

Package ‘ICAMS’

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Type Package

Title In-depth Characterization and Analysis of Mutational Signatures

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Description A toolkit for analysis and visualization of experimentally elucidated mutational signatures -- the kind of analysis and visualization presented in Boot et al., "In-depth characterization of the cisplatin mutational signature in human cell lines and in esophageal and liver tumors", 2018, <https://genome.cshlp.org/content/28/5/654.short>. This package has functions to read in variant call files and to collate and plot the mutational spectra.

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R topics documented:

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CatalogRowOrder	<i>Canonical order of row names in a catalog</i>
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Description

Canonical order of row names in a catalog

Usage

catalog.row.order

Format

A list which contains string of characters indicating the canonical order of row names in a catalog.

Note

In the ID (insertion and deletion) catalog, deletion repeat size ranges from 0 to 5+, but for plotting and end user documentation it ranges from 1 to 6+.

CollapseCatalog	<i>Collapse catalog functions</i>
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Description

Collapse a catalog matrix

Usage

Collapse192To96(catalog)

Collapse1536To96(catalog)

Collapse144To78(catalog)

Arguments

catalog A catalog as defined in [ICAMS](#)

Details

Collapse192To96 Collapse a SNS 192 catalog matrix to a SNS 96 catalog matrix.
Collapse1536To96 Collapse a SNS 1536 catalog matrix to a SNS 96 catalog matrix.
Collapse144To78 Collapse a DNS 144 catalog matrix to a DNS 78 catalog matrix.

Value

A catalog as defined in [ICAMS](#)

FindDelMH	<i>Return the length of microhomology at a deletion</i>
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Description

Return the length of microhomology at a deletion

Usage

FindDelMH(context, deleted.seq, pos, trace = 0)

Arguments

context The deleted sequence plus ample surrounding sequence on each side (at least as long as del . sequence).
deleted.seq The deleted sequence in context. #'
pos The position of del . sequence in context.
trace If > 0, cat various messages.

Details

This function is primarily for internal use, but we export it so that the logic behind it will be documented for users.

Example:

GGCTAGTT aligned to GGCTAGAACTAGTT with a deletion represented as:

```
GGCTAGAACTAGTT
GG-----CTAGTT GGCTAGTT GG[CTAGAA]CTAGTT
                        ----  ----
```

Presumed repair mechanism leading to this:

```
....
GGCTAGAACTAGTT
CCGATCTTGATCAA
```

=>

```
....
GGCTAG      TT
CC      GATCAA
      ....
```

=>

```
GGCTAGTT
CCGATCAA
```

The deletion caller can represent the same deletion in several different, but completely equivalent, ways.

```
GGC-----TAGTT GGCTAGTT GGC[TAGAAC]TAGTT
                        *  ---  *  ---
```

```
GGCT-----AGTT GGCTAGTT GGCT[AGAACT]AGTT
                        **  --  **  --
```

```
GGCTA-----GTT GGCTAGTT GGCTA[GAACTA]GTT
                        ***  -  ***  -
```

```
GGCTAG-----TT GGCTAGTT GGCTAG[AACTAG]TT
                        ****  ****
```

A deletion in a *repeat* can also be represented in several different ways. A deletion in a repeat is abstractly equivalent to microhomology that spans the entire deleted sequence. For example;

```
GACTAGCTAGTT
GACTA----GTT GACTAGTT GACTA[GCTA]GTT
                        ***  -***  -
```

is really a repeat

```

TODO(Steve): add check in code
GACTAG-----TT GACTAGTT GACTAG[CTAG]TT
               ****  ----

GACT-----AGTT GACTAGTT GACT[AGCT]AGTT
               **  ----*  --

```

But the function only flags this with a -1 return; it does not figure out the repeat extent.

In the implementation, the function finds:

1. The maximum match of undeleted sequence on left that is identical to the right end of the deleted sequence, and
2. The maximum match of undeleted sequence on the right this is identical to the left end of the deleted sequence.

