Manuscript Draft

Manuscript Number:

Title: Comparing age and growth estimates from Bayesian and integrative data approaches for the deepwater snapper Pristipomoides filamentosus in the Hawaiian Islands

Article Type: Research Paper

Keywords: Growth, Pristipomoides filamentous, Hawaii, life history

Corresponding Author: Dr. Stephen Scherrer, Ph.D.

Corresponding Author's Institution:

First Author: Stephen Scherrer, Ph.D.

Order of Authors: Stephen Scherrer, Ph.D.; Don Kobayashi; Kevin Weng;

Francis Oishi; Henry Okamoto; Erik Franklin

Manuscript Region of Origin: USA

Abstract: Pristipomoides filamentosus is an economically and culturally important species of deep-water snapper in the Hawaiian archipelago. From 1989 to 1993, the State of Hawaii initiated a fisher participation markrecapture study to quantify growth and other life history parameters for the species. Over a span of approximately 10 years, 10.5% of 4,179 tagged fish were recaptured. We compared Bayesian and maximum likelihood approaches to estimate von Bertalanffy growth parameters from the tagging data. Direct aging and length frequency data previously used in other published regional growth studies were incorporated to produce integrated estimates of growth. Results from our preferred integrated model reconcile 30+ years of effort from various methods to estimate growth parameters (L ∞ = 67.6 cm FL and K = 0.22) and demonstrate the importance of individual variability in P. filamentosus due primarily to the asymptotic length parameter L ∞ . These results have management implications as growth is often an input for age-based stock assessment models and used as a proxy for other life history traits.

Dear Editors and Reviewers.

We are excited to submit for your consideration a manuscript entitled "Comparing age and growth estimates from Bayesian and integrative data approaches for the deepwater snapper Pristipomoides filamentosus in the Hawaiian Islands". We hope that this manuscript provides a valuable contribution as a *Research Paper*.

Deep-water snappers are an important resource throughout the Indo-Pacific. These species are particularly susceptible to overfishing due to their longevity, delayed maturity, and slow growth. *Pristipomoides filamentosus* is a key component of Hawaii's deep-water bottomfish complex, accounting for over half of the revenue of the region's commercial bottomfish fishery.

Starting in the 1980s, a great deal of effort has been put forth to understand how these fish grow for the purpose of assessment and management. In this manuscript, we report growth parameters estimated using a previously unpublished mark-recapture dataset using Bayesian and maximum likelihood methods. These estimates are fit with nearly four times as many observations as those previously reported. In addition to reporting these growth estimates, we also report estimates obtained using models that incorporate six additional data sources from four prior studies to produce integrative growth estimates. The integrative parameters better predict growth observed in recaptured fishes than those fit using tagging data exclusively. They also better predict observed growth than growth estimates from previous studies.

Growth is a key life history parameter often used directly or indirectly in the management process. By refining estimates for *P. filamentosus* in Hawaii, our integrative estimates can improve assessment models for this species.

Please contact us if you require further information, and we look forward to hearing from you.

Sincerely,

Stephen R. Scherrer

Comparing age and growth estimates from Bayesian and integrative data approaches for the deepwater snapper Pristipomoides filamentosus in the Hawaiian Islands

Scherrer, Stephen R. 1* Kobayashi, Donald R. 2 Weng, Kevin C. 3 Okamoto, Henry Y. 4 Oishi, Francis G. 4 Franklin, Erik C. 5

¹University of Hawaii Oceanography Department 1000 Pope Rd Honolulu, Hawaii 96822 scherrer@hawaii.edu

²National Marine Fisheries Service Pacific Islands Fisheries Science Center 1845 Wasp Blvd Honolulu, Hawaii 96818 donald.kobayashi@noaa.gov

³Virginia Institute of Marine Science College of William and Marry 1375 Greate Road Gloucester Point, Virginia 23062 kevinweng@vims.edu

⁴Division of Aquatic Resources State of Hawaii Department of Land and Natural Resources 1151 Punchbowl Street, Room 330 Honolulu, Hawaii 96813

⁵University of Hawaii School of Ocean and Earth Science and Technology Hawaii Institute of Marine Biology 46-007 Lilipuna Rd Kaneohe, Hawaii 96744 erik.franklin@hawaii.edu

^{*} Corresponding Author

Highlights (for review)

Highlights

- Growth parameters were estimated for *P. filamentosus* using Bayesian and integrative maximum likelihood methods.
- The best parameters were $L_{\infty} = 67.6$ cm FL, k = 0.22, $t_0 = -0.37$ yr.
- These estimates support those derived from using radioisotopic analysis of bomb-carbon.
- Growth parameters estimated with length-at-age, length-frequency, and growth-increment data consistently outperform estimates from any single method.
- Negatively biased growth predictions for the largest individuals suggests the potential for sexually dimorphic growth.

Abstract

1

23

2 Pristipomoides filamentosus is an economically and culturally important species of deep-water snapper in the Hawaiian archipelago. From 1989 to 1993, the State of Hawaii initiated a fisher 3 4 participation mark-recapture study to quantify growth and other life history parameters for the species. Over a span of approximately 10 years, 10.5% of 4,179 tagged fish were recaptured. We 5 6 compared Bayesian and maximum likelihood approaches to estimate von Bertalanffy growth 7 parameters from the tagging data. Direct aging and length frequency data previously used in other published regional growth studies were incorporated to produce integrated estimates of 8 9 growth. Results from our preferred integrated model reconcile 30+ years of effort from various methods to estimate growth parameters ($L_{\infty} = 67.6$ cm FL and K = 0.22) and demonstrate the 10 importance of individual variability in *P. filamentosus* due primarily to the asymptotic length 11 parameter L_{∞} . These results have management implications as growth is often an input for age-12 13 based stock assessment models and used as a proxy for other life history traits. 14 1. Introduction 15 16 Pristipomoides filamentosus (Valenciennes, 1830) is a species of long-lived deep-water snapper 17 distributed throughout the tropical Pacific and Indian Oceans (Allen, 1985; Gaither et al., 2011). 18 Known as opakapaka in Hawaii, the species constitutes a significant fraction of the region's 19 commercial bottomfish fishery (Ralston and Polovina 1982, Langseth et al. 2018). The current 20 stock assessment for this fishery used a surplus production model fit for the 6 species of snapper 21 and one endemic grouper (Langseth et al., 2018). However there is interest in the potential use of species-specific, age-structured assessments for this fishery which require accurate life history 22

studies of age and growth (Langseth et al., 2018; Mauder et al., 2015).

In Hawaii and elsewhere in the Indo-Pacific, growth parameters have been estimated for P. filamentosus using a variety of methods (Table 2). Direct aging approaches using length-atage data from otolith growth increments have been used to derive growth parameters for P. filamentosus (Ralston & Miyamoto, 1983; Uchiyama & Tagami, 1984; Radtke, 1987; DeMartini, Landgraf & Ralston, 1994, Ralston & Williams, 1988). However, this method relies on the integration of daily otolith bands may be biased due to episodic growth and/or poor increment resolution in early (< 5 years) life stages (Andrews et al., 2012; Wakefield et al., 2017). A length frequency study focused on early growth in juvenile fish but individual variability was not considered when extrapolating growth for larger size classes which can result in biased parameter estimates (Moffitt and Parrish, 1996; Sainsbury, 1980). An ongoing mark-recapture study has attempted to estimate growth parameters using a growth increment approach but preliminary results have been limited by the size distribution of recaptured individuals and use model parameterizations incompatible with other methods for determining growth (R. I. C. C. Francis, 1988; O'Malley, 2015). While the methods of the aforementioned studies produced individual estimates of growth parameters, none of them holistically integrated across the three classes of data (direct aging, modal progression, growth increment) to explicitly evaluate the parameter values and sources of uncertainty. Analytical and statistical advances to methods for estimating growth have been developed to account for sources of variability and permit parameter comparisons across lengthat-age, length frequency, and tagging based approaches (Eveson et al., 2004; R. I.C.C. Francis,

1988; Wang et al., 1995). Structural modifications to Fabens (1965) parameterization of the von

Bertalanffy growth model address issues of compatibility between growth parameters derived

from tagging studies and other methods (Maller and Deboer 1988, James 1991, Palmer et al.

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

2

1991, Laslett et al. 2002, Eveson et al. 2004, 2007, Zhang et al. 2009). Maximum likelihood and Bayesian model fitting procedures accommodate individual variability by describing population parameters using probability distributions (R. I.C.C. Francis, 1988; Kimura et al., 1993; Wang et al., 1995; Zhang et al., 2009). Bayesian approaches allow K and L_{∞} to be sampled in this manner and can account for prior information when estimating parameters (Zhang et al. 2009). Maximum likelihood approaches typically treat K as a fixed effect but flexibility in their implementation has allowed for the development of model structures that can estimate a single set of growth parameters from direct aging, length frequency, and growth increment data simultaneously (Wang et al. 1995, Laslett et al. 2002, Eveson et al. 2004).

Here we derive growth parameters using Bayesian and maximum likelihood methods applied to a previously unreported tagging data of *P. filamentosus* maked and recaptured in the Main Hawaiian Islands (MHI). Parameters estimated from the tagging data using a bayesian framework are compared to estimates integrating tagging data with previous length-at-age and length frequency data collected from the MHI and Northwestern Hawaiian Islands (NWHI) derived through maximum likelihood. We then compare these new growth estimates to those previously reported for *P. filamentosus* the Hawaiian Archipelago.

