

# Package ‘aveytoolkit’

March 6, 2015

**Title** Compilation of functions for data analysis

**Version** 0.1

**Description** Provides functionality for simple functions like Pause(), and resetPar() as well as complex functions like sigHeatmap, PlotTimeCourse, ComputeTimeLag, ggSmartBoxplot, etc.

**Depends** R (>= 3.0.2)

**License** Stefan Avey wrote most of these functions

**LazyData** true

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AverageReplicates	<i>AverageReplicates</i>
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**Description**

This function averages replicates in a matrix or data.frame

**Usage**

```
AverageReplicates(eSubSet, numRep)
```

**Arguments**

eSubSet	a matrix or data.frame of values with samples as columns
numRep	the number of replicates

**Value**

a data.frame of averaged values with column names coming from the first of each of the replicates with .avg appended

**Note**

Assumes that the replicates are all next to each other

**Author(s)**

Stefan Avey

**Examples**

```
mat <- matrix(rnorm(1000), ncol=10) ## 10 columns of random uniform numbers
avgMat <- AverageReplicates(mat, numRep=2) ## average adjacent pairs of columns
```

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aveytoolkit	<i>aveytoolkit.</i>
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**Description**

aveytoolkit.

---

barplotCI

*barplotCI*


---

**Description**

Create a barplot from the list x with one bar for each element of x

**Usage**

```
barplotCI(x, CIs = NULL, compareTo = 1, ...)
```

**Arguments**

x	list of values to create a boxplot from
CIs	the confidence intervals. Default is NULL and they will be calculated as the 95% confidence interval.
compareTo	non-negative integer specifying to which element of x should comparisons be made for significance. If 0, no significance will be added.
...	other arguments passed to barplot2 function

**Value**

Same as return from barplot2. A numeric vector (or matrix, when `beside = TRUE`), say mp, giving the coordinates of `_all_` the bar midpoints drawn, useful for adding to the graph. If `beside` is true, use `colMeans(mp)` for the midpoints of each `_group_` of bars, see example.

**Author(s)**

Christopher Bolen (creator) ; Stefan Avey (modified)

---

browseIndex

*browseIndex*


---

**Description**

Simple function to open HTML page of index in the default browser

**Usage**

```
browseIndex(package = NULL, lib.loc = NULL)
```

**Arguments**

package	a string with the name of package to use. If no name is supplied, the most recently loaded package is used.
lib.loc	a string of the directory name of the R library, or <code>&lt;e2&gt;&lt;80&gt;&lt;98&gt;NULL&lt;e2&gt;&lt;80&gt;&lt;99&gt;</code> . The default value of <code>&lt;e2&gt;&lt;80&gt;&lt;98&gt;NULL&lt;e2&gt;&lt;80&gt;&lt;99&gt;</code> corresponds to all libraries currently known.

**Details**

Only works for a single package. Could improve to list an HTML page with multiple packages that you could then choose from. Not currently implemented. Borrows the concepts from `utils::browseVignettes`

**Value**

invisibly, the URL passed to `browseURL` (i.e. "file:" + `<index_filename>`)

**Author(s)**

Stefan Avey

**Examples**

```
browseIndex("utils")
```

---

<code>cbind.fill</code>	<i>resetPar</i>
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**Description**

Simple function to combine multiple objects by column while filling in NAs into extra rows created from differing lengths

**Usage**

```
cbind.fill(...)
```

**Arguments**

... the objects, that will be converted to a list, to bind column-wise

**Value**

the cbind'ed objects passed in

**Author(s)**

Dimitris Rizopoulos and Tyler Rinker

**References**

<http://stackoverflow.com/questions/7962267/cbind-a-df-with-an-empty-df-cbind-fill>

**See Also**

[cbind](#)

**Examples**

```
x<-matrix(1:10,5,2)
y<-matrix(1:16, 4,4)
z<-matrix(1:12, 2,6)

cbind.fill(x,y)
cbind.fill(x,y,z)
cbind.fill(mtcars, mtcars[1:10,])
```

collapseDataset

*collapseDataset***Description**

Collapses a dataset from probes to gene symbols.