The microhomology sequence is the concatenation of items (1) and (2).

Value

The length of the maximum microhomology of del. sequence in context.

GetVAF

Extract the VAFs (variant allele frequencies) from a VCF file.

Description

Extract the VAFs (variant allele frequencies) from a VCF file.

Usage

GetStrelkaVAF(vcf)

GetMutectVAF(vcf)

Arguments

vcf said VCF as a data.frame.

Value

A vector of VAFs, one for each row of vcf.

Description

A toolkit for analysis and visualization of experimentally elucidated mutational signatures – the kind of analysis and visualization presented in Boot et al., "In-depth characterization of the cisplatin mutational signature in human cell lines and in esophageal and liver tumors", *Genome Research*, 2018, <https://genome.cshlp.org/content/28/5/654.short>.

Details

A key data type in ICAMS is a "catalog" of mutation counts, of mutation densities, or of mutational signatures. A catalog is one of the following:

1. Matrix of mutation counts (one column per sample),
2. Matrix of mutation densities, i.e. mutations per occurrences of source sequences (one column per sample), or
3. Mutational signatures (proportions of different mutations, summing to 1, one column per signature).

ICAMS can read in variant call files (VCFs) generated by Strelka or Mutect, and collate the mutations into "catalogs" of mutational spectra. ICAMS can plot the catalogs of mutational spectra and signatures.

ICAMS can build and plot catalogs of mutational spectra for single nucleotide substitutions (SNS), double nucleotide substitutions (DNS), and small insertions and deletions (ID). It can also read and write these catalogs.

Creating catalogs from variant call files (VCF files)

1. [StrelkaSNSVCFFilesToCatalog](#), which creates 3 SNS catalogs (96, 192, 1536) and 3 DNS catalogs (78, 136, 144) from the Strelka SNS VCFs.
2. [StrelkaIDVCFFilesToCatalog](#), which creates ID (indel) catalog from the Strelka ID VCFs.
3. [MutectVCFFilesToCatalog](#), which creates 3 SNS catalogs (96, 192, 1536), 3 DNS catalogs (78, 136, 144) and ID (indel) catalog from the Mutect VCFs.

Plotting catalogs

Functions for plotting catalogs of mutational spectra or of mutational signatures to a PDF file. Mutational *signatures* are similar to spectra, but where spectra consist of counts of mutations in each mutation class (e.g. ACA > AAA, ACA > AGA, ACA > ATA, ACC > AAC, ...) signatures consist of the proportions of mutations in each class (with all the proportions summing to 1). [PlotCatalogToPdf](#)

Writing catalogs

Functions for writing a catalog of mutational spectra or of mutational signatures to a file on disk. [WriteCatalog](#)

Reading catalogs

Functions for reading files that contain catalogs of mutational spectra or of signatures in standardized format. [ReadCatalog](#)

Transforming catalogs

There is a function to transform catalogs of mutational spectra or signatures to account for differing abundances of the source sequence of the mutations in the genome. For example, mutations from ACG are much rarer in the human genome than mutations from ACC simply because CG dinucleotides are rare in the genome. This function can also transform spectra based on observed genome-wide counts to "density"-based catalogs. In density-based catalogs mutations are expressed as mutations per source sequences. For example, a density-based catalog represents the proportion of ACCs mutated to ATCs, the proportion of ACGs mutated to ATGs, etc. This is opposed to count-based catalogs, which contain the number of ACC-to-ATC mutations, the number of ACG-to-ATG mutations, etc. This function can also transform observed-count based spectra or signatures from genome to exome based counts, or between different species (since the abundances of source sequences vary between genome and exome and between species). [TransformCatalog](#)

Collapsing catalogs

Functions for collapsing a mutational spectrum or signature catalog based on a fined-grained set of features (for example, single-nucleotide substitutions in the context of the preceding and following 2 bases) to a catalog based on a coarser-grained set of features (for example, single-nucleotide substitutions in the context of the immediately preceding and following bases). [CollapseCatalog](#)

Exported data

1. [CatalogRowOrder](#) Canonical order of row names in a catalog.
2. [TranscriptRanges](#) Transcript ranges and strand information for a particular organism.