2. Materials and Methods

- 65 2.1 Opakapaka Tagging Program
- Tagging data used for this analysis were obtained by biologists from Hawaii's Division of
- 67 Aquatic Resources (DAR) within the state's Department of Land and Natural Resources
- 68 (DLNR). Between 1989 and 1993 the Opakapaka Tagging Program (OTP), led by staff biologist

Henry Okamoto and operating from fishing vessels contracted out of Honolulu Harbor, tagging and releasing 4,179 *P. filamentosus* in total.

All tagging occurred in the Main Hawaiian Islands (MHI). Coarse location data was provided by the commercial statistical reporting grid areas in a given fish was tagged and recaptured (Table 1, Figure 1). Effort was concentrated primarily around the island of Oahu and the Maui Nui complex consisting of the islands of Maui, Molokai, Lanai and Kahoolawe. Since 1990, these areas have accounted for approximately 67.7% of Hawaii's commercial bottomfish harvest. Less than 1% of fish in this study were tagged off the islands of Niihau or Hawaii (Big Island).

Fish were caught with hook-and-line gear and brought to the surface at a rate of 2-5 feet per second. Prior to tagging, each fish was placed in a holding container with aerated seawater to assess survival likelihood. Fish appearing lively and upright were deemed likely to survive and thus suitable candidates for tagging. If the stomach was inverted and full of gas, it was punctured using a small sharp instrument (e.g., scalpel, hypodermic needle, fishhook). A few scales were carefully removed and a small (~1 cm) incision was made near the fish's anal opening to assist in expelling gas from the body cavity. Fish were then surgically implanted with unique identifiable internal anchor tags with a monofilament streamer protruding from the incision in the peritoneal cavity. The fork length of each fish measured to the nearest ¼ inch, as well as the location and time of capture were recorderd before the fish was returned headfirst to sea with downward momentum to assist in counteracting buoyancy caused by any residual gas.

Reports of recaptured *P. filamentosus* were reported up to a decade after they were tagged, most recently in October of 2003 (Kobayashi et al., 2008; Okamoto, 1993). There were 487 recaptures recorded for 439 unique individuals for a recapture rate of 10.5% of tagged fish.

Local commercial and recreational fishers were made aware of the program through fliers distributed at the local fish markets, to fish dealers, at fishing supply outlets, and posted at small boat harbors. Fishers were incentivized to report the location, depth, fork length, and date that tagged fish were landed with a \$10 reward. Tagged fish recaptured by OTP personnel were fit with an additional tag and released again.

2.2 Tagging Data Management

The data collected by OTP was entered into a spreadsheet with subsequent analysis performed using R (R Core Team, 2014), the Bayesian statistical software JAGS (Plummer, 2003), and the R package R2Jags (Su and Yajima, 2012). Fish were removed from the dataset if they were not the correct species of interest, if no recapture was reported, or if there was no record of the tag identification number. Fork lengths for the remaining fish recorded at tagging and recapture were linearly transformed from inches to centimeters prior to model fitting. Observed growth (Δl) and time at liberty (Δt) were calculated for each fish. If an individual was recaptured on more than one occasion, Δl and Δt were only calculated between the first marking event and the last recapture so as to not violate assumptions of independence. Fish with Δt less than 60 days were excluded from the dataset.

2.3 Parameter Estimation from Tagging Data: Bayesian Approach

Growth parameters were estimated for the *P. filamentosus* tagging data following the Bayesian methodology of Zhang et al. (2009). This approach uses a Fabens version of the von Bertalanffy growth curve but allows the parameters to vary among individuals. Hence the predicted length of a captured individual is expressed as:

(E1)
$$L_{i,j} = L_{\infty,i}(1 - e^{-K_i(A_i + t_{i,i})})$$

This is parameterized such that L_i , is the length of individual i for the jth time the individual is captured, $t_{i,j}$ is the time-at-liberty for individual i for the jth capture, A_i is the relative age of individual i at tagging (age minus t_0), and K_i and $L_{\infty,i}$ are the von Bertalanffy growth parameters for the ith individual. These individual parameters were drawn from Gaussian distributions defining the population means for K and L_{∞} . Uninformative priors were used for all input parameters, using Gaussian, gamma, beta, and uniform distributions following the approach of Zhang et al. (2009). The JAGS code for specifying these parameters and performing this analysis is provided in Appendix 1.

The model which allowed both the K and L_{∞} parameters to vary across individuals as described above is henceforth referred to as Model 1. Three additional models were run with various constraints to K and L_{∞} . Model 2 used a fixed K parameter while allowing the L_{∞} parameter to vary across individuals. The term "fixed" in this context does not imply a user-specified constant value, but instead refers to the value that is estimated by the Bayesian modeling approach from a single distribution used to represent the mean growth process across all individuals. Model 3 used a fixed L_{∞} parameter while allowing the K parameter to vary across individuals and both parameters were fixed under Model 4. Model 4 would A priori be most similar to the Fabens approach, with both fixed K and L_{∞} , but with the added feature of estimating ages at initial tagging, A_i . It is the inclusion of this term that models growth as a function of age, rather than length, which allows for direct comparison between parameters estimated using tagging data and those obtained from direct-age and length frequency approaches (Wang et al., 1995).

Evaluating the restrictive assumptions of models 2-4 was accomplished by comparing growth parameters to those estimated by Model 1. Model 1 is the presumptive best estimate for *P. filamentosus* von Bertalanffy growth curve parameters, since it allows the most flexible incorporation of individual variability in the parameter estimation process. If a given parameter is relatively stable when the parameter was allowed to be variable across individuals and when it was treated as fixed for the population, then it might be inferred that treating this parameter on an individual basis is not warranted. However, if parameter estimates differed when the parameter was fixed, then it might be inferred that treating this parameter on an individual basis is

For each Bayesian hierarchical model run, the first 150,000 samples from the posterior distribution were treated as burn-in and discarded from the Monte Carlo simulation. Every 50th sample from the following 1,400,000 samples (number kept = 28,000) was tabulated into the posterior distributions to reduce potential autocorrelation between sequential values or strings of values. The mean K and L_{∞} values from the posterior distribution were used as metrics of population mean values. Median values deviated from mean values by less than one half of 1 percent (Table 3), indicative of symmetrical distributions easily characterized by any descriptor of value tendency (i.e., mean, median, or mode). Initial starting estimates of K and L_{∞} were obtained by fitting Fabens (1965) approach using non-linear least squares (Table 2). Two additional chains were run simultaneously with initial starting values 50% lower and 100% higher than the Fabens' estimates. This resulted in nearly identical solutions as shown in Table 3. Convergence was also ascertained by examination of the Gelman-Rubin statistic (Gelman and Rubin, 1992).

The fit of each model was assessed by calculating its Bayesian p-value from the posterior predictive distribution and comparing DIC values for each model. Bayesian p-values were simulated using the model's posterior distribution and test whether simulated data is more extreme than the observed data. Bayesian p-values range between 0 and 1 where values approaching 0.5 indicate the model is a good fit to the data, while extreme values near 0 or 1 indicate that the model does not adequately represent the data (Meng, 1994).

2.4 Parameter Estimation from Tagging Data: Maximum Likelihood Approach

Model 5 was fit using the maximum likelihood approach of Laslett, Everson, & Polacheck

167 (2002) using Equation 2.

(E2)
$$l_{ij} = \mu_{\infty} \left(1 - e^{-K(a_i + \Delta t_i)}\right) + \varepsilon_{ij}$$

This method derived growth parameters from the joint distribution of an individual's length at tagging and recapture to estimate growth parameters. This approach is most similar to Model 2 of the Bayesian approach in that asymptotic length, L_{∞} , is treated as a normal random effect $N(\mu_{\infty}, \sigma_{\infty}^2)$ while K is treated as a fixed unknown parameter. The distribution of L_{∞} is assumed to be normal with a mean μ_{∞} and standard deviation σ_{∞}^2 , accounting for differences between an individual the total population. Rather than using length increments to fit observed growth, a bivariate normal joint distribution of lengths recorded at marking and recapture was is to estimate each individual's age at tagging a_i . The distribution of individual a_i s is A and is treated as a random effect with a lognormal distribution $L(\mu_{logA}, \sigma_{logA}^2)$. Measurement error was also treated as a random normal distribution $N(0, \sigma^2)$. An unconditional joint density was then derived for each individual by integrating their individual joint distribution with respect to a. A detailed description of this process is described by Laslett et. al. (2002).

Growth function parameters were estimated through minimizing of the negative loglikelihood cost function obtained by summing the unconditional joint density $h(l_1, l_2)$ of each individual (E3).

$$(E3) - \ln (\lambda_1) = -\sum_{i} \ln h(l_{m,i}, l_{r,i})$$

This approach was used to estimate values of the parameters μ_∞, σ_∞², K, μ_{log A}, σ_{log A}², and σ².
 Two-sided 95% confidence intervals (2.5%, Median, 97.5%) for each parameter were then
 estimated from 10,000 successful bootstrap iterations. During each iteration, the model was refit

using data randomly resampled from the original tagging data with replacement.

