

# **BIOL334 Conservation & Ecological Genetics**

## LECTURE 9: MUTATION, MIGRATION & SELECTION II

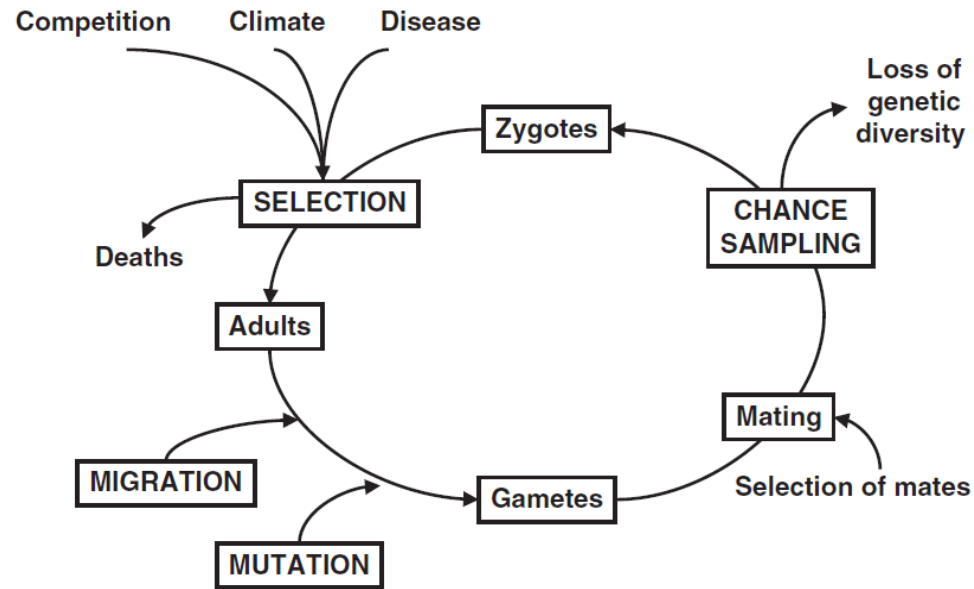


# Mutation, Migration & Selection

Assignment #1 grades on iLearn

Continuing from Lecture 8...

**Fig. 6.1** An evolving population as a complex system.



# Genetics & Population size

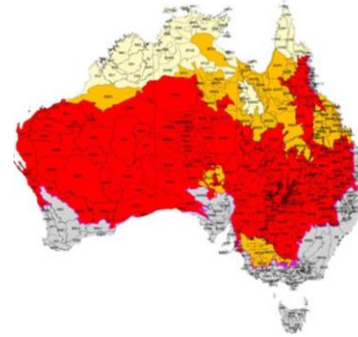
## LARGE vs SMALL POPS

### Large N

High levels of  $V_G$   
= “variant” alleles  
= **resilience** to change

High mutation potential  
= **NEW**  $V_G$

Power for selection (all forms)  
= **adaptive potential**



Rabbit  
N~10 Billion (1920)



Water buffalo  
N~100,000

# Genetics & Population size

## LARGE vs SMALL POPS

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### Large N

High levels of  $V_G$   
= “variant” alleles  
= **resilience** to change

High mutation potential  
= **NEW**  $V_G$  (over time)

Power for selection (all forms)  
= **adaptive potential**

### Small N

1. Low  $V_G$   
= **vulnerable** to change
2. few new mutations, **BUT**  
existing mutations exposed
3. **Selection** overpowered  
by **genetic drift...**

# Genetics & Population size

## GENETIC DRIFT

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Random **luck of the draw**...

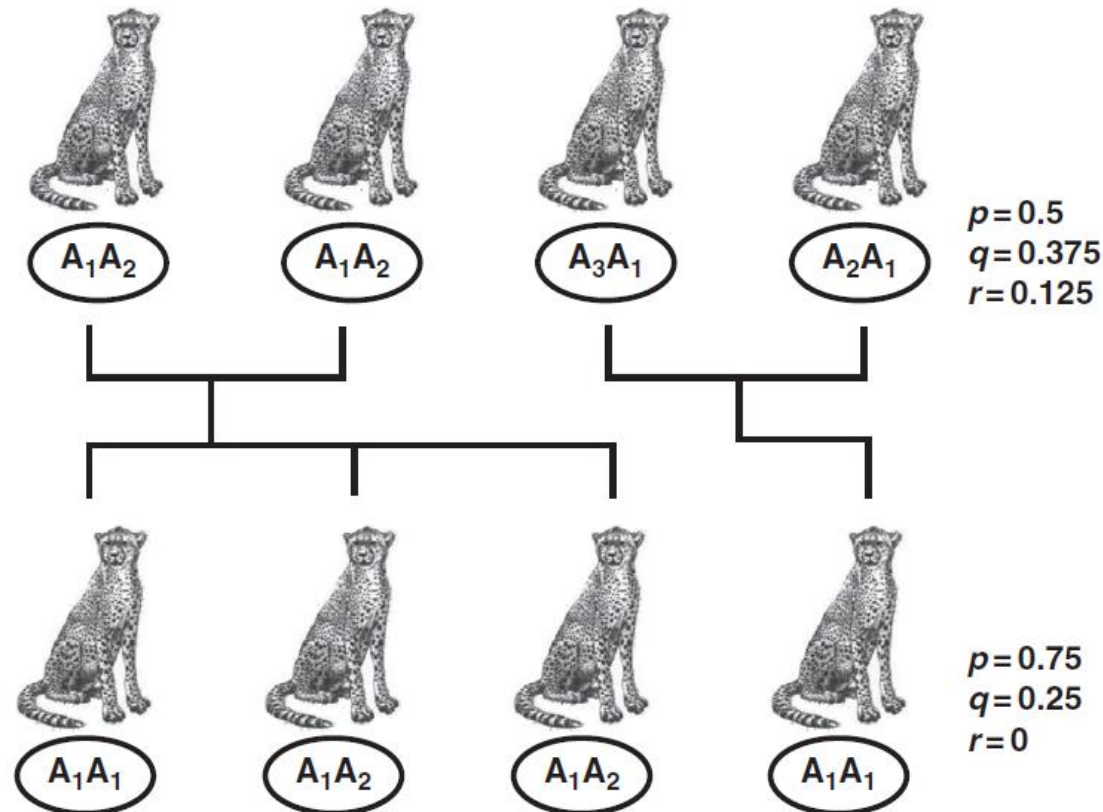
Chance decides allele frequencies  
from one generation to the next;

These effects accumulate – once a rare gene is lost, it  
is gone forever!

Causes small populations to evolve in ‘random’  
directions – i.e. to **drift**...