MutectVCFFilesToCatalog

Create SNS and DNS catalogs from Mutect VCF files

Description

Create 3 SNS catalogs (96, 192, 1536) and 3 DNS catalogs (78, 136, 144) from the Mutect VCFs specified by `vector.of.file.paths`

Usage

```
MutectVCFFilesToCatalog(vector.of.file.paths, genome, trans.ranges)
```

Arguments

`vector.of.file.paths`

A vector containing the paths of the Mutect VCF files.

genome	A particular reference genome(without quotation marks). Use available.genomes to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing genome = "GRCh38" or genome = "hg38". User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing genome = "GRCh37" or genome = "hg19".
trans.ranges	A data.table which contains transcript range and strand information.

Details

This function calls [VCFsToSNSCatalogs](#), [VCFsToDNSCatalogs](#) and [VCFsToIDCatalogs](#)

Value

A list of 3 SNS catalogs (one each for 96, 192, and 1536) , 3 DNS catalogs (one each for 78, 136, and 144) and ID catalog.

Note

SNS 192 and DNS 144 catalog only contains mutations in transcribed regions.

PlotCatalog	<i>Plot catalog functions</i>
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Description

Plot the catalog of one sample which has mutations

Usage

```
PlotCatSNS96(catalog, type, id = colnames(catalog), cex = 0.8,
  grid = TRUE, upper = TRUE, xlabels = TRUE)
```

```
PlotCatSNS192(catalog, type, id = colnames(catalog), cex = 0.8)
```

```
PlotSNSClassStrandBias(catalog, type, id = colnames(catalog), cex = 1)
```

```
PlotCatSNS1536(catalog, type, id = colnames(catalog))
```

```
PlotCatDNS78(catalog, type, id = colnames(catalog))
```

```
PlotDNSClassStrandBias(catalog, type, id = colnames(catalog), cex = 1)
```

```
PlotCatDNS136(catalog, type, id = colnames(catalog))
```

```
PlotCatID(catalog, type, id = colnames(catalog))
```


Arguments

catalog	A catalog as described in ICAMS . The input catalog must be in matrix format, you may use data.matrix to convert a data frame to a numeric matrix. This catalog matrix must have rownames to facilitate sorting in the plotting functions. You may use CatalogRowOrder to give row names to your catalog matrix.
type	A character specifying type of the input catalog, one of "counts", "signature" or "density". If type = "counts", the graph will plot the occurrences of the mutation types in the sample. If type = "signature", the graph will plot mutation signatures of the sample. If type = "density", the graph will plot the rates of mutations per million nucleotides for each mutation type. (Please take note there is no "signature" type for PlotCatDNS136 function, no "density" type for PlotCatID function and the option of type = "density" is not implemented for function PlotCatSNS192, PlotSNSClassStrandBias and PlotDNSClassStrandBias at the current stage.)
id	The identifier of the sample which has mutations.
cex	A numerical value giving the amount by which mutation class labels, mutation counts(if it exists), y axis and its labels, x axis labels and its annotations(if it exists) sample name and legend(if it exists) should be magnified relative to the default.
grid	If TRUE, draw grid lines in the graph.
upper	If TRUE, draw horizontal lines and the names of major mutation class on top of graph.
xlabels	If TRUE, draw x axis labels.

Details

PlotCatSNS96 Plot the SNS 96 mutation catalog of one sample.

PlotCatSNS192 Plot the SNS 192 mutation catalog of one sample.

PlotSNSClassStrandBias Plot the transcription strand bias graph of 6 SNS mutation types ("C>A", "C>G", "C>T", "T>A", "T>C", "T>G") in one sample.