2.5 Estimation of Integrative Growth Parameters using sources of growth data

Datasets previously used to estimate regional growth for *P. filamentosus* in the MHI and NWHI and our tagging data exclusively from the MHI were used to produce a single set of parameter estimates using a modified form of the integrated method proposed by Eveson, Laslett, and Polachek (2004). Additional datasets that were included represent both direct aging and length frequency approaches and are briefly described.

2.6 Parameter Estimation: Length Frequency Data

Length frequency data consisted of the size distributions of juvenile *P. filamentosus* sampled over 13 months between October 1989 and February 1991 reported by Moffitt and Parrish (1996). The reported fork length of captured fish was binned by 1 cm increments and presented in 13 histograms corresponding to each month of sampling. The number of fish of a given fork length captured during each month of sampling was determined by overlaying a series of evenly spaced horizontal lines across the Y-axis of each histogram corresponding to the addition of a

single fish. Using this method to reconstruct monthly length frequency data resulted in a total count of 1,048, individuals while in the original study reports 1,047 (Moffitt and Parrish, 1996).

The reconstructed length frequency data were incorporated into integrative models using the two-step method described in Laslett et al 2004. During the first step, a Gaussian mixture model was fit using maximum likelihood and used to decompose the distribution of fork lengths from individuals sampled during discrete time periods for each cohort present in the data. This was accomplished using the normalmixEM function from the mixtools package in R (Benaglia et al., 2009) by assuming the mean of each distribution corresponded to the observed mode. A bimodal Gaussian mixture model was fit for the data collected between the months of October-February, as the original study reported that two cohorts were present during this period. A single cohort was present the remainder of the year. The estimated mean fork length, $\hat{\mu}_{ijk}$, and standard error, s_{ijk} , of each cohort during each sampling period was used to estimate growth parameters (E4).

$$(E4) \quad \hat{\mu}_{ijk} = \mu_{\infty} \left(1 - e^{-K(a_{ijk} - a_0)} \right) + e_{ijk} + \varepsilon_{ijk}$$

With this model, i, j, and k reflect the fishing year, month, and age cohort, respectively. The estimated age of each cohort during a sampling period is denoted by a_{ijk} . Ages were estimated relative to the month of July when peak peak spawning of P. filamentosus occurs (Luers et al., 2017). This produced age estimates between 3 and 19 months. Sampling and residual model errors were described using random normal distributions $e_{ijk} \sim N(0, s_{ijk}^2)$ and $\varepsilon_{ijk} \sim N(0, \sigma_{\varepsilon}^2)$ respectively. In contrast to tagging and direct aging methods, length frequency approaches lack the information to estimate the variance component of asymptotic length (L_{∞}) , so this term was modeled as fixed effect, μ_{∞} . From this, the expected mode fork length of each cohort (E6), and associated variability during each sampling period (E7) were used to minimize the model's

negative log-likelihood cost function (E8). The rationale for these approximations is discussed to greater depth in Eveson et al. 2004.

$$(E6) \quad E(\hat{\mu}_{ijk}) = \mu_{\infty} \left(1 - e^{-K(a_{ijk} - a_0)} \right)$$

$$(E7) \quad V(\hat{\mu}_{ijk}) = s_{ijk}^2 + \sigma_{\varepsilon}^2$$

$$(E8) \quad -\ln(\lambda_2) = \frac{1}{2} \sum_{i} \sum_{j} \sum_{k} \left[\ln\left(2\pi V(\hat{\mu}_{ijk})\right) + \frac{(\hat{\mu}_{ijk} - E(\hat{\mu}_{ijk}))^2}{V(\hat{\mu}_{ijk})} \right]$$

227 2.7 Parameter Estimation: Direct Aging Data

- Sources of direct ageing data included four length-at-age datasets from three prior growth studies. Each dataset estimated age differently. Methods for estimating age included analytical integration of otolith bands (Ralston and Miyamoto, 1983, n=65), counts of otolith micro increments (DeMartini et al., 1994, n=35), comparison of otolith derived bomb radiocarbon ratios (Δ^{14} C) relative to a standard reference obtained from hermatypic coral cores from the Hawaiian Archipelago (Andrews et al., 2012, n=33), and otolith derived lead-radium ratios pooled by size class (Andrews et al., 2012, n=3).
- The details of the method for estimating growth parameters from direct aging data are described in Eveson et al. 2004. Briefly summarized, parameters are modeled using the VBGF model described by equation E9.

(E9)
$$l_i = l_{\infty i} (1 - e^{-K(a_i - a_0)}) + \gamma_i$$

Expected length for each individual and the variance of the measurement error is described by equations E10 and E11.

(E10)
$$E(l_i) = \mu_{\infty} (1 - e^{-K(a_i - a_0)})$$

(E11) $V(l_i) = \sigma_{\infty}^2 (1 - e^{-K(a_i - a_0)})^2 + \sigma_{\gamma}^2$

Where l_i denotes the length of the i^{th} fish, at age a_i and a_0 was a fixed parameter analogous to t_0 when a fish has a hypothetical length of zero. As with the model for tagging data, $l_{\infty i}$ is the individual asymptotic length of the i^{th} fish drawn from the random normal distribution $L_{\infty} = N(\mu_{\infty}, \sigma_{\infty})$. γ_i represents the distribution of individual measurement error and is similarly random, drawn from the distribution $\gamma = N(0, \sigma_{\gamma})$. Equation 12 describes the log-likelihood cost function derived from these equations.

$$(E12) - \ln(\lambda_2) = \frac{1}{2} \sum_{i} \left[\ln(2\pi V(l_i)) + \frac{(l_i - E(l_i))^2}{V(l_i)} \right]$$

2.8 Defining an objective cost function and estimating integrative growth parameters

To derive integrated growth parameters across tag-recapture, direct aging, length frequency, and growth increment data sources, an appropriate overall likelihood cost function (E13) is defined from the sum of the negative log-likelihood cost functions for each source, each with their own scaling constant, β . The single set of growth parameters best describing all data sources is obtained through minimization of the overall likelihood cost function, Λ (E13).

(E13)
$$\Lambda = \beta_1 \ln(\lambda_1) + \beta_2 \ln(\lambda_2) + \beta_3 \ln(\lambda_3) \dots + \beta_n \ln(\lambda_n)$$

By manipulating the value of scaling constants, how similar datasets were treated, and which datasets were included, six additional model structures for estimating growth in P. filamentosus were developed and evaluated (Table 4). Two approaches were used to define the scaling constants (β) within each model structures overall cost function. The first weighted β scaling constants for every data source so that each source had equal influence on the resulting parameter estimates. This was achieved by setting the β constant for each data source equal to the inverse of the number of observations for the data. The second approach weighed β for each

data source relative to the number of observations of within the data set ($\beta_1 = \beta_2 = \beta_3$). This meant that data sources with more observations had a greater influence during parameter estimation.

The structure of model 5 fit only tagging data from the OTP study. The remaining models 6-11 incorporated the additional length-at-age and length frequency data sources and differed in their treatment of β coefficients. Other differences between these models included whether the four direct aging data sources were considered independently and assigned their own log-likelihood cost function or if the four data sources were pooled with a single log-likelihood cost function. Omission of direct aging data where ages were estimated by integrating daily growth increments was also considered as this method is likely to underestimate age (Table 4; Wakefield et al 2017).

The six candidate integrative model structures (Models 6-11) were evaluated against one another using a repeated train-test cross validation procedure (Burman, 1989) to determine the combination of model weighting, data pooling, and data sources parameter estimates that consistently best predicted the growth observed in the tagging dataset. Each model structure was trained using two-thirds of the tagging data (n = 258) selected at random while the remaining one-third (n = 129) was reserved for testing model performance. Model performance was evaluated by calculating the mean sum of squared residuals (mssr) using the test dataset. Length at recapture was compared between predictions for the test data made using parameters fit with the training data using Equation 2 and observed length reported in the test set (E14).

$$(E14) \quad s^2 = \frac{1}{n} \sum_{i} \left(L_{ri} - \widehat{L_{ri}} \right)^2$$

The preferred model structure was the one whose estimated parameters most frequently resulted in the lowest mssr over 10,000 iterations. To determine if incorporating additional data sources

improved predictive performance, mssr for the preferred model structure were compared to those calculated using a model structure identical to Model 5, calculated including only tagging data.

The integrative model structure that best predicted observed growth most frequently was refit using the entire data set. Two-sided 95% confidence intervals were estimated for each parameter from the results of 10,000 bootstrap iterations. As with tagging data, the procedure for resampling direct aging data was straightforward and involved random sampling with replacement from the dataset to construct pseudo-datasets with an equal number of observations as the original data. Bootstrapping length frequency data was slightly more complicated with each study period in the pseudo-dataset resampled from the corresponding period of the reconstructed study data. Each study period in the pseudo-dataset contained the same number of observations as in the corresponding time period of the original study data.

3. Results

3.1 Opakapaka Tagging Program

Of the 4,179 *P. filamentosus* tagged 439 individuals were recaptured at least once (10.5%, Table 1). Mortality of fish upon release appeared to be generally low, facilitated by the strong tagging selectivity for healthy fish in good condition. Some immediate mortality was observed due to sharks and cetaceans or capture stress (4 individuals). Long-term mortality was thought to be relatively low based upon the high rates of tag return spanning many years. Hydra (small cnidarian polyps) biofouling of the tags was observed for some individuals with large times at liberty, with some lesions apparent around the opening where the tag exited the body cavity. This was not thought to be a serious health issue since the fish appeared to be feeding and swimming normally.

