# Name of dataset - Breast cancer analysis

### **About Dataset**

Features are computed from a digitized image of a fine needle aspirate (FNA) of a breast mass. They describe characteristics of the cell nuclei present in the image. n the 3-dimensional space is that described in: [K. P. Bennett and O. L. Mangasarian: "Robust Linear Programming Discrimination of Two Linearly Inseparable Sets", Optimization Methods and Software 1, 1992, 23-34].

This database is also available through the UW CS ftp server: ftp ftp.cs.wisc.edu cd math-prog/cpo-dataset/machine-learn/WDBC/

Also can be found on UCI Machine Learning

Repository: <a href="https://archive.ics.uci.edu/ml/datasets/Breast+Cancer+Wisconsin+%28D">https://archive.ics.uci.edu/ml/datasets/Breast+Cancer+Wisconsin+%28D</a> iagnostic%29

Attribute Information:

1) ID number 2) Diagnosis (M = malignant, B = benign) 3-32)

Ten real-valued features are computed for each cell nucleus:

a) radius (mean of distances from center to points on the perimeter) b) texture (standard deviation of gray-scale values) c) perimeter d) area e) smoothness (local variation in radius lengths) f) compactness (perimeter^2 / area - 1.0) g) concavity (severity of concave portions of the contour) h) concave points (number of concave portions of the contour) i) symmetry j) fractal dimension ("coastline approximation" - 1)

The mean, standard error and "worst" or largest (mean of the three largest values) of these features were computed for each image, resulting in 30 features. For instance, field 3 is Mean Radius, field 13 is Radius SE, field 23 is Worst Radius.

All feature values are recoded with four significant digits.

Missing attribute values: none

Class distribution: 357 benign, 212 malignant

## import liberties

```
library(party)
library(psych)
library(dplyr)
library(data.table)
library(ggplot2)
library(plotly)
library(expss)
library(pander)
library(forcats)
library(stringr)
library(caTools)
library(VIM)
library(caret)
require(reshape2)
library(GGally)
library(corrplot)
library(factoextra)
library(gridExtra)
library(C50)
library(highcharter)
library(rpart)
library(e1071)
library(ranger)
library(epiR)
library(randomForest)
library(party)
library(class)
library(kknn)
library(gbm)
library(ada)
library(c3)
```

# See The Data and Data Type:

```
> head(data)
8.43e5 M
8.43e7 M
8.43e7 M
8.44e7 M
                                                       0.0702
                20.6
                      17.8
                           133.
                                      0.084<u>7</u> 0.078<u>6</u> 0.086<u>9</u>
                                                              0.181 0.0567
                                 1326
                                 <u>1</u>203
                                            0.160
                                                       0.128
                                                  0.197
                                                              0.207
                      21.2
                19.7
                           130
                                      0.110
                           77.6
                                                              0.260 0.0974
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                      20.4
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                      15.7
                           82.6
                                      0.128
                                            0.17
                                                  0.158
                                                       0.0809
                                                              0.209 0.0761
```

<dh7> 0.242 0.0787

0.0600

> The dataset has 33 columns, but one is completely empty, so I removed it...

## **Missing Data**

### > There are no Missing Data

To design a machine learning algorithm that is able to correctly classify whether the tumor is benign or malignant.

## **Target variable**

> Target variable is a character-type variable, so I convert it into a factor...