PlotCatSNS1536 Plot the pentanucleotide sequence contexts for one sample, normalized by pentanucleotide occurrence in the genome. The mutation types are in six-letters like CATTAT, first 2-letters CA refers to (-2, -1) position, third letter T refers to the base which has mutation, next second 2-letters TA refers to (+1, +2) position, last letter T refers to the base after mutation.

PlotCatDNS78 Plot the DNS 78 mutation catalog of one sample.

PlotDNSClassStrandBias Plot the transcription strand bias graph of 10 major DNS mutation types ("AC>NN", "AT>NN", "CC>NN", "CG>NN", "CT>NN", "GC>NN", "TA>NN", "TC>NN", "TG>NN", "TT>NN") in one sample.

PlotCatDNS136 Plot the tetranucleotide sequence context of 10 major DNS mutation types ("AC>NN", "AT>NN", "CC>NN", "CG>NN", "CT>NN", "GC>NN", "TA>NN", "TC>NN", "TG>NN", "TT>NN") for one sample.

PlotCatID Plot the insertion and deletion catalog of one sample. (Please take note that deletion repeat size ranges from 0 to 5+ in the catalog, but for plotting and end user documentation it ranges from 1 to 6+.)

Value

invisible(TRUE)

PlotCatalogToPdf	<i>Plot catalog to pdf functions</i>
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Description

Plot mutation catalogs of various samples to a PDF file

Usage

```
PlotCatSNS96ToPdf(catalog, name, type, id = colnames(catalog),
  grid = TRUE, upper = TRUE, xlabel = TRUE)
```

```
PlotCatSNS192ToPdf(catalog, name, id = colnames(catalog),
  type = "counts", cex = 0.8)
```

```
PlotSNSClassStrandBiasToPdf(catalog, name, type, id = colnames(catalog),
  cex = 1)
```

```
PlotCatSNS1536ToPdf(catalog, name, type, id = colnames(catalog))
```

```
PlotCatDNS78ToPdf(catalog, name, type, id = colnames(catalog))
```

```
PlotDNSClassStrandBiasToPdf(catalog, name, type, id = colnames(catalog),
  cex = 1)
```

```
PlotCatDNS136ToPdf(catalog, name, type, id = colnames(catalog))
```

```
PlotCatIDToPdf(catalog, name, type, id = colnames(catalog))
```

Arguments

catalog	A catalog as described in ICAMS . The input catalog must be in matrix format, you may use data.matrix to convert a data frame to a numeric matrix. This catalog matrix must have rownames to facilitate sorting in the plotting functions. You may use CatalogRowOrder to give row names to your catalog matrix.
name	The name of the PDF file to be produced.
type	A character specifying type of the input catalog, one of "counts", "signature" or "density". If type = "counts", the graph will plot the occurrences of the mutation types in the sample. If type = "signature", the graph will plot mutation signatures of the sample. If type = "density", the graph will plot the rates of mutations per million nucleotides for each mutation type. (Please take note there is no "signature" type for PlotCatDNS136ToPdf function, no "density" type for PlotCatIDtoPdf function and the option of type = "density" is not implemented for function PlotCatSNS192ToPdf, PlotSNSClassStrandBiasToPdf and PlotDNSClassStrandBiasToPdf at the current stage.)
id	A vector containing the identifiers of the samples in catalog.
grid	If TRUE, draw grid lines in the graph.
upper	If TRUE, draw horizontal lines and the names of major mutation class on top of graph.

xlabels	If TRUE, draw x axis labels.
cex	A numerical value giving the amount by which mutation class labels, y axis labels, sample name and legend (if it exists) should be magnified relative to the default.

Details

PlotCatSNS96ToPdf Plot the SNS 96 mutation catalog of various samples to a PDF file.

PlotCatSNS192ToPdf Plot the SNS 192 mutation catalog of various samples to a PDF file.

