

STEPHANIE L. JOHNSON, PH.D.

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PROFILE

Interdisciplinary scientist with 12 years of experience designing and managing research projects at the interface of experimental biophysics, biochemistry, and computation, for the study of DNA-protein interactions and genome structure-function relationships.

- Exceptional quantitative and critical reasoning skills, trained through Ph.D. work in an applied physics lab at the California Institute of Technology, and postdoctoral work in quantitative enzymology at the University of California, San Francisco.
- Accustomed to mastering new techniques from diverse fields quickly and routinely, and to communicating clearly and efficiently between fields, due to a highly interdisciplinary career.
- Strong record of productivity, with 8 publications, one recognized by the Faculty of 1000, and a Best Poster Prize at an American Society for Biochemistry and Molecular Biophysics symposium.

EDUCATION

California Institute of Technology

Ph.D. in Biochemistry and Molecular Biophysics 2012

Stanford University

B.S. with Distinction and Honors in Biology 2006

SKILLS

Communication: Strong verbal and written communication skills, especially across disciplines; wrote three successful grants and have given talks at both biophysics and biology conferences.

Management: Extensive experience teaching and mentoring students, and guiding students' research projects both one-on-one and in small teams; and in taking projects from concept to completion independently, with minimal oversight.

Reagent preparation: Cloning, protein purification from *E. coli* and *S. cerevisiae*, site-specific protein and DNA labeling, surface attachment chemistry, and standard *in vitro* chromatin reconstitution techniques, including large-scale nucleosome assembly from purified octamer and DNA.

Assays: Gel-based and fluorescence-based analytical techniques, and single-molecule approaches (tethered particle motion, single-molecule FRET, optical trapping).

Microscopy: Light microscopy (including differential interference contrast) and total internal reflection fluorescence microscopy. Experience with microscopy automation via Micro-Manager programming.

Programming: Proficient in Matlab. Experience with Python (numpy, scipy, matplotlib).

Computation/Modeling: Training in statistical mechanics, and in analyzing reaction kinetics at both ensemble and single molecule levels. Experience with statistical modeling and inference, including Expectation-Maximization and Markov chain Monte Carlo fitting methods for hidden Markov models.

RESEARCH EXPERIENCE

University of California, San Francisco

San Francisco, CA

Postdoctoral Scholar in Biochemistry and Biophysics

February 2013 – present

Advisor: Geeta Narlikar

Project: Biophysical and biochemical characterization of chromatin remodeling enzymes.

- Assay development: Established the biophysical assay of single molecule FRET in Dr. Narlikar's biochemistry lab, which had no prior experience with such techniques; developed fast, robust, open source analysis software for this assay.
- Mechanism of INO80: Reported the first mechanistic description of the INO80 chromatin remodeler, with implications for its essential roles in transcriptional regulation and DNA damage repair. INO80's mechanism had remained opaque to traditional ensemble approaches.

- Regulation by both substrate and self: Demonstrated how the human SNF2h chromatin remodeler, which has roles in transcriptional regulation and genome architecture, is regulated by both chromatin epitopes and internal conformational changes.
- Mentorship: Direct supervisor for two San Francisco State masters students.
- Wrote a successful Leukemia and Lymphoma Society fellowship and an NIH R01 grant.
- Resulted in a Best Poster Prize at a 2016 American Society for Biochemistry and Molecular Biophysics special symposium, and 2 publications, plus 2 in preparation/revision.

California Institute of Technology
Postdoctoral Scholar in Applied Physics
Ph.D. in Biochemistry and Molecular Biophysics

Pasadena, CA
 May 2012 – December 2012
 September 2006 – April 2012

Advisor: Rob Phillips

Thesis: DNA mechanics and transcriptional regulation in the *E. coli lac* operon.

- Assay development: Extended the single molecule technique of tethered particle motion, with a statistical mechanical model of the system, to make robust, quantitative measurements of DNA-protein binding constants and of the mechanical properties of DNA.
- Sequence-dependent DNA flexibility: Showed that DNA flexibility is highly context dependent, in that DNA sequence flexibility rules do not carry over between biological contexts.
- Wrote a successful NSF Graduate fellowship to support this work.
- Culminated in five publications, one recognized by the Faculty of 1000 as being of special significance in its field.