Initial fork length at capture across all individuals ranged in size from 16.5 to 53.3 cm (mean = 31.9 cm, standard deviation (s.d.) = 5.5) and ranged from 19.1 cm and 52.8 cm (mean = 32.8, s.d. = 5.1) for fish that were later recaptured. For those fish that were later recaptured, fork length at recapture ranged between 22.9 cm and 76.2 cm (mean = 41.9, s.d. = 8.7). The minimum time at liberty for any fish between tagging and recapture was a single day while the maximum time at liberty was 10.3 years (3.748 days) (Figure 2). The mean time at liberty was 1.82 years or 666 days (s.d. = 625).

One fish was excluded from further analysis as its fork length at capture was not recorded. Seven fish were removed because the recapture date was not properly recorded. Of the remaining 432 fish recaptured, 351 were recaptured a single time, 33 fish were recaptured a total of two times, one fish recaptured 3 times, and two fish were recaptured 4 times. We also excluded from analysis 45 individuals for whom time at liberty was less than 60 days yielding a data set of 387 unique individuals.

3.2 Estimating Growth Parameters from Tagging Data: Bayesian Approach

The Bayesian hierarchical approach using the JAGS software yielded mean estimates of L_{∞} and K for each of the Models 1–4 examined (Table 2). Model 1, which incorporated individual variability in both L_{∞} and K yielded mean parameter estimates of L_{∞} = 59.9 cm (coefficient of variation [c.v.] = 2.59) and K = 0.32 (c.v. = 8.57). L_{∞} and K parameter estimates for Model 2, where K was fixed, were 60.1 cm (c.v. = 2.74) and 0.35 (c.v. = 45.7) respectively. Under Model 3, where L_{∞} was fixed and K was fit freely L_{∞} = 76.9 cm (c.v. = 42.2) and K = 0.17 (c.v. = 8.62) and L_{∞} = 77.3 cm (c.v. = 43.1) and K = 0.24 (c.v. = 73.1) for Model 4, where both parameters were fixed. Additional parameters for each of the four models are presented in Table 3. The

Gelman-Rubin convergence criteria indicated that the model solutions were credible, with asymptotic convergence clearly occurring after ~4000 iterations, well within the burn-in phase of the Bayesian modeling runs. All 4 models appeared to fit the the data well; the mean Bayesian P-values from all retained posterior samples for all models ranged between 0.500 and 0.501. Model 1 had the largest DIC score (10582.86) followed by model 2 (10490.96), model 3 (5033.42), and model 4 (4874.83), however treating model parameters as fixed under models 2-4 resulted in excessively large coefficients of variation suggesting that individual variability in L_{∞} and K is important, with perhaps variability in L_{∞} being more important based upon the response of L_{∞} standard deviation from the base case of Model 1 to the constrained individual variability in Model 3 and Model 4 (Figure 3).

3.3 Parameter estimation using maximum likelihood

The maximum likelihood approach used for Model 5 successfully converged to produce estimates of μ_{∞} , σ_{∞}^2 , K, $\mu_{\log A}$, $\sigma_{\log A}^2$, and σ^2 (Table 5). Bootstrap confidence intervals of parameters μ_{∞} and K overlapped L_{∞} and K parameters from Bayesian models 1 and 2 (Table 2). From these results, it was concluded that estimates produced by maximum likelihood were satisfactorily similar to estimates from the Bayesian approach. Model residuals were distributed around zero fairly consistently for all but the largest fish. For fish with recapture lengths exceeding 60 cm, growth models underestimated observed recapture lengths (Figures 3).

3.4 Comparing model performance

Across all 10,000 cross validation iterations to determine model structure, mssr ranged between 7.29 and 24.96 (mean = 14.20, s.d. = 2.20) where a lower predictive variance indicates a better

model fit. From all candidate likelihood models, the structure of Model 11 best predicted growth observed in the test data for 3,486 of 10,000 cross validation iterations. The mssr for Model 11 ranged between 7.29 and 20.10 (mean = 13.64, s.d. = 1.91). The structure of Model 5, fit exclusively using tagging data, ranged in predictive variance between 7.17 and 26.09 (mean = 14.35, s.d. = 2.44). The structure of Model 11 performed better than the structure of Model 5 in 6,351 of 10,000 cross validation iterations. Differences in mssr between these two competing structures ranged between -1.60 and 10.80 (mean = 0.72, s.d. = 1.37) and indicated that the inclusion of additional growth data improved the predictive capability of growth models compared to tagging data alone. Bootstrapped parameter estimates refit to the structure of the prefered model (Model 11) and Model 5's tagging only data are summarized in Table 2 and all parameters for models 5-11 are reported in full in Table 5. When fit to the entire tagging data set, the residual pattern of Model 11 underestimated lengths at recapture length for the largest individuals.

4. Discussion

Our integrative model results reconcile 30+ years of efforts to determine growth for P. filamentosus in the Hawaiian Archipelago and provides robust support for some observed life history parameters. Growth parameters derived using integrative models that incorporated additional length frequency and length-at-age data were better able to predict observed growth in recaptured fish. These parameters were in agreement with those derived from; 1) the fit of only integrated daily growth increments from otoliths collected in the NWHI without constraining L_{∞} (Ralston and Miyamoto, 1983), 2) integrated daily growth increments and microincrement counts (DeMartini et al. 1994), and 3) the radioisotopic composition of otolith material and

counts of otolith increments from the MHI and NWHI (Andrews et al. 2012) and support the implicit assumption that tagging individuals did not disrupt their growth trajectory. Integrative parameters differed from estimates from an ongoing mark recapture study in the MHI which reported faster growth and smaller asymptotic lengths (O'Malley, 2015). These differences could arise from real changes in growth rate between the periods fish were collected, methodological differences in model interpretation, and/or that thus far, none of the fish recaptured during the ongoing study have been of the largest size classes (maximum size reported = 47.6 cm FL).

Compared to their broader distribution, *P. filamentosus* from the Hawaiian archipelago were generally slower growing but obtained a larger asymptotic length than those from the Mariana Archipelago (Ralston and Williams, 1988) and Papua New Guinea (Fry et al. 2006, Andrews et al. 2012) but were faster growing and smaller in their asymptotic length when compared to estimates from the Seychelles (Hardman-Mountford et al., 1997; Mees, 1993; Mees and Rousseau, 1997; Pilling, 2000).

Of the Bayesian models, Model 1 was presumed optimal because it incorporated individual variability in both L_{∞} and K parameters. Models 2-4 suggest that individual variability in both K and L_{∞} is important, with perhaps variability in L_{∞} being more important based upon the response of L_{∞} standard deviation from the base case of Model 1 to the constrained individual variability in Model 3 and Model 4 (Figure 3). While Models 3 and 4 had lower DIC values, based upon parameter estimates and patterns of standard deviation, it is likely that these models were not credible. Similar parameter estimates obtained from Models 1 and 2 suggested that the primary source of individual variability was due to variability in the L_{∞} parameter. This is consistent with other studies where the best models accounted for individual variability in both terms but accounting for individual variation in the L_{∞} term alone was

sufficient to describe growth while significantly reducing computational complexity (Eveson et al. 2007, Zhang et al. 2009).

Comparing growth parameter estimates fit exclusively with tagging data indicate that Bayesian and maximum likelihood fitting methods performed similarly. The treatement of individual variability in parameters for Model 2 were identical to those used to fit Model 5 (OTP data only). Parameter estimates for Models 1 and 2 were contained within the 95% confidence intervals of Model 5. These results suggest that treatment of *K* as a fixed effect was unlikely to bias estimates in integrative models which were evaluated under the same assumptions as models 2 and 5.

Across all candidate growth models, the parameters estimated by Model 11 best predicted length at recapture across cross validation iterations and therefore represents the best estimated parameter set. Information from older/larger fish was very important for grounding the upper end of integrative growth curves resulting in parameters that better predicted length at recapture. Omission of the largest individuals from Models 1-5 resulted in lower estimates of L_{∞} , causing growth curves to asymptote prematurely. When Included, these additional data sources resulted in growth parameters that were better able to predict the length of fish recaptured from the MHI in the OTP study.

Additional data sources included here represent collections spanning several decades from the MHI and NWHI. When incorporating these additional data sources, it is an assumption that growth within the population did not differ significantly with time or region. With the exception of Ralston and Miyamoto (1983), all subsequent studies of growth for *P. filamentosus* in the Hawaiian archipelago have included data or parameter estimates from one or more previous studies in their calculations regardless of time and place of collection (DeMartini et al.

1994, Moffitt and Parrish 1996, Andrews et al. 2012). Genetic homogeneity between NWHI and MHI stocks provides some justification pooling data across both regions (Gaither et al. 2010, Gaither et al. 2011). However, these spatial and temporal assumptions may not reflect phenotypic realities and further work is required to resolve whether differences in growth exist between the two regions.