PlotSNSClassStrandBiasToPdf Plot the transcription strand bias graph of 6 SNS mutation types ("C>A", "C>G", "C>T", "T>A", "T>C", "T>G") of various samples to a PDF file.

PlotCatSNS1536ToPdf Plot the 1536 mutation catalog of ≥ 1 samples to a PDF file. The mutation types are in six-letters like CATTAT, first 2-letters CA refers to (-2, -1) position, third letter T refers to the base which has mutation, next second 2-letters TA refers to (+1, +2) position, last letter T refers to the base after mutation.

PlotCatDNS78ToPdf Plot the DNS 78 mutation catalog of various samples to a PDF file.

PlotDNSClassStrandBiasToPdf Plot the transcription strand bias graph of 10 major DNS mutation types ("AC>NN", "AT>NN", "CC>NN", "CG>NN", "CT>NN", "GC>NN", "TA>NN", "TC>NN", "TG>NN", "TT>NN") of various samples to a PDF file.

PlotCatDNS136ToPdf Plot the tetranucleotide sequence contexts of 10 major DNS mutation types ("AC>NN", "AT>NN", "CC>NN", "CG>NN", "CT>NN", "GC>NN", "TA>NN", "TC>NN", "TG>NN", "TT>NN") of various samples to a PDF file.

PlotCatIDToPdf Plot the insertion and deletion catalog of various samples to a PDF file. (Please take note that deletion repeat size ranges from 0 to 5+ in the catalog, but for plotting and end user documentation it ranges from 1 to 6+.)

Value

invisible(TRUE)

ReadAndSplitMutectVCFs

Read and split Mutect VCF files from paths

Description

Read and split Mutect VCF files from paths

Usage

```
ReadAndSplitMutectVCFs(vector.of.file.paths)
```

Arguments

vector.of.file.paths

A vector containing the paths of the VCF files.

Value

A list with 3 in-memory VCFs and two left-over VCF-like data frames with rows that were not incorporated into the first 3 VCFs, as follows:

1. SNS VCF with only single nucleotide substitutions.
2. DNS VCF with only doublet nucleotide substitutions as called by Mutect.
3. ID VCF with only small insertions and deletions.
4. `other.subs` VCF like `data.frame` with rows for coordinate substitutions involving 3 or more nucleotides, e.g. `ACT > TGA` or `AACT > GGTA`.
5. `multiple.alternative.alleles` VCF like `data.frame` with rows for variants with multiple alternative alleles, for example `ACT` mutated to both `AGT` and `ACT` at the same position.

See Also

[MutectVCFFilesToCatalog](#)

ReadAndSplitStrelkaSNSVCFs

Read and split Strelka SNS VCF files from paths

Description

Read and split Strelka SNS VCF files from paths

Usage

```
ReadAndSplitStrelkaSNSVCFs(vector.of.file.paths)
```

Arguments

```
vector.of.file.paths
```

A vector containing the paths of the VCF files.

Value

A list of 3 in-memory objects with the elements: `SNS.vcfs`: List of Data frames of pure SNS mutations – no DNS or 3+BS mutations `DNS.vcfs`: List of Data frames of pure DNS mutations – no SNS or 3+BS mutations `ThreePlus`: List of Data tables with the key `CHROM`, `LOW.POS`, `HIGH.POS` and additional information (reference sequence, alternative sequence, context, etc.) Additional information not fully implemented at this point because of limited immediate biological interest.

See Also

[StrelkaSNSVCFFilesToCatalog](#)

ReadCatalog

Read Catalog Functions

Description

Read a catalog in standardized format from path

Usage

```
ReadCatSNS96(path, strict = TRUE)
```

```
ReadCatSNS192(path, strict = TRUE)
```

```
ReadCatSNS1536(path, strict = TRUE)
```

```
ReadCatDNS78(path, strict = TRUE)
```

```
ReadCatDNS144(path, strict = TRUE)
```

```
ReadCatDNS136(path, strict = TRUE)
```

```
ReadCatID(path, strict = TRUE)
```

Arguments

`path` Path to a catalog on disk in the standardized format.

