Finding cis-eQTLs

Linear model for eQTL testing

```
library(dplyr)
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
      filter, lag
  The following objects are masked from 'package:base':
##
##
      intersect, setdiff, setequal, union
e <- read.csv("data/geuv_chr22_100genes_expression.csv", row.names=1) # Gene expressions
head(e)[,1:3] # Rows represent genes
                                                HG00099
##
                        HG00096
                                   HG00097
                     ## ENSG00000249263.2
## ENSG00000224688.1
                     4.19482678 3.36943992 2.335469635
## ENSG00000075240.12 3.53180308 3.63554131 1.251434011
## ENSG0000099937.6
                     ## ENSG00000099998.12 0.07363038 0.04110881 0.017493349
## ENSG00000093072.10 13.34694664 13.49828357 2.517851727
start <- read.csv("data/geuv_chr22_100genes_position.csv", row.names=1) # Start site of genes
head(start)
                                  Gene position
##
## ENSG00000249263.2 ENSG00000249263.2 17134782
## ENSG00000224688.1 ENSG00000224688.1 21495913
## ENSG00000075240.12 ENSG00000075240.12 46971909
## ENSG00000099937.6 ENSG00000099937.6 21128167
## ENSG00000099998.12 ENSG00000099998.12 24615622
## ENSG00000093072.10 ENSG00000093072.10 17660194
snp <- read.csv("data/chr22_snp.csv", row.names=1) # SNP genotypes</pre>
head(snp)[,1:3]
           HG00096 HG00097 HG00099
## snp_1590
                0
## snp_1591
                0
                        0
                                1
## snp_1592
                0
                        0
                                0
## snp_1593
                0
                       1
                                1
## snp_1594
                0
## snp_1595
                0
```

```
snp.position <- read.csv("data/chr22_snp_position.csv", row.names=1) # SNP position
head(snp.position)</pre>
```

```
## CHROM POS
## snp_1590 22 17048696
## snp_1591 22 17049115
## snp_1592 22 17049827
## snp_1593 22 17049907
## snp_1594 22 17050088
## snp_1595 22 17050565
```

For each gene, we'll identify which SNPs are within 1000 bp of the gene start position (either upstream or downstream).

```
eqtl <- list()
for (i in 1:nrow(start)) { # For each gene
   tmp <- vector()
   for (j in 1:nrow(snp.position)) { # For each SNP
      if (snp.position[j,2] >= start[i,2]-1000 & snp.position[j,2] <= start[i,2]+1000) {
        tmp <- append(tmp, rownames(snp.position)[j])
      }
   }
   eqtl[[i]] <- tmp
}
head(eqtl) # For each of the 100 genes, a list of SNPs +-1000 bp of the gene's start site</pre>
```

```
## [1] "snp_1828" "snp_1829" "snp_1830" "snp_1831" "snp_1832" "snp_1833"
##
## [[2]]
## logical(0)
##
## [[3]]
## [1] "snp_81122" "snp_81123" "snp_81124" "snp_81125" "snp_81126" "snp_81127"
## [7] "snp_81128" "snp_81129" "snp_81130"
##
## [[4]]
## [1] "snp_12759" "snp_12761" "snp_12762"
##
## [[5]]
## [1] "snp_23834" "snp_23835" "snp_23836"
## [[6]]
## [1] "snp_3373" "snp_3374" "snp_3375" "snp_3376" "snp_3377" "snp_3378"
## [7] "snp_3379" "snp_3380" "snp_3381" "snp_3382" "snp_3383" "snp_3384"
## [13] "snp_3385" "snp_3386" "snp_3387" "snp_3388" "snp_3389" "snp_3390"
```

Now, for each SNP within 1000 bp of gene start position, we're going to check if the SNP is associated with expression of the gene using the linear regression model

$$E_i = \mu + \beta_i G_{ij} + \epsilon_{ij}$$

where E_i is expression of gene in individual i and G_{ij} is genotype of SNP j.

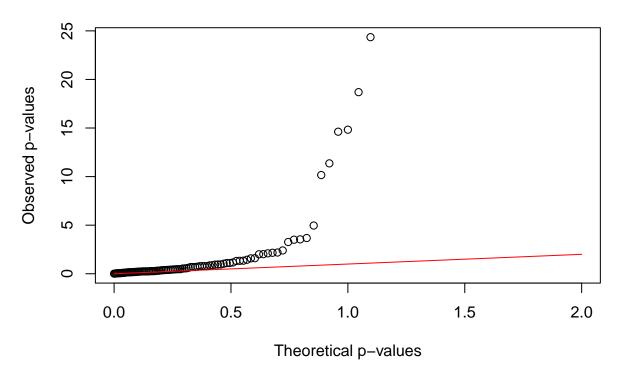
We're going to do this for each gene.

```
# Setup gene expression matrix so that it's an appropriate input for lm()
E <- as.data.frame(t(e)) # Rows now represent individuals</pre>
obs_p <- vector()</pre>
all.M <- list()
for (i in 1:ncol(E)) { # For each gene
  if (length(eqtl[[i]]) != 0) {
    # Prepare genotype matrix for the given gene
    g <- snp[eqtl[[i]],]</pre>
    G <- as.data.frame(t(g))</pre>
    M <- lm(as.matrix(E[,i]) ~ as.matrix(G)) # Apply linear regression
    all.M[[i]] <- M # Save result for each gene
    obs_p <- append(obs_p, anova(M)$'Pr(>F)'[1])
 else {
    all.M[[i]] <- NULL
    obs_p <- append(obs_p, 0)</pre>
}
# anova(M)$'Pr(>F)'[1]
# summary(M)$coefficients[,4]
```

Q-Q plot of p-values

```
# Since these tests can give very small p-values, we transform by taking -log10(p-value)
obs_p = -log10(sort(obs_p))
# Creating theoretical p-values that follow a uniform distribution
th_p = -log10(seq(1/100,1,1/100))
plot(th_p,obs_p, xlab="Theoretical p-values", ylab="Observed p-values", main="Q-Q Plot")
lines(th_p,th_p,col='red')
```

Q-Q Plot



How many genes have at least one SNP with an association p-value less than 0.01?

```
##
         result.genes
                              result.snps result.pvals
##
    [1,] "ENSG00000040608.9"
                              "snp 11154" "0.00306879873900085"
##
    [2,] "ENSG00000131100.8"
                              "snp_4948"
                                          "0.00578227284560896"
    [3,] "ENSG00000128185.5"
                              "snp_11289" "4.42895505988439e-06"
   [4,] "ENSG00000233995.1"
                              "snp 1602"
                                           "0.00094072367419325"
##
##
    [5,] "ENSG00000228274.1"
                              "snp 60228" "0.000837432198472403"
   [6,] "ENSG00000099940.6"
##
                              "snp_12924" "8.3018005042306e-10"
   [7,] "ENSG00000167074.9"
                              "snp_65276" "1.85809411106647e-20"
   [8,] "ENSG00000075234.12" "snp_80092" "2.54963223389296e-15"
   [9,] "ENSG00000128218.7"
                              "snp_22399" "0.000307118325005016"
```