***Revised growth estimates for Pristipomoides filamentosus in the Hawaiian Islands using mark-recapture studies, Bayesian analysis, and integrative data approaches***

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***Abstract (175 Words Max)***

*Pristipomoides filamentosus* is an economically and culturally important species of deep-water snapper in the Hawaiian archipelago. From 1989 to 1993, the State of Hawaii initiated a fisher participation mark-recapture study to quantify growth and other life history parameters for the species. Over a span of approximately 10 years, 10.5% of 4,179 tagged fish were recaptured. We compared Bayesian and maximum likelihood approaches to estimate von Bertalanffy growth parameters from the tagging data. Direct aging and length frequency data previously used in other published regional growth studies were incorporated to produce integrated estimates of growth. Results from our preferred integrated model reconcile 30+ years of effort from various methods to estimate growth parameters ( = 67.6 cm FL and K = 0.22) and demonstrate the importance of individual variability in *P. filamentosus* due primarily to the asymptotic length parameter . These results have management implications as growth is often an input for age-based stock assessment models and used as a proxy for other life history traits.

***Introduction (500 Words)***

*Pristipomoides filamentosus* (Valenciennes, 1830) is a species of long-lived deep-water snapper distributed throughout the tropical Pacific and Indian Oceans (Allen 1985, Andrews et al. 2012). Known as opakapaka in Hawaii, the species constitutes a significant fraction of the Hawaiian commercial bottomfish fishery, a complex of 6 snapper and 1 grouper species (Ralston and Polovina 1982, Langseth et al. 2018). While the current stock assessment for this fishery used a surplus production model for the entire complex, there is interest in the potential use of species-specific, age-structured assessments that require improved life history studies of age and growth of bottomfish (Langseth et al. 2018).

Growth parameters have been estimated for *P. filamentosus* using a variety of methods in Hawaii and elsewhere (Table 2). Parameter estimates were determined using direct aging approaches from length-at-age data using otolith growth increments (Ralston & Miyamoto, 1983; Uchiyama & Tagami, 1984; Radtke, 1987; DeMartini, Landgraf & Ralston, 1994, Ralston & Williams, 1988). However, age estimates relying on the integration of daily otolith bands may be biased due to episodic growth and/or poor increment resolution in early (< 5 years) life stages (Andrews et al. 2012, Wakefield et al. 2017). Growth was also estimated using a modal progression approach during a length frequency study targeting juvenile fish (< 2 years) but did not consider individual variability when extrapolating growth to larger size classes (Moffitt and Parrish 1996). Preliminary results of an ongoing tagging study have been limited by the size distribution of recaptured individuals and use model parameterizations incompatible with other methods for determining growth (O’Malley 2015). While these studies produced individual estimates of growth parameters, none of them holistically integrated across the three classes of data (direct aging, modal progression, growth increment) to explicitly evaluate the parameter values and sources of uncertainty.

Analytical and statistical advances to methods for estimating growth have been developed to account for sources of variability and permit parameter comparisons across length-at-age, length frequency, and tagging based approaches (Francis 1988, Wang et al. 1995, Eveson et al. 2004). Structural modifications to Fabens (1965) parameterization of the von Bertalanffy growth model address issues of compatibility between growth parameters estimated from tagging studies and other methods, and can reduce bias through the accommodation of modest measurement errors ( Maller and Deboer 1988, James 1991, Palmer et al. 1991, Laslett et al. 2002, Eveson et al. 2004, 2007, Zhang et al. 2009). Maximum likelihood and Bayesian model fitting procedures accommodate individual growth variability by describing population parameters using probability distributions (Francis 1988, Kimura et al. 1993, Wang et al. 1995, Zhang et al. 2009). The flexibility of Bayesian approaches allows and to be sampled in this manner and can account for prior information when estimating parameters. Maximum likelihood approaches typically treat as a fixed effect but flexibility in their implementation has allowed for the development of model structures that can estimate a single set of growth parameters from direct aging, length frequency, and growth increment data simultaneously (Wang et al. 1995, Laslett et al. 2002, Eveson et al. 2004, Zhang et al. 2009).

Here previously unreported tagging data collected in the Main Hawaiian Islands (MHI) are used to estimate growth parameters for *P. filamentosus* usingBayesian and maximum likelihood procedures. A series of models integrating previous length-at-age and length frequency data collected from the MHI and Northwestern Hawaiian Islands (NWHI) with the tagging data are developed to describe growth across most of the species’ life history. Models are tested to determine a preferred model structure. New growth parameters are estimated and compared to those previously reported for the Hawaiian Archipelago.

***Methods***

*Opakapaka Tagging Program*

Tagging data used for this analysis were obtained by biologists from Hawaii’s Division of Aquatic Resources (DAR) within the state’s Department of Land and Natural Resources (DLNR). Between 1989 and 1993 the Opakapaka Tagging Program (OTP), led by staff biologist Henry Okamoto and operating from fishing vessels contracted out of Honolulu Harbor, targeted and tagged *P. filamentosus*.

All tagging effort occurred in the Main Hawaiian Islands (MHI) and was concentrated primarily around the island of Oahu and the Maui Nui complex consisting of the islands of Maui, Molokai, Lanai and Kahoolawe. Since 1990, these areas have accounted for approximately 67.7% of Hawaii’s commercial bottomfish harvest. Coarse location data was provided in the form of the commercial statistical reporting grid areas in which individuals were tagged and recaptured (Table 1, Figure 1). Less than 1% of fish in this study were tagged off the islands of Niihau or Hawaii (Big Island). Adult bottomfish occupy depths between 100 and 400 m along undersea shelves and banks (Parke 2007). In total, the OTPtagged 4,179 juvenile and adult *P. filamentosus*.

Fish were caught with hook-and-line gear and brought to the surface at a rate of 2-5 feet per second. Prior to tagging, each fish was placed in a holding container with aerated seawater to ascertain survival likelihood. If the stomach was inverted and full of gas, it was punctured using a small sharp instrument (e.g., scalpel, hypodermic needle, fish hook). A few scales were carefully removed and a small (~1 cm) incision was made near the fish’s anal opening to assist in expelling gas from the body cavity. Fish appearing lively and upright were deemed likely to survive and thus suitable candidates for tagging. These fish were surgically implanted with unique identifiable internal anchor tags with a monofilament streamer protruding from the incision in the peritoneal cavity. The fork length of each fish measured to the nearest ¼ inch was recorded before the fish was returned headfirst to sea with enough downward momentum to assist in counteracting buoyancy caused by any residual gas.