Stanford University
Undergraduate Research Assistant

Stanford, CA
 June 2004 – May 2006

Advisor: Steven Block

Honors thesis project: Worked toward a single-molecule optical trapping assay for studying the ability of *E. coli* RNA Polymerase to transcribe through DNA-bound factors.

SOFTWARE

Traces (github.com/stephlj/Traces), for extracting time course data from microscopy images of single-molecule fluorescence resonance energy transfer (FRET).

Slopey (github.com/stephlj/slopey), continuous-time hidden Markov model for quantifying non-instantaneous state transitions in noisy time course data with sub-camera-frame resolution.

**AWARDS/
HONORS**

University of California, San Francisco

Leukemia and Lymphoma Society Career Development Program Fellow (2014-2017)

Best Poster Prize at the American Society for Biochemistry and Molecular Biology special symposium, Transcriptional Regulation: Chromatin and RNA Polymerase II (2016)

California Institute of Technology

First-author publication Johnson et al (2012) recommended by the Faculty of 1000 (2012)

National Science Foundation Graduate Research Fellow (2008-2011)

Virginia Gilloon Fellowship for Women in Science and Engineering (2006)

Stanford University

Phi Beta Kappa (2006)

**TEACHING /
MENTORING**

2015 - present: Mentored a younger female graduate student as she transitions from a cell biology background to a biophysics lab.

2014 - 2016: Direct supervisor for two San Francisco State masters students as part of a joint UCSF-SF State program to recruit under-represented minorities into post-graduate STEM programs. One student is now a Clinical Trials Associate at ProTrials Research Inc and is applying to medical school; the other is an Associate Scientist at SutroVax.

2007 - 2012: Mentored three younger graduate students at Caltech representing minorities in the sciences who are now in postdoctoral positions at the NIH and at two University of California schools.

2009, 2010, 2012: Teaching assistant for BE/APh 161: Physical Biology of the Cell at the California Institute for Technology (graduate course in the application of statistical mechanics to problems in biology). Head TA 2009 and 2010.

2007, 2008, 2009, 2011: Teaching assistant for BE/APh 262: Physical Biology Bootcamp at the California Institute for Technology (week-long intensive introductory biophysics lab course). Responsibilities included giving lectures on Matlab and basic molecular biology, and leading 3-4 person teams in exploratory biophysics projects.

2008: Teaching assistant for Bi 1: Biology and Biophysics of Viruses at the California Institute for Technology (introductory biology for non-majors).

2006: Teaching assistant for Bi/Ch 110: Introduction to Biochemistry at the California Institute for Technology.

FUNDING

National Institutes of Health 2016 – 2018
 PI: Dr. Geeta Narlikar R01GM073767
Wrote substantial portions of this R01, based on my postdoctoral work in the Narlikar lab.

Leukemia and Lymphoma Society Career Development Program 2014 – 2017
 PI: Dr. Stephanie Johnson
Postdoctoral fellowship; \$55,000 for each of three years.

National Science Foundation 2008 – 2011
 PI: Stephanie Johnson
Graduate fellowship; \$40,500 for each of three years.

Virginia Gilloon Fellowship for Women in Science and Engineering 2006
California Institute of Technology internal fellowship for graduate students; \$53,595 for one year.

PRESENTATIONS Oral Presentations

“Regulation of rapid nucleosome sliding by the INO80 chromatin remodeling complex,” at the 2018 Biophysics of Nuclear Organization and Function QB3 Symposium, Berkeley, CA, July 24, 2018.

“Mechanism of nucleosome sliding in yeast INO80 as revealed by single-molecule FRET,” at the EMBO conference The Nucleosome: From Atoms to Genomes, at EMBL in Heidelberg, Germany, August 31st, 2017.

“Protomer coordination in a dimeric chromatin remodeling motor,” Best Poster Prize talk at the 2016 American Society for Biochemistry and Molecular Biology special symposium on Transcriptional Regulation: Chromatin and RNA Polymerase II, Snowbird, UT, October 10th, 2016.

