Examples**

```
#\dontrun{
#qqman_manhattan(gwasResults)
#}
```

qqman\_qq

Creates a Q-Q plot

# **Description**

Creates a quantile-quantile plot from p-values from a GWAS study.

# Usage

```
qqman_qq(pvector, ...)
```

# **Arguments**

pvector A numeric vector of p-values.
... Other arguments passed to plot()

#### Value

A Q-Q plot.

#### **Examples**

```
## Not run:
qqman_qq(gwasResults$P)
## End(Not run)
```

sp.gwas	Selection probabilities using generalized linear model with regular-
	ization for a SNP data in the hapmap format.

# **Description**

For analysis of high-dimensional genomic data, penalized regression can be a solution to accomodate correlations between predictors. Moreover, selection probabilities do not depend on tuning parameter selection so that it produces a stablity selection. Thresholds are also generated to control the false positives(errors). Thresholds vary with the expected number of false positives to be controlled by the user. Input data is the hapmap formatted SNP data and phenotype data corresponding to SNP data. Output includes files of three types: (1) Matched data files (genotype, numerical, snp info, and phenotype), (2) Results file (selection probabilities and thresholds), (3) Circular manhattan plot with (blue dotted) significant line corresponding to the largest value among user-defined false discoveries.

# Usage

```
sp.gwas(genotype = NULL, phenotype = NULL, input.type = c("object",
   "path"), save.path = "./sp.folder", y.col = 2, y.id.col = 1,
   normalization = TRUE, method = "lasso", family = "gaussian",
   Falsediscovery = c(1, 5, 10), plot.ylim = NULL,
   lambda.min.quantile = 0.5, n.lambda = 10, K = 100, psub = 0.5,
   manhattan.type = "c", plot.name = "", plot.type = "jpg",
   plot.dpi = 300)
```

# Arguments

genotype	Either R object or file path can be considered. A genotype data is a p by (n+11) matrix. It is formatted by hapmap which has (rs, allele, chr, pos) in the first four(1-4) columns, (strand, assembly, center, protLSID, assayLSID, panel, Qcode) in the following seven(5-11) columns. If NULL, user can choose a path in interactive use.
phenotype	Either R object or file path can be considered. A phenotype data is an n by p matrix. Since the first some columns can display attributes of the phenotypes, you should enter the arguments, y.col and y.id.col, which represent the columns of phenotypes to be analyzed and the column of sample ID. If NULL, user can choose a path in interactive use.
input.type	Default is "object". If input.type is "object", obejects of genotype/phenotype will be entered, and if "path", paths of genotype/phenotype will be enterd. If you want to use an object, you have to make sure that the class of each column of genotype data is equal to "character".
save.path	A save.path which has all output files. If there exists save.path, sp.gwas will check if there is an output file. Note that if there is an output RData file in "save.path", sp.gwas will just load the output files(.RData) in there, thereby not providing the results for new "genotype" and "phenotype".
y.col	The columns of phenotypes. At most 4 phenotypes can be considered, because the plot of them will be fine. Default is 2.
y.id.col	The column of sample ID in the phenotype data file. Default is 1.

normalization If TRUE, phenotypes are converted to be normal-shape using box-cox transformation when all phenotypes are positive.

A method of penalized regression. It includes "lasso" for the lasso and "enet" method

for the elastic-net.

family A family of response variable(phenotype). It is "gaussian" for continuous re-

sponse variable, "binomial" for binary, "poisson" for count, etc. Now you can use only the same family for the multi phenotypes. For more details, see the

function(stats::glm). Default is "gaussian".

The expected number of false discovery to be controlled. The larger it is, the Falsediscovery

higher threshold becomes. Default is c(1, 5, 10).

A range of the y-axis. If NULL, automatic range in the y-axis will be provided. plot.ylim

For plot.ylim=c(0,1), the y-axis has a range of 0 and 1.

lambda.min.quantile

A range of lambda sequence. Default is 0.5 (median). If the range is so small that it can have many tied selection probabilities which is 1. To handle with this

problem, you should increase the value of "lambda.min.quantile".

n.lambda The length of lambda sequence. The larger n.lambda, the more detailed lambda

sequence will be.