There were 487 recaptures recorded for 439 unique individuals for a recapture rate of 10.5% of tagged fish. Recaptures of marked *P. filamentosus* were reported up to a decade after tagging with the most recent fish reported in October of 2003 (Okamoto 1993, Kobayashi et al. 2008). Individuals recaptured by OTP personnel were outfitted with an additional tag following procedures similar to their initial capture. For each individual, the location of capture (DAR statistical reporting grid), length at tagging, and date of capture were recorded. Local commercial and recreational fishers were made aware of the program through fliers distributed at the local fish markets, to fish dealers, at fishing supply outlets, and posted at small boat harbors. Fishers were incentivized to report the location, depth, fork length, and date that tagged fish were landed with a $10 reward.

*Tagging Data Management*

The data collected by OTP was entered into an Excel spreadsheet with subsequent analysis performed in R (R Core Team 2014), the Bayesian statistical software JAGS (Plummer 2003), and the R package R2Jags (Su and Yajima 2012). Fish were removed from the dataset if they were not the correct species of interest, if no recapture was reported, or if there was no record of the tag identification number. Fork lengths for the remaining fish recorded at tagging and recapture were linearly transformed from inches to centimeters prior to model fitting. Observed growth () and time at liberty () were calculated for each fish. If an individual was recaptured on more than one occasion, and were only calculated between the first marking event and the last recapture so as to not violate assumptions of independence. Fish with less than 60 days were excluded from the dataset.

*Parameter Estimation from Tagging Data: Bayesian Approach*

Growth parameters were estimated for the *P. filamentosus* tagging data following the Bayesian methodology of Zhang et al. (2009). This approach uses a Fabens version of the von Bertalanffy growth curve but allows the parameters to vary among individuals. Hence the length upon recapture is expressed as:

This is parameterized such that is the length of individual *i* for the *jth* recapture, is the time-at-liberty for individual *i* for the *jth* recapture, is the relative age of individual *i* at tagging (age minus ), and and are the von Bertalanffy growth parameters for the *ith* individual. These individual parameters were drawn from Gaussian distributions defining the population mean values for and . Uninformative priors were used for all input parameters, using Gaussian, gamma, beta, and uniform distributions following the approach of Zhang et al. (2009). The JAGS code for specifying these parameters and performing this analysis is provided in Appendix 1.

The model which allowed both the parameter and parameter to vary across individuals as described above is henceforth referred to as Model 1. Three additional models were run in modified versions of the JAGS code. Model 2 used a fixed parameter while allowing the parameter to vary across individuals. Model 3 used a fixed parameter while allowing the parameter to vary across individuals. Lastly, Model 4 used both a fixed parameter and a fixed parameter. The term “fixed” in this context does not imply a user-specified constant value, but instead refers to the value that is estimated by the Bayesian modeling approach from a single distribution used to represent the mean growth process across all individuals. Model 4 would *a priori* be most similar to the Fabens approach, with both fixed and , but with the added feature of estimating ages at initial tagging, *,* within the Bayesian framework. Inclusion of the term represents a significant improvement over prior methods by modeling growth as a function of age, rather than observed length, allowing growth parameters to be compared between models using tagging data and length-at-age methods (Wang et al. 1995). Model 1 is the presumptive best estimate for *P. filamentosus* von Bertalanffy growth curve parameters, since it would allow the most flexible incorporation of individual variability in the parameter estimation process.

For each Bayesian hierarchical model run, the first 10,000 samples from the posterior distribution were treated as burn-in and discarded from the Monte Carlo simulation. Every 50th sample from the following 500,000 samples (number kept = 10,000) was tabulated into the posterior distributions to reduce potential autocorrelation between sequential values or strings of values. The mean and values from the 10,000 kept samples were used as metrics of population mean values. Median values deviated from mean values by less than one half of 1 percent (Table 3), indicative of symmetrical distributions easily characterized by any descriptor of value tendency (i.e., mean, median, or mode). The results from the Fabens (1965) approach fit using non-linear least squares provided estimates of and (Table 2), which were used as initial starting points in the Bayesian hierarchical approach. Two additional chains were run starting with initial values 50% lower and 100% higher than the initial estimates which resulted in nearly identical solutions as shown in Table 3. Convergence was also ascertained by examination of the Gelman-Rubin statistic (Gelman and Rubin 1992).

The fit of each model was assessed by calculating its Bayesian p-value (pB) from the posterior predictive distribution and the best model was selected using the DIC criterion. Bayesian p-values were of data simulated from model parameters and test whether simulated data is more extreme than the observed data. Bayesian P-values approaching 0.5 indicate the model is a good fit to the data, while extreme Bayesian p-values near 0 or 1 indicate that a given model does not adequately represent the data (Meng 1994). Comparison between models 1-4 were accomplished using DIC. The model with the lowest DIC value was the one that best fits the data with the fewest parameters (CITE).

*Parameter Estimation from Tagging Data: Maximum Likelihood Approach*

Model 5 was fit using the maximum likelihood approach of Laslett, Everson, & Polacheck (2002) using Equation 2.

This method derived growth parameters from the joint distribution of an individual’s length at tagging and recapture to estimate growth parameters. This approach was most similar to model 2 of the Bayesian approach in that asymptotic length, , was treated as a normal random effect while was treated as a fixed unknown parameter. The distribution of was treated as normal with a mean and standard deviation , accounting for individual deviation from the population mean. Rather than using length increments to fit observed growth, a bivariate normal joint distribution of lengths recorded at marking and recapture was used to estimate each individual’s age at tagging . The distribution of individual s is and is treated as a random effect with a lognormal distribution . Measurement error was also treated as a random normal distribution . An unconditional joint density was then derived for each individual by integrating their individual joint distribution with respect to . A detailed description of this process is described by Laslett et. al. (2002).

Growth function parameters were estimated through minimizing of the negative log-likelihood function obtained by summing the unconditional joint density of each individual (E3).

This approach was used to estimate values of the parameters , , , , , and . Two-sided 95% confidence intervals (2.5%, Median, 97.5%) were then estimated from the distribution of each parameter following 10,000 successful bootstrap iterations to obtain population parameters. For each bootstrap iteration, the model was refit on data randomly resampled from the original tagging data with replacement.

*Estimation of Integrative Growth Parameters using sources of growth data*

Datasets previously used to estimate regional growth for *P. filamentosus* in the MHI and NWHI and our tagging data exclusively from the MHI were used to produce a single set of parameter estimates using a modified form of the integrated method proposed by Eveson, Laslett, and Polachek (2004). Additional datasets that were included represent both direct aging and length frequency approaches.

*Parameter Estimation: Length Frequency Data*

Length frequency data consisted of the size distributions of juvenile *P. filamentosus* sampled over 13 months between October 1989 and February 1991 reported by Moffitt and Parrish (1996). The reported fork length of captured fish was binned by 1 cm increments and presented in 13 histograms corresponding to each month of sampling. The number of fish of a given fork length captured during each month of sampling was determined by overlaying a series of evenly spaced horizontal lines across the Y-axis of each histogram corresponding to the addition of a single fish. Using this method to reconstruct monthly length frequency data resulted in a total count of 1,048, individuals while in the original study reported 1,047 (Moffitt and Parrish 1996).