**Usage**

```
collapseDataset(exprsVals, platform = NULL, mapVector = NULL, oper = max,
  prefer = c("none", "up", "down"), singleProbeset = FALSE,
  returnProbes = FALSE, deProbes = NULL)
```

**Arguments**

exprsVals	a matrix or data.frame of expression values with rownames denoting the probes.
platform	the microarray platform the data comes from for extracting the gene symbols
mapVector	a named character vector with names specifying the current identifiers (probes matching the rownames of exprsVals) and the values of the vector specifying the gene symbols (or other identifier to collapse to).
oper	the operation used to choose which probe when multiple probes map to the same gene. Default is max which will calculate the maximum of the average.
prefer	one of "none", "up", or "down", can be abbreviated.
singleProbeset	If TRUE, the operation applies to the average of each sample. Otherwise, if FALSE, the operation applies to the probesets over all samples and only one probeset will be selected. Default is FALSE.
returnProbes	if TRUE, a list of the collapsed expression matrix and the probes are both returned (see return).
deProbes	a list with named vectors "up" and "down" giving the names of up and downregulated probes

**Details**

If singleProbeset is set to TRUE, untested and not recommended, the values for each sample will be taken from the maximum across any probe that maps to that gene. This means that a gene's expression values may be a composition of values from different probes rather than a single probe. if prefer is "up", when multiple deProbes match the same gene, the upregulated will be chosen. Similary for "down". Default is "none" and the probe with the oper will be chosen.

**Value**

If returnProbes is TRUE, a list containing the collapsed dataset in \$exprsVals and the probes chosen in \$probeSets. Otherwise, if returnProbes is FALSE, only the expression matrix is returned.

**Author(s)**

Christopher Bolen, Modified by Stefan Avey

**Examples**

```
## ??
```

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fishersMethod	<i>fishersMethod</i>
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---

**Description**

This function combines multiple p-values according to Fisher's Method

**Usage**

```
fishersMethod(x)
```

**Value**

a single combined p-value

**Author(s)**

Mike Love

**References**

<http://mikelove.wordpress.com/2012/03/12/combining-p-values-fishers-method-sum-of-p-values-bino>

**Examples**

```
x <- c(runif(1000, 0, 1),runif(100,.1,.2))
fishersMethod(x)
```

---

*FoldChange**FoldChange*

---

**Description**

Calculate the fold change between pairs of conditions in a matrix or data frame

**Usage**

```
FoldChange(x, condNum, condDen, conditions, grouping, preserveOrder = FALSE,  
           log2Transform = FALSE)
```

**Arguments**

x	matrix or data.frame from which to calculate fold changes with samples in columns.
condNum	a vector of condition(s) to be used as the numerator in the fold change calculation
condDen	a vector of condition(s) to be used as the denominator in the fold change calculation
conditions	a vector with length equal to the number of columns of x containing the condition labels between which to find the fold changes.
grouping	a vector with length equal to the number of columns of x containing a grouping of the samples (e.g. subjects, cell lines, strains).
preserveOrder	if TRUE, the same ordering of the columns in x will be kept after columns in condDen are removed. If FALSE (the default for backwards compatability), the ordering is changed to sort by group, then by condNum in order passed in.
log2Transform	when 'TRUE', log2 transformation will be applied to x before taking the FC. If 'FALSE' (default) no transformation is applied and x is ASSUMED to be already log transformed.

**Details**

FoldChange takes the fold change of log2 transformed data by subtracting columns of the x dataframe or matrix depending on the conditions passed in.

