

# Malaria-transmitting *Anopheles* in the Colombian Pacific region

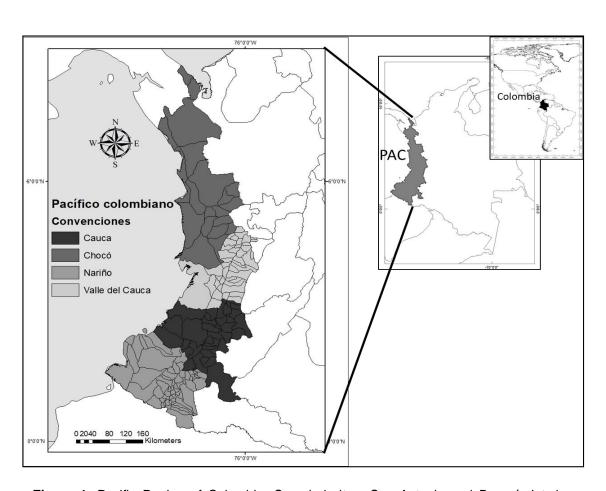
#### Estefani Piedrahita<sup>1</sup>, Laura López<sup>1</sup>, Santiago Pemberty<sup>1</sup>, Margarita M. Correa<sup>1</sup>

<sup>1</sup> Grupo de Microbiología Molecular, Escuela de Microbiología, Universidad de Antioquia, Medellín, Colombia.

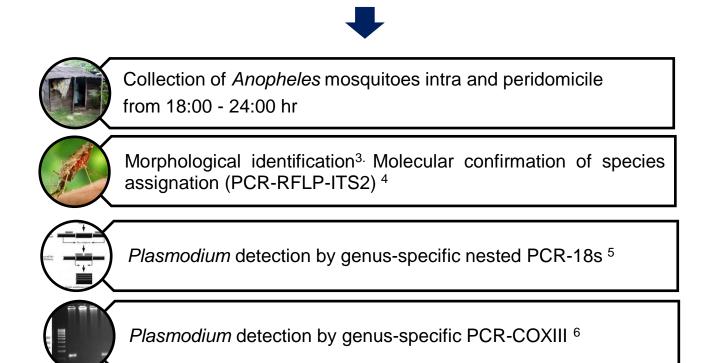
#### **INTRODUCTION**

Malaria remains a significant public health concern in Colombia, and particularly, in the most malaria-endemic Pacific region<sup>1</sup>. The detection of *Plasmodium* infected mosquitoes is an important parameter for the incrimination of vectors in malaria transmission<sup>2</sup>. Therefore, the aim of this study was to detect natural infection by *Plasmodium* parasites in *Anopheles* mosquitoes collected in localities of the Colombian Pacific region.

## **METHODOLOGY**



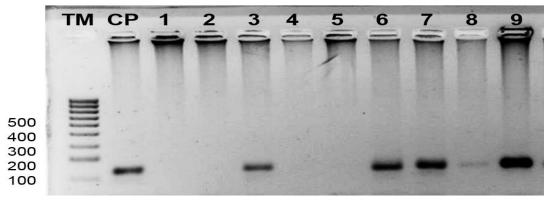
**Figure 1**. Pacific Region of Colombia. Sampled sites: San Antonio and Basurú, Istmina (Department of Chocó); Buchely and Inguapí del Carmen, Tumaco (Department of Nariño); La Playa and Salahondita, Francisco Pizarro (Department of Nariño); Córdoba and Zacarías, Buenaventura (Department of Valle del Cauca)



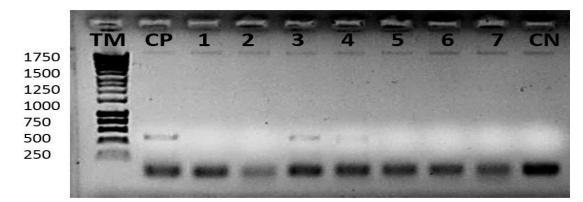
#### **RESULTS**

Table 1. Rate of Anopheles natural infection by Plasmodium spp.

Locality, Municipality (Department)	Species	n (%)	Parasite	Infection rate (%)
Córdoba, Buenaventura (Valle del Cauca)	Anopheles nuneztovari	715 (100%)	Plasmodium sp. ª	0.14
Bucheley, Tumaco (Nariño)	Anopheles albimanus	191 (90.1%)	P. falciparum <sup>a,b</sup>	0.52
Salahondita, Francisco Pizarro (Nariño)	Anopheles calderoni	49 (40.2%)	Plasmodium sp.a,b	2.04
Basurú, Istmina (Chocó)	Anopheles darlingi	84 (47%)	P. vivax a,b	1.2



**Nested PCR. CP:** Positive control. **Samples 1-9. Lane 6.** *An. albimanus* positive for *P. falciparum* from Tumaco. **Lane 8.** *An. calderoni* positive for *Plasmodium* spp. **Lane 9.** *An. darlingi* positive for Istmina *P. vivax*. Electrophoresis, 0.8% agarose gel. **TM:** 1 Kb GeneRuler molecular marker



**COXIII PCR. CP:** Positive control. CN: Negative control. **Samples 1-7. Lane 3.** *An. albimanus* positive para *P. falciparum* from Tumaco. **Lane 4.** *An. darlingi* positive para *P. vivax* from Istmina. Electrophoresis, 0.8% agarose gel. **TM:** 1 Kb GeneRuler molecular marker

Figure 2. Anopheles natural infection by Plasmodium spp. using molecular methods.

### **CONCLUSIONS**

The findings suggest that the main malaria vectors, *An. albimanus*, *An. darlingi* and *An. nuneztovari*, drive transmission in this region, but their distribution varies spatially.

The detection of *Plasmodium* natural infection in *An.* calderoni suggests its role as a local vector in specific areas.

These findings indicate that malaria transmission and vector roles vary spatially, emphasizing the need for locally adapted vector control interventions.

#### ACKNOWLEDGMENT

This work received support from Escuela de Microbiología, Universidad de Antioquia, Project code 2023-66350 and includes specimenes from Projects codes 596-2013 of COLCIENCIAS (Now MINCIENCIAS). E. Piedrahita received funding from Minciencias conv. 933-2023. To members of the Grupo de Microbiología Molecular, Escuela de Microbiología, Universidad de Antioquia

#### **REFERENCES**

- 1. Instituto Nacional de Salud. Sistema de Vigilancia en Salud Pública SIVIGILA. 2024. Semana Epidemiológica 52.
- 2. Gutiérrez et al., 2009. Species composition and natural infectivity of anthropophilic Anopheles (Diptera: Culicidae) in the states of Córdoba and Antioquia, Northwestern Colombia. Mem Inst Oswaldo Cruz. 104: 1117-24
- 3. González y Carrejo (2009). Introducción al estudio taxonómico de Anopheles de Colombia: claves y notas de distribución. segunda edición. Universidad del Valle, Cali, Colombia.
- 4. Zapata et al., 2007. Discrimination of seven Anopheles species from San Pedro de Uraba, Antioquia, Colombia, by polymerase chain reaction-restriction fragment length polymorphism analysis of its sequences.
- 5. Singh B, et al 1999. A genus and species-specific nested polymerase chain reaction malaria detection assay for epidemiologic studies. Am J Trop Med Hyg 1999 vol: 60 (4) pp: 687-692.

  6. Echeverry, D. F. et al., 2016. Human malaria diagnosis using a single-step direct-PCR based on the Plasmodium cytochrome oxidase III Gene. Malar J, 15, 128.