The reconstructed length frequency data were incorporated into integrative models using the two-step method described in Laslett et al 2004. During the first step, a Gaussian mixture model was fit using maximum likelihood and used to decompose the distribution of fork lengths from individuals sampled during discrete time periods for each cohort present in the data. This was accomplished using the normalmixEM function from the mixtools package in R (Benaglia et al. 2009) by constraining the mean of each distribution to the observed mode. A bimodal Gaussian mixture model was fit for the months of October-February, as the original study reported that two cohorts were present during this period, while a single cohort was present the remainder of the year. The estimated mean fork length, and standard error, , of each cohort during each sampling period was used to estimate growth parameters (E4).

With this model, and reflect the fishing year, month, and age cohort, respectively. The estimated age of each cohort during a sampling period is denoted by . July is the month of peak spawning for *P. filamentosus* (Luers et al. 2017) which resulted in age estimates between 3 and 19 months. Sampling and residual model errors were described using random normal distributions and respectively. In contrast to tagging and direct aging components, there is a dearth of information available to estimate the variance component of asymptotic length, using length frequency methods, so this term was modeled as fixed effect, . From this, the expected mode fork length of each cohort (E6), and associated variability during each sampling period (E7) were calculated and used to construct the negative log likelihood function (E8). The rationale for these approximations is discussed to greater depth in Eveson et al. 2004.

*Parameter Estimation: Direct Aging Data*

Sources of direct ageing data consisted of four previously reported length-at-age datasets from three studies. Age estimates for length at age data were obtained through analytical integration of otolith bands (Ralston and Miyamoto, 1983, n = 65), counts of otolith micro increments (DeMartini et al., 1994, n = 35), comparison of bomb radiocarbon (Δ14C) derived from otoliths relative to a standard reference obtained from hermatypic coral cores from the Hawaiian Archipelago (Andrews et al., 2012, n = 33), and the lead-radium ratios of individuals pooled by size class (Andrews et al., 2012, n = 3).

The details of the method for estimating growth parameters from direct aging data components are described in Eveson et al. 2004. Briefly, parameters were modeled using the VBGF model described by equation E9.

Expected length for each individual and the variance of the measurement error was described by equations E10 and E11.

Where denoted the length of the fish*,* at age and was a fixed parameter analogous to when a fish has a hypothetical length of zero. As with the model for tagging data, was the individual asymptotic length of thefish drawn from the random normal distribution . represented the distribution of individual measurement error and was similarly random, drawn from the distribution . Equation 12 describes the log-likelihood function derived from these equations.

An appropriate overall objective likelihood function (E13) was then defined from the sum of the negative log-likelihood functions for tag-recapture, direct aging, length frequency, and growth increment approaches, each with its own scaling constant, .

*Defining an objective function and estimating integrative growth parameters*

A single set of growth parameters best describing the data was obtained by minimizing the objective likelihood function (E13).

By manipulating the value of scaling constants, how similar datasets were treated, and which datasets were included, six additional model structures were developed and evaluated (Table 4). Two approaches were used to define the scaling constants within each model’s objective likelihood function. The first equally weighted each likelihood function so that each data source had equal influence on the resulting parameter estimates. This was achieved by selecting a for each data source equal to the inverse of the number of observations for the data. The second weighted each data source relative to the number of observations of that particular data set ().

The structure of model 5 fit only tagging data from the OTP study while models 6-11 incorporated the additional length-at-age and length frequency data and differed from one another in the treatment of coefficients, whether direct aging data sources were considered independently and assigned their own log-likelihood function or if these data sources were pooled and contributed to estimation of a single log-likelihood function. Omission of direct aging data where ages were estimated by integrating daily growth increments was also considered as this method is likely to result in underestimations of age (Newman et al. 2015)(Table 4).

The six candidate integrative model structures (Models 6-11) were evaluated against one another using the following repeated training-testing cross validation procedure (Burman 1989) to determine the combination of modei

l weighting, data pooling, and data sources parameter estimates that consistently best predicted observed growth from tagging data. Each model structure was trained using two-thirds of the tagging data (n = 258) selected at random while the remaining one-third (n = 129) was reserved for evaluating each model’s predictive ability. Model performance was evaluated using the parameters and estimated from training data, applied to the length at tagging and time at liberty of each individual in the validation set to predict length at recapture using Equation 2. The variance (between the predicted ( and observed (length of each fish recapture was used as a metric for comparing the performance of competing model structures (E14).

The preferred model structure was the one whose estimated parameters most frequently produced the smallest variance. This procedure was repeated 10,000 times. The preferred model structure was the one that most frequently reported the lowest variance across all of these iterations. To determine if incorporating additional data sources improved predictive performance, cross validation variances for the preferred model structure were compared to those calculated using a model structure identical to Model 5, calculated including only tagging data.

The integrative model structure that best predicted observed growth most frequently was refit using the entire data set. Two-sided 95% confidence intervals were estimated for each parameter from the results of 10,000 bootstrap iterations. As with tagging data, the procedure for resampling direct aging data was straightforward and involved random sampling with replacement from the dataset to construct pseudo data sets with an equal number of observations as the original data. Bootstrapping length frequency data was slightly more complicated with each study period in the pseudo data resampled from the corresponding period of the reconstructed study data. Each study period in the pseudo dataset contained the same number of observations as in the corresponding time period of the original study data.

***Results***

*Opakapaka Tagging Program*

Of the 4,179 *P. filamentosus* tagged 439 individuals were recaptured at least once (10.5%, Table 1). Mortality of fish upon release appeared to be generally low, facilitated by the strong tagging selectivity for healthy fish in good condition. Some immediate mortality was observed due to sharks and cetaceans or capture stress (4 individuals). Long-term mortality was thought to be relatively low based upon the high rates of tag return spanning many years. Hydra (small cnidarian polyps) biofouling of the tags was observed for some individuals with large times at liberty, with some lesions apparent around the opening where the tag exited the body cavity. This was not thought to be a serious health issue since the fish appeared to be feeding and swimming normally.

Initial fork length at capture across all individuals ranged in size from 16.5 to 53.3 cm (mean = 31.9 cm, standard deviation (s.d.) = 5.5) and ranged from 19.1 cm and 52.8 cm (mean = 32.8, s.d. = 5.1) for fish that were later recaptured. For those fish that were later recaptured, fork length at recapture ranged between 22.9 cm and 76.2 cm (mean = 41.9, s.d. = 8.7). The minimum time at liberty for any fish between tagging and recapture was a single day while the maximum time at liberty was 10.3 years (3,748 days) (Figure 2). The mean time at liberty was 1.82 years or 666 days (s.d. = 625).