**Value**

a data.frame of the fold changes with one column for each fold change

**Author(s)**

Stefan Avey

geomMean

*geomMean*

---

**Description**

Calculate the geometric mean

**Usage**

```
geomMean(x, na.rm = TRUE)
```

**Arguments**

x	a vector of positive numeric values.
na.rm	(optional) whether to remove NA values before calculation. Default is TRUE

**Details**

This function handles negative or 0 values by warning that they are ignored and calculating the geometric mean without them

**Value**

the geometric mean of x

**Author(s)**

Paul McMurdie

**References**

<http://stackoverflow.com/questions/2602583/geometric-mean-is-there-a-built-in>

**See Also**

[exp](#) [sum](#) [log](#)

**Examples**

```
x <- 1:10
x2 <- x^2
x3 <- -5:5

geomMean(x)
mean(x)

geomMean(x2)
mean(x2)

## Warning because x3 contains negative values ##
geomMean(x3)
mean(x3)
```



---

getLoginDetails	<i>getLoginDetails</i>
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---

**Description**

Uses tcltk to display a prompt for a loginID and password

**Usage**

```
getLoginDetails()
```

**Details**

This function displays a window for a user to enter a loginID and password without showing the password.

**Value**

an invisible named vector of loginID and password

**Author(s)**

Markus Gesmann, Barry Rowlingson

**References**

<http://www.r-bloggers.com/simple-user-interface-in-r-to-get-login-details/> <http://r.789695.n4.nabble.com/tkentry-that-exits-after-RETURN-tt854721.html#none>

**See Also**

[tcltk](#)

**Examples**

```
credentials <- getLoginDetails()
## Do what needs to be done with loginID and password
rm(credentials) # Delete credentials
```

---

ggSmartBoxplot	<i>ggSmartBoxplot</i>
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**Description**

Boxplot wrapper for ggplot

**Usage**

```
ggSmartBoxplot(x, mat, splitRowBy = NA, splitColBy = NA, colorBy = NULL,
  rows, cols = NA, whichCols = NA, sep = ".", outlier.shape = 17,
  outlier.color = NULL, ylab = NULL, space = "fixed", scales = "fixed",
  fileName = NA, plot = TRUE, ...)
```

**Arguments**

<code>x</code>	the variable to group by for boxplots
<code>mat</code>	data.frame or matrix of values to plot with samples in columns
<code>splitRowBy</code>	a factor used to split the data by row in <code>facet_grid</code>
<code>splitColBy</code>	a factor used to split the data by col in <code>facet_grid</code>
<code>colorBy</code>	a factor used for coloring. No coloring will be done if NULL (default)
<code>rows</code>	row names or row indices of the items to be plotted
<code>cols</code>	substring to search for with "grep" in column names to be plotted
<code>whichCols</code>	the column indices or full column names
<code>sep</code>	a separator used in searching for cols in the column names
<code>outlier.shape</code>	shape of outliers (default is 17, filled triangle)
<code>outlier.color</code>	color of outliers (default is NULL, i.e. they will not be colored differently)
<code>ylab</code>	if NULL, default is to use rownames. Can specify a string instead to use
<code>space</code>	If "fixed", the default, all panels have the same size. If "free_y" their height will be proportional to the length of the y scale; if "free_x" their width will be proportional to the length of the x scale; or if "free" both height and width will vary. This setting has no effect unless the appropriate scales also vary.
<code>scales</code>	Are scales shared across all facets (the default, "fixed"), or do they vary across rows ("free_x"), columns ("free_y"), or both rows and columns ("free")
<code>fileName</code>	
<code>plot</code>	logical specifying whether or not to plot the plot(s). Default is TRUE.
<code>...</code>	other arguments that are passed to <code>qplot</code>
<code>filename</code>	the name of a file to write a PDF to or NA to plot in standard graphics device.

**Value**

invisibly returns a list with 2 elements: `ggplot`: the `ggplot` object to be plotted (this can be added to `dat`: a named list of the data frame(s) passed to data in `ggplot`. The names come from converting the rows argument to a character vector.

**Author(s)**

Stefan Avey

**See Also**

[ggplot2](#), [qplot](#)