`strict` If TRUE, do additional checks on the input, and stop if the checks fail.

Details

`ReadCatSNS96` Read a 96 SNS catalog from path

`ReadCatSNS192` Read a 192 SNS catalog from path

`ReadCatSNS1536` Read a 1536 SNS catalog from path

`ReadCatDNS78` Read a 78 DNS catalog from path

`ReadCatDNS144` Read a 144 DNS catalog from path

`ReadCatDNS136` Read a 136 DNS catalog from path

`ReadCatID` Read a ID (insertion/deletion) catalog from path. (Please take note that deletion repeat size ranges from 0 to 5+ in the catalog, but for plotting and end user documentation it ranges from 1 to 6+.)

See also [WriteCatalog](#)

Value

A catalog in canonical in-memory format.

ReadStrelkaIDVCFs	<i>Read Strelka ID (insertion and deletion) VCF files from paths</i>
-------------------	--

Description

Read Strelka ID (insertion and deletion) VCF files from paths

Usage

```
ReadStrelkaIDVCFs(vector.of.file.paths)
```

Arguments

```
vector.of.file.paths
```

A vector containing the paths of the VCF files.

Value

A list of vcfs from vector.of.file.paths.

Note

In the ID (insertion and deletion) catalog, deletion repeat size ranges from 0 to 5+, but for plotting and end user documentation it ranges from 1 to 6+.

See Also

[StrelkaIDVCFFilesToCatalog](#)

revc	<i>Reverse complement every string in string.vec.</i>
------	---

Description

Reverse complement every string in string.vec.

Usage

```
revc(string.vec)
```

Arguments

```
string.vec
```

a vector of type character.

Value

A vector of type characters with the reverse complement of every string in string.vec.

`StrelkaIDVCFFilesToCatalog`*Create ID (indel) catalog from Strelka ID VCF files*

Description

Create ID (indel) catalog from the Strelka ID VCFs specified by `vector.of.file.paths`

Usage

```
StrelkaIDVCFFilesToCatalog(vector.of.file.paths, genome)
```

Arguments

`vector.of.file.paths`

A vector containing the paths of the Strelka ID VCF files.

`genome`

A particular reference genome(without quotation marks). Use [available.genomes](#) to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing `genome = "GRCh38"` or `genome = "hg38"`. User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing `genome = "GRCh37"` or `genome = "hg19"`.

Details

This function calls [VCFsToIDCatalogs](#)

Value

An ID (indel) catalog

Note

In the ID (insertion and deletion) catalog, deletion repeat size ranges from 0 to 5+, but for plotting and end user documentation it ranges from 1 to 6+.

`StrelkaSNSVCFFilesToCatalog`*Create SNS and DNS catalogs from Strelka SNS VCF files*

Description

Create 3 SNS catalogs (96, 192, 1536) and 3 DNS catalogs (78, 136, 144) from the Strelka SNS VCFs specified by `vector.of.file.paths`

Usage

```
StrelkaSNSVCFFilesToCatalog(vector.of.file.paths, genome, trans.ranges)
```

Arguments

<code>vector.of.file.paths</code>	A vector containing the paths of the Strelka SNS VCF files.
<code>genome</code>	A particular reference genome(without quotation marks). Use available.genomes to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing <code>genome = "GRCh38"</code> or <code>genome = "hg38"</code> . User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing <code>genome = "GRCh37"</code> or <code>genome = "hg19"</code> .
<code>trans.ranges</code>	A data.table which contains transcript range and strand information.