# **Descriptive analysis**

```
> psych::describeBy(data[3:32], group=data$diagnosis)
    Descriptive statistics by group group: B
                                                                                                                                                                                                                                                                                                                                                                                                                               by group

vars n mean sd median

1 356 12.16 1.77 12.20

2 356 17.90 3.99 17.38

3 356 78.16 11.71 78.22

4 356 463.58 133.64 458.55

5 356 0.09 0.01 0.09

6 356 0.08 0.03 0.02

9 356 0.17 0.02 0.17

10 356 0.05 0.04 0.04

8 356 0.17 0.02 0.17

10 356 0.08 0.01 0.02

11 356 0.28 0.11 0.26

12 356 1.22 0.59 1.11

13 356 2.20 0.77 1.85

14 356 2.00 0.77 0.04

16 356 0.02 0.00 0.01

17 356 0.01 0.00 0.01

18 356 0.02 0.00 0.01

18 356 0.02 0.00 0.02

19 356 1.22 0.00 0.00

20 0.02

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22 356 0.00 0.00 0.00

23 356 0.00 0.00 0.00

24 356 18.39 1.97 13.35

25 356 2.35 0.5 4.8

25 356 87.08 13.47 86.94

25 356 0.13 0.02 0.13

24 356 550.71 62.10 547.60

25 356 0.13 0.02 0.17

77 356 0.18 0.09 0.17

77 356 0.18 0.09 0.17

77 356 0.17 0.14 0.14

28 356 0.07 0.04 0.07

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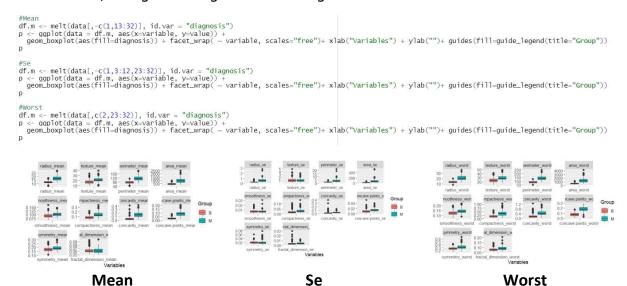
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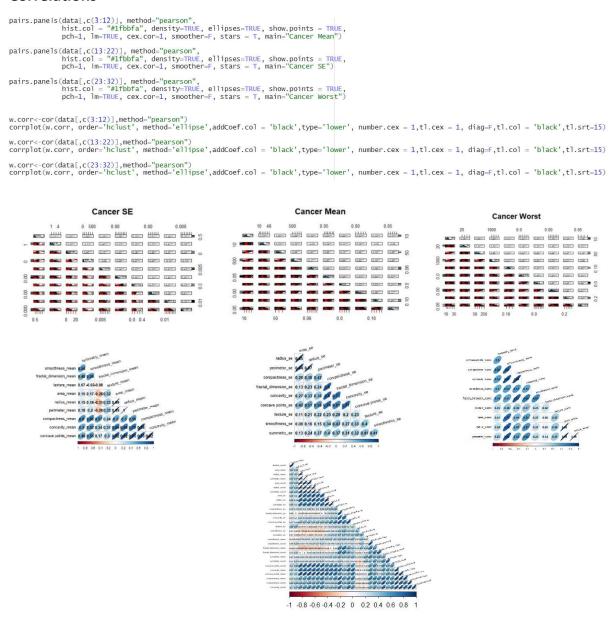
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2.67 61.30
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13.42
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26.68
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3.56
3.56
3.80
0.26
1.06
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2.54
0.73
1.60
1.31
0.08
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3.77
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fractal_dimension_worst 1.34
```

## > In General, Malignant Diagnoses Have Higher Scores in All Variables

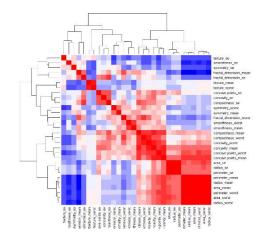


## **Correlations**



> We see that there are extremely high correlations between some of the variables...

## **Heatmap & clustering**



# **Training and testing datasets**

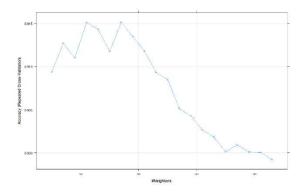
```
dataset=data
head(dataset)
```

# Dataset is divided into two datasets: training (70%) and testing (30%)

```
set.seed(123)
smp_size <- floor(0.70 * nrow(dataset))
train_ind <- sample(seq_len(nrow(dataset)), size = smp_size)
train <- dataset[train_ind, ]
test <- dataset[-train_ind, ]</pre>
```

# Let's check the target variable.

### K-NN



# Testing the model

```
> knnPredict <- predict(knnFit,newdata = test )
> cm_knn
confusion Matrix and Statistics

Reference
Prediction B M
B 97 7
M 1 66

Accuracy : 0.9532
95% CI : (0.9099, 0.9796)
No Information Rate : 0.5731
P-Value [Acc > NIR] : <2e-16
Kappa : 0.9034

Mcnemar's Test P-Value : 0.0771

Sensitivity : 0.9898
Specificity : 0.99041
Pos Pred Value : 0.9327
Neg Pred Value : 0.9327
Neg Pred Value : 0.9327
Neg Pred Value : 0.9351
Prevalence : 0.5731
Detection Prevalence : 0.5673
Detection Prevalence : 0.6062
Balanced Accuracy : 0.9470
'Positive' Class : B
```