**Examples**

```
data(OrchardSprays)
## Example of functionality
ggSmartBoxplot(x=OrchardSprays$treatment,
               mat=t(OrchardSprays[,1]),
               rows=1, whichCols=1:ncol(t(OrchardSprays)),
               colorBy=factor(OrchardSprays$rowpos+OrchardSprays$colpos > 9),
               xlab="Treatment")
```

```
## NOT RUN:
cellType <- "PBMC"
geneSub <- grep("HLA-A29.1", rownames(expr))
age <- "Young"
ages <- c("Young", "Old")
responses <- c("NR", "R")
subset <- targetFClst[[cellType]]$Age %in% ages &
  targetFClst[[cellType]]$Response %in% responses
ggSmartBoxplot(x=targetFClst[[cellType]][subset, "Time"],
  mat=exprFClst[[cellType]], ylim=c(-1,1),
  rows=geneSub, whichCols=which(subset),
  colorBy=targetFClst[[cellType]][subset, "Response"],
  splitRowBy=targetFClst[[cellType]][subset, "Age"],
  xlab="Days (Post Vaccination)",
  fileName=NA)
```

ggSmoothExprPlot

*ggSmoothExprPlot*

## Description

Wrapper around ggplot to transform data and plot profiles (e.g. expression or activity) over time

## Usage

```
ggSmoothExprPlot(x, mat, rows, method = "auto", formula = formula("y ~ x"),
  splitRowBy = NA, splitColBy = NA, colorBy = NULL, cols = NA,
  whichCols = NA, sep = ".", colorByLabel = "Response", ggtitle = TRUE,
  xlab = "Time (Post-Vaccination)", ylab = "Expression",
  colors = c("#1B9E77", "#D95F02", "#7570B3", "#E7298A", "#66A61E", "#E6AB02",
    "#A6761D", "#666666"), space = "fixed", scales = "fixed", fileName = NA,
  plot = TRUE)
```

## Arguments

x	the numeric x-axis variable for the plot (usually time)
mat	data.frame or matrix of values to plot with samples in columns
rows	row names or row indices of the items to be plotted
method	smoothing method (function). See <code>stat_smooth</code>
formula	a formula to use for smoothing in <code>stat_smooth</code> (e.g. the default "y ~ x" or "y ~ ns(x, 3)").
splitRowBy	a factor used to split the data by row in <code>facet_grid</code>
splitColBy	a factor used to split the data by col in <code>facet_grid</code>
colorBy	a factor used for coloring. No coloring will be done if NULL (default)
cols	substring to search for with "grep" in column names to be plotted
whichCols	the column indices or full column names
sep	a separator used in searching for cols in the column names
colorByLabel	the labels used for the color legend
ggtitle	logical. If TRUE, rows is coerced to character and passed to ggtitle

xlab	passed to xlab. Defaults to "Time (Post-Vaccination)"
ylab	passed to ylab. Defaults to "Expression"
colors	The colors to use. Defaults to the colors given by using RColorBrewer's "Dark2" palette (but RColorBrewer is not called directly so is not required).
space	If "fixed", the default, all panels have the same size. If "free_y" their height will be proportional to the length of the y scale; if "free_x" their width will be proportional to the length of the x scale; or if "free" both height and width will vary. This setting has no effect unless the appropriate scales also vary.
scales	Are scales shared across all facets (the default, "fixed"), or do they vary across rows ("free_x"), columns ("free_y"), or both rows and columns ("free")
fileName	the name of a file to write a PDF to or NA to plot in standard graphics device.
plot	logical specifying whether or not to plot the plot(s). Default is TRUE.

### Value

invisibly returns a list with 2 elements: ggplot: the ggplot object to be plotted (this can be added to dat: a named list of the data frame(s) passed to data in ggplot. The names come from converting the rows argument to a character vector. if plot=FALSE: invisibly returns the

### Author(s)

Stefan Avey

### See Also

[ggplot2](#)

### Examples

```
data(OrchardSprays)
## Example of functionality
library(Biobase)
data(sample.ExpressionSet, package="Biobase")
dat <- sample.ExpressionSet
## Normally x-axis is time but in this dataset there is no time
## so we will use the score as the x-axis
genderF <- dat$sex == "Female"
ggSmoothExprPlot(x=dat$score[genderF],
                 mat=exprs(dat),
                 rows="31345_at",
                 whichCols=which(genderF), # females only
                 colorBy=as.factor(dat$type)[genderF],
                 colorByLabel="Condition",
                 xlab="score")

## NOT RUN:
tmp <- ggSmoothExprPlot(x=times[subset], mat=expr, rows=gene,
                       formula=formula("y ~ ns(x,3)"),
                       whichCols=subset, colorBy=target[subset,respType],
                       splitColBy=splitby,
                       splitRowBy=as.factor(target[subset,"Study"]),
                       ggtitle=TRUE, colorByLabel=respType, plot=TRUE)

## End NOT RUN
```

---

MakeDF	<i>MakeDF</i>
--------	---------------

---

**Description**

Creates a data frame from a list. Useful for when the list elements have unequal lengths and `as.data.frame` fails.