Details

This function calls [VCFsToNSCatalogs](#) and [VCFsToDNSCatalogs](#)

Value

A list of 3 SNS catalogs (one each for 96, 192, and 1536) and 3 DNS catalogs (one each for 78, 136, and 144)

Note

SNS 192 and DNS 144 catalog only contains mutations in transcribed regions.

TranscriptRanges	<i>Transcript ranges data</i>
------------------	-------------------------------

Description

Transcript ranges and strand information for a particular organism

Usage

```
trans.ranges.GRCh37
```

```
trans.ranges.GRCh38
```

Format

A data.table which contains transcript range and strand information for a particular organism.

Details

`trans.ranges.GRCh37` A data.table which contains transcript range and strand information for **Human** GRCh37. It is derived from a raw **GFF3** format file, from which only the following four gene types are kept to facilitate transcriptional strand bias analysis: `protein_coding`, `retained_intron`, `processed_transcript` and `nonsense_mediated_decay`. It contains chromosome name, start, end position, strand information and gene name and is keyed by `chrom`, `chromStart`, and `chromEnd`. It can be used in function [StrelkaSNSVCFFilesToCatalog](#).

`trans.ranges.GRCh38` A data.table which contains transcript range and strand information for **Human** GRCh38. It is derived from a raw **GFF3** format file, from which only the following four gene types are kept to facilitate transcriptional strand bias analysis: `protein_coding`, `retained_intron`, `processed_transcript` and `nonsense_mediated_decay`. It contains chromosome name, start, end position, strand information and gene name and is keyed by `chrom`, `chromStart`, and `chromEnd`. It can be used in function [StrelkaSNSVCFFilesToCatalog](#).

TransformCatalog	<i>Transform between count and density catalogs and signatures and between different source sequence abundances.</i>
------------------	--

Description

Transform between count and density catalogs and signatures and between different source sequence abundances.

Usage

```
TransformCatalog(catalog, which.n, source.type,
  target.type = source.type, source.abundance = NULL,
  target.abundance = NULL)
```

Arguments

<code>catalog</code>	An SNS or DNS catalog as described in ICAMS . The input catalog can not be an ID (indel) catalog.
<code>which.n</code>	The length of the source sequences, one of 2:5.
<code>source.type</code>	A character specifying type of the input catalog, one of "counts", "signature" or "density".
<code>target.type</code>	A character specifying type of the output catalog, with the same possible values as <code>source.type</code> .
<code>source.abundance</code>	Either NULL or a numeric vector with one element for each source sequence for the mutation types in <code>catalog</code> or a string specifying such a vector, one of "GRCh37.genome", "GRCh37.exome", "GRCh38.genome", or "GRCh38.exome". This is the abundance upon which the counts, densities, or proportions in <code>catalog</code> are based. For example, for SNS in trinucleotide context, e.g. ACT > AGT or TAC > TTC, the source sequences are ACT and TAC.
<code>target.abundance</code>	Same possibilities as <code>source.abundance</code> .

Details

Only certain transformations are legal.

1. The type "density" must always be associated with a NULL abundance.
2. The type "signature" can be associated with a NULL abundance.
3. The type "counts" must **not** be associated with the NULL abundance.
4. Otherwise, the following are legal:

- (a) counts -> counts
- (b) counts -> density
- (c) counts -> signature
- (d) density -> counts (in which case the semantics are to infer the genome-wide or exome wide counts based on the densities.)
- (e) density -> signature
- (f) signature -> signature

Value

A catalog as defined in [ICAMS](#)

VCFsToDNSCatalogs	<i>Create DNS catalogs from VCFs</i>
-------------------	--------------------------------------

Description

Create a list of 3 catalogs (one each for DNS78, DNS144 and DNS136) out of the contents in list.of.DNS.vcfs. The VCFs must not contain any type of mutation other than DNSs.