# **Naive Bayes**

### **Decision Tree**

```
> #Training the model
> learn_ct <- ctree(diagnosis~., data=train[,-1], controls=ctree_control(maxdepth=2))</pre>
> #Testing the model
> cm ct
Confusion Matrix and Statistics
          Reference
Prediction B M
         B 95 10
         M 3 63
               Accuracy: 0.924
95% CI: (0.8735, 0.9589)
    No Information Rate : 0.5731
P-Value [Acc > NIR] : < 2e-16
                  Kappa : 0.8427
 Mcnemar's Test P-Value: 0.09609
            Sensitivity: 0.9694
            Specificity: 0.8630
         Pos Pred Value: 0.9048
         Neg Pred Value: 0.9545
            Prevalence: 0.5731
  Detection Rate : 0.5556
Detection Prevalence : 0.6140
      Balanced Accuracy : 0.9162
       'Positive' Class : B
```

## K-means

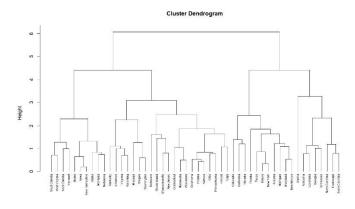
```
> #Training the model
> predict.kmeans <- function(newdata, object){
     centers <- object$centers
     n_centers <- nrow(centers)
     dist_mat <- as.matrix(dist(rbind(centers, newdata)))</pre>
     dist_mat <- dist_mat[-seq(n_centers), seq(n_centers)]</pre>
     max.col(-dist_mat)
+ }
> learn_kmeans <- kmeans(train[,-c(1,2)], centers=2)
> pre_kmeans <- predict.kmeans(test[,-c(1,2)],learn_kmeans)
> pre_kmeans <- factor(ifelse(pre_kmeans == 1,"B","M"))
> cm_kmeans <- confusionMatrix(pre_kmeans, test$diagnosis)</pre>
> cm_kmeans
Confusion Matrix and Statistics
             Reference
Prediction B M
            B 98 21
            M 0 52
                   Accuracy: 0.8772
                      95% CI : (0.8184, 0.9223)
     No Information Rate : 0.5731
P-Value [Acc > NIR] : < 2.2e-16
                       Kappa : 0.7395
 Mcnemar's Test P-Value: 1.275e-05
               Sensitivity: 1.0000
Specificity: 0.7123
            Pos Pred Value: 0.8235
            Neg Pred Value : 1.0000
                Prevalence: 0.5731
            Detection Rate: 0.5731
    Detection Prevalence : 0.6959
Balanced Accuracy : 0.8562
         'Positive' Class : B
```

## **Hierarchical**

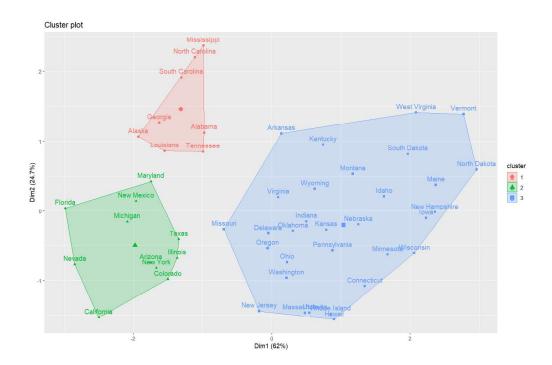
```
# Dissimilarity matrix
d <- dist(df, method = "euclidean")

# Hierarchical clustering using Complete Linkage
hc1 <- hclust(d, method = "complete")

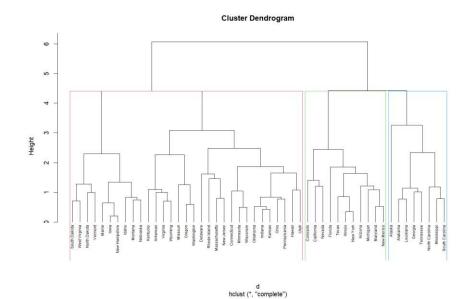
# Plot the obtained dendrogram
plot(hc1, cex = 0.6, hang = -1)</pre>
```



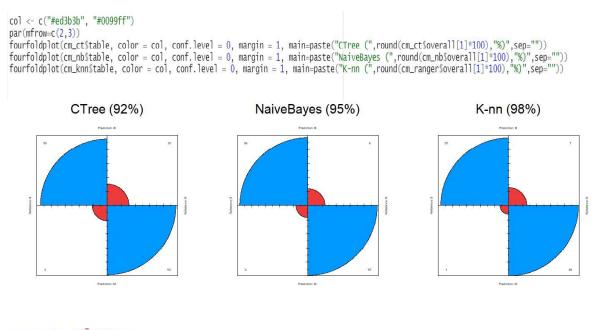
```
# Plot the obtained dendrogram with
# rectangle borders for k clusters
plot(hcl, cex = 0.6, hang = -1)
rect.hclust(hcl, k = 3, border = 2:4)
```



```
# Cut tree into 3 groups
sub_grps <- cutree(hc1, k = 3)
# Visualize the result in a scatter plot
fviz_cluster(list(data = df, cluster = sub_grps))</pre>
```



# **Best ML Model Out Of Them:**



> cm\_knn\$table Reference Prediction B M B 97 7 M 1 66

> out of them, k-nn is best predictor for this dataset... it has 98% accuracy....