**Usage**

```
MakeDF(list, names)
```

**Arguments**

<code>list</code>	the list to convert
<code>names</code>	the names of the list

**Value**

a data frame of the converted list.

**Author(s)**

Josh O'Brien

**References**

<http://stackoverflow.com/questions/15753091/convert-mixed-length-named-list-to-data-frame>

**See Also**

[gsub](#)

**Examples**

```
## Test timing with a 50k-item list
ll <- createList(50000)
nms <- c("a", "b", "c")

system.time(makeDF(ll, nms))
# user  system elapsed
# 0.47    0.00    0.47
```

---

makeTransparent	<i>makeTransparent</i>
-----------------	------------------------

---

**Description**

Simple function to make some colors transparent

**Usage**

```
makeTransparent(alpha = 0.5, ...)
```

**Arguments**

alpha	transparency factor in range [0,1]
...	vector or list of colors

**Value**

a vector of new colors made transparent

**Author(s)**

Ricardo Oliveros-Ramos

**References**

<http://stackoverflow.com/questions/8047668/transparent-equivalent-of-given-color>

**See Also**

[rgb](#), [col2rgb](#)

**Examples**

```
makeTransparent("red", "blue")
##[1] "#FF00007F" "#0000FF7F"
makeTransparent("red", "blue", alpha=0.8)
## [1] "#FF0000CC" "#0000FFCC"
```

---

Pause	<i>Pause</i>
-------	--------------

---

**Description**

This function prompts for return key and waits until the return is pushed to continue execution. It is used often to view plots coded in a loop one at a time allowing the user to control when the next plot should be displayed

**Usage**

```
Pause(str = "continue", quiet = FALSE)
```

**Arguments**

<code>str</code>	optional string to display. Defaults to "continue".
<code>quiet</code>	if TRUE, no prompt is displayed. Default is FALSE

**Details**

The Pause function uses `readline` to wait until a newline character (produced by the Enter key) is given. Instead of pressing Enter, a newline character can be used to automate this waiting time.

**Value**

NULL is returned by invisible

**Author(s)**

Stefan Avey

**See Also**

[readline](#), [invisible](#)

**Examples**

```
for(p in 1:10) {
  plot(-10:10, (-10:10)^p, type=b)
  Pause(paste0("see plot of x^",p+1))
}
```

---

PlotTimeCourse

*PlotTimeCourse*


---

**Description**

Plot helper function for PlotPCATimeCourse

**Usage**

```
PlotTimeCourse(x, y, colors, groups, sampleNames, pch = 19, plotTitle = "",
  legend.loc = "topleft", plotType = c("times", "points"), alpha = 0.15,
  cex.pt = 1, cex.time = 2, time.adj = c(-0.3, -0.3), arrLen = 0.1,
  lwd = 3, numRep = 3, plotFont = NULL, ctrl = TRUE, hourMarks = TRUE,
  legend.cex = 2, ...)
```

**Arguments**

<code>x</code>	x-values for plotting
<code>y</code>	y-values for plotting
<code>colors</code>	named vector specifying colors for each sample
<code>groups</code>	the virus strain names for the conditions of interest
<code>sampleNames</code>	names of the samples