One fish was excluded from further analysis as its fork length at capture was not recorded. Seven fish were removed because the recapture date was not properly recorded. Of the remaining 432 fish recaptured, 351 were recaptured a single time, 33 fish were recaptured a total of two times, one fish recaptured 3 times, and two fish were recaptured 4 times. We also excluded from analysis 45 individuals for whom time at liberty was less than 60 days yielding a data set of 387 unique individuals.

*Estimating Growth Parameters from Tagging Data: Bayesian Approach*

The Bayesian hierarchical approach using the JAGS software yielded mean estimates of and for each of the Models 1–4 examined (Table 2). The presumptive optimal model which incorporated individual variability in both and (Model 1) yielded mean parameter estimates of = 58.8 cm (coefficient of variation [c.v.] = 2.59) and = 0.32 (c.v. = 8.57). and parameter estimates for Model 2, where was fixed, were 60.2 cm (c.v. = 2.74) and 0.35 (c.v. = 45.7) respectively. Under Model 3, where was fixed and was fit freely = 76.8 cm (c.v. = 42.2) and = 0.17 (c.v. = 8.52). = 77.3 cm (c.v. = 43.1) and = 0.24 (c.v. = 73.1) for Model 4, where both parameters were fixed. Additional parameters for each of the four models are presented in Table 3. The Gelman-Rubin convergence criteria indicated that the model solutions were credible, with asymptotic convergence clearly occurring after ~4000 iterations, well within the burn-in phase of the Bayesian modeling runs.

*Parameter estimation using maximum likelihood*

The maximum likelihood approach used for Model 5 successfully converged to produce estimates of , , , , , and (Table 5). Bootstrap confidence intervals of parameters and overlapped and parameters from Bayesian models 1 and 2 (Table 2). From these results, it was concluded that estimates produced by maximum likelihood were satisfactorily similar to estimates from the Bayesian approach. Model residuals were distributed around zero fairly consistently for all but the largest fish. For fish with recapture lengths exceeding 60 cm, growth models underestimated observed recapture lengths (Figures 3).

*Comparing model performance*

Across all 10,000 cross validation iterations to determine model structure, the mean predictive variance metric ranged between 7.29 and 24.96 (mean = 14.20, s.d. = 2.20) where a lower predictive variance indicates a better model fit. From all candidate likelihood models, the structure of Model 11 best predicted cross validation data in 3,486 of 10,000 iterations. The predictive variance for Model 11 ranged between 7.29 and 20.10 (mean = 13.64, s.d. = 1.91). The structure of Model 5, fit exclusively using tagging data, ranged in predictive variance between 7.17 and 26.09 (mean = 14.35, s.d. = 2.44). The structure of Model 11 performed better than the structure of Model 5 in 6,351 of 10,000 cross validation iterations. Differences in predictive variance between these two competing structures ranged between -1.60 and 10.80 (mean = 0.72, s.d. = 1.37) and indicated that the inclusion of additional growth data did improve the predictive capability of growth models compared to tagging data alone. Bootstrapped parameter estimates that were refitted(?) using the prefered model structure and Model 5’s tagging only data are summarized in Table 2 and all parameters for models 5-11 are reported in full in Table 5. When fit to the entire tagging data set, the residual pattern of Model 11 also underestimated lengths at recapture length for the largest individuals.

***Discussion***

Our integrative model results reconcile 30+ years of efforts to determine growth for *P. filamentosus* in the Hawaiian Archipelago and provide robust support for some observed life history parameters. Growth parameters derived using integrative models that incorporated additional length frequency and length-at-age data were better able to predict observed growth in recaptured fish. These parameters were in agreement with those derived from; 1) the fit of only integrated daily growth increments from otoliths collected in the NWHI without constraining (Ralston and Miyamoto 1983), 2) integrated daily growth increments and microincrement counts of from the MHI and NWHI (DeMartini et al. 1994), and 3) the radioisotopic composition of otolith material and counts of otolith increments from the MHI and NWHI (Andrews et al. 2012). These findings are consistent with broad genetic homogeneity in the species throughout the Hawaiian Archipelago (Shaklee and Samollow 1984, Gaither et al. 2010, 2011) and support the implicit assumption that tagging individuals did not disrupt their growth trajectory. Integrative parameters differed from estimates from an ongoing mark recapture study in the MHI which reported faster growth and smaller asymptotic lengths (O’Malley 2015). These differences could arise from real changes in growth between the periods fish were collected, methodological differences, as well as that thus far, none of the fish recaptured during the ongoing study have been of the largest size classes (maximum size reported = 47.6 cm FL).

Compared to their broader distribution, *P. filamentosus* from the Hawaiian archipelago were slower growing but obtained larger asymptotic lengths than those from the Mariana Archipelago (Ralston and Williams 1988) and Papua New Guinea (Fry et al. 2006, Andrews et al. 2012), and were faster growing but ultimately smaller in their asymptotic length when compared to estimates from the Seychelles (Mees 1993, Hardman-Mountford et al. 1997, Mees and Rousseau 1997, Pilling 2000).

Comparing growth parameter estimates fit exclusively with OTP data indicate that Bayesian and maximum likelihood fitting methods performed similarly. The treatement of individual variability in parameters estimated in Model 2 were identical to those used to fit Model 5 (OTP data only). Parameters estimated by Models 1 and 2 were contained within the 95% confidence intervals of Model 5. Integrative Models 6-11 were evaluated under the same assumptions of parameter variability as models 2 and 5.

Of the Bayesian models, Model 1 was the presumed best. This model allowed both and to vary for each fish. Models 2-4 suggested that individual variability in both and was important, with perhaps variability in being more important based upon the response of standard deviation from the base case of Model 1 to the constrained individual variability in Model 3 and Model 4 (Figure 3). Based upon parameter estimates and patterns of standard deviation, it is likely that Model 3 and Model 4 were not credible. Similar performance between Model 1 and Model 2 suggested that the primary source of individual variability was due to variability in the parameter. This is consistent with other studies where the best models accounted for individual variability in both terms but accounting for individual variation in the term alone was sufficient to describe growth while significantly reducing computational complexity (Eveson et al. 2007, Zhang et al. 2009).

Across all models, the parameters from Model 11 best predicted length at recapture across validation iterations and therefore represents the best estimated parameter set. Information from older/larger fish was very important for grounding the upper end of integrative growth curves resulting in parameters that better predicted length at recapture. Omission of the largest individuals from Models 1-5 resulted in lower estimates of , causing growth curves to asymptote prematurely. The OTP data described here includes *P. filamentosus* with fork lengths at capture spanning 19.1 - 52.8 cm (mean = 32.9, s.d. = 5.1). Using parameters from Model 11, this corresponded to mean ages between 1.5 and 7 years (Figure 5).

