# Network Learning

Our goal is to contruct gene co-expression networks of types WGCNA and Graphical Lasso [from the given gene expression data] and compare them.

```
library(igraph)
##
## Attaching package: 'igraph'
## The following objects are masked from 'package:stats':
##
##
       decompose, spectrum
  The following object is masked from 'package:base':
##
##
       union
library(proxy) # For similarity matrix
##
## Attaching package: 'proxy'
## The following objects are masked from 'package:stats':
##
##
       as.dist, dist
  The following object is masked from 'package:base':
##
##
       as.matrix
e <- read.delim("data/expr_ceph_utah_1000.txt", row.names=1)
head(t(e))[,1:5] # Rows are genes and columns are samples
##
                   GM06986
                              GM12342
                                         GM07016
                                                    GM07017
                                                                GM12146
                -2.7317300 0.6767469 0.1738114 0.3578004 -0.05128586
## X207245_at
## X201215 at
                -0.9004577 -0.9270311 0.6892607
                                                  0.8609533 -1.49116258
                -3.1137361 -3.1137361
## X200602_at
                                      1.0623280 0.2845590
                                                             0.34568798
## X202609_at
                -0.8803472 1.6654487 0.0627374 -1.2088007
                                                             1.01824344
## X203828 s at 0.9423669 0.3734892 -0.3160627 -0.3472103 0.08644669
## X213194 at
                 1.4307468 2.1298571 1.6143684 -0.2839742 -0.57423760
```

#### Weighted Gene Co-Expression Network Analysis (WGCNA)

The process flow for creating a WGCNA is straightforward. Starting with a **gene expression matrix**, we create a **similarity matrix** by carrying out a similarity measure selection between genes i and j. From the similarity matrix, we construct an **adjacency matrix** using a threshold measure tau. We then plot the graph from the adjacency matrix.

We will use absolute pearson correlation as our measure of similarity between genes i and j, that is S(i,j) = |corr(i,j)|. The similarity matrix S is a symmetrix matrix with values in range [0,1] and a unit diagonal.

```
s <- abs(as.matrix(simil(t(e)), FUN=NULL, diag=1)) # Similarity matrix
s[1:5,1:5]</pre>
```

```
## X207245_at X201215_at X200602_at X202609_at X203828_s_at ## X207245_at 1.00000000 0.04304491 0.12104940 0.10970442 0.04130138 ## X201215_at 0.04304491 1.00000000 0.10109481 0.17051288 0.09339149 ## X200602_at 0.12104940 0.10109481 1.00000000 0.06033583 0.13198330 ## X202609_at 0.10970442 0.17051288 0.06033583 1.00000000 0.01280272 ## X203828_s_at 0.04130138 0.09339149 0.13198330 0.01280272 1.00000000
```

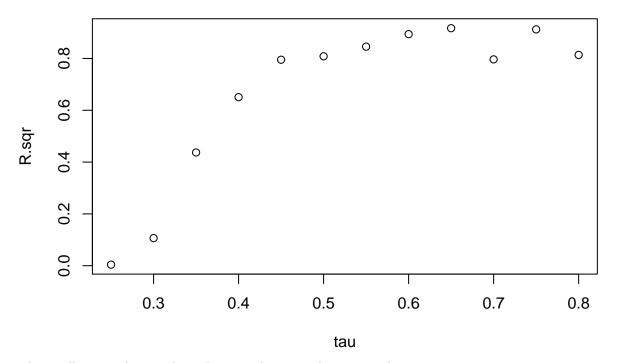
Next, we perform "soft" thresholding via scale-free topology à la Zhang and Horvath 2005; since gene co-expression networks have been found to at least approximately display scale-free topology, so thresholding values that do not give rise to networks displaying scale-free topology should not be considered.

```
sft.power.fit <- function(x) {</pre>
  a \leftarrow s * (s >= x) # Adjacency matrix with threshold x
  g <- graph.adjacency(a, mode="undirected", weighted=TRUE, diag=TRUE) # Construct WGC network
  # Degree of each vertex, not counting self-loops
  d <- degree(g, v=V(g), loops=FALSE)</pre>
  # Continuous distribution for linear model fitting
  deg \leftarrow 1:max(d)
  # Proportion of nodes with degree d
  d.dist <- degree.distribution(g, v=V(g), loops=FALSE, cumulative=FALSE)</pre>
  # Only account for frequencies of non-zero degree vertices
  d.dist <- d.dist[2:length(d.dist)]</pre>
  # Remove all zero frequency positions
  non.zero.positions <- which( d.dist!=0 )</pre>
  deg <- deg[non.zero.positions]</pre>
  d.dist <- d.dist[non.zero.positions]</pre>
  # Linear model fitting
  SFT.model <- lm(log(d.dist) ~ log(deg))
  SFT.R.sqr <- summary(SFT.model)$r.squared</pre>
  SFT.alpha <- -coef(SFT.model)[[2]]</pre>
  return(SFT.R.sqr)
}
tau \leftarrow c(0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.55, 0.6, 0.65, 0.7, 0.75, 0.8)
R.sqr <- sapply(tau, sft.power.fit)</pre>
cbind(tau, R.sqr) # Display tau vs R-squared values
```

```
## tau R.sqr
## [1,] 0.25 0.004106849
## [2,] 0.30 0.106532695
## [3,] 0.35 0.437114635
## [4,] 0.40 0.650661556
## [5,] 0.45 0.794968940
## [6,] 0.50 0.808327761
## [7,] 0.55 0.845574982
```

```
## [8,] 0.60 0.893740743
## [9,] 0.65 0.916673300
## [10,] 0.70 0.796389129
## [11,] 0.75 0.912218243
## [12,] 0.80 0.813450199
```

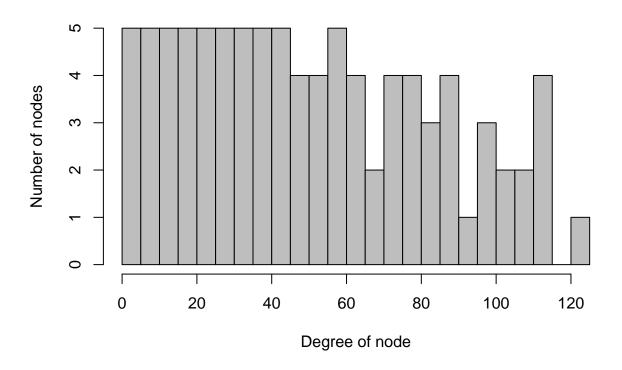
```
plot(x=tau, y=R.sqr) # Scatter plot
```



The smallest tau that produces R-squared greater than or equal to 0.8 is tau = 0.5.

Let us plot the node degree distribution plot for our weighted gene co-expression network with tau = 0.5.

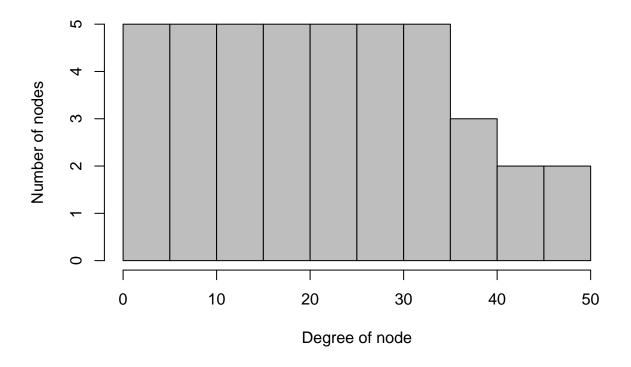
## Node degree distribution for WGCN with tau = 0.5



#### **Graphical Lasso**

```
c <- cov(e) # Sample covariance matrix</pre>
c[1:5,1:5]
##
                X207245_at X201215_at X200602_at X202609_at X203828_s_at
               1.00000000 0.04304491 0.12104940 0.10970442
## X207245_at
                                                                 0.04130138
## X201215_at 0.04304491 1.00000000 0.10109481 -0.17051288
                                                                 0.09339149
## X200602_at
               0.12104940 0.10109481 1.00000000 -0.06033583
                                                                 0.13198330
## X202609_at
              0.10970442 -0.17051288 -0.06033583 1.00000000
                                                                 0.01280272
## X203828_s_at 0.04130138 0.09339149 0.13198330 0.01280272
                                                                 1.00000000
library(glasso)
p <- glasso(c, rho=0.55, thr=1e-4, penalize.diagonal=FALSE) # Precision matrix
colnames(p$wi) <- colnames(e)</pre>
rownames(p$wi) <- colnames(e)</pre>
glasso_net <- graph.adjacency(p$wi, mode="undirected", weighted=TRUE, diag=TRUE) # Construct glasso net
# Histogram plot of node degrees in glasso_net
hist(x=deg, breaks=length(deg)/4, freq=TRUE,
    xlab="Degree of node", ylab="Number of nodes",
    main="Node degree distribution for Graphical Lasso network", col="gray")
```

### Node degree distribution for Graphical Lasso network



#### Comparing WGCNA network and Graphical Lasso network

```
gsize(glasso_net) - ncol(e) # Edge count for glasso_net, discounting self-loops

## [1] 2769

gsize(wgcna_net) - ncol(e) # Edge count for wgcna_net, discounting self-loops

## [1] 7690

# Number of edges common to both networks, discounting self-loops
length(intersection(E(glasso_net), E(wgcna_net))) - ncol(e)
```

## [1] 2769

All edges present in glasso\_net are present in wgcna\_net, that is, glasso\_net is a subset of wgcna\_net. However, the revserse is not true: wgcna\_net has some extra edges.

The proportion of edges present in wgcna\_net also present in glasso\_net is 2769/7690 = 0.360078. The proportion of edges present in glasso\_net also present in wgcna\_net is 2769/2769 = 1.0.

Ideally, we want to separate indirection relationships from direct relationships. Graphical Lasso does so [with the regularization term, encouraging sparsity], but WGCNA does not.