<code>pch</code>	the plotting character. Default is 19 (a closed circle).
<code>plotTitle</code>	a string used for the plotting title
<code>legend.loc</code>	location of the legend. Default is topleft.
<code>plotType</code>	one of "times" or "points". See Details
<code>alpha</code>	transparency factor passed to the alpha function (scales library)
<code>cex.pt</code>	size of points. Default is 1
<code>cex.time</code>	size of time labels. Default is 2
<code>time.adj</code>	the ammount to adjust the time labels. Default is c(-.3, -.3) which moves them to the lower left
<code>arrLen</code>	length of the arrows plotted at the average of each time point. Default is 0.1
<code>lwd</code>	line width. Default is 3
<code>numRep</code>	the number of replicates. Default is 3
<code>plotFont</code>	which font to use for plotting text
<code>ctrl</code>	should the control time points be included?
<code>hourMarks</code>	should the 4 and 8 hour time points be marked on the plot?
<code>legend.cex</code>	size expansion for the legened. Default is 2.
<code>...</code>	other arguments passed to heatmap.2

### Details

If `plotType` is "times", ??? Also used to plot 2 genes expression against each other over time. If `legend.loc` is "none", no legend is plotted. `ctrl` flag indicates whether or not first `numRep` values in `x` and `y` are from a control measurement

### Value

Nothing is returned

### Note

Colors are assumed to have as the names attribute some part of the `sampleName` which can uniquely identify it.

### Author(s)

Stefan Avey



---

PrepareExpression	<i>PrepareExpression</i>
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---

## Description

Takes in an expression matrix or data frame and prepares it for further analysis

## Usage

```
PrepareExpression(eset, target, returnProbes = TRUE, labelColumn = "Label",
  select = colnames(target), collapse = ".")
```

## Arguments

eset	expression information and (potentially) other columns
target	target file where the column names of eset can be matched to 'Label'
returnProbes	whether probe mapping should be returned along with expression values in a list. This will only be returned correctly if there is a column of eset matching SYMBOL in any case.
labelColumn	the column name in the target file to use for matching the column names of eset. Default is "Label"
select	the column names of target to select and merge as the new column names of eset
collapse	

## Details

Wrote this to automate the few lines I always perform to "prepare" an expression set for further processing. I always want to remove the symbols column, rename the column names based on the target file, and (usually) change the rownames to be gene symbols. This function takes in the matrix format that I use to store processed expression files (in a package or file).

## Value

if returnProbes is FALSE: a list of the prepared expression data frame (exprDat) and the (potentially modified) target data frame (target) . if returnProbes is TRUE (default): a list of three elements including the two above and probeMap (a vector mapping from gene symbols to probe names).

## Author(s)

Stefan Avey

## Examples

```
## Creating fake expression matrix
dat <- matrix(rnorm(1000, mean=8, sd=1), nrow=100, ncol=10)
colnames(dat) <- sample(letters[1:10], size=10)
fakeGenes <- as.vector(outer(LETTERS[1:26], LETTERS[1:26], paste0))
x <- data.frame(symbol=fakeGenes[1:nrow(dat)], dat, row.names=paste0("Probe_", 1:nrow(dat)))
head(x) # look at first 6 rows of toy data set
target <- data.frame(Label=letters[1:26], Class=rep(1:3, length.out=26))
head(target)
```

```

preList <- PrepareExpression(x, target, select="Class")
head(preList$exprDat)
head(preList$target)
head(preList$probeMap)

## NOT RUN:
library(HIPC)
data(y3ExprPBM, y3Target)
preList <- PrepareExpression(y3ExprPBM, y3Target,
                             select=c("Response", "SubjectID", "Age", "Time"))

```

---

ProcessNames	<i>ProcessNames</i>
--------------	---------------------

---

## Description

Cleans up strings to make them pretty names by removing punctuation, whitespace, and specified substrings

## Usage

```
ProcessNames(strs, stringsToRm = NULL, rmPunct = TRUE, sep = "_")
```

## Arguments

<code>strs</code>	vector or strings to process
<code>stringsToRm</code>	a vector or list of strings to search for and remove from <code>strs</code>
<code>rmPunct</code>	should punctuation be removed? Default is TRUE.
<code>sep</code>	character to replace whitespace

## Details

`stringsToRm` are replaced by " " in the order they are given using `gsub`. After this, punctuation is removed if `rmPunct` is TRUE. Then, leading and/or trailing whitespace will be removed and the `sep` will be used to separate words. This function is useful when reading in other people's data and you want to change the row or column names to legal R names or just shorten the names.

