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Summary

- I have added a Gaussian mixture option to Paul's MATLAB code.
- I have run this model on the BCC TCGA data. This consists of 4 datasets for breast cancer from the TCGA. Gene expression, protein, methylation and miRNA data.
 - I have followed the pre-processing from the paper and then mean-centred and standardised the variance before running MDI (due to prior assuming a standard normal).

Gaussian mixture model

The MATLAB code assumes a standard normal distribution with exchangability of variables (i.e. independence of variables). The code uses a collapsed Gibbs sampler. This means that instead of calculating the probability of membership via:

$$p(z_i = k|z_{-i}, x_i, \pi_k, \theta_k) \propto \pi_k f(x_i|\theta_k)$$

one must compare the marginal likelihoods. In this case as the π_k and θ_k are not sampled one uses:

$$p(z_i = k|z_{-i}, x_i, \pi_k, \theta_k) \propto \pi_k \int f(x|\theta) p(\theta) d\theta$$
$$= \pi_k F(x)$$

Here F(x) is the marginal likelihood. One computes the marginal likelihood for the model considering all allocations except that of the i^{th} , $F(x_{-i})$ (call this the marginal likelihood for the **excluded model**). One then considers the ratio of marginal likelihoods for proposed model with $z_i = k$ and the excluded model; this enables comparison of models as the ratio in the above equation moves to an equality.

This is what the MATLAB code implements.

BCC data pre-processing

The pre-processing from the BCC paper involves the following steps:

- Gene expression data:
 - 1. impute missing values using impute.knn from the impute Bioconductor package;
 - 2. select only the genes with a standard deviation greater than 1.
- Methylation data:
 - 1. Remove rows with missing values in more than 50% of entries;
 - 2. Take the square root of the remaining values.
- miRNA data:

- 1. Take the log of the data.
- Protein data:
 - 1. Mean centre and standardise (in original paper separate to MDI requirements).

BCC data analysis

I have applied consensus inference to the data with 1,000 seeds for 500 iterations and both of 50 and 250 components (as a maximum). The number of components had no impact upon the results. I have to compare the clustering to the labels proposed by TGCA. TGCA has 4 prominent types; I have to find the labels for these but there's some encouraging result in that 4 clusters emerge from the miRNA dataset:

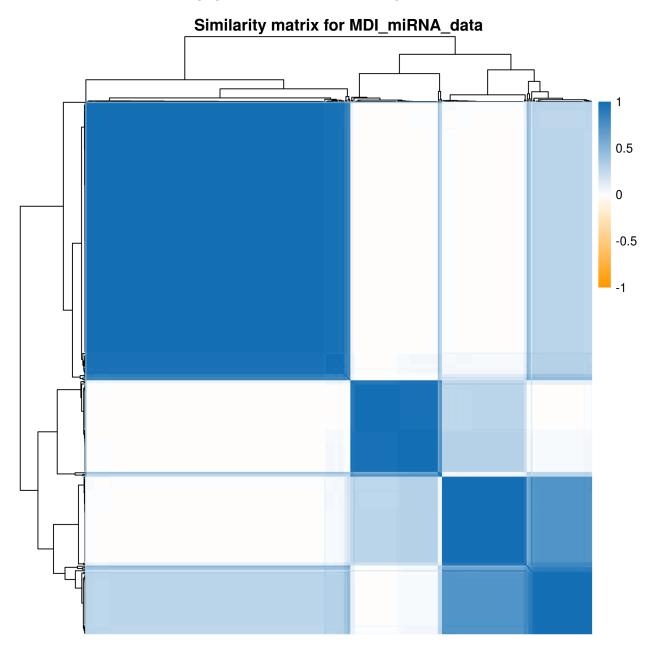


Figure 1: PSM for miRNA data with 1,000 seeds for 500 iterations and a maximum of 50 components.