Usage

```
VCFsToDNSCatalogs(list.of.DNS.vcfs, genome, trans.ranges)
```

Arguments

- | | |
|------------------|--|
| list.of.DNS.vcfs | List of in-memory data frames of pure DNS mutations – no SNS or 3+BS mutations. The list names will be the sample ids in the output catalog. |
| genome | A particular reference genome(without quotation marks). Use available.genomes to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing genome = "GRCh38" or genome = "hg38". User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing genome = "GRCh37" or genome = "hg19". |
| trans.ranges | A data frame containing transcript ranges. |

Value

A list of 3 DNS catalogs, one each for 78, 144, 136: catDNS78 catDNS144 catDNS136

Note

DNS 144 catalog only contains mutations in transcribed regions.

VCFsToIDCatalogs	<i>Create ID (insertion and deletion) catalog from ID VCFs</i>
------------------	--

Description

Create ID (insertion and deletion) catalog from ID VCFs

Usage

```
VCFsToIDCatalogs(list.of.vcfs, genome)
```

Arguments

list.of.vcfs	List of in-memory VCFs. The list names will be the sample ids in the output catalog.
genome	A particular reference genome(without quotation marks). Use available.genomes to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing genome = "GRCh38" or genome = "hg38". User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing genome = "GRCh37" or genome = "hg19".

Value

An ID (indel) catalog

VCFsToNSNCatalogs	<i>Create SNS catalogs from SNS VCFs</i>
-------------------	--

Description

Create a list of 3 catalogs (one each for 96, 192, 1536) out of the contents in list.of.SNS.vcfs. The SNS VCFs must not contain DNSs, indels, or other types of mutations.

Usage

```
VCFsToNSNCatalogs(list.of.SNS.vcfs, genome, trans.ranges)
```

Arguments

list.of.SNS.vcfs	List of in-memory data frames of pure SNS mutations – no DNS or 3+BS mutations. The list names will be the sample ids in the output catalog.
genome	A particular reference genome(without quotation marks). Use available.genomes to get the list of "BSgenome data packages" currently available. There are 2 types of predefined reference genome which are incorporated in this function. User can invoke a predefined human GRCh38/hg38 BSgenome data package by typing genome = "GRCh38" or genome = "hg38". User can invoke a predefined human GRCh37/hg19 BSgenome data package by typing genome = "GRCh37" or genome = "hg19".
trans.ranges	A data frame containing transcript ranges.

Value

A list of 3 SNS catalogs, one each for 96, 192, 1536: catSNS96 catSNS192 catSNS1536

Note

SNS 192 catalog only contains mutations in transcribed regions.

WriteCatalog	<i>Write Catalog Functions</i>
--------------	--------------------------------

Description

Write a mutation catalog to a file on disk

Usage

```
WriteCatSNS96(ct, path, strict = TRUE)

WriteCatSNS192(ct, path, strict = TRUE)

WriteCatSNS1536(ct, path, strict = TRUE)

WriteCatDNS78(ct, path, strict = TRUE)

WriteCatDNS144(ct, path, strict = TRUE)

WriteCatDNS136(ct, path, strict = TRUE)

WriteCatID(ct, path, strict = TRUE)
```

Arguments

ct	A catalog as defined in ICAMS .
path	The path of the file to be written on disk.
strict	If TRUE, do additional checks on the input, and stop if the checks fail.

Details

WriteCatSNS96 Write a SNS 96 mutation catalog to a file on disk

WriteCatSNS192 Write a SNS 192 mutation catalog to a file on disk

WriteCatSNS1536 Write a SNS 1536 mutation catalog to a file on disk

WriteCatDNS78 Write a DNS 78 mutation catalog to a file on disk

WriteCatDNS144 Write a DNS 144 mutation catalog to a file on disk

WriteCatDNS136 Write a 136 DNS catalog from path

WriteCatID Write a ID (insertion/deletion) catalog to a file on disk. (Please take note that deletion repeat size ranges from 0 to 5+ in the catalog, but for plotting and end user documentation it ranges from 1 to 6+.)

See also [ReadCatalog](#)

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