## Value

a vector of modified strings from `strs`

## Author(s)

Stefan Avey

## See Also

[gsub](#)

**Examples**

```
badNames <- c("Whos Birthday?", "[Date]", "gift Received")
## Remove the string "Whos", remove punctuation, and separate words by _
goodNames <- ProcessNames(badNames, stringsToRm="Whos", rmPunct=TRUE, sep=_)
goodNames
## Remove the string "Whos", dont remove punctuation, and put no separation between words
goodNames <- ProcessNames(badNames, stringsToRm="Whos", rmPunct=FALSE, sep=)
goodNames
```

---

RepeatBefore

*RepeatBefore*


---

**Description**

Replaces NAs with the latest non-NA value

**Usage**

```
RepeatBefore(x)
```

**Arguments**

`x` a vector of values

**Details**

NA values will be replaced by the most recent value with a lower index. If there is no non-NA value before the NA appears, it will remain NA.

**Value**

a vector of values

**Author(s)**

Ruben

**References**

<http://stackoverflow.com/questions/7735647/replacing-nas-with-latest-non-na-value>

**See Also**

[rep](#)

**Examples**

```
x = c(NA, NA, a, NA, NA, NA, NA, NA, NA, NA, NA, b, c, d, NA, NA, NA, NA, NA, e)
newX <- RepeatBefore(x)
show(newX)
```

---

resetPar	<i>resetPar</i>
----------	-----------------

---

## Description

Simple function to reset plotting parameters for when things get wonky

## Usage

```
resetPar()
```

## Details

This function resets the graphical parameters from the `par` function. It flashes a new device on the screen but works to reset parameters. Meant to be used when things get hairy and not coded in scripts

## Value

an invisible named list of parameters returned by calling `par`

## Author(s)

Gavin Simpson

## References

<http://stackoverflow.com/questions/5789982/reset-par-to-the-default-values-at-startup>

## See Also

[par](#)

## Examples

```
par(oma=c(4,10,2,1))
plot(1,1)
## paramter settings werent saved so do a reset
resetPar()
plot(1,1)
```

---

runLimma	<i>runLimma</i>
----------	-----------------

---

## Description

A wrapper around limma functions to perform a basic analysis on the given expression matrix

## Usage

```
runLimma(eset, labels, contrasts, block = NULL, covariates = NULL,
  filterReplicateGenes = TRUE, min.fold.change = 1, min.intensity = 4,
  p.cutoff = 0.05, fitOnly = FALSE)
```

## Arguments

eset	the expression matrix
labels	the labels for each column of the eset
contrasts	Vector of contrasts to make
block	vector or factor specifying a blocking variable on the arrays. Has length equal to the number of arrays. Must be <code>NULL</code> if <code>ndups &gt; 2</code> (Not extensively tested, use with caution)
covariates	data frame of covariates (of same length as labels) to include in the model. Use this if there are paired samples, etc.
filterReplicateGenes	Only include one probeset for each gene (determined by symbol)
min.fold.change	Minimum log2 fold change to be differentially expressed. Default is 1.
min.intensity	Minimum log2 intensity (at any time) to be differentially expressed. Default is 4.
p.cutoff	FDR corrected cutoff for significant differential expression. Default is 0.05.
fitOnly	If true, will return fit2, rather than the matrix of significant genes. Default is FALSE.

## Details

Generally, an expression matrix is made up of rows of genes (or any other features) and columns of samples. The matrix has data for multiple classes (which are denoted with the 'labels' parameter) and the classes are compared using the vector of contrasts. Block can be used for biological (or technical) replicates or for separate subjects (in which case it will determine the inter-subject correlation). See `?duplicateCorrelation` for more information.

