

Automated ChIP-Seq Analysis and Reporting Pipeline

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Objective

Develop a reproducible ChIP-Seq pipeline that can adapt to varying numbers of samples, treatment groups, and parameter sets while allowing for user expansion with custom programs and analysis tasks.

Pipeline usage summary

- Create new directory for analysis from a clone of the pipeline repository
- ②Set input files (fastq, bam)
- Generate sample sheet with pipeline-provided scripts
- (Optional) Modify parameters as needed, add custom tasks
- 6 Run pipeline
- 6 Compile report

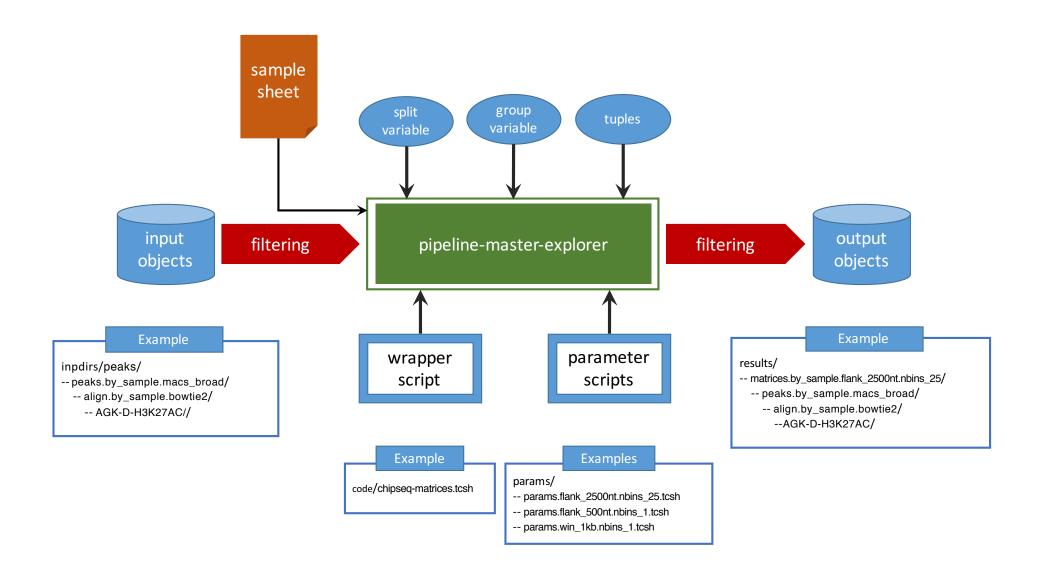


Figure 1: ChIP-Seq pipeline programmatic workflow. All sets of parameters are evaluated for each pipeline task in a combinatorial fashion

ChIP-Seq Pipeline tasks **Programs Used** Alignment bowtie2 bowtie2, R Alignment Stats

Quality Control FastQC, deepTools

MACS Peak Calling

GenomicTools Matrix Correlation

Principal Component Analysis **R** Heatmap Clustering DiffBind Differential Peak Binding Visualization

developed methods listed in bold.