Κ The number of iterations in resampling when calculating the selection probabil-

ities.

The subsampling proportion. For efficiency, default is 0.5. psub

manhattan.type A type of manhattan plot to be drawn includes circular('c') and rectangular('m').

A name of plot file. plot.name

A type of plot file which includes "jpg", "pdf", "tiff", etc. plot.type

A resolution of plot. If you want to get a high-resolution image, plot.dpi should plot.dpi

be large.

#### **Details**

The penalty function of elastic-net is defined as

$$\alpha ||\beta||_1 + (1-\alpha)||\beta||_2/2,$$

where  $\alpha$  is a mixing proportion of ridge and the lasso, and  $\beta$  is regression coefficients. This penalty is equivalent to the Lasso penalty if alpha=1.

An algorithm of selection probabilities with elastic-net.

0: Let us assume that a genomic data has n samples and p variables.

1: For all  $\Lambda = (\alpha, \lambda)$ , where  $\alpha in[0, 1], \lambda > 0$ .

2: for k=1 to K do.

3: ——— Subsample  $I_k$  with size [n/2]. 4: ——— Compute  $\hat{\beta}_{j}^{\Lambda}(I_k)$  with regularization model.

5: end for  $S_{j}$  (\*\*\*) \*\*Marrogana\*\* 5: end for  $SP_{j}^{\Lambda} = \frac{1}{K} \# \{k \leq K : \hat{\beta}_{j}^{\Lambda}(I_{k}) \neq 0\}.$  7:  $SP_{j} = \max_{\Lambda} SP_{j}^{\Lambda}, \ j = 1, \cdots, p.$  8: return  $SP = (SP_{1}, \cdots, SP_{p}).$ 

#### Value

 $\label{thm:continuous} \mbox{Histogram of original and transformed phenotypes}$ 

Histogram of phenotypes with p-value by Shapiro-test on the top right corner.

myDATA A list of myX, myGD, myGM, myGT, myY, and myY.original(for "gaussian").

sp.res A list of sp.df and threshold.

Circular Manhattan plot

Manhattan plot for the first phenotype is the innermost circle. Colors for chromosome is fixed, so that if you want to change colors, you would edit the R code of sp.manhattan function.

The dataframe with new mean and sum columns

#### Author(s)

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#### **Examples**

```
genotype <- sp.gwas::genotype # load("genotype.rda")</pre>
phenotype <- sp.gwas::phenotype # load("phenotype.rda")</pre>
sp.gwas(genotype = genotype,
        phenotype = phenotype,
        input.type = c("object", "path")[1],
        save.path = "./EXAMPLE",
        y.id.col = 1,
        y.col = 2:4,
        normalization = FALSE,
        method="lasso",
        family="gaussian",
        Falsediscovery = c(1,5,10),
        plot.ylim = NULL,
        lambda.min.quantile = 0.5,
        n.lambda = 10,
        K = 100,
        psub = 0.5,
        manhattan.type = c("c", "r")[1],
        plot.name = "Test",
        plot.type = "jpg",
        plot.dpi = 300)
results <- readxl::read_xlsx("./EXAMPLE/[2]sp.results.xlsx")</pre>
class(results$chr) <- "numeric"</pre>
class(results$pos) <- "numeric"</pre>
thresholds <- readxl::read_xlsx("./EXAMPLE/[2]sp.thresholds.xlsx")</pre>
highlight1 <- results$rs[results$v1>thresholds$v1[1]]
highlight10 <- setdiff( results$rs[results$v1>thresholds$v1[3]], highlight1 )
jpeg("./EXAMPLE/Manhattan_from_qqman.jpeg", width=12, height=5, unit="in", res=600)
qqman_manhattan(results,
                chr="chr", bp="pos", snp="rs", p="v1", logp=FALSE,
                suggestiveline = thresholds$v1[1],
                genomewideline = thresholds$v1[3],
                highlight = list(highlight1, highlight10),
                col.highlight = c("blue", "red"),
                ylab="Selection probabilities", ylim=c(0,1))
```

dev.off()

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