## Value

depends on fitOnly

## Author(s)

Christopher Bolen, Stefan Avey

**See Also**[limma](#)**Examples**

```
## Example:
## If you have a m X 10 matrix eset, with 5 samples of class A and 5 of class B,
## you could compare class A to class B using the following code:
##
## results = runLimma(eset, c(A,A,A,A,A,B,B,B,B,B), "B-A")
##
## This will return to you a matrix with columns for each comparison and rows for each gene.
## The value in each cells will either be -1, 0, or 1, depending on whether the gene is
## significantly higher in B, not significant, or significantly higher in A, respectively.
## If you want information on p-values and fold changes, set "fitOnly=T", and you can access
## the fit object to get the information.
##
## For other comparisons, you can look at the LIMMA user guide: limmaUsersGuide()
```

---

sigHeatmap*sigHeatmap*

---

**Description**

Draw heatmap with significance indicated on boxes

**Usage**

```
sigHeatmap(hm, pvals, pvalDisplayName = "P-value", cutoff = 0.05,
  showOnly = c("both", "positive", "negative", "all"), main = "",
  mainNewlines = 0, sigChar = "*", Rowv = T, hclustMethod = "ward.D",
  ...)
```

**Arguments**

hm	a matrix of values used for drawing the heatmap
pvals	a list or data frame of (possibly FDR corrected but this is not handled by the function) positive p-values
pvalDisplayName	is printed on the heatmap as a legend. Default is "P-value" but might want to change to "Q-value", "FDR", etc.
cutoff	is threshold for significance of pvals. Default is 0.05
showOnly	one of "both", "positive", "negative", or "all" can be abbreviated.
main	a string giving the plot main title. Default is "" (i.e. no title is plotted).
mainNewlines	a non-negative integer specifying the number of newline characters to plot before the main title. Used to make the title appear lower on the page. Default is 0
sigChar	the character used for plotting on top of significant boxes
Rowv	should the rows be reordered, passed into heatmap.2

`hclustMethod` passed to the function `stats::hclust`. The agglomeration method to be used. This should be (an unambiguous abbreviation of) one of "ward.D", "ward.D2", "single", "complete", "average" (= UPGMA), "mcquitty" (= WPGMA), "median" (= WPGMC) or "centroid" (= UPGMC). Default is "ward.D".

... other arguments passed to `heatmap.2`

### Details

Only rows with at least one significant column are plotted. If `showOnly` is "both", plots both positive and negative significant changes. If `showOnly` is "positive" or "negative", plots only rows of `hm` with significant positive or negative values respectively. If `showOnly` is "all", all rows of `hm` are shown.

### Value

a vector indicating which of the rows of `hm` were determined to be significant and subsequently plotted

### Author(s)

Stefan Avey

### Examples

```
data(mtcars)
x <- as.matrix(mtcars)
alpha <- 10^-7 # significance threshold
## Caculate whether difference from mean is significant
## This is not done correctly but just to have some sort of significance
diffMean <- mtcars-matrix(colMeans(mtcars),
                          ncol=ncol(mtcars), nrow=nrow(mtcars), byrow=TRUE)
stdErr <- matrix(sapply(mtcars, sd)/sqrt(nrow(mtcars)),
                 ncol=ncol(mtcars), nrow=nrow(mtcars), byrow=TRUE)
tstats <- diffMean/stdErr
pvals <- pt(as.matrix(tstats), nrow(mtcars)-2, lower=FALSE)
op <- par(oma=c(4,0,0,20))
sel <- sigHeatmap(x, pvals=pvals, cutoff=alpha, showOnly="b",
                 main="mtcars Example Heatmap", sigChar="*", notecol=black,
                 notecex=2, Colv=T, Rowv=T, dendrogram="row", trace="none")
par(op)
## Which cars werent selected
rownames(mtcars)[setdiff(1:nrow(mtcars), sel)]
```

---

VennDiagram

VennDiagram

---

### Description

Draw a venn diagram of 2 or 3 sets

### Usage

```
VennDiagram(setList, mar = c(0, 0, 1, 0), ...)
```

**Arguments**

`setList` a (named) list of the sets to be plotted. The names will be used on the plot. If the list is unnamed, the default names in [vennDiagram](#)

**Details**

Wrapper around the [limma vennDiagram](#) function to make it simpler.

**Value**

a data frame of binary values indicating membership in each set with rownames giving the set entries.

**Author(s)**

Stefan Avey

**References**

Code modified from <http://research.stowers-institute.org/mcm/venn.R>

**See Also**

[vennDiagram](#)



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