#### #CODING PAGE

```
#import liberies
library(party)
library(psych)
library(dplyr)
library(data.table)
library(ggplot2)
library(plotly)
library(expss)
library(pander)
library(forcats)
library(stringr)
library(caTools)
library(VIM)
library(caret)
require(reshape2)
library(GGally)
library(corrplot)
library(factoextra)
library(gridExtra)
library(C50)
library(highcharter)
library(rpart)
library(e1071)
library(ranger)
library(epiR)
library(randomForest)
library(party)
library(class)
library(kknn)
library(gbm)
library(ada)
library(c3)
#The dataset has 33 columns, but one is completely empty, so I removed
it...
head(data)
str(data)
data=data[,-33]
str(data)
head(data)
#To design a machine learning algorithm that is able to correctly classify
whether the tumor is benign or malignant.
#missing data
missing values = data %>% summarize all(funs(sum(is.na(.))/n()))
missing values
aggr(data, prop = FALSE, combined = TRUE, numbers = TRUE, sortVars = TRUE,
sortCombs = TRUE)
#Target variable
table(data$diagnosis)
prop.table(table(data$diagnosis))*100
#Target variable is a character-type variable, so I convert it into a
factor.
```

```
data$diagnosis=factor(data$diagnosis, labels=c('B','M'))
prop.table(table(data$diagnosis))*100
#Descriptive analysis
psych::describeBy(data[3:32], group=data$diagnosis)
'in general, malignant diagnoses have higher scores in all variables.'
#Mean
df.m \leftarrow melt(data[,-c(1,13:32)], id.var = "diagnosis")
p <- ggplot(data = df.m, aes(x=variable, y=value)) +</pre>
  geom boxplot(aes(fill=diagnosis)) + facet wrap( ~ variable,
scales="free") + xlab("Variables") + ylab("") +
guides(fill=guide legend(title="Group"))
#Se
df.m \leftarrow melt(data[,-c(1,3:12,23:32)], id.var = "diagnosis")
p <- ggplot(data = df.m, aes(x=variable, y=value)) +</pre>
  geom boxplot(aes(fill=diagnosis)) + facet wrap( ~ variable,
scales="free") + xlab("Variables") + ylab("") +
guides(fill=guide legend(title="Group"))
#Worst
df.m \leftarrow melt(data[,c(2,23:32)], id.var = "diagnosis")
p <- ggplot(data = df.m, aes(x=variable, y=value)) +</pre>
  geom boxplot(aes(fill=diagnosis)) + facet wrap( ~ variable,
scales="free") + xlab("Variables") + ylab("") +
quides(fill=quide legend(title="Group"))
#Correlations
pairs.panels(data[,c(3:12)], method="pearson",
             hist.col = "#1fbbfa", density=TRUE, ellipses=TRUE, show.points
= TRUE,
             pch=1, lm=TRUE, cex.cor=1, smoother=F, stars = T, main="Cancer
Mean")
pairs.panels(data[,c(13:22)], method="pearson",
             hist.col = "#1fbbfa", density=TRUE, ellipses=TRUE, show.points
= TRUE,
             pch=1, lm=TRUE, cex.cor=1, smoother=F, stars = T, main="Cancer
SE")
pairs.panels(data[,c(23:32)], method="pearson",
             hist.col = "#1fbbfa", density=TRUE, ellipses=TRUE, show.points
= TRUE,
             pch=1, lm=TRUE, cex.cor=1, smoother=F, stars = T, main="Cancer
Worst")
w.corr<-cor(data[,c(3:12)],method="pearson")</pre>
corrplot(w.corr, order='hclust', method='ellipse',addCoef.col =
'black', type='lower', number.cex = 1, tl.cex = 1, diag=F,tl.col =
'black',tl.srt=15)
w.corr<-cor(data[,c(13:22)],method="pearson")</pre>
```

```
corrplot(w.corr, order='hclust', method='ellipse',addCoef.col =
'black', type='lower', number.cex = 1, tl.cex = 1, diag=F,tl.col =
'black',tl.srt=15)
w.corr<-cor(data[,c(23:32)],method="pearson")</pre>
corrplot(w.corr, order='hclust', method='ellipse',addCoef.col =
'black', type='lower', number.cex = 1, tl.cex = 1, