

Data exploratory analysis

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## ##	The following object is masked _byGlobalEnv:	
##		

Project: Comprehensive Omics Catalogue for Hartwell

St. Jude Children's Research Hospital Bio
Hackathon Team ${\bf 1}$

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1 Information about this notebook

This is an exploratory analysis of the data availability in terms of assays in the Comprehensive Omics Catalogue for Hartwell. This is critical for mitigating duplicate sequencing requests and efforts through Hartwell. This notebook aims to showcase: (1) which samples have already been sequenced by Hartwell, and (2) what omics data are available per sample.

For demo purposes, we use dummy data cohort and subset by human brain tumor samples. Finally, we investigate the number of samples per cancer_type_brain and Assay.

2 Set up

```
suppressPackageStartupMessages({
  library(tidyverse)
})
```

3 Directories and paths to file Inputs/Outputs

```
attach(params)
## The following object is masked _by_ .GlobalEnv:
##
##
       root_dir
analysis_dir <- file.path(root_dir, "analyses", "data-exploratory-analysis")
input_dir <- file.path(analysis_dir, "input")</pre>
# We will first read in metadata file as we need to define sample_name
metadata_file <- file.path(input_dir, input_file) # metadata input file</pre>
palette_file <- file.path(root_dir, "figures", "palettes", "assay_color_palette.tsv")</pre>
tumor_palette_file <- file.path(root_dir, "figures", "palettes", "tumor_color_palette.tsv")</pre>
# File path to `plots` directory
plots_dir <- file.path(analysis_dir, "plots")</pre>
if (!dir.exists(plots_dir)) {
  dir.create(plots_dir)}
figures_plots_dir <- file.path(plots_dir, "figures")</pre>
if (!dir.exists(figures_plots_dir)) {
  dir.create(figures_plots_dir)}
source(paste0(analysis_dir, "/util/function-create-barplot.R"))
source(paste0(root_dir, "/figures/scripts/theme_plot.R"))
```

4 Read metadata file

We will subset by human brain tumor samples.

```
# Read metadata
df <- read.csv(metadata_file, stringsAsFactors=FALSE)
# Number of samples per cancer_type_brain</pre>
```

4.1 Color palette for plotting

```
# Read color palette
palette_df <- readr::read_tsv(palette_file, guess_max = 100000, show_col_types = FALSE)

# Define and order palette
palette <- palette_df$hex_codes
names(palette) <- palette_df$color_names

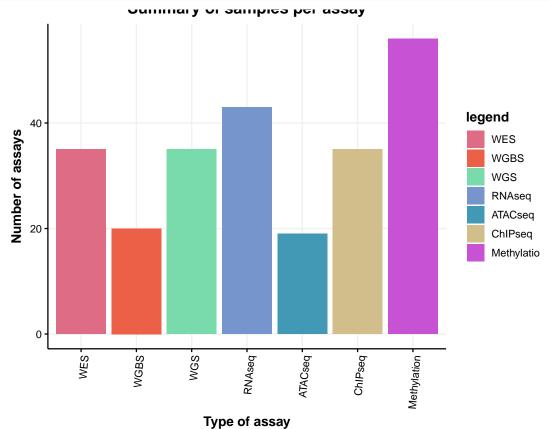
# Read color palette for tumor type
tumor_palette_df <- readr::read_tsv(tumor_palette_file, guess_max = 100000, show_col_types = FALSE)

# Define and order palette
tumor_palette <- tumor_palette_df$hex_codes
names(tumor_palette) <- tumor_palette_df$color_names</pre>
```

5 Number of samples with assay information

Table 1: Summary of samples per assay

Assay	n
WES	35
WGBS	20
WGS	35
RNAseq	43
ATACseq	19
ChIPseq	35
Methylation	56



```
pdf(file = fname, width = 6, height = 5)
print(p)
dev.off()
```

pdf ## 2

6 Number of samples per brain cancer type and assay

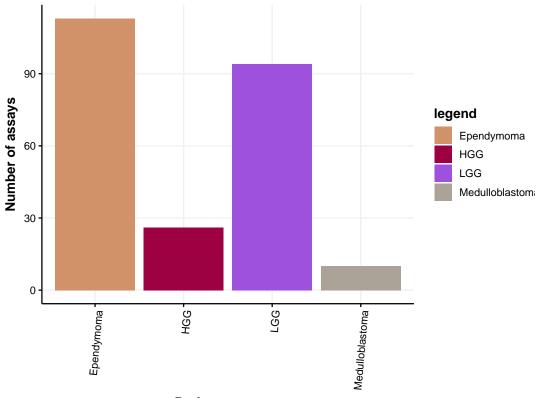
6.1 Overall assays

There are 59 brain tumor samples with 243 assays in total.

Table 2: Summary of samples and assays per brain cancer type

cancer_type_brain	n
Ependymoma	113
HGG	26
LGG	94
Medulloblastoma	10

ounniary or samples and assays per brain cancer type



Brain tumor type

```
pdf(file = fname, width = 6, height = 5)
print(p)
dev.off()
```

pdf ## 2

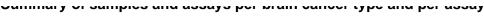
6.2 Per assay

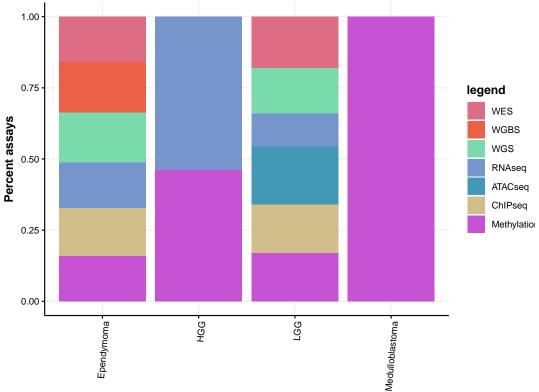
Table 3: Summary of samples and assays per brain cancer type and per assay

cancer_type_brain	WES	WGBS	WGS	RNAseq	ChIPseq	Methylation	ATACseq
Ependymoma	18	20	20	18	19	18	0

cancer_type_brain	WES	WGBS	WGS	RNAseq	ChIPseq	Methylation	ATACseq
HGG	0	0	0	14	0	12	0
LGG	17	0	15	11	16	16	19
Medulloblastoma	0	0	0	0	0	10	0

```
tables1 <- df %>% count(cancer_type_brain, Assay) %>%
  as.data.frame() %>%
  mutate_all(funs(replace_na(.,0)))
## Warning: `funs()` was deprecated in dplyr 0.8.0.
## i Please use a list of either functions or lambdas:
##
## # Simple named list: list(mean = mean, median = median)
##
## # Auto named with `tibble::lst()`: tibble::lst(mean, median)
##
## # Using lambdas list(~ mean(., trim = .2), ~ median(., na.rm = TRUE))
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was generated.
# Plot stacked barplot
fname <- paste0(figures_plots_dir, "/", "cancer-type-brain-per-assay.pdf")</pre>
p <- create_stacked_barplot(plot_df = tables1,</pre>
                             x_value = tables1$cancer_type_brain,
                             use_palette = palette,
                            xtitle ="Brain tumor type",
                             legend = tables1$Assay,
                             title_value = caption_value)
```





Brain tumor type

```
pdf(file = fname, width = 6, height = 5)
print(p)
dev.off()
```

7 Number of samples per brain cancer type, assay, and SJUID

Table 4: Summary of samples and assays per brain cancer type, per assay and per SJUID

can-	CHID			maa	DMA	CI ID	26.1.1.1	ATAC-
cer_type_brain	SJUID V	WES	WGBS	WGS	RNAseq	ChIPseq	Methylation	seq
Ependymoma	SJH0H5WYREP		1	1	0	1	1	0
Ependymoma	SJH2HPPEEKM	[1	1	1	1	1	1	0
Ependymoma	SJH51B396IW	1	1	1	1	1	1	0
Ependymoma	SJH9YML-	1	1	1	1	1	1	0
	FOTI							
Ependymoma	SJHA6KC56J6	1	1	1	1	1	1	0
Ependymoma	SJH-	1	1	1	1	1	1	0
	BKS7QSFO							
Ependymoma	SJHC70DZRJS	1	1	1	1	1	1	0
Ependymoma	SJHCN03RCVD	1	1	1	1	1	1	0
Ependymoma	SJHD1KI-	1	1	1	1	1	1	0
	UMM6							
Ependymoma	SJHFG-	1	1	1	0	1	1	0
	GJRHYO							
Ependymoma	SJH-	1	1	1	1	0	1	0
	HZH67WMF							
Ependymoma	SJHI8CC7QT8	1	1	1	1	1	1	0
Ependymoma	SJHIQT-	1	1	1	1	1	1	0
	NAYIF							
Ependymoma	SJHKVQE-	1	1	1	1	1	0	0
	BOCP							
Ependymoma	SJHKYUIYAME	1	1	1	1	1	0	0
Ependymoma	SJHPVXLA-	1	1	1	1	1	1	0
	CLM							
Ependymoma	SJHUMKP2L6V	1	1	1	1	1	1	0
Ependymoma	SJHXTBTQ5ZT	1	1	1	1	1	1	0
Ependymoma	SJH5HCKPC97	0	1	1	1	1	1	0
Ependymoma	SJH98QNKIUU	0	1	1	1	1	1	0
HGG	SJH5QHZM4US	0	0	0	1	0	1	0
HGG	SJHADL5OJME	0	0	0	1	0	1	0
HGG	SJH-	0	0	0	1	0	1	0
	BISK5KBU							
HGG	SJHBL7CDYRN	0	0	0	1	0	1	0
HGG	SJHD-	0	0	0	1	0	0	0
	DTE0SYL							
HGG	SJHEOVURBM.	J 0	0	0	1	0	1	0
HGG	SJHHPN-	0	0	0	1	0	1	0
	QERSQ							
HGG	SJHHW23UJ9P	0	0	0	1	0	1	0
HGG	SJHJ59RLHSU	0	0	0	1	0	1	0
HGG	SJHKKDDX-	0	0	0	1	0	1	0
	OYH							
HGG	SJHMI643DMD	0	0	0	1	0	1	0

can- cer_type_brain	SJUID V	VES	WGBS	WGS	RNAseq	ChIPseq	Methylation	ATAC- seq
HGG	SJHQBOPS9FD	0	0	0	1	0	0	0
HGG	SJHUT-	0	0	0	1	0	0	0
1100	PISXFQ	O	Ü	Ü	1	Ü	· ·	V
HGG	SJHVIS5Q8HX	0	0	0	1	0	0	0
HGG	SJHEQG3P4FK	0	0	0	0	0	1	0
HGG	SJHRO-	0	0	0	0	0	1	0
	JUQZAP							
LGG	SJH2W47P7DG	1	0	1	1	1	1	1
LGG		1	0	1	1	1	1	1
LGG	SJHBN-	1	0	1	0	1	1	1
	JSZHW6							
LGG	SJHBV3Q6UVR	1	0	1	1	1	1	1
LGG	SJH-	1	0	1	1	1	1	1
	CLGFJTIG							
LGG	SJHD-	1	0	1	0	1	0	0
	DTE0SYL							
LGG	SJHI52BLNWK	1	0	1	1	1	1	1
LGG	SJHN-	1	0	1	0	1	1	1
	PJTQHIT							
LGG	SJHO-	1	0	1	1	1	1	1
	FORRR7C							
LGG	SJHOY05OJJN	1	0	0	1	0	1	1
LGG	SJHQBOPS9FD	1	0	1	0	1	1	1
LGG	SJHUT-	1	0	1	0	1	1	1
	PISXFQ							
LGG	SJHVIS5Q8HX	1	0	1	0	1	1	1
LGG	SJHWS0NRZVA		0	0	1	1	1	1
LGG	SJHXIKCWNKY	1	0	1	1	1	1	1
LGG	SJHYP-	1	0	1	1	1	1	1
	KTG3P5							
LGG	SJHZW7GYEF9		0	1	1	1	1	1
LGG	SJH5HCKPC97	0	0	0	0	0	0	1
LGG	SJHC70DZRJS	0	0	0	0	0	0	1
LGG	SJHIQT-	0	0	0	0	0	0	1
	NAYIF							
Medulloblastoma	SJH77NRD-	0	0	0	0	0	1	0
	WUX							
Medulloblastoma	SJHAF-	0	0	0	0	0	1	0
	TIOMPQ					_		
Medulloblastoma	SJHC1QST5GR	0	0	0	0	0	1	0
Medulloblastoma	SJHF-	0	0	0	0	0	1	0
3.5 1 11 11	CYGKSDY							
Medulloblastoma	SJHJAC-	0	0	0	0	0	1	0
3.5 1 11 11	CGA3S							
Medulloblastoma	SJHQS0D51KH	0	0	0	0	0	1	0
Medulloblastoma	SJHXVMEU21L		0	0	0	0	1	0
Medulloblastoma	SJHY4W1ZWCY		0	0	0	0	1	0
Medulloblastoma	SJHZ40PT-	0	0	0	0	0	1	0
36 1 11 12	СҮН	0			0			^
Medulloblastoma	SJHZZLR-	0	0	0	0	0	1	0
	CGJ6							

8 Future directions

The current exploratory data analysis module can be expanded by investigating samples with paired assays. Moreover, if other metadata are available, e.g., disease_stage, treatment, this will build large, longitudinal cohorts with multi-omic sequencing data. Such an analysis permits consideration of samples according to the condition(s) of the experiment and research aims. In addition, it can be used to refine research questions and/or generate new ones.

This will facilitate collaboration across departments at St. Jude, expedite discoveries, and find cures for children with cancer and other catastrophic diseases.

9 Session Info

```
## R version 4.4.0 (2024-04-24)
## Platform: x86_64-pc-linux-gnu
## Running under: Red Hat Enterprise Linux 8.8 (Ootpa)
##
## Matrix products: default
           /research/rgs01/applications/hpcf/authorized_apps/rhel8_apps/lapack/3.10.1/install/lib64/lib
## LAPACK: /research/rgs01/applications/hpcf/authorized_apps/rhel8_apps/lapack/3.10.1/install/lib64/lib
## locale:
##
  [1] LC_CTYPE=en_US.UTF-8
                                   LC_NUMERIC=C
  [3] LC_TIME=en_US.UTF-8
                                   LC_COLLATE=en_US.UTF-8
   [5] LC_MONETARY=en_US.UTF-8
                                   LC_MESSAGES=en_US.UTF-8
   [7] LC_PAPER=en_US.UTF-8
                                   LC_NAME=C
##
  [9] LC_ADDRESS=C
                                   LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## time zone: America/Chicago
## tzcode source: system (glibc)
## attached base packages:
## [1] grid
                 stats
                           graphics grDevices utils
                                                          datasets methods
## [8] base
## other attached packages:
  [1] ggthemes_5.1.0
                        lubridate_1.9.3 forcats_1.0.0
##
                                                         stringr_1.5.1
   [5] dplyr_1.1.4
                        purrr_1.0.2
                                        readr_2.1.5
                                                         tidyr_1.3.1
## [9] tibble_3.2.1
                        ggplot2_3.5.1
                                        tidyverse_2.0.0
## loaded via a namespace (and not attached):
## [1] sass_0.4.9
                          utf8_1.2.4
                                             generics_0.1.3
                                                               stringi_1.8.4
## [5] hms_1.1.3
                          digest_0.6.37
                                             magrittr_2.0.3
                                                               evaluate_0.24.0
## [9] timechange_0.3.0 fastmap_1.2.0
                                             jsonlite_1.8.8
                                                               tinytex_0.52
## [13] fansi_1.0.6
                          scales_1.3.0
                                             jquerylib_0.1.4
                                                               cli_3.6.3
## [17] rlang_1.1.4
                          crayon_1.5.3
                                             bit64_4.0.5
                                                               munsell_0.5.1
## [21] withr_3.0.1
                                             yaml_2.3.10
                                                               tools_4.4.0
                          cachem_1.1.0
## [25] parallel_4.4.0
                          tzdb_0.4.0
                                             colorspace_2.1-1
                                                               vctrs_0.6.5
## [29] R6_2.5.1
                          mime_0.12
                                             lifecycle_1.0.4
                                                               bit_4.0.5
## [33] vroom_1.6.5
                                             pillar_1.9.0
                          pkgconfig_2.0.3
                                                               bslib_0.8.0
## [37] gtable_0.3.5
                          glue_1.7.0
                                             xfun 0.47
                                                               tidyselect 1.2.1
                                                               htmltools_0.5.8.1
## [41] highr_0.11
                          knitr_1.48
                                             farver_2.1.2
## [45] rmarkdown_2.28
                          labeling_0.4.3
                                             compiler_4.4.0
```