Additional data sources included in integrative models represent collections spanning several decades and were collected across both the MHI and NWHI. When incorporating these additional data sources, it is assumed that growth within the population did not differ significantly with time or region. Genetic homogeneity between NWHI and MHI stocks (Gaither et al. 2010, Gaither et al. 2011) justified incorporating data from both regions and with the exception of Ralston and Miyamoto (1983), all subsequent studies of growth for *P. filamentosus* in the Hawaiian archipelago have included data or parameter estimates from one or more previous studies in their calculations regardless of time and place of collection (DeMartini et al. 1994, Moffitt and Parrish 1996, Andrews et al. 2012). However, these spatial and temporal assumptions may not reflect phenotypic realities and further work is required to resolve whether differences in growth exist between the two regions.

Parameters obtained from our models and those published elsewhere underestimate the size at recapture for the largest fish in the OTP dataset (approximate fork length > 50 cm) (Figure 4). We suspect sexual dimorphism may explain this poor predictive ability. Dimorphic growth patterns have been observed in a number of other lutjanid species for *P. filamentosus* elsewhere in distribution, with males attaining greater asymptotic lengths (Grimes 1987, Mees 1993, Newman et al. 2000, Newman and Dunk 2002, Williams et al. 2017, Taylor et al. 2018). Estimations of growth parameters for *P. filamentosus* in the Central Pacific have been historically sex agnostic and methods for non-invasive sexing of this species was unknown until recently (Luers et al. 2017). If one sex attains a greater asymptotic length than the other, that sex is likely to be over represented in the largest size classes relative to the total population. At sizes where the sex ratio of individuals is similar to the sex ratio of the total sampled population, averaging of model parameters between sexes results in excess model deviation. However, for the largest sizes where sex ratios are not representative of the population as a whole, estimated growth parameters represent an average of both sexes and will underestimate recapture lengths for largest individuals from one sex while overestimating the recapture length of the largest individuals of the other.

Accurate estimates of von Bertalanffy growth parameters are very important for management. Growth parameters are often used directly or indirectly in stock assessment and fisheries management (Polovina et al. 1987, Haight et al. 1993). These efforts are sensitive to both growth parameters and the model used to estimate those parameters. For example, the rate of instantaneous natural mortality *M* is a value of interest often inferred using empirical relationships between *M* and (Ralston 1987, Jensen 1996, Thorson et al. 2017). Underestimating will underestimate *M*, characterizing a stock as less productive than it actually is while overestimating K will overestimate M. If the management regime is linked to such a flawed estimate of stock productivity, then the stock is likely to be mismanaged and under or over harvested, respectively, relative to its true biological potential. Future work to refine growth estimates for *P. filamentosus* should consider that growth trajectories may differ between males and females.

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**References**

Allen, G.R. 1985. Fao Species Catalogue Vol . 6 . Snappers of the World. Fao Fish. Synopsis **6**(125): 208. doi:10.1016/0025-326X(92)90600-B.

Andrews, A.H., DeMartini, E.E., Brodziak, J., Nichols, R.S., and Humphreys, R.L. 2012. A long-lived life history for a tropical, deepwater snapper (Pristipomoides filamentosus): bomb radiocarbon and lead-radium dating as extensions of daily increment analyses in otoliths. Can. J. Fish. Aquat. Sci. **69**: 1850–1869. doi:10.1139/f2012-109.

Andrews, A.H., Humphreys, R.L., DeMartini, E.E., Nichols, R.S., and Brodziak, J. 2011. Bomb Radiocarbon and Lead-Radium Dating of Opakapaka (Pristipomoides filamentosus). *In* NOAA Technical Memorandum NMFS-PIFSC. Honolulu, HI. Available from http://www.pifsc.noaa.gov/library/pubs/admin/PIFSC\_Admin\_Rep\_11-07.pdf.

Benaglia, T., Chauveau, D., Hunter, D.R., and Young, D. 2009. Mixtools: An R package for analyzing finite mixture models. J. Stat. Softw. **32**(6): 1–29. Available from http://www.jstatsoft.org/v32/i06/.

Burman, P. 1989. A comparative study of ordinary cross-validation, v-fold cross-validation and the repeated learning-testing methods. Biometrika **76**(3): 503–514. doi:10.1093/biomet/76.3.503.

DeMartini, E.E., Landgraf, K.C., and Ralston, S. 1994. A recharacterizatinon of the age-length and growth relationships of Hawaiian snapper Pristipomoides filamentosus. U.S. Department of Commerce, National Oceanic and Atmospheric Administration, National Marine Fisheries Service, Southwest Fisheries Science Center, Honolulu, HI.

Eveson, J.P., Laslett, G.M., and Polacheck, T. 2004. An integrated model for growth incorporating tag–recapture, length–frequency, and direct aging data. Can. J. Fish. Aquat. Sci. **61**(2): 292–306. doi:10.1139/f03-163.

Eveson, J.P., Polacheck, T., and Laslett, G.M. 2007. Consequences of assuming an incorrect error structure in von Bertalanffy growth models: a simulation study. Can. J. Fish. Aquat. Sci. **64**(4): 602–617. doi:10.1139/f07-036.

Francis, R.I.C.C. 1988. Maximum likelihood estimation of growth and growth variability from tagging data. New Zeal. J. Mar. Freshw. Res. **22**(1): 43–51. doi:10.1080/00288330.1988.9516276.

Fry, G.C., Brewer, D.T., and Venables, W.N. 2006. Vulnerability of deepwater demersal fishes to commercial fishing: Evidence from a study around a tropical volcanic seamount in Papua New Guinea. Fish. Res. **81**(2–3): 126–141. doi:10.1016/j.fishres.2006.08.002.

Gaither, M.R., Jones, S. a., Kelley, C., Newman, S.J., Sorenson, L., and Bowen, B.W. 2011. High connectivity in the deepwater snapper Pristipomoides filamentosus (lutjanidae) across the indo-pacific with isolation of the Hawaiian archipelago. PLoS One **6**(12): 1–13. doi:10.1371/journal.pone.0028913.

Gaither, M.R., Toonen, R.J., Sorenson, L., and Bowen, B.W. 2010. Isolation and characterization of microsatellite markers for the crimson jobfish, pristipomoides filamentosus (Lutjanidae). Conserv. Genet. Resour. **2**: 169–172. doi:10.1007/s12686-009-9119-3.

Gelman, A., and Rubin, D.B. 1992. lnference from Iterative Simulation Using Multiple Sequences. Stat. Sci. **7**(4): 457–472. doi:10.2307/2246093.

Grimes, C.B. 1987. Reproductive biology of the Lutjanidae: A review. *In* Tropical snappers and groupers: Biology and fisheries management. *Edited by* J.J. Polovina and S. Ralston. Westview Press, Boulder, Colorado. pp. 239–294.

Haight, W.R., Kobayashi, D.R., and Kawamoto, K.E. 1993. Biology and Management of Deepwater Snappers of the Hawaiian Archipelago. Mar. Fish. Rev. **55**(2): 20–27.

Hardman-Mountford, N.J., Polunin, N.V.C., and Boulle, D. 1997. Can the age of the tropical species be determined by otolith measurement?: a study using Pristipomoides filamentosus (Pisces: Lutjanidae) from the Mahe Plateau, Seychelles. Naga, ICLARM Q. **20**(2): 27–31.

Hawaii Reported Landing Tables. 2016. Available from https://www.pifsc.noaa.gov/wpacfin/hi/dar/Pages/hi\_data\_3.php [accessed 23 March 2018].

James, I.R. 1991. Estimation of von Bertalanffy growth curve parameters from recapture data. Biometrics **47**: 1519–1530. doi:10.2307/2532403.

Jensen, A.L. 1996. Beverton and Holt life history invariants result from optimal trade-off of reproduction and survival. Can. J. Fish. Aquat. Sci. **53**(4): 820–822. doi:10.1139/f95-233.

Kimura, D.K., Shimada, A.M., and Lowe, S.A. 1993. Estimating von Bertalanffy growth parameters of sablefish Anoplopoma fimbria and Pacific cod Gadus macrocephalus using tag-recapture data. Fish. Bull. **91**(2): 271–280.

Kobayashi, D.R., Okamoto, H.Y., and Oishi, F.G. 2008. Movement of the deepwater snapper opakapaka, Pristipomoides filamentosus, in Hawaii: Insights from a large-scale tagging program and computer simulation.

Langseth, B., Syslo, J., Yau, A., Kapur, M., and Brodziak, J. 2018. Stock assessment for the main Hawaiian Islands Deep 7 bottomfish complex in 2018, with catch projections through 2022. NOAA Tech. Memo. NMFS-PIFSC **69**(February): 217.

Laslett, G.M., Eveson, J.P., and Polacheck, T. 2002. A flexible maximum likelihood approach for fitting growth curves to tag-recapture data. Can. J. Fish. Aquat. Sci. **59**(6): 976–986. doi:10.1139/f02-069.

Luers, M.A., DeMartini, E.E., and Humphreys, R.L.J. 2017. Seasonality, sex ratio, spawning frequency and sexual maturity of the opakapaka Pristipomoides filamentosus (Perciformes: Lutjanidae) from the Main Hawaiian Islands: fundamental input to size-at-retention regulations. Mar. Freshw. Res. **69**(2): 325–335. doi:10.1071/MF17195.

Maller, R.A., and Deboer, E.S. 1988. An analysis of two methods of fitting the von Bertalanffy curve to capture-recapture data. Mar. Freshw. Res. **39**(4): 459–466. doi:10.1071/MF9880459.

Mees, C.C. 1993. Population biology and stock assessment of Pristipomoides filamentosus on the Mahe Plateau, Seychelles. J. Fish Biol. **43**(5): 695–708. doi:10.1111/j.1095-8649.1993.tb01147.x.

Mees, C.C., and Rousseau, J.A. 1997. The potential yield of the lutjanid fish Pristipomoides filamentosus from the Mahe Plateau, Seychelles: Managing with uncertainty. Fish. Res. **33**(1–3): 73–87. doi:10.1016/S0165-7836(97)00069-6.

Meng, X.-L. 1994. Posterior predictive p-values. Ann. Stat. **22**(3): 1142–1160.

Moffitt, R.B., and Parrish, F.A. 1996. Habitat and life history of juvenile Hawaiian pink snapper, Pristipomoides filamentosus. Pacific Sci. **50**(4): 371–381.

Newman, S.J., Cappo, M., and Williams, D.M.B. 2000. Age, growth and mortality of the stripey, Lutjanus carponotatus (Richardson) and the brown-stripe snapper, L. vitta (Quoy and Gaimard) from the central Great Barrier Reef, Australia. Fish. Res. **48**(3): 263–275. doi:10.1016/S0165-7836(00)00184-3.

Newman, S.J., and Dunk, I.J. 2002. Growth, age validation, mortality, and other population characteristics of the red emperor snapper, Lutjanus sebae (Cuvier, 1828), off the Kimberley coast of north-western Australia. Estuar. Coast. Shelf Sci. **55**(1): 67–80. doi:10.1006/ecss.2001.0887.

Newman, S.J., Wakefield, C.B., Williams, A.J., Nicol, S.J., O’Malley, J.M., DeMartini, E.E., Humphreys, R.L., Andrews, A.H., Nichols, R.S., Halafihi, T., Kaltavara, J., and Taylor, B.M. 2015. International workshop on methodological evolution to improve estimates of life history parameters and fisheries management of data-poor deep-water snappers and groupers. Mar. Policy **60**: 182–185.

O’Malley, J. 2015. A Review of the Cooperative Hawaiian Bottomfish Tagging Program of the Pacific Islands Fisheries Science Center and the Pacific Islands Fisheries Group. Honolulu, HI. doi:10.7289/V59W0CF7.

Okamoto, H.Y. 1993. Develop opakapaka (pink snapper) tagging techniques to assess movement behavior. Honolulu, HI.

Palmer, M.J., Phillips, B.F., and Smith, G.T. 1991. Application of nonlinear models with random coefficients to growth data. Biometrics **47**: 623–635. doi:10.2307/2532151.

Parke, M. 2007. Linking Hawaii Fisherman Reported Commercial Bottomfish Catch Data to Potential Bottomfish Habitat and Proposed Restricted Fishing Areas using GIS and Spatial Analysis. Fish. Sci. (September): 1–45.

Pilling, G.M. 2000. Validation of annual growth increments in the otoliths of the lethrinid Lethrinus mahsena and the lutjanid Aprion virescens from sites in the tropical Indian Ocean, with notes on the nature of growth increments in Pristipomoides filamentosus. Fish. Bull. **98**(3): 600–611.

Plummer, M. 2003. JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing. *In* Proceedings of the 3rd international workshop on distributed statistical computing. Vol. 124.

Polovina, J.J., Ralston, S., and Ralston, S. 1987. Assessment and management of deepwater bottom fishes in Hawaii and the Marianas. Trop. snappers groupers Biol. Fish. Manag.: 505–532.

R Core Team. 2014. R: A Language and Environment for Statistical Computing. Vienna, Austria. Available from http://www.r-project.org/.

Radtke, R.L. 1987. Age and growth information available from the otoliths of the Hawaiian snapper, Pristipomoides filamentosus. Coral Reefs **6**(1): 19–25. doi:10.1007/BF00302208.

Ralston, S., and Miyamoto, G.T. 1983. Analyzing the width of daily otolith increments to age the Hawaiian snapper, Pristipomoides filamentosus. Fish. Bull. **81**: 523–535.

Ralston, S. V., and Williams, H.A. 1988. Depth distributions, growth, and mortality of deep slope fishes from the Mariana archipelago.

Ralston, S.V.D. 1987. Mortality rates of snappers and groupers. Trop. snappers groupers Biol. Fish. Manag.: 375–404.

Ralston, S.V.D., and Polovina, J. 1982. A multispecies analyis of the commercial deep-sea handline fishery in Hawaii. Fish. Bull. **80**(3): 435–448.

Shaklee, J.B., and Samollow, P.B. 1984. Genetic variation and population structure in a deepwater snapper, Pristipomoides filamentosus, in the Hawaiian Archipelago. Fish. Bull. **82**(4): 703–713.

Su, Y., and Yajima, M. 2012. R2jags: A Package for Running JAGS from R. Available from http://cran.r-project.org/package=R2jags.

Taylor, B.M., Oyafuso, Z.S., Pardee, C.B., Ochavillo, D., and Newman, S.J. 2018. Comparative demography of commercially-harvested snappers and an emperor from American Samoa. PeerJ **6**: e5069. doi:10.7717/peerj.5069.

Thorson, J.T., Munch, S.B., Cope, J.M., and Gao, J. 2017. Predicting life history parameters for all fishes worldwide. Ecol. Appl. **27**(8): 2262–2276. doi:10.1002/eap.1606.

Uchiyama, J.H., and Tagami, D.T. 1984. Life history, distribution, and abundance of bottomfishes in the Northwestern Hawaiian Islands. *In* Proceedings of the Second Symposium on Resource Investigations in the Northwestern Hawaiian Islands. *Edited by* R.W. Grigg and K.Y. Tanoue. pp. 229–247.

Wakefield, C.B., O’Malley, J.M., Williams, A.J., Taylor, B.M., Nichols, R.S., Halafihi, T., Humphreys, R.L., Kaltavara, J., Nicol, S.J., and Newman, S.J. 2017. Ageing bias and precision for deep-water snappers: Evaluating nascent otolith preparation methods using novel multivariate comparisons among readers and growth parameter estimates. ICES J. Mar. Sci. **74**(1): 193–203. doi:10.1093/icesjms/fsw162.

Wang, Y.-G., Thomas, M.R., and Somers, I.F. 1995. A maximum likelihood approach for estimating growth from tag–recapture data. Can. J. Fish. Aquat. Sci. **52**(2): 252–259. doi:10.1139/f95-025.

Williams, A.J., Wakefield, C.B., Newman, S.J., Vourey, E., Abascal, F.J., Halafihi, T., Kaltavara, J., and Nicol, S.J. 2017. Oceanic, Latitudinal, and Sex-Specific Variation in Demography of a Tropical Deepwater Snapper across the Indo-Pacific Region. Front. Mar. Sci. **4**(December). doi:10.3389/fmars.2017.00382.

Zhang, Z., Lessard, J., and Campbell, A. 2009. Use of Bayesian hierarchical models to estimate northern abalone, Haliotis kamtschatkana, growth parameters from tag-recapture data. Fish. Res. **95**(2–3): 289–295. doi:10.1016/j.fishres.2008.09.035.

**Appendix 1**. JAGS code for Bayesian hierarchical growth model. Model 1 incorporates both L∞ and K individual variability; Model 2 incorporates L∞ individual variability; Model 3 incorporates K individual variability; and Model 4 incorporates no individual variability. Methodology from Zhang et al. (2009).

**# Model 1**

**model{**

**for (i in 1:387) {**

**for (j in 2:n[i]) {**

**L[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**L\_Exp[i, j] <- Linf[i] \*(1.0 - exp(-k[i]\*(A[i]+t[i, j -1])))**

**L.pred[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**p.value[i, j] <- step(L.pred[i, j] - L[i, j])**

**}**

**L[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**L\_Exp[i, 1] <- Linf[i] \*(1.0 - exp(-k[i]\*A[i]))**

**L.pred[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**p.value[i, 1] <- step(L.pred[i, 1]- L[i, 1])**

**Linf[i] ~ dnorm(Linf\_mu, Linf\_tau)**

**k[i] ~ dnorm(k\_mu, k\_tau) I(0,1)**

**A[i] ~ dgamma(shape, rate)**

**}**

**Linf\_std <- sqrt(1/Linf\_tau)**

**k\_std <- sqrt(1/k\_tau)**

**var <- 1/tau**

**Linf\_mu ~ dnorm(100, 0.0001)**

**Linf\_tau ~ dgamma(0.001, 0.0001)**

**shape ~ dunif(0, 100)**

**rate ~ dunif(0, 100)**

**k\_mu ~ dbeta(1, 1)**

**k\_tau ~ dgamma(0.001, 0.0001)**

**tau ~ dgamma(0.001, 0.0001)**

**}**

**# Model 2**

**model{**

**for (i in 1: 387) {**

**for (j in 2:n[i]) {**

**L[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**L\_Exp[i, j] <- Linf[i] \*(1.0 - exp(-k\*(A[i]+t[i, j -1])))**

**L.pred[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**p.value[i, j] <- step(L.pred[i, j] - L[i, j])**

**}**

**L[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**L\_Exp[i, 1] <- Linf[i] \*(1.0 - exp(-k\*A[i]))**

**L.pred[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**p.value[i, 1] <- step(L.pred[i, 1]- L[i, 1])**

**Linf[i] ~ dnorm(Linf\_mu, Linf\_tau)**

**A[i] ~ dgamma(shape, rate)**

**}**

**Linf\_std <- sqrt(1/Linf\_tau)**

**k\_std <- sqrt(1/k\_tau)**

**var <- 1/tau**

**k ~ dnorm(k\_mu, k\_tau) I(0,1)**

**Linf\_mu ~ dnorm(100, 0.0001)**

**Linf\_tau ~ dgamma(0.001, 0.0001)**

**shape ~ dunif(0, 100)**

**rate ~ dunif(0, 100)**

**k\_mu ~ dbeta(1, 1)**

**k\_tau ~ dgamma(0.001, 0.0001)**

**tau ~ dgamma(0.001, 0.0001)**

**}**

**# Model 3**

**model{**

**for (i in 1: 387) {**

**for (j in 2:n[i]) {**

**L[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**L\_Exp[i, j] <- Linf\*(1.0 - exp(-k[i]\*(A[i]+t[i, j -1])))**

**L.pred[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**p.value[i, j] <- step(L.pred[i, j] - L[i, j])**

**}**

**L[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**L\_Exp[i, 1] <- Linf \*(1.0 - exp(-k[i]\*A[i]))**

**L.pred[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**p.value[i, 1] <- step(L.pred[i, 1]- L[i, 1])**

**k[i] ~ dnorm(k\_mu, k\_tau) I(0,1)**

**A[i] ~ dgamma(shape, rate)**

**}**

**Linf\_std <- sqrt(1/Linf\_tau)**

**k\_std <- sqrt(1/k\_tau)**

**var <- 1/tau**

**Linf ~ dnorm(Linf\_mu, Linf\_tau)**

**Linf\_mu ~ dnorm(100, 0.0001)**

**Linf\_tau ~ dgamma(0.001, 0.0001)**

**shape ~ dunif(0, 100)**

**rate ~ dunif(0, 100)**

**k\_mu ~ dbeta(1, 1)**

**k\_tau ~ dgamma(0.001, 0.0001)**

**tau ~ dgamma(0.001, 0.0001)**

**}**

**# Model 4**

**model{**

**for (i in 1: 387) {**

**for (j in 2:n[i]) {**

**L[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**L\_Exp[i, j] <- Linf\*(1.0 - exp(-k\*(A[i]+t[i, j -1])))**

**L.pred[i, j] ~ dnorm(L\_Exp[i, j], tau)**

**p.value[i, j] <- step(L.pred[i, j] - L[i, j])**

**}**

**L[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**L\_Exp[i, 1] <- Linf \*(1.0 - exp(-k\*A[i]))**

**L.pred[i, 1] ~ dnorm(L\_Exp[i, 1], tau)**

**p.value[i, 1] <- step(L.pred[i, 1]- L[i, 1])**

**A[i] ~ dgamma(shape, rate)**

**}**

**Linf\_std <- sqrt(1/Linf\_tau)**

**k\_std <- sqrt(1/k\_tau)**

**var <- 1/tau**

**k ~ dnorm(k\_mu, k\_tau) I(0,1)**

**Linf ~ dnorm(Linf\_mu, Linf\_tau)**

**Linf\_mu ~ dnorm(100, 0.0001)**

**Linf\_tau ~ dgamma(0.001, 0.0001)**

**shape ~ dunif(0, 100)**

**rate ~ dunif(0, 100)**

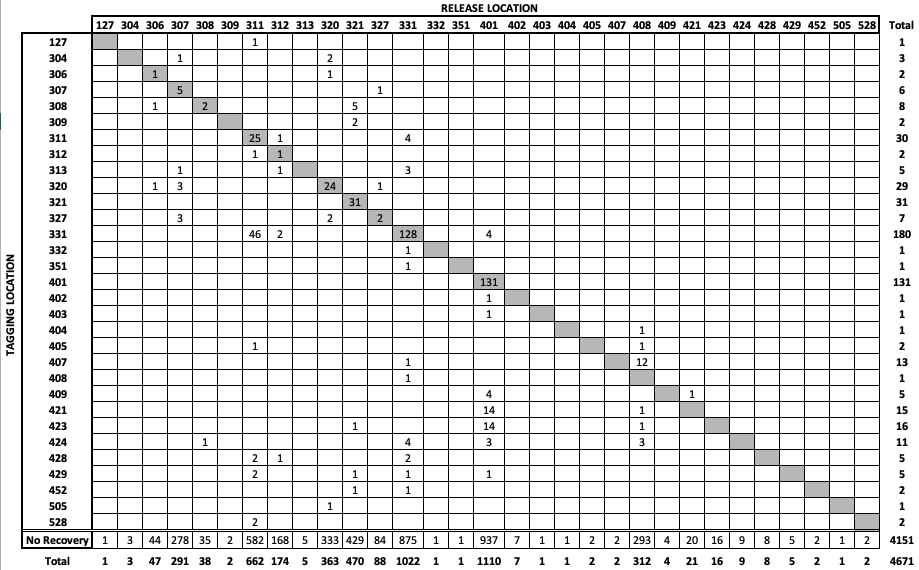
**k\_mu ~ dbeta(1, 1)**

**k\_tau ~ dgamma(0.001, 0.0001)**

**tau ~ dgamma(0.001, 0.0001)**

**}**

**Table 1.** Summary of OTP tagging and recapture data for fish with valid locations. Release and recapture location numbers correspond to the State of Hawaii’s statistical reporting grids (Figure 1). Adapted from Kobayashi, Okamoto & Oishi (2008).

****

**Table 2.** Estimates of vonBertalanffy parameters including average asymptotic length () the Brody growth coefficient (*k),* and theoretical age at length zero t0 for *P. filamentosus* estimated in the Main Hawaiian Islands (MHI), Northwestern Hawaiian Islands (NWHI) and pooled across the Hawaii Archipelago. When available in the literature, 95% confidence intervals for parameter estimates are presented in brackets next to parameter point estimates. A screenshot of a cell phone

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**Table 3**. Bayesian hierarchical growth model specifications for Bayesian models. Monte Carlo simulation was burned in for n=10,000 runs with every 50th of the following 500,000 runs retained for tabulation into the posterior distributions. Variable names are kept consistent with the Appendix 1 JAGS code and are not consistent with text references to von Bertalanffy growth parameters but remain intuitively similar (e.g., K=k\_mu, L∞=Linf\_mu).

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**Table 4.** A reference for the candidate model structures used to determine the preferred integrative model structure.



**Table 5.** Sample and population parameter estimates from maximum likelihood growth models. Model 5 was fit to only the tagging data and Model 11 is the preferred model. For both models, parameter estimates fit to the full data set are reported in the Sample Estimate columns while bootstrapped parameter estimates (Median, 2.5%, 97.5%) are reported under the Population CI column.

**A picture containing text, receipt

Description automatically generatedFigure 1:** Map showing the location and number of the State of Hawaii’s statistical reporting grids. The number of fish tagged and recaptured in each reporting grid are summarized in Table 1.

**A close up of a map

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**Figure 2.** The length of *P. filamentosus* recaptured and included in analysis of OTP tagging data and the distribution of times at liberty. The fork length of fish during tagging is highlighted in red while length at recapture is shown in blue.



**Figure 3:** Coefficient of variation for 2 von Bertalanffy growth curve parameters K and L∞ for *P. filamentosus*. Individual variability was examined incorporating individual variability in both parameters, in either one of the parameters in series, or in neither parameter.

****

**Figure 4.** Plots comparing observed and predicted recapture lengths fit using parameter point estimates from Bayesian Models 1 and 2, as well as population parameter estimates from Maximum likelihood Models 5 and 11. Length at recapture was predicted as a function of length at marking and time at liberty. The 1:1 line indicates where points would fall if model parameters perfectly predicted length at recapture.



**Figure 5.** Comparison of von Bertalanffy growth function curves produced from parameters during this study. Horizontal dashed lines indicate the minimum (min Lm) and maximum length (max Lm) of individuals at the time of marking recorded in the OTP dataset. Growth curves were estimated for two best Bayesian (Models 1 and 2) and maximum likelihood models (Models 5 and 11). Note that while the species may live to 40+ years, this plot has been truncated to 20 years to highlight differences between early growth under each model.