“Transcription Factor Mediated Looping and the Role of DNA Flexibility,” at the Coarse-Grain Mechanics of DNA: Part II From Electrons to Oligomers CECAM conference at the Swiss Federal Institute of Technology Lausanne (EPFL), Switzerland, September 1st, 2011.

Recent Poster Presentations

“Regulation of rapid nucleosome sliding by substrate cues and accessory subunits in the yeast INO80 chromatin remodeling complex,” with Zhou C.Y., Lee L.J., Longhurst A.D., Johnson M.J., and Narlikar G.J., at the 2018 Genome Biophysics: Integrating Genomics and Biophysics to Understand Structural and Functional Aspects of Genomes meeting, Santa Cruz, CA, August 22, 2018.

“Mechanism of nucleosome sliding in yeast INO80 as revealed by single-molecule FRET,” with Zhou C.Y., Lee L.J., Longhurst A., Johnson M.J., and Narlikar G.J., at the 2018 annual Biophysical Society Meeting, San Francisco, CA, February 2th, 2018.

“Single-molecule mechanistic dissection of ATP-dependent chromatin remodeling,” with Johnson M.J., Gamarra N.I., and Narlikar G.J., at the 2016 American Society for Biochemistry

and Molecular Biology special symposium on Transcriptional Regulation: Chromatin and RNA Polymerase II, Snowbird, UT, October 7th, 2016. *Winner of one of two Best Poster awards.*

“Single-molecule mechanistic dissection of a chromatin remodeling motor,” with Johnson M.J., Gamarra N.I., and Narlikar G.J., at the 2016 annual Biophysical Society Meeting, Los Angeles, CA, February 28th, 2016.

PUBLICATIONS Peer-reviewed publications

Gamarra N., **Johnson S.L.**, Trnka, M.J., Burlingame, A.L., and Narlikar G.J. (2018) The nucleosomal acidic patch relieves auto-inhibition by the ISWI remodeler SNF2h. *eLIFE* 7: e35322.

Zhou C.Y.*, **Johnson S.L.***, Lee L.J., Longhurst A.D., Beckwith S., Johnson M.J., Morrison A., and Narlikar G.J. (2018) The yeast INO80 complex operates as a tunable DNA length-sensitive switch to regulate nucleosome sliding. *Mol Cell* 69:677-688. (*** equal contribution**)

Zhou C.Y., **Johnson S.L.**, Gamarra N.I., and Narlikar G.J. (2016) Mechanisms of ATP-dependent chromatin remodeling motors. *Annu Rev Biophys* 45: 153-81.

Chen Y.-J.*, **Johnson S.***, Mulligan P.*, Spakowitz A., and Phillips R. (2014) Modulation of DNA loop lifetimes by the free energy of loop formation. *Proc Natl Acad Sci* 111(49): 17402-17407. PMCID: PMC4267329 (*** equal contribution**)

Johnson S., van de Meent J.-W., Phillips R., Wiggins C., and Lindén M. (2014) Multiple Lac-mediated loops revealed by Bayesian statistics and tethered particle motion. *Nucleic Acids Res* 42(16): 10265-10277. PMCID: PMC4176382

Johnson S.*, Chen Y.-J.*, and Phillips R. (2013) Poly(dA:dT)-rich DNAs are highly flexible in the context of DNA looping. *PLOS ONE* 8(10): e75799. PMCID: PMC3795714 (*** equal contribution**)

Boedicker J., Garcia H.G., **Johnson S.**, and Phillips R. (2013) DNA sequence-dependent mechanics and protein-assisted bending in repressor-mediated loop formation. *Physical Biology* 10(6): 066005. PMCID: 3915735.

Johnson S., Lindén M., and Phillips R. (2012) Sequence dependence of transcription factor-mediated DNA looping. *Nucleic Acids Res* 40(16): 7728-7738. PMCID: PMC3439888. *Recommended by the Faculty of 1000 as being of special significance in its field.*

Manuscripts in revision

Armache J.-P.*, Gamarra N.*, **Johnson S.L.**, Leonard J.D., Wu S., Narlikar G.J., and Cheng Y. (2018) Electron cryo-microscopy structure of remodeler-nucleosome intermediates suggest allosteric control through the nucleosome. *in revision*. (*equal contribution)