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

Sexual size dimorphism may explain the tendency of parameters obtained here and elsewhere to underestimate the length at recapture observed for the largest fish in the OTP dataset (approximate fork length > 50 cm) (Figure 4). If one sex attains a greater asymptotic length than the other, that sex is likely to be overrepresented in the largest size classes relative to the total population. In smaller sizes where the sex ratio of individuals is similar to the sex ratio of the total sampled population, averaging of model parameters between sexes results in increased estimates of variance. However, for the largest size classes where sex ratios are not representative of the population as a whole, estimated growth parameters continue to represent an average of both sexes and will result in underestimates of length at recapture for the larger, overrepresented sex while overestimating the length at recapture for the largest individuals of the other. While not pronounced, dimorphic size differences have been observed in a number of lutjanid species (Grimes, 1987; Mees, 1993; Newman et al., 2000; Newman and Dunk, 2002; Nichols, 2019; Taylor et al., 2018; Williams et al., 2017). Elsewhere in their distribution, larger asymptotic lengths have been reported for male *P. filamentosus* in the Seychelles while during research fishing in the Northwestern Hawaiian Islands, the number of females outnumbered males almost 2:1 in the largest size classes, and in Guam no differences between sexes were observed (Kami, 1973; Kikkawa, 1984; Mees, 1993). However, estimations of growth

parameters for *P. filamentosus* in the Central Pacific are sex agnostic and a method for non-invasive sexing of this species was unknown until recently (Luers et al. 2017).

Accurate estimates of von Bertalanffy growth parameters are very important for management. Growth parameters are often used directly or indirectly in stock assessment and fisheries management (Haight et al., 1993; Polovina et al., 1987). These efforts are sensitive to both growth parameters and the model used to estimate those parameters. For example, the rate of instantaneous natural mortality M is a value of interest often inferred from K using empirical relationships (Jensen, 1996; Ralston, 1987; Thorson et al., 2017). Underestimating K will underestimate M, characterizing a stock as less productive than it actually is while overestimating K will have the opposite effect. If a management regime is linked to such a flawed estimate of stock productivity, then the stock is likely to be mismanaged and under or over harvested, respectively, relative to its true biological potential. Future work to refine these estimates for P. filamentosus should consider that differences in life history may exist between males and females.

Acknowledgements

The authors would like to thank everyone involved with OTP. We are also thankful to the authors of the other studies that collected the data presented here including Frank Parrish, Robert Moffitt, Stephen Ralston, Garret Miyamoto, Allen Andrews, Edward DeMartini, Jon Brodziak, Ryan Nichols, and Robert Humphreys. Thanks to Annette Tagawa for providing the OTP mark recapture data used in this analysis. We would also like to thank Zane Zhang and Paige Eveson for providing code used to fit Bayesian and maximum likelihood models. Finally, we would like to thank Joe O'Malley and Frank Parrish for their advice and feedback during this project and Allen Andrews for reviewing our manuscript. Funding was provided by NOAA award

465 #NA10NMF4520163 (to ECF). This is SOEST contribution ###### and HIMB contribution 466 ####. 467 468 **Competing Interests** The authors declare that they have no competing interests. 469 470 References 471 Allen, G.R., 1985. Fao Species Catalogue Vol. 6. Snappers of the World. Fao Fish. Synopsis 6, 472 473 208. https://doi.org/10.1016/0025-326X(92)90600-B Andrews, A.H., DeMartini, E.E., Brodziak, J., Nichols, R.S., Humphreys, R.L., 2012. A long-474 475 lived life history for a tropical, deepwater snapper (Pristipomoides filamentosus): bomb radiocarbon and lead-radium dating as extensions of daily increment analyses in otoliths. 476 477 Can. J. Fish. Aquat. Sci. 69, 1850–1869. https://doi.org/10.1139/f2012-109 478 Andrews, A.H., Humphreys, R.L., DeMartini, E.E., Nichols, R.S., Brodziak, J., 2011. Bomb Radiocarbon and Lead-Radium Dating of Opakapaka (Pristipomoides filamentosus), 479 480 NOAA Technical Memorandum NMFS-PIFSC. Honolulu, HI. 481 Benaglia, T., Chauveau, D., Hunter, D.R., Young, D., 2009. Mixtools: An R package for 482 analyzing finite mixture models. J. Stat. Softw. 32, 1–29. Burman, P., 1989. A comparative study of ordinary cross-validation, v-fold cross-validation and 483 the repeated learning-testing methods. Biometrika 76, 503–514. 484 https://doi.org/10.1093/biomet/76.3.503 485 DeMartini, E.E., Landgraf, K.C., Ralston, S., 1994. A recharacterization of the age-length and 486 growth relationships of Hawaiian snapper Pristipomoides filamentosus. U.S. Department of 487 488 Commerce, National Oceanic and Atmospheric Administration, National Marine Fisheries

- Service, Southwest Fisheries Science Center, Honolulu, HI.
- Eveson, J.P., Laslett, G.M., Polacheck, T., 2004. An integrated model for growth incorporating
- 491 tag-recapture, length-frequency, and direct aging data. Can. J. Fish. Aquat. Sci. 61, 292-
- 492 306. https://doi.org/10.1139/f03-163
- Eveson, J.P., Polacheck, T., Laslett, G.M., 2007. Consequences of assuming an incorrect error
- structure in von Bertalanffy growth models: a simulation study. Can. J. Fish. Aquat. Sci. 64,
- 495 602–617. https://doi.org/10.1139/f07-036
- 496 Francis, R. I. C. C., 1988. Are Growth Parameters Estimated from Tagging and Age–Length
- 497 Data Comparable? Can. J. Fish. Aquat. Sci. 45, 936–942. https://doi.org/10.1139/f88-115
- 498 Francis, R. I.C.C., 1988. Maximum likelihood estimation of growth and growth variability from
- tagging data. New Zeal. J. Mar. Freshw. Res. 22, 43–51.
- 500 https://doi.org/10.1080/00288330.1988.9516276
- 501 Fry, G.C., Brewer, D.T., Venables, W.N., 2006. Vulnerability of deepwater demersal fishes to
- commercial fishing: Evidence from a study around a tropical volcanic seamount in Papua
- 503 New Guinea. Fish. Res. 81, 126–141. https://doi.org/10.1016/j.fishres.2006.08.002
- Gaither, M.R., Jones, S. a., Kelley, C., Newman, S.J., Sorenson, L., Bowen, B.W., 2011. High
- connectivity in the deepwater snapper Pristipomoides filamentosus (lutjanidae) across the
- indo-pacific with isolation of the Hawaiian archipelago. PLoS One 6, 1–13.
- 507 https://doi.org/10.1371/journal.pone.0028913
- 508 Gelman, A., Rubin, D.B., 1992. Inference from iterative simulation using multiple sequences.
- 509 Stat. Sci. 7, 457–472. https://doi.org/10.2307/2246093
- Grimes, C.B., 1987. Reproductive biology of the Lutjanidae: A review, in: Polovina, J.J.,
- Ralston, S. (Eds.), Tropical Snappers and Groupers: Biology and Fisheries Management.

512 Westview Press, Boulder, Colorado, pp. 239–294. 513 Haight, W.R., Kobayashi, D.R., Kawamoto, K.E., 1993. Biology and management of deepwater snappers of the Hawaiian archipelago. Mar. Fish. Rev. 55, 20–27. 514 515 Hardman-Mountford, N.J., Polunin, N.V.C., Boulle, D., 1997. Can the age of the tropical species be determined by otolith measurement?: a study using Pristipomoides filamentosus (Pisces: 516 517 Lutjanidae) from the Mahe Plateau, Seychelles. Naga, ICLARM Q. 20, 27–31. Hawaii Reported Landing Tables [WWW Document], 2016. URL 518 https://www.pifsc.noaa.gov/wpacfin/hi/dar/Pages/hi data 3.php (accessed 3.23.18). 519 520 James, I.R., 1991. Estimation of von Bertalanffy growth curve parameters from recapture data. 521 Biometrics 47, 1519–1530. https://doi.org/10.2307/2532403 Jensen, A.L., 1996. Beverton and Holt life history invariants result from optimal trade-off of 522 523 reproduction and survival. Can. J. Fish. Aquat. Sci. 53, 820–822. https://doi.org/10.1139/f95-233 524 Kami, H.T., 1973. The Pristipomoides (Pices: lutjanidae) of Guam with notes on their biology. 525 526 Micronesica 9, 97–117. https://doi.org/10.1080/15235882.2014.934485 Kikkawa, B.S., 1984. Maturation, spawning, and fecundity of Opakapaka, Pristipomoides 527 528 filamentosus, in the Northwest Hawaiian Islands. Proc. Second Symp. Resour. Investig. Northwest. Hawaiian Islands. 529 Kimura, D.K., Shimada, A.M., Lowe, S.A., 1993. Estimating von Bertalanffy growth parameters 530 of sablefish Anoplopoma fimbria and Pacific cod Gadus macrocephalus using tag-recapture 531 data. Fish. Bull. 91, 271–280. 532 Kobayashi, D.R., Okamoto, H.Y., Oishi, F.G., 2008. Movement of the deepwater snapper 533

opakapaka, Pristipomoides filamentosus, in Hawaii: Insights from a large-scale tagging