# Loss of rare alleles via drift

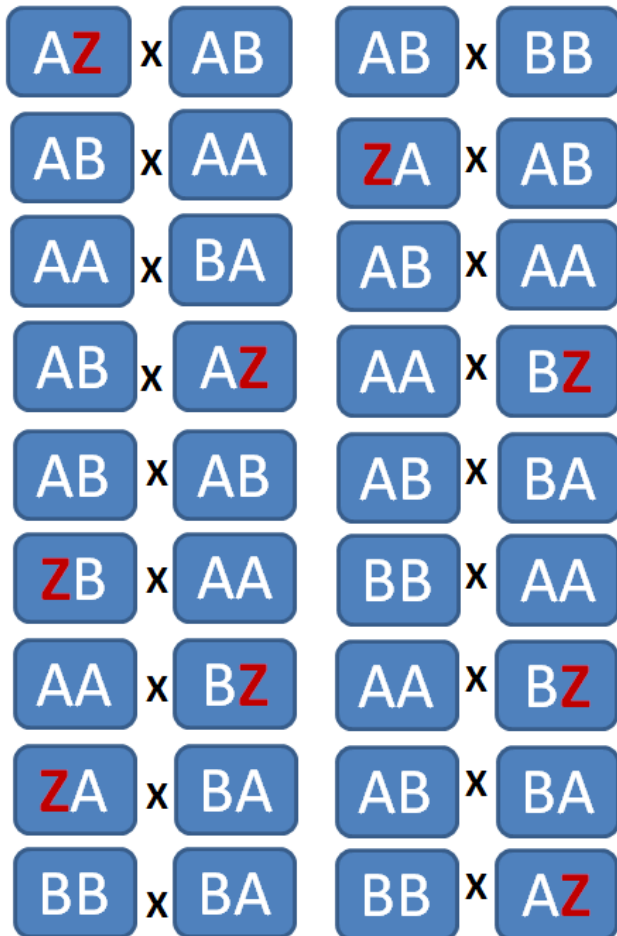


**Fig. 8.1** Genetic drift in allele frequencies in a small population of cheetahs.  $p$ ,  $q$  and  $r$  are the frequencies of alleles  $A_1$ ,  $A_2$  and  $A_3$ , respectively. Allele  $A_3$  is lost by chance. Further, the frequencies of  $A_1$  and  $A_2$  change from one generation to the next, with  $A_1$  rising and  $A_2$  falling.

# Genetics & Population size

## GENETIC DRIFT

Population N=36



Allele frequencies

A: 36 (50.0%)  
B: 27 (37.5%)  
**Z**: 9 (12.5%)

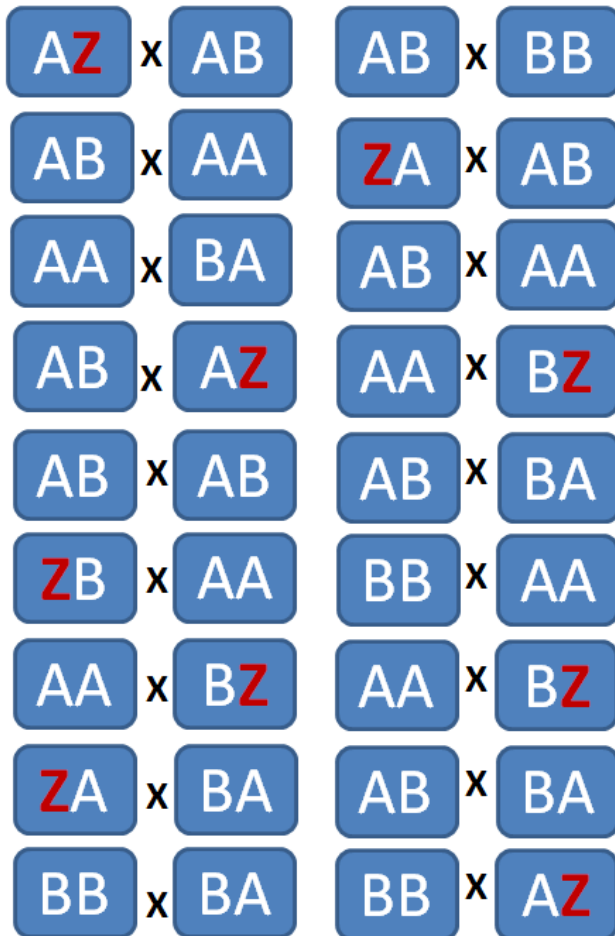
Gen  
+1

**Nine** 50:50  
chances of  
retaining  
the **Z** allele

# Genetics & Population size

## GENETIC DRIFT

Population N=36

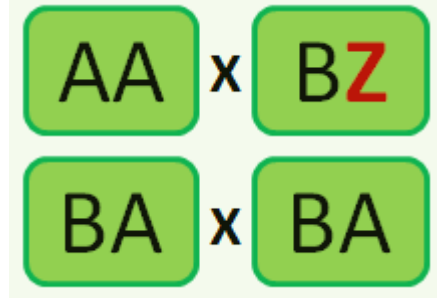


Allele frequencies

A: 36 (50.0%)  
B: 27 (37.5%)  
Z: 9 (12.5%)

Gen  
+1

**Nine** 50:50  
chances of  
retaining  
the **Z** allele



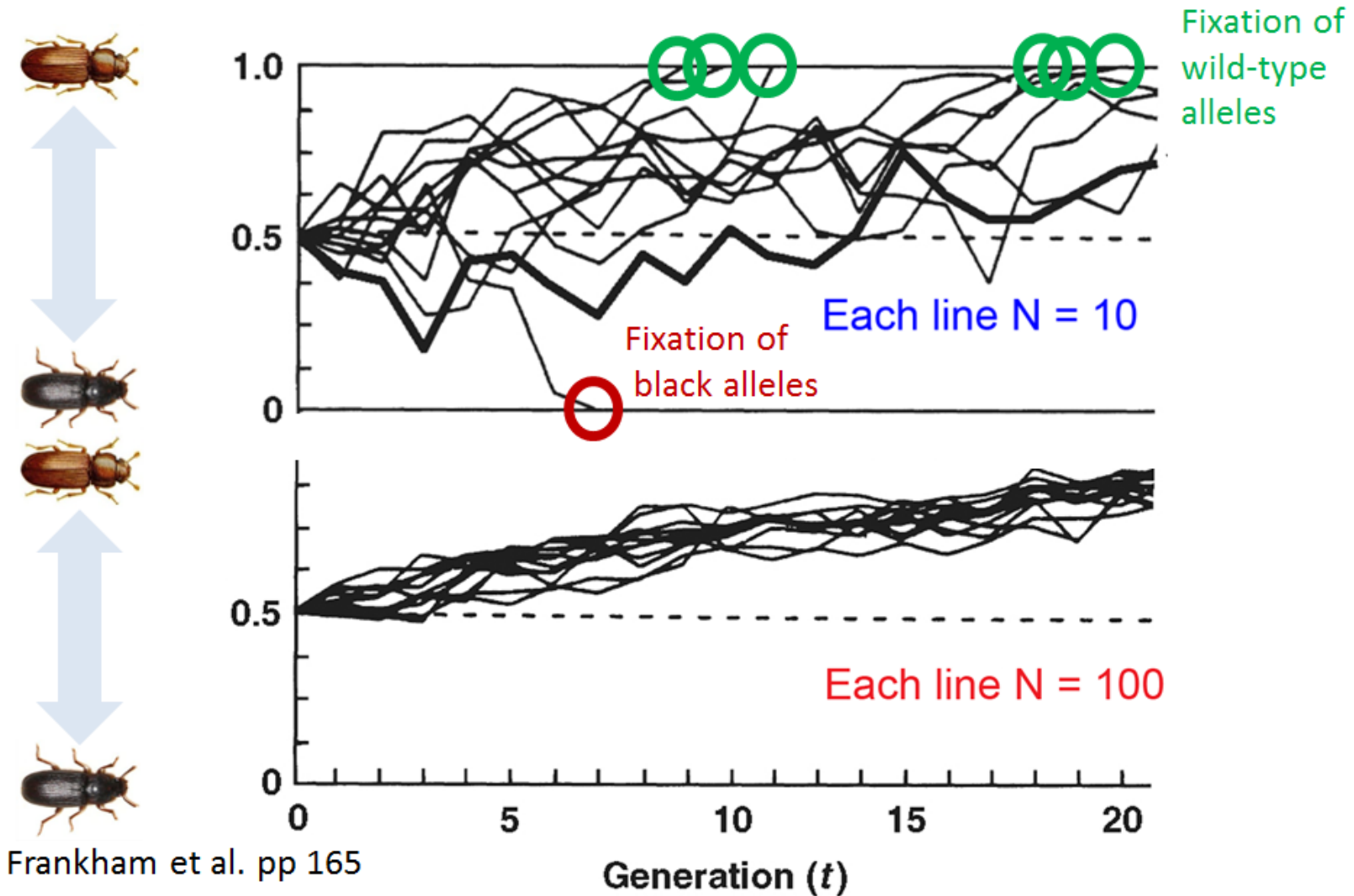
N = 4 (8 alleles)  
A: 8 (50.0%)  
B: 7 (37.5%)  
Z: 1 (12.5%)

**ONE** 50:50  
chance of  
retaining  
the **Z** allele



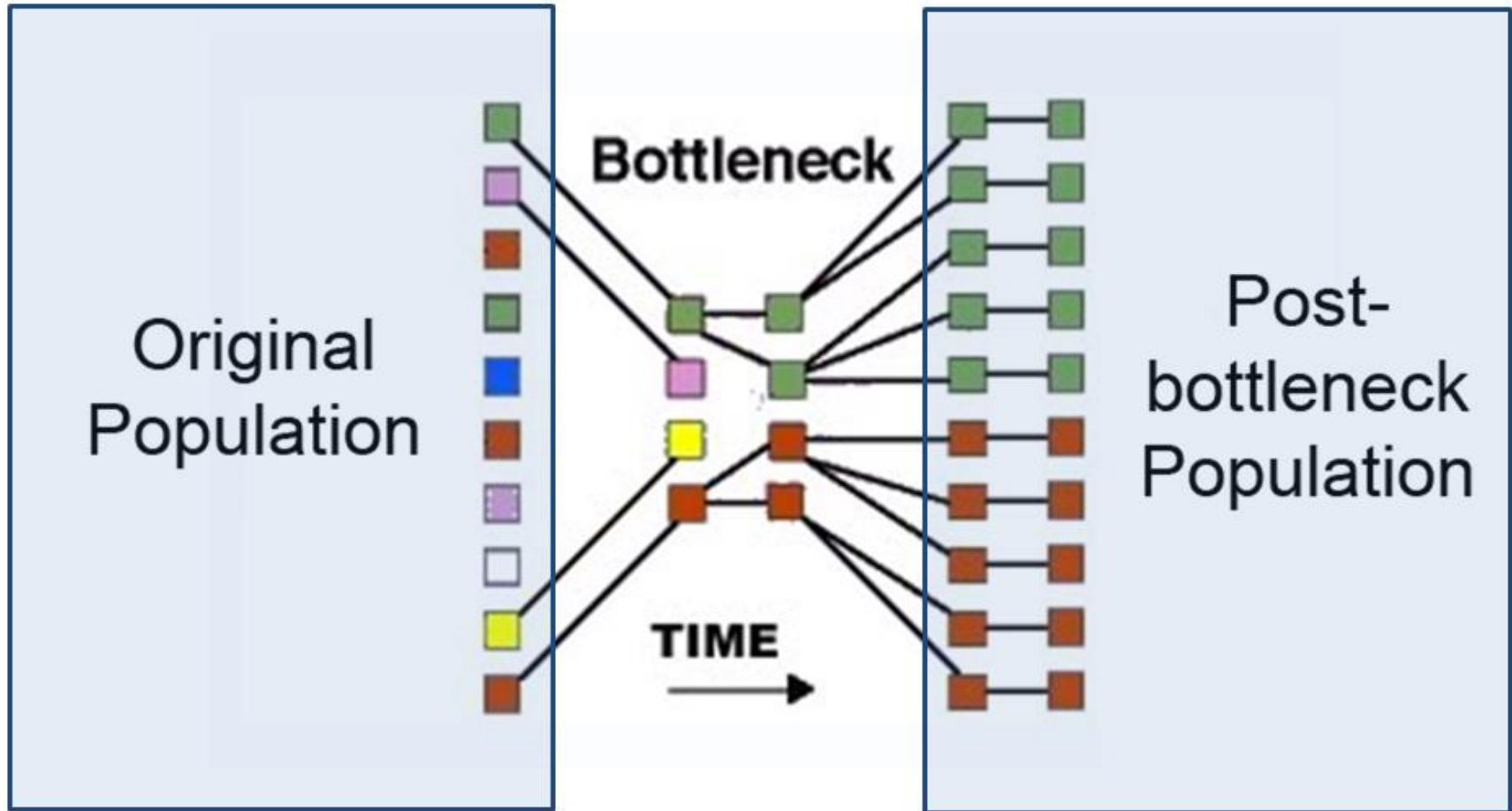
# Genetics & Population size

## GENETIC DRIFT vs SELECTIVE POWER



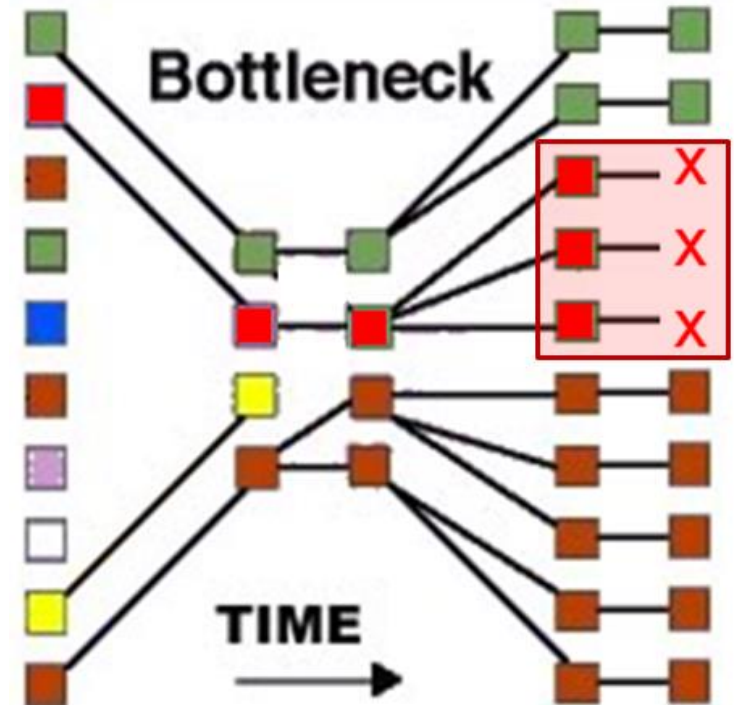
# Genetics & Population size

## BOTTLENECKS IN THE PAST...



# Genetics & Population size

## SUBHEADING

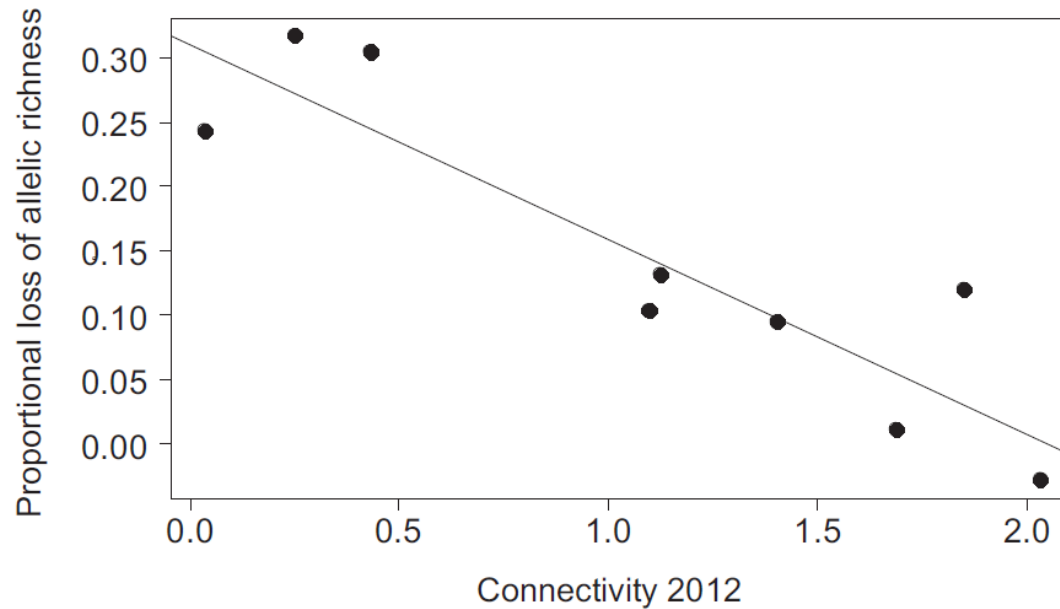


## Northern Elephant Seal:

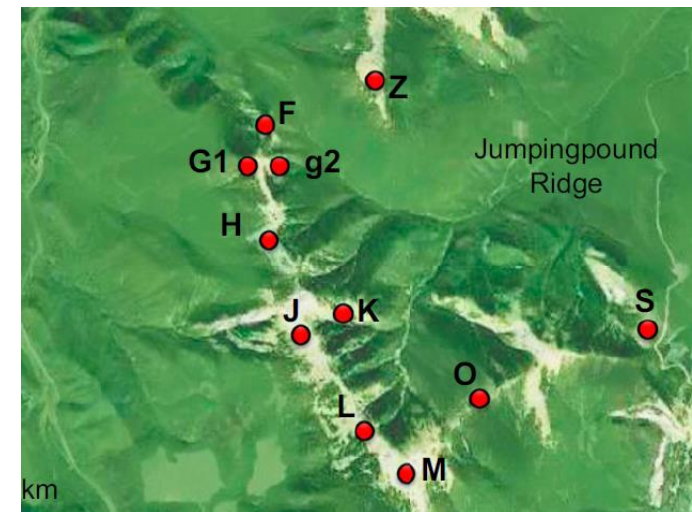
20-30 individuals in late 1800s

>175,000 now

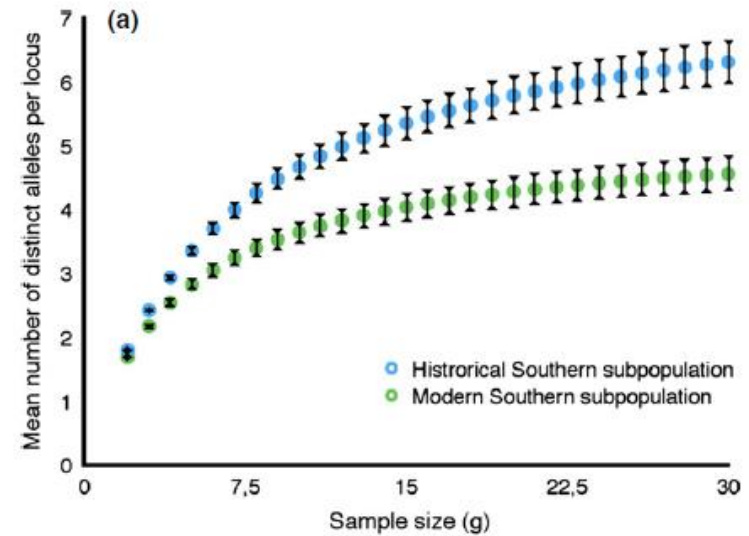
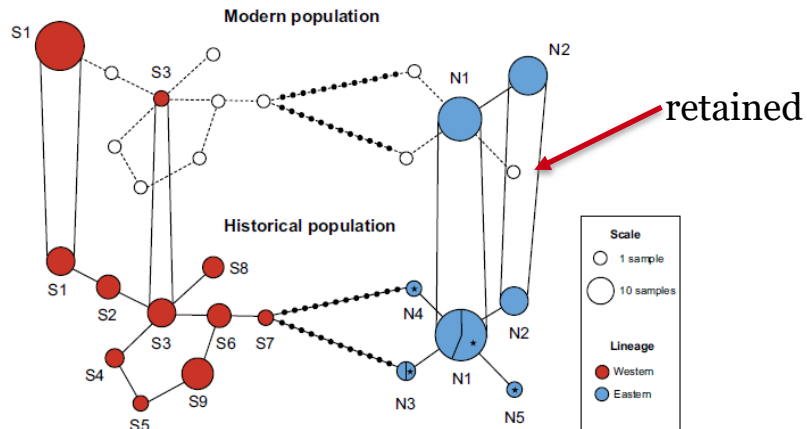
# Importance of connectivity



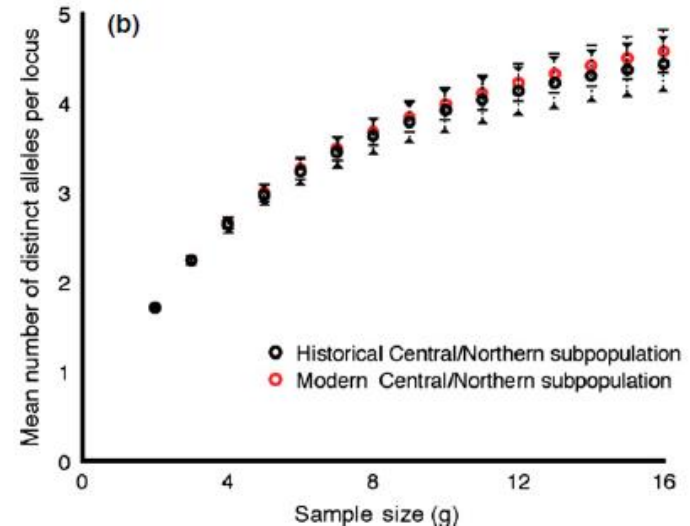
Jangjoo et al. 2016 PNAS



# Magnitude of bottleneck effects vary across a distribution



Xenidoudakis et al.  
2015 Mol Ecol





# Genetics & Population size

## LARGE vs SMALL POPS

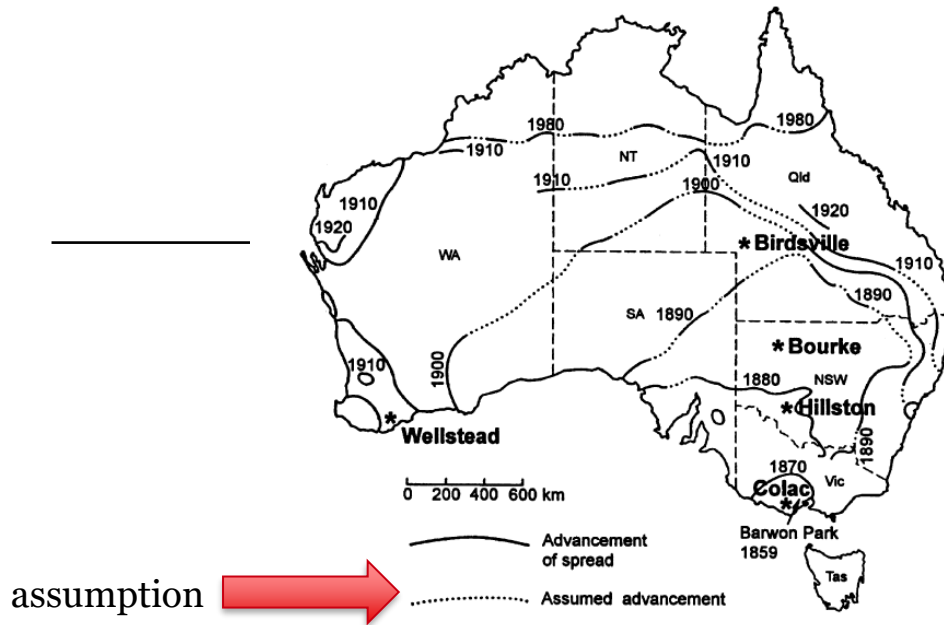
Original release by Thomas Austin in 1859 (Vic).

Austin hunted rabbits on his weekends in England. Upon arriving in Australia, Austin requested **12 grey rabbits, 5 hares, 72 partridges and some sparrows** to fuel his hobby.

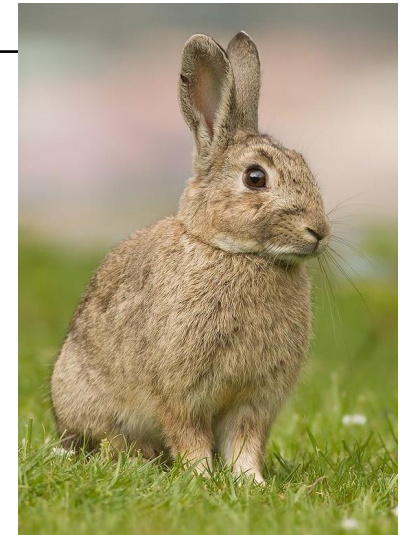
His nephew sent **grey & domestic rabbits** to meet this order.

Many other releases of rabbits following Austin.





**Fig. 1** The summary spread of the rabbit over mainland Australia (after Stodart & Parer 1988). Map also shows geographical location of sample sites.



**Table 1** Estimates of genetic diversity (Mean  $\pm$  SE) for rare alleles (rA; = 0.05 frequency), number of alleles (A), observed and expected heterozygosity ( $H_O$  and  $H_E$ ) and unique alleles (uA) across all loci for all countries (bold) and across individual Australian populations. Results of heterozygosity excess test (HET), mode shift and  $M$  ratio analysis where ‘-’ indicates no significant reduction in  $N_e$

Sample location	<i>n</i>	rA	A	$H_O$	$H_E$	uA	HET TPM	Mode shift	$M$ ratio§ (M)
<b>Spain†</b>	110	$4.10 \pm 0.32$	$9.62 \pm 0.39$	$0.66 \pm 0.03$	$0.79 \pm 0.02$	$4.86 \pm 0.70$	—	—	—
<b>France‡</b>	257	$1.23 \pm 0.20$	$5.03 \pm 0.26$	$0.63 \pm 0.03$	$0.64 \pm 0.03$	$0.86 \pm 0.26$	—	—	0.477–0.554*
<b>England‡</b>	340	$1.68 \pm 0.22$	$5.05 \pm 0.29$	$0.44 \pm 0.06$	$0.64 \pm 0.04$	$1.00 \pm 0.60$	—	—	—
<b>Australia</b>	252	$0.97 \pm 0.17$	$5.03 \pm 0.21$	$0.66 \pm 0.02$	$0.67 \pm 0.01$	$0.29 \pm 0.18$	—	—	—
Wellstead	49	$1.84 \pm 0.55$	$5.57 \pm 0.61$	$0.64 \pm 0.06$	$0.65 \pm 0.03$	$0.71 \pm 0.42$	—	—	0.519*
Birdsville	49	$1.14 \pm 0.26$	$5.00 \pm 0.53$	$0.64 \pm 0.04$	$0.66 \pm 0.03$	$0.43 \pm 0.20$	—	—	0.514*
Bourke	47	$1.00 \pm 0.22$	$4.86 \pm 0.26$	$0.69 \pm 0.04$	$0.68 \pm 0.02$	$0.14 \pm 0.14$	—	—	0.491*
Hillston	57	$0.43 \pm 0.30$	$5.14 \pm 0.46$	$0.72 \pm 0.04$	$0.72 \pm 0.03$	$0.14 \pm 0.14$	*	—	0.525*
Colac	50	$0.43 \pm 0.20$	$4.43 \pm 0.30$	$0.63 \pm 0.05$	$0.65 \pm 0.01$	0	*	*	0.437*

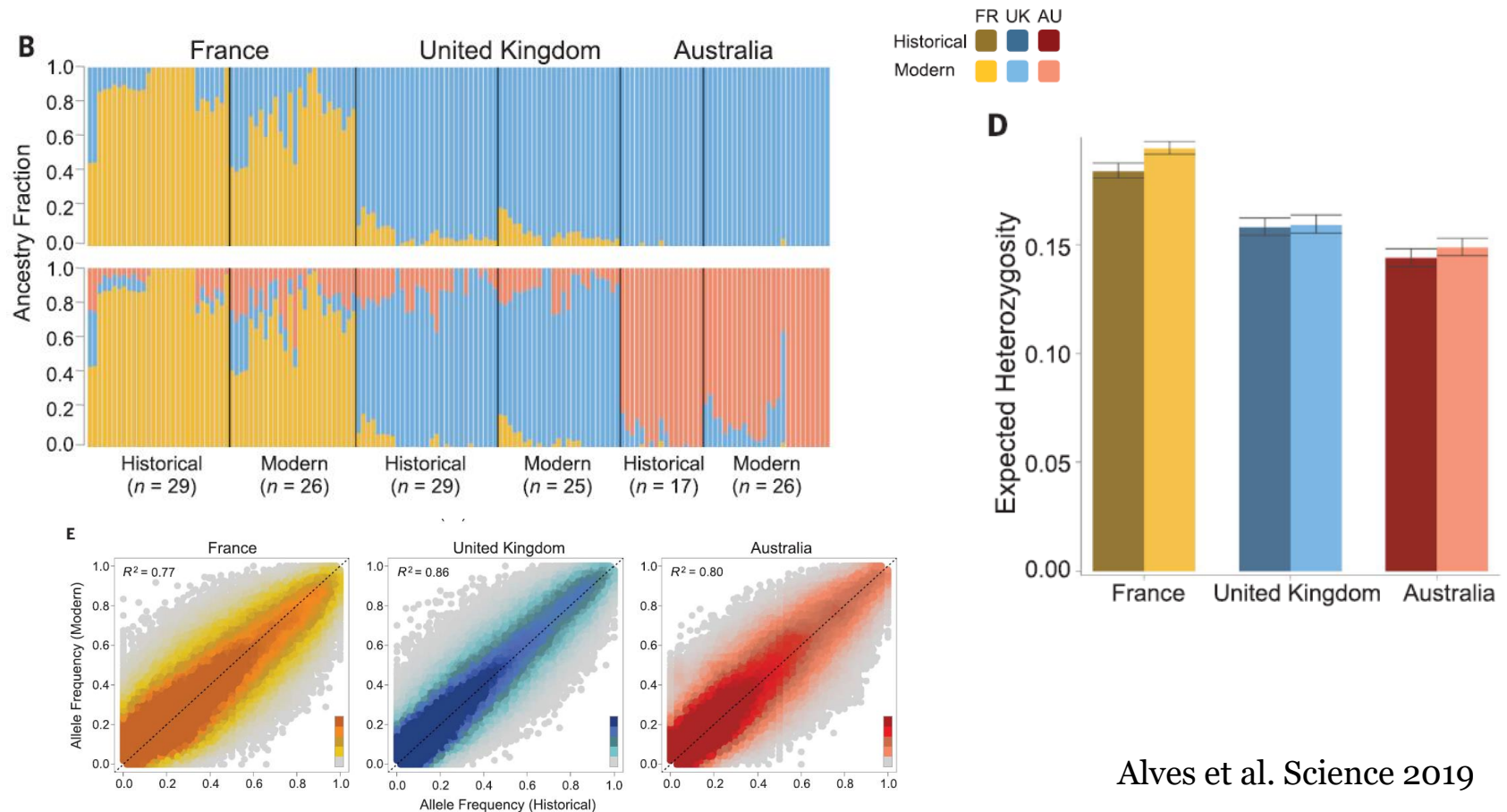
†Data obtained from Queney *et al.* (2001).

‡Data based on sat5, sat7 and sat8 from Surridge *et al.* (1999).

§Significance based on  $M < M_C$  where  $M_C = 0.712-0.828$ ; \* $P < 0.05$ .

No sequential  
loss in  
variation?

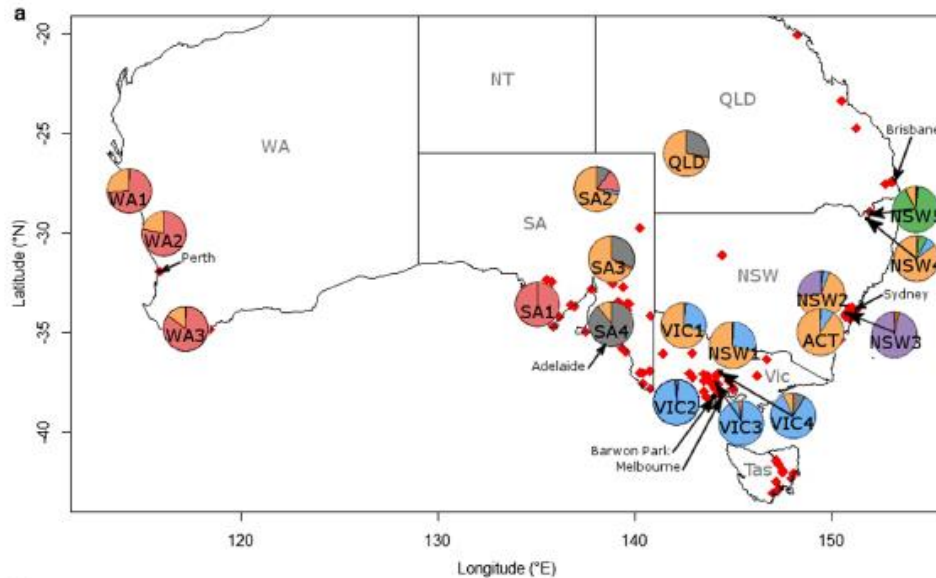
# Lower Australian Vg supported by large scale SNP analysis



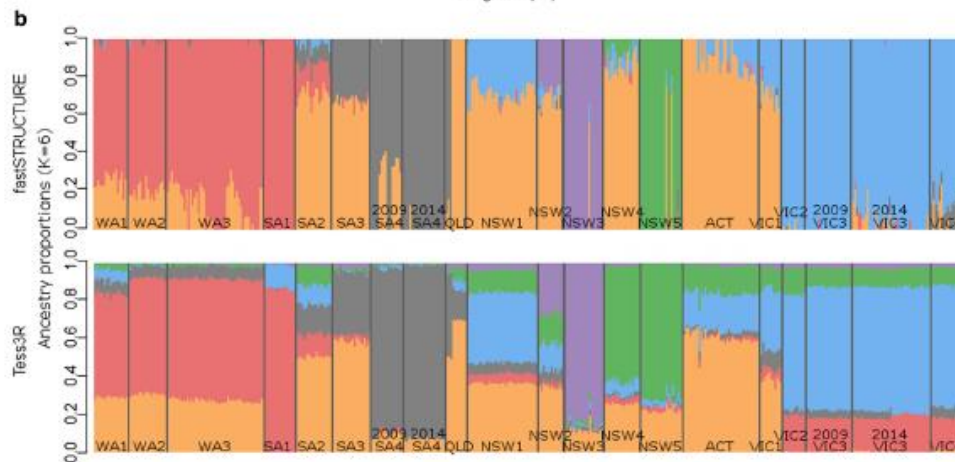
Alves et al. Science 2019



# Multiple Introductions

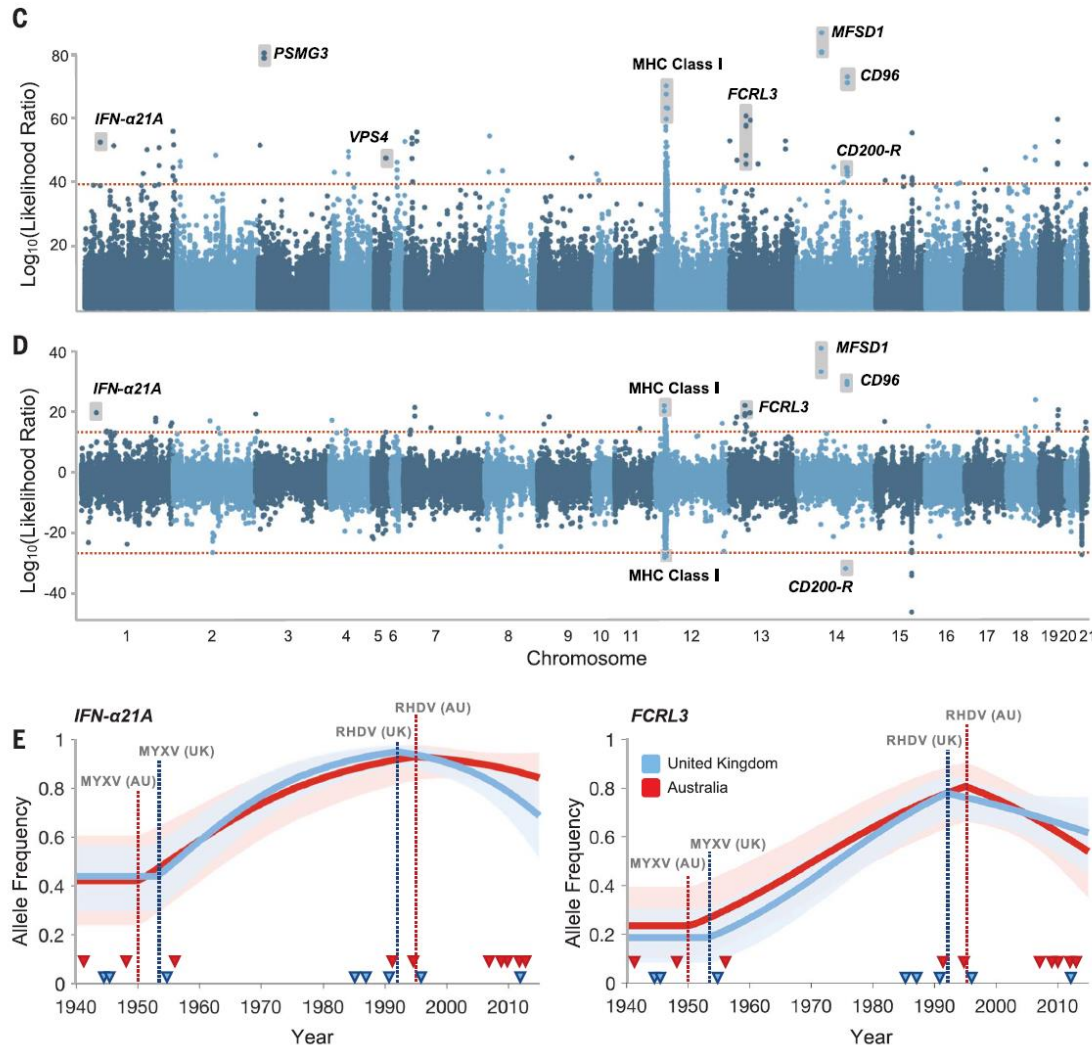


~ 40000 SNPs



Iannella et al.  
Biological Invasions 2019

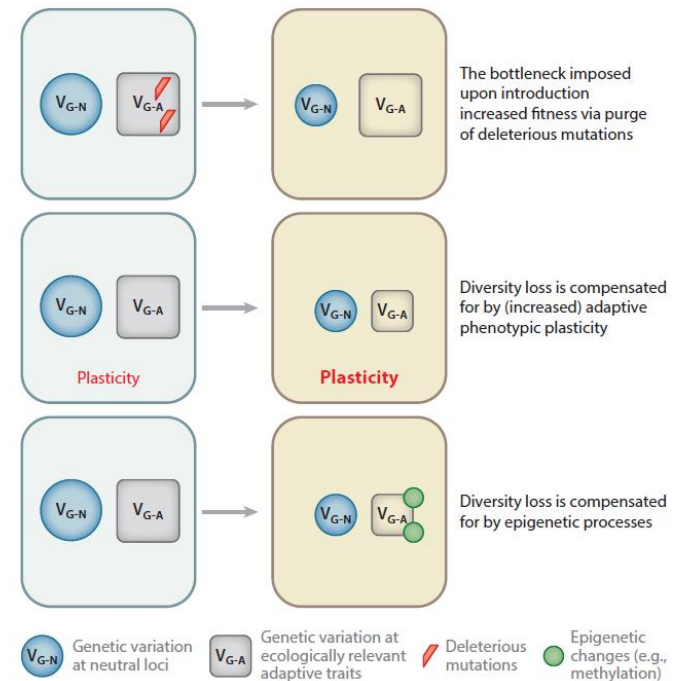
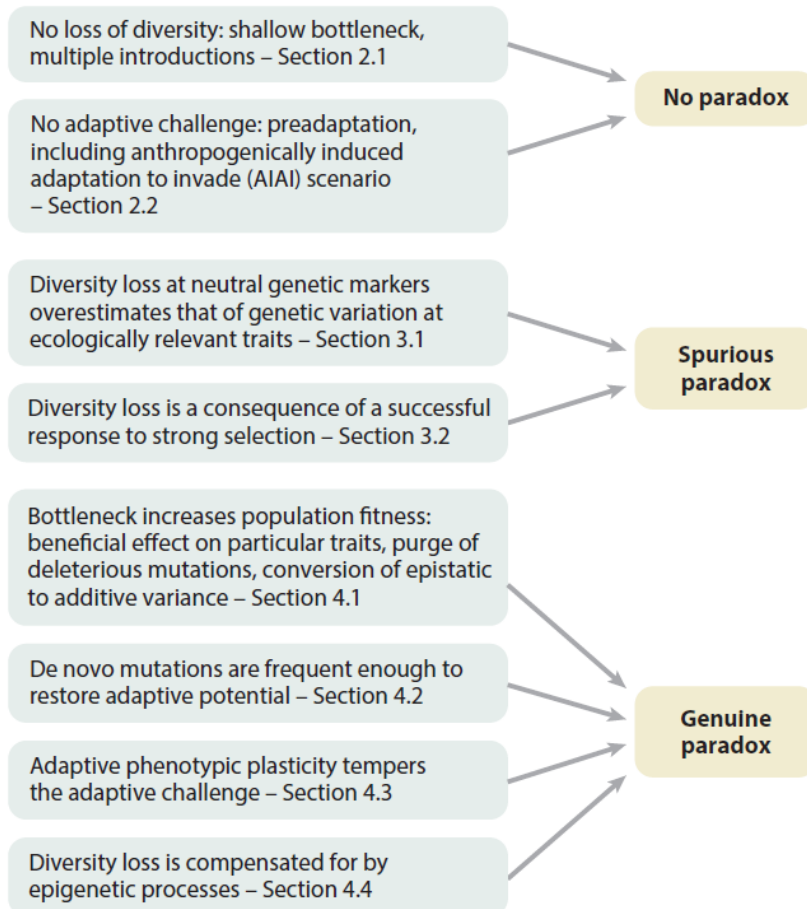
# Enough standing genetic variation for evolution



Alleles selected for  
by myxoma virus

Decrease in  
Myxoma virulence

# Invasive species success- a genetic paradox?



Estoup et al. 2016 Annual Reviews

# Decrease in methylation expected in new invasives

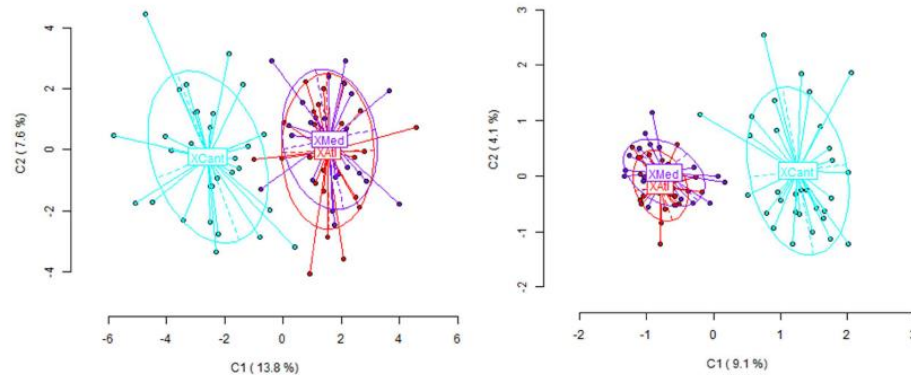


Figure 2. Two-dimensional visualization of the Principal Component Analysis (PCoA) of the detected methylation patterns in *Xenostrobus securis*, with the epigenetic variation (methylation-sensitive loci) on the left and the genetic variation (no methylated loci, NML) on the right. The individuals of each population are represented by the acronyms XAtl, XCant and XMed for the Atlantic international port, Cantabric international port and Mediterranean lagoon populations respectively.

New

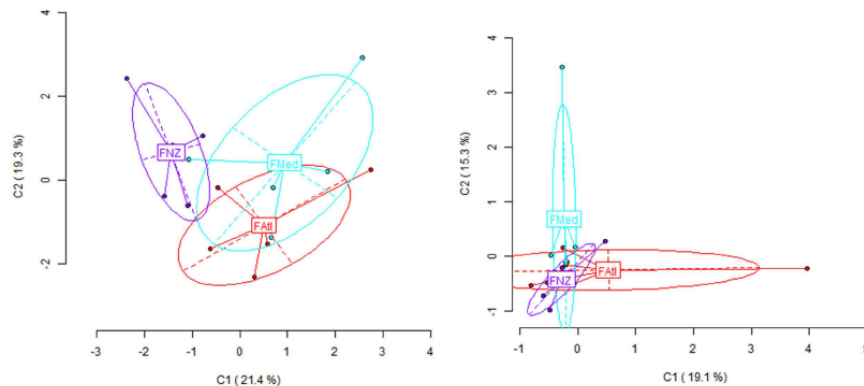


Figure 4. Two-dimensional visualization of the Principal Component Analysis (PCoA) of the detected methylation patterns in *Ficopomatus enigmaticus*, with the epigenetic variation (methylation-sensitive loci) on the left and the genetic variation (no methylated loci, NML) on the right. Each population is represented by the acronyms FNZ, FCant and FMed for samples, respectively, from the international Napier port in New Zealand, Cantabric fishing port and Mediterranean lagoon locations.

old



Next Lecture

# Inbreeding