R, μ T_EX 2_{ε} Automatic Reporting

Table 1: ChIP-Seq pipeline standard components. Internally

Auto-report Sample Output

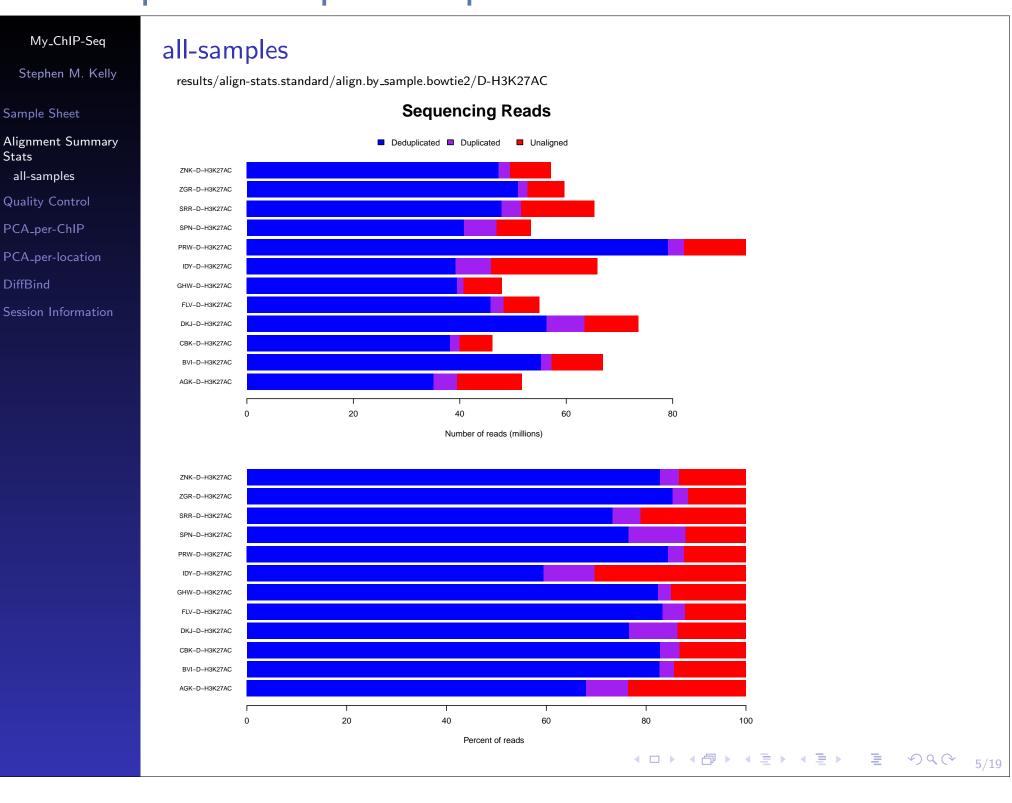


Figure 2: Alignment summary statistics

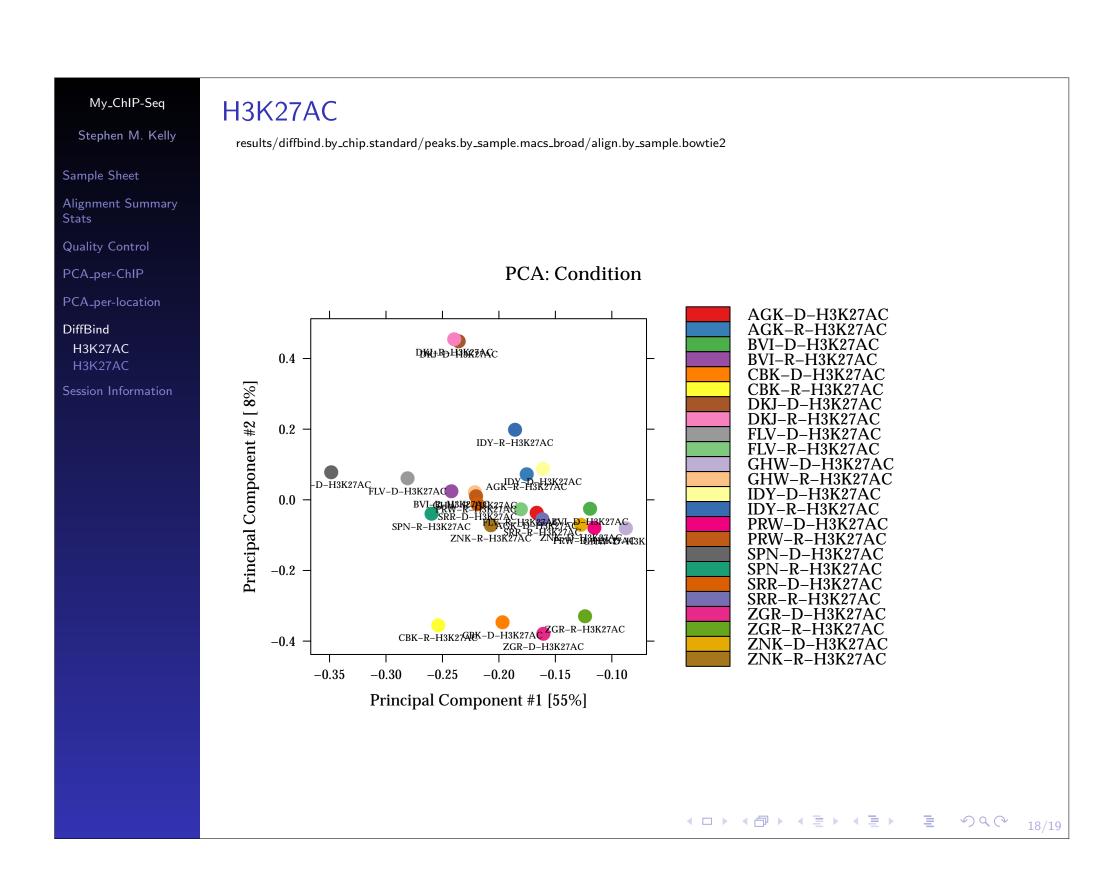


Figure 4: Principal component analysis

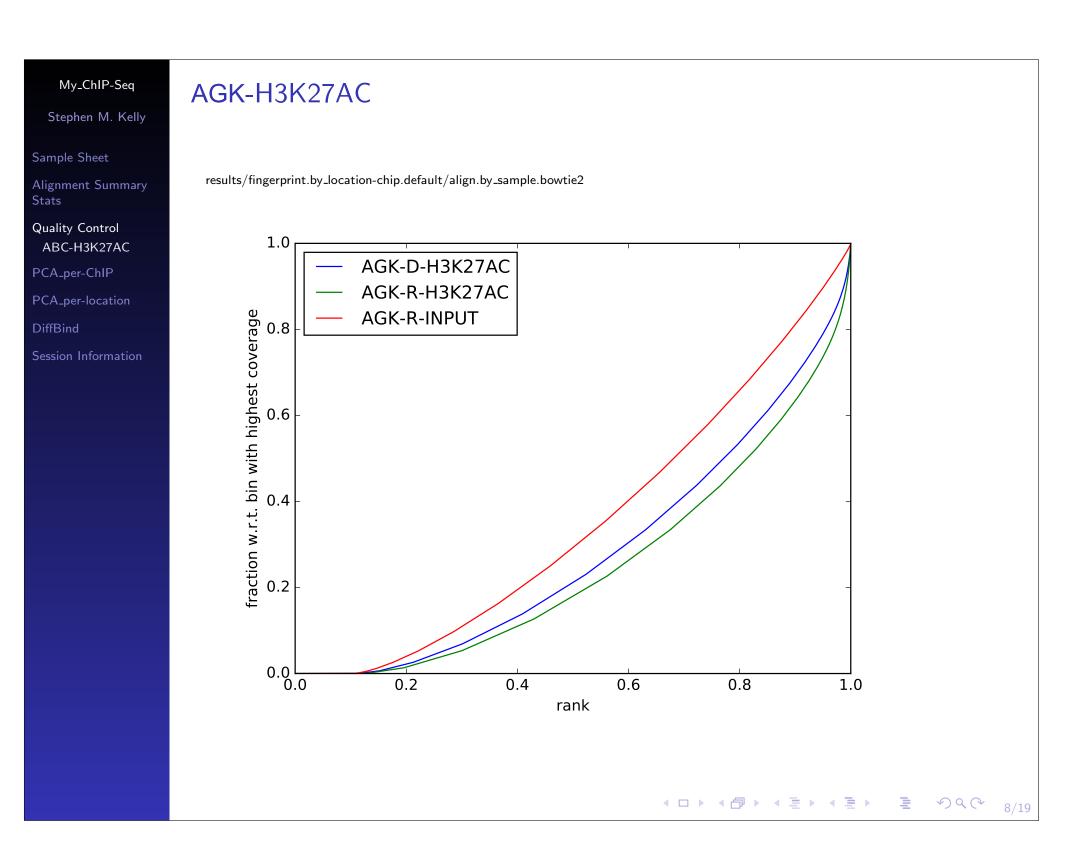


Figure 3: Quality control metrics

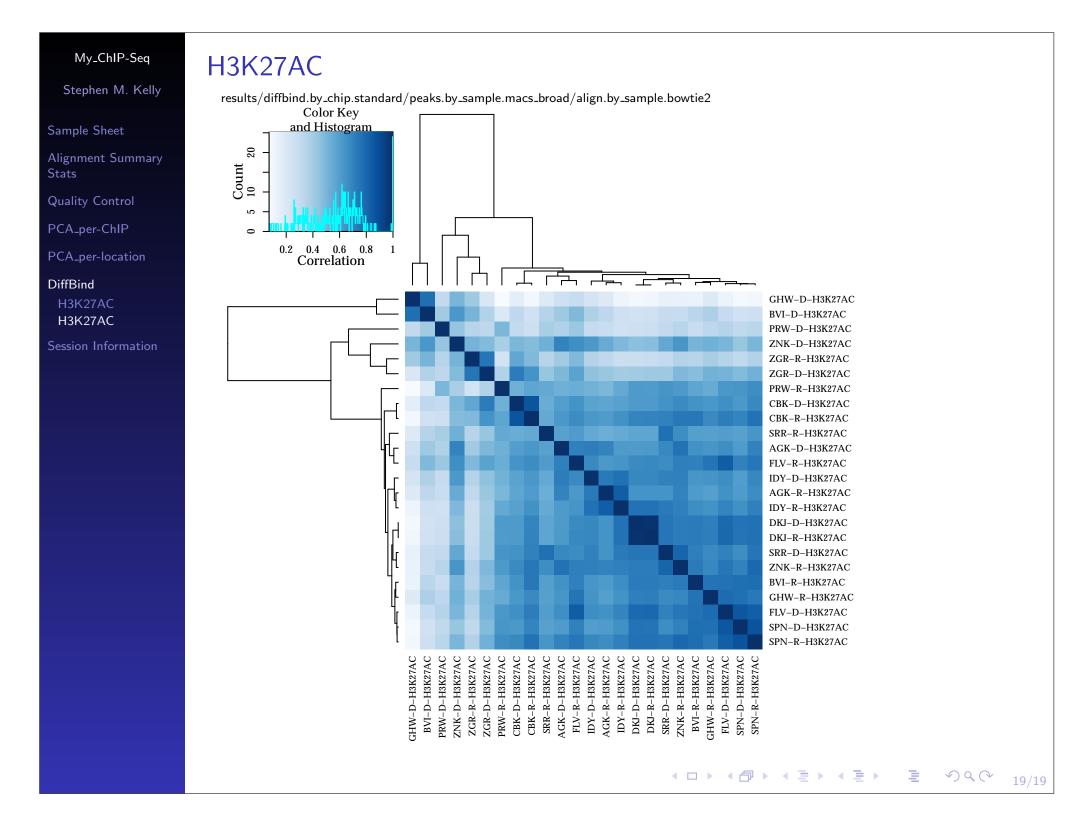


Figure 5: Differential binding heatmaps

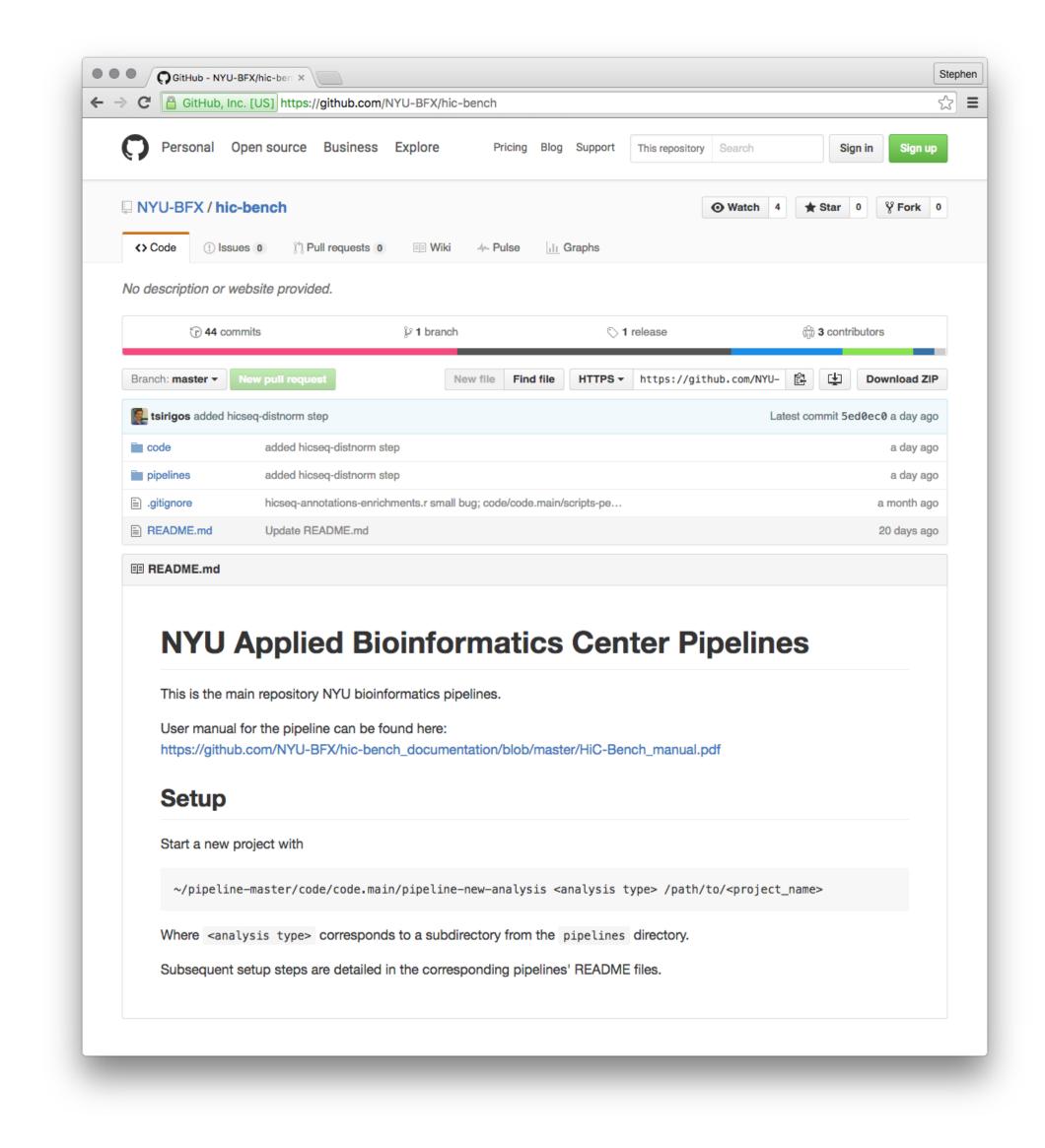


Figure 6: Our ChIP-Seq pipeline is part of the HiC-Bench software package, available on GitHub

Future Developments

- Automated motif analysis
- Peak overlapping and filtering

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Software

- Web: http://www.med.nyu.edu/ocs/applied-bioinformatics-center
- GitHub: https://github.com/NYU-BFX/hic-bench
- Zenodo: https://zenodo.org/record/47676
- Contact: stephen.kelly@nyumc.org, aristotelis.tsirigos@nyumc.org