534

535 program and computer simulation. 536 Langseth, B., Syslo, J., Yau, A., Kapur, M., Brodziak, J., 2018. Stock assessment for the main Hawaiian Islands Deep 7 bottomfish complex in 2018, with catch projections through 2022. 537 538 NOAA Tech. Memo. NMFS-PIFSC 69, 217. Laslett, G.M., Eveson, J.P., Polacheck, T., 2002. A flexible maximum likelihood approach for 539 fitting growth curves to tag-recapture data. Can. J. Fish. Aquat. Sci. 59, 976–986. 540 https://doi.org/10.1139/f02-069 541 Luers, M.A., DeMartini, E.E., Humphreys, R.L.J., 2017. Seasonality, sex ratio, spawning 542 543 frequency and sexual maturity of the opakapaka Pristipomoides filamentosus (Perciformes: 544 Lutjanidae) from the Main Hawaiian Islands: fundamental input to size-at-retention regulations. Mar. Freshw. Res. 69, 325–335. https://doi.org/10.1071/MF17195 545 546 Maller, R.A., Deboer, E.S., 1988. An analysis of two methods of fitting the von Bertalanffy curve to capture-recapture data. Mar. Freshw. Res. 39, 459–466. 547 https://doi.org/10.1071/MF9880459 548 549 Mauder, M.N., Crone, P.R., Valero, J.L., Semmens, B.X. (Eds.), 2015. Growth: theory, estimation, and application in fishery stock assessment, in: CAPAM Workshop Series 550 551 Report 2. Center for the Advancement of Population Assessment Methodology, La Jolla, CA, p. 55. 552 Mees, C.C., 1993. Population biology and stock assessment of Pristipomoides filamentosus on 553 the Mahe Plateau, Seychelles. J. Fish Biol. 43, 695–708. https://doi.org/10.1111/j.1095-554 8649.1993.tb01147.x 555 556 Mees, C.C., Rousseau, J.A., 1997. The potential yield of the lutjanid fish Pristipomoides 557 filamentosus from the Mahe Plateau, Seychelles: Managing with uncertainty. Fish. Res. 33,

558 73–87. https://doi.org/10.1016/S0165-7836(97)00069-6 559 Meng, X.-L., 1994. Posterior predictive p-values. Ann. Stat. 22, 1142–1160. 560 Moffitt, R.B., Parrish, F.A., 1996. Habitat and life history of juvenile Hawaiian pink snapper, 561 Pristipomoides filamentosus. Pacific Sci. 50, 371–381. Newman, S.J., Cappo, M., Williams, D.M.B., 2000. Age, growth and mortality of the stripey, 562 Lutjanus carponotatus (Richardson) and the brown-stripe snapper, L. vitta (Quoy and 563 Gaimard) from the central Great Barrier Reef, Australia. Fish. Res. 48, 263–275. 564 https://doi.org/10.1016/S0165-7836(00)00184-3 565 566 Newman, S.J., Dunk, I.J., 2002. Growth, age validation, mortality, and other population characteristics of the red emperor snapper, Lutjanus sebae (Cuvier, 1828), off the 567 Kimberley coast of north-western Australia. Estuar. Coast. Shelf Sci. 55, 67–80. 568 569 https://doi.org/10.1006/ecss.2001.0887 570 Nichols, R.S., 2019. Sex-specific growth and longevity of "Ehu", Etelis carbunculus (Family lutianidae) within the Hawaiian archipelago. University of Hawaii. 571 572 O'Malley, J., 2015. A Review of the Cooperative Hawaiian Bottomfish Tagging Program of the Pacific Islands Fisheries Science Center and the Pacific Islands Fisheries Group. Honolulu, 573 574 HI. https://doi.org/10.7289/V59W0CF7 Okamoto, H.Y., 1993. Develop opakapaka (pink snapper) tagging techniques to assess 575 movement behavior. Honolulu, HI. 576 577 Palmer, M.J., Phillips, B.F., Smith, G.T., 1991. Application of nonlinear models with random coefficients to growth data. Biometrics 47, 623–635. https://doi.org/10.2307/2532151 578 Parke, M., 2007. Linking Hawaii Fisherman Reported Commercial Bottomfish Catch Data to 579 580 Potential Bottomfish Habitat and Proposed Restricted Fishing Areas using GIS and Spatial

581 Analysis, NOAA Technical Memorandum NMFS-PIFSC-11. Honolulu, HI. 582 Pilling, G.M., 2000. Validation of annual growth increments in the otoliths of the lethrinid 583 Lethrinus mahsena and the lutjanid Aprion virescens from sites in the tropical Indian Ocean, 584 with notes on the nature of growth increments in Pristipomoides filamentosus. Fish. Bull. 98, 600–611. 585 Plummer, M., 2003. JAGS: A program for analysis of Bayesian graphical models using Gibbs 586 587 sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, in: Proceedings of the 3rd International Workshop on Distributed Statistical 588 589 Computing. Vol. 124. Polovina, J.J., Ralston, S., Ralston, S., 1987. Assessment and management of deepwater bottom 590 fishes in Hawaii and the Marianas. Trop. snappers groupers Biol. Fish. Manag. 505–532. 591 R Core Team, 2014. R: A Language and Environment for Statistical Computing. 592 593 Radtke, R.L., 1987. Age and growth information available from the otoliths of the Hawaiian snapper, Pristipomoides filamentosus. Coral Reefs 6, 19–25. 594 595 https://doi.org/10.1007/BF00302208 596 Ralston, S., Miyamoto, G.T., 1983. Analyzing the width of daily otolith increments to age the Hawaiian snapper, Pristipomoides filamentosus. Fish. Bull. 81, 523–535. 597 Ralston, S.V.D., 1987. Mortality rates of snappers and groupers. Trop. snappers groupers Biol. 598 Fish. Manag. 375–404. 599 600 Ralston, S.V.D., Polovina, J., 1982. A multispecies analysi of the commercial deep-sea handline fishery in Hawaii. Fish. Bull. 80, 435–448. 601 Ralston, S. V., Williams, H.A., 1988. Depth distributions, growth, and mortality of deep slope 602

fishes from the Mariana archipelago. Southwest Fish. Sci. Cent., Natl. Mar. Fish. Serv.,

603

- NOAA, Honolulu, HI 96822-2396. Southwest Fish. Sci. Cent. Admin. Technical
- Memorandum. 113
- Sainsbury, K.J., 1980. Effect of individual variability on the von Bertalanffy growth equation.
- 607 Can. J. Fish. Aquat. Sci. 37, 241–247. https://doi.org/10.1139/f80-031
- Su, Y., Yajima, M., 2012. R2jags: A Package for Running JAGS from R.
- Taylor, B.M., Oyafuso, Z.S., Pardee, C.B., Ochavillo, D., Newman, S.J., 2018. Comparative
- demography of commercially-harvested snappers and an emperor from American Samoa.
- 611 PeerJ 6, e5069. https://doi.org/10.7717/peerj.5069
- Thorson, J.T., Munch, S.B., Cope, J.M., Gao, J., 2017. Predicting life history parameters for all
- fishes worldwide. Ecol. Appl. 27, 2262–2276. https://doi.org/10.1002/eap.1606
- 614 Uchiyama, J.H., Tagami, D.T., 1984. Life history, distribution, and abundance of bottomfishes in
- the Northwestern Hawaiian Islands, in: Grigg, R.W., Tanoue, K.Y. (Eds.), Proceedings of
- the Second Symposium on Resource Investigations in the Northwestern Hawaiian Islands.
- 617 pp. 229–247.
- Wakefield, C.B., O'Malley, J.M., Williams, A.J., Taylor, B.M., Nichols, R.S., Halafihi, T.,
- Humphreys, R.L., Kaltavara, J., Nicol, S.J., Newman, S.J., 2017. Ageing bias and precision
- for deep-water snappers: Evaluating nascent otolith preparation methods using novel
- multivariate comparisons among readers and growth parameter estimates. ICES J. Mar. Sci.
- 622 74, 193–203. https://doi.org/10.1093/icesjms/fsw162
- Wang, Y.-G., Thomas, M.R., Somers, I.F., 1995. A maximum likelihood approach for estimating
- 624 growth from tag-recapture data. Can. J. Fish. Aquat. Sci. 52, 252–259.
- 625 https://doi.org/10.1139/f95-025
- Williams, A.J., Wakefield, C.B., Newman, S.J., Vourey, E., Abascal, F.J., Halafihi, T.,

627	Kaltavara, J., Nicol, S.J., 2017. Oceanic, Latitudinal, and Sex-Specific Variation in
628	Demography of a Tropical Deepwater Snapper across the Indo-Pacific Region. Front. Mar.
629	Sci. 4. https://doi.org/10.3389/fmars.2017.00382
630	Zhang, Z., Lessard, J., Campbell, A., 2009. Use of Bayesian hierarchical models to estimate
631	northern abalone, Haliotis kamtschatkana, growth parameters from tag-recapture data. Fish
632	Res. 95, 289–295. https://doi.org/10.1016/j.fishres.2008.09.035
633	
634	

Tables 635 **Table 1.** Summary of tagging and recapture locations for *P. filamentosus* used for growth 636 increment approaches as referenced to reporting grids in Figure 1. Adapted from Kobayashi, 637 638 Okamoto & Oishi (2008). 639 Table 2. Estimates (and 95% confidence intervals when available) of von Bertalanffy growth 640 641 parameters L_{∞} , K, and t_o reported by the present and prior studies. 642 **Table 3.** Von Bertalanffy growth parameter estimates from Bayesian hierarchical growth models 643 (Models 1-4). 644 645 **Table 4.** A reference for the candidate model structures used to determine the preferred model 646 structure from integrative maximum likelihood growth models (Models 5-11). 647 648 **Table 5.** Sample and Population Parameter Estimates (with bootstrapped confidence intervals) 649 from Maximum Likelihood Growth Models (Models 5-11). 650 651

Figure Captions 652 Figure 1. Reporting Grid Map. 653 Map showing the location and number of the State of Hawaii's statistical reporting grids 654 corresponding to the reported location of tagging and recaptured for fish summarized in 655 *Table 5.2.* 656 657 Figure 2. Length and Time at Liberty for OTP Data. 658 The length of P. filamentosus recaptured and included in analysis of OTP tagging data and 659 the distribution of times at liberty. The fork length of fish during tagging is highlighted in red 660 while length at recapture is shown in blue. 661 662 663 Figure 3. Coefficient of Variation for von Bertalanffy Growth Function Parameters. Coefficient of variation for 2 von Bertalanffy growth function parameters (Brody growth 664 coefficient, K) and (Mean asymptotic length $L\infty$) for P. filamentosus. Individual variability 665 666 was examined incorporating individual variability in both parameters, in either one of the parameters in series, or in neither parameter. 667 668 Figure 4. Plots Comparing Predicted and Observed Length at Recapture. 669 Predicted lengths at recapture fit using parameter point estimates from Bayesian Models 1 670 671 and 2 and population parameter estimates from Maximum likelihood Models 5 and 11 672 compared to observed length at recapture. Length at recapture was predicted as a function of length at marking and time at liberty. The 1:1 line indicates where points would fall if model 673 674 parameters perfectly predicted length at recapture.

RELEASE LOCATION

		127	304	306	307	308	309	311	312	313	320	321	327	331	332	351	401	402	403	404	405	407	408	409	421	423	424	428	429	452	505	528	Total
	127							1																									1
	304				1						2																						3
	306			1							1																						2
	307				5								1																				6
	308			1		2						5																					8
	309											2																					2
	311							25	1					4																			30
	312							1	1																								2
	313				1				1					3																			5
	320			1	3						24		1																				29
	321											31																					31
	327				3						2		2																				7
	331							46	2					128			4																180
	332													1																			1
2	351													1																			1
LOCATION	401																131																131
	402																1																1
Adding	403																1																1
90	404																						1										1
-	405							1															1										2
	407													1									12										13
	408													1																			1
	409																4								1								5
	421																14						1										15
	423											1					14						1										16
	424					1								4			3						3										11
	428							2	1					2																			5
	429							2				1		1			1																5
	452											1		1																		$oxed{oxed}$	2
	505										1																						1
	528							2																									2
L	No Recovery	1	3	44	278	35	2	582	168	5	333	429	84	875	1	1	937	7	1	1	2	2	293	4	20	16	9	8	5	2	1	2	4151

TAGGING LOCATION

				ŝ	win Bair	and Cal	iths Lev	etelli kiel			
	Method	Region	Oki	dith Gro	with Batt. Britis Por	oled Or	iths Lev	the Linf (95% CI)	K (95% CI)	to (95% CI)	Source
	Daily Increments	NWHI	17	-	-	-	-	-	-	-	Moffitt (1980)
	Daily Increments	NWHI	N.R.	-	-	-	-	80.5	0.16	-	Ralston (1980)
	Daily Growth Integration	NWHI	64	-	-	-	-	78	0.146	-1.67	Ralston & Miyamoto (1983)*
ging	Daily Growth Integration	NWHI	64	-	-	-	-	66.4	0.235	-0.81	Ralston & Miyamoto (1983)
Direct Aging	Daily Increments & Integration	NWHI	N.R.	-	-	-	-	69.8	0.534	0.18	Radtke (1987)
Dire	Daily Increments & Integration	MHI & NWHI	92	-	-	-	-	70.4 (63.9 - 76.9)	0.25 (0.20, 0.31)	-0.22 (-0.39, -0.06)	DeMartini et al. (1994)
	Annual Increments	NWHI	N.R.	-	-	-	-	97.1	0.31	0.02	Uchiyama & Tagami (1984)
	Daily Increments, Integration, & Radioisotopes	MHI & NWHI	100	33	3	-	-	67.5 (65.7, 69.3)	0.242 (0.185, 0.299)	-0.29 (-0.38, -0.20)	Andrews et. al (2012)
	Modal Progression	MHI	-		-	13		78	0.21	0	Moffit & Parrish (1996)*
	Mark Recapture	MHI	-		-	-	96	71.55	0.15	-	O'Malley (2015) - Gulland and Holt
	Mark Recapture	МНІ	-		-	-	96	57.80 (55.97, 58.67)	0.28 (0.25, 0.31)	-	O'Malley (2015) - Francis
	Mark Recapture	MHI	-		-	-	387	65.92 (60.9 - 71.6)	0.24 (0.19 - 0.30)	-	Present Study - Francis
ement	Mark Recapture	МНІ	-		-	-	387	59.7 (56.9 - 63.0)	0.32 (0.27 - 0.38)	-	Present Study - Bayesian Model 1
Growth Increment	Mark Recapture	МНІ	-		-	-	387	60.2 (57.3 - 61.2)	0.35 (0.07 - 0.39)	-	Present Study - Bayesian Model 2
Grow	Mark Recapture	MHI	-		-	-	387	76.8 (13.7 - 162.7)	0.17 (0.14 - 0.20)	-	Present Study - Bayesian Model 3
	Mark Recapture	МНІ	-		-	-	387	77.3 (12.36 - 162.7)	0.24 (0.04 - 0.76)	-	Present Study - Bayesian Model 4
	Mark Recapture	МНІ	-		-	-	387	61.0 (56.1, 66.7)	0.30 (0.23, 0.39)	-	Present Study - Maximum Likelihood Model 5
	Integrative	MHI & NWHI	113	33	3	13	378	67.6 (65.4, 69.6)	0.22 (0.12, 0.25)	-0.37 (-0.47, -0.28)	Present Study - Integrative Model 11

^{*} Linf parameter constrained during fit

Table 3: Von Bertalanffy growth parameter estimates from Bayesia

	Parameter	Mean	SD	2.50%	Median	97.50%	Rî	n eff
	L∞µ	59.89	1.58	57.10	59.80	63.24	1.00	2000
	$L_{\infty\sigma}$	5.47	0.34	4.80	5.46	6.15	1.00	36000
	$L_{\infty T}$	0.03	0.00	0.03	0.03	0.04	1.00	36000
	Shape	26.85	4.40	19.63	26.36	36.86	1.00	2500
7	deviance	3351.95	120.29	3106.84	3355.69	3575.02	1.00	3700
Model 1	k_{μ}	0.32	0.03	0.27	0.32	0.37	1.00	1800
2	k_{σ}	0.01	0.00	0.01	0.01	0.02	1.00	38000
	$k_{\scriptscriptstyle T}$	10741.60	7970.11	2241.98	8492.59	32025.77	1.00	38000
	rate	10.52	1.59	7.88	10.35	14.10	1.00	2800
	τ	0.27	0.04	0.20	0.26	0.36	1.00	5700
	σ	3.85	0.60	2.77	3.82	5.11	1.00	5700
	L∞µ	60.12	1.62	57.21	60.04	63.52	1.00	30000
	$L_{\infty\sigma}$	5.50	0.34	4.85	5.50	6.17	1.00	84000
	L _{∞T}	0.03	0.00	0.03	0.03	0.04	1.00	84000
	Shape	26.64	4.21	19.63	26.24	36.04	1.00	7900
2	deviance	3356.96	119.45	3110.59	3361.29	3578.22	1.00	28000
Model 2	k_{μ}	0.35	0.16	0.07	0.32	0.79	1.00	63000
2	k_{σ}	1.14	85.41	0.01	0.10	3.24	1.00	84000
	k _T	1294.41	3430.44	0.10	105.30	10874.77	1.00	84000
	rate	10.36	1.49	7.85	10.22	13.67	1.00	5200
	τ	0.26	0.04	0.19	0.26	0.36	1.00	21000
	σ	3.88	0.60	2.77	3.85	5.14	1.00	21000
	L∞µ	76.89	31.95	17.19	74.42	160.86	1.00	84000
	$L_{\infty\sigma}$	88.68	4772.30	0.01	1.32	364.22	1.00	84000
	$L_{\infty T}$	570.00	2323.38	0.00	0.57	6238.55	1.00	84000
	Shape	62.32	13.32	41.03	60.49	92.62	1.00	1400
<u>~</u>	deviance	3937.75	46.81	3847.12	3937.23	4030.55	1.00	41000
Model 3	k_{μ}	0.17	0.01	0.14	0.17	0.20	1.00	3200
2	k_{σ}	0.02	0.00	0.01	0.02	0.03	1.00	8000
	$k_{\scriptscriptstyle T}$	2543.07	807.09	1398.95	2400.92	4491.85	1.00	8000
	rate	17.93	3.84	11.85	17.41	26.63	1.00	1600
	τ	0.13	0.01	0.11	0.13	0.15	1.00	84000
	σ	7.92	0.61	6.80	7.89	9.20	1.00	84000
	L∞µ	77.32	33.39	13.20	74.63	166.14	1.00	25000
	$L_{\infty\sigma}$	132.25	9204.30	0.01	1.65	429.54	1.00	84000
	$L_{\infty T}$	529.45	2244.18	0.00	0.37	5947.71	1.00	84000
	Shape	32.76	3.60	26.33	32.55	40.48	1.00	13000
4	deviance	4090.50	39.61	4016.26	4089.51	4171.26	1.00	34000
Model 4	\mathbf{k}_{μ}	0.24	0.18	0.04	0.18	0.77	1.00	84000
Σ	k_σ	5.93	1357.98	0.01	0.09	3.67	1.00	84000
	$k_{\scriptscriptstyle T}$	1293.51	3336.91	0.07	115.99	10767.12	1.00	84000

Table 4: Structure of integrative models (Models 5-11)

Data Source	Model 5	Model 6	Model 7	Model 8	Model 9	Model 10	Model 11
Growth Increment OTP Mark Recapture	X	X	X	X	X	X	X
Direct Aging Ralston & Miyamoto (1983) Integrated Daily Otolith Counts	-	X	X	X	X	-	-
Direct Aging Demartini et al. (1994) Otolith Microincrements	-	X	X	X	X	X	X
Direct Aging Andrews et al. (2012) Bomb Carbon	-	X	X	X	X	X	X
Direct Aging Andrews et al. (2012) Lead:Radium	-	X	X	X	X	X	X
Length Frequency Moffitt & Parrish (1996) Modal Progression	-	X	X	X	X	X	X
Weighting	NA	Equal	By n	Equal	By n	Equal	By n
Pooled Within Data Types?	NA	Yes	Yes	No	No	No	No

Table 5: Sample and Population Parameter Estimates

Parameter Estiamtes for Integrated Growth Models

		Model 5		Model 6			
Parameter	Parameter Sample Population			Population	Sample	Population	Sample
Linf_mu	60.92	60.98 (56.17,66.67)	77.96	66.79 (70.27, 78.69)	64.74	64.80 (61.91, 67.17)	66.87
Linf_std	5.32	5.3 (4.53,6.07)	6.02	5.256 (4.00, 6.83)	5.62	5.57 (4.72, 6.36)	5.53
K	0.300	0.299 (0.229,0.393)	0.122	0.189 (0.121, 0.235)	0.262	0.260 (0.231, 0.302)	0.253
A_mu	0.95	0.95 (0.8,1.09)	1.5	1.21 (1.06, 1.50)	1	1.00 (0.92, 1.08)	0.99
A_sig	0.19	0.19 (0.15,0.24)	0.13	0.16 (0.12, 0.19)	0.18	0.18 (0.14, 0.22)	0.18
Sig	2.1	2.08 (1.50,2.55)	2.97	2.51 (2.05, 3.11)	2.2	2.20 (1.74, 2.62)	2.32
t0	-	-	-0.86	-0.50 (-0.90, -0.34)	-0.31	-0.32 (-0.44, -0.20)	-0.27
oto_sig	-	-	6.79	3.93 (1.31, 7.09)	1.82	1.76 (0.68, 3.03)	1.33
lf_sig	-	-	1.33	3.06 (1.31, 4.06)	4.07	4.39 (3.86, 4.98)	3.93

	Model 9			Model 10	Model 11			
Parameter	arameter Sample Population		Sample	Sample Population		Population		
Linf_mu	64.74	64.80 (62.22, 67.03)	69.34	68.72 (65.23, 71.68)	68.52	67.55 (65.42,69.55)		
Linf_std	5.62	5.58 (4.74, 6.37)	4.26	4.08 (3.00, 5.11)	4.22	5.00 (4.26,5.68)		
К	0.261	0.26 (0.23, 0.30)	0.146	0.17 (0.13, 0.21)	0.173	0.219 (0.198,0.245)		
A_mu	1	1.00 (0.925, 1.08)	1.5	1.37 (1.19, 1.60)	1.34	1.11 (1.03,1.19)		
A_sig	0.18	0.18 (0.15, 0.22)	0.14	0.155 (0.119, 0.184)	0.16	0.17 (0.14,0.2)		
Sig	2.2	2.20 (1.75, 2.61)	3.29	2.99 (2.45, 3.62)	2.9	2.39 (2,2.77)		
t0	-0.31	-0.32 (-0.43, -0.21)	-0.8	-0.65 (-0.96, -0.43)	-0.63	-0.37 (-0.47,-0.28)		
oto_sig	1.82	1.74 (0.66, 2.94)	1.61	1.42 (0.97, 1.84)	1.4	0.96 (0.49,1.31)		
lf_sig	4.07	4.38 (3.88, 4.94)	1.43	2.41 (1.43, 3.29)	3.09	4.63 (4.15,5.15)		

Figure 1: Reporting Grid Map

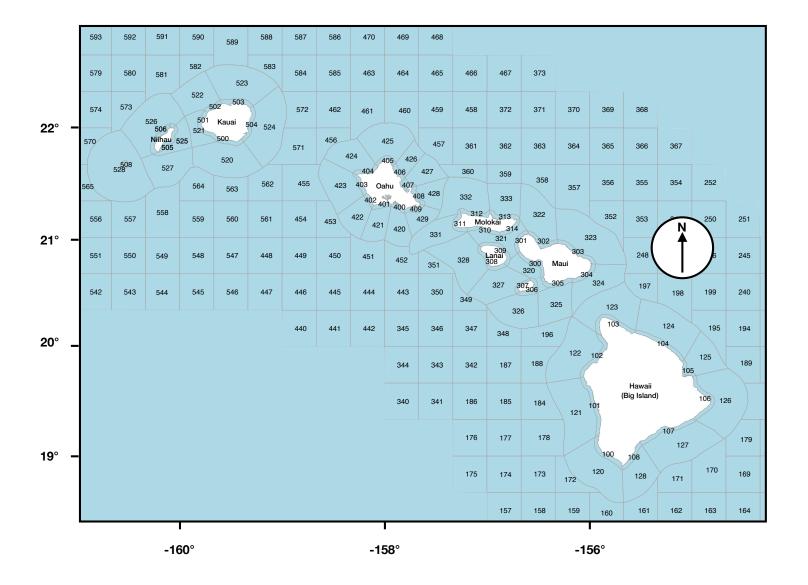
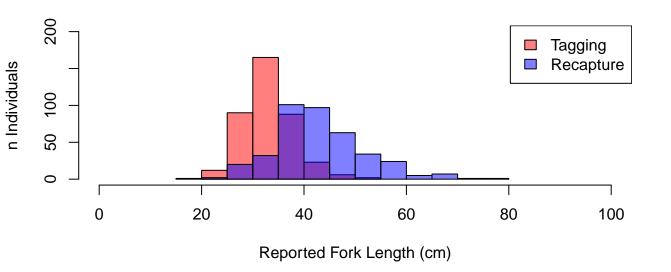


Figure 2: Length and Time at Liberty for OTP Data



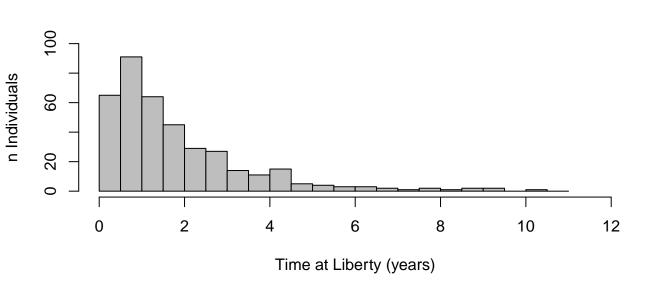


Figure 3: Coefficient of variation of growth function parameters

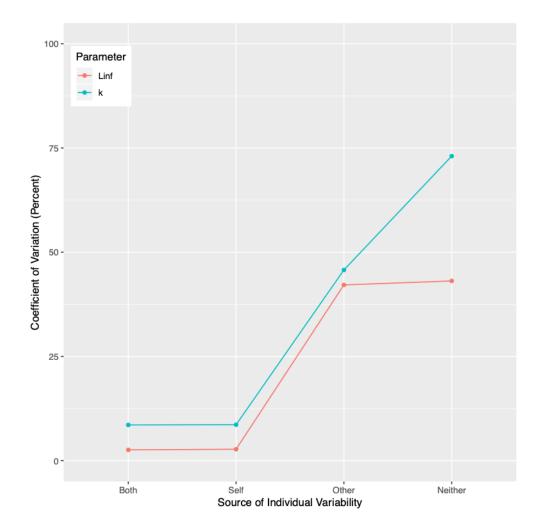


Figure 4: Plots comparing predicted and observed length at recap 7 Model 8 Model 9 Linf = 70.9k = 0.18Linf = 65.01k = 0.26Linf = 66.91k = 0.25Linf = 65.02k = 0.26Line of 1:1 agreement Line of 1:1 agreement Line of 1:1 agreement Line of 1:1 agreement Predicted Length at Recapture (cm) Predictive Variance: 14.073 Predictive Variance: 12.592 Predictive Variance: 12.589 Predictive Variance: 13.092 Observed Length at Recapture (cm) Model 10 Model 11 Model 12 Model 5 Linf = 68.79k = 0.16Linf = 61.45k = 0.29Linf = 60.94k = 0.3Linf = 68.41k = 0.21Line of 1:1 agreement Line of 1:1 agreement Line of 1:1 agreement Line of 1:1 agreement Predicted Length at Recapture (cm) Predictive Variance: 13.287 Predictive Variance: 17.79 Predictive Variance: 12.792 Predictive Variance: 12.725 Observed Length at Recapture (cm) Observed Length at Recapture (cm) Observed Length at Recapture (cm) Observed Length at Recapture (cm)

*Declaration of Interest Statement

Declaration of interests
\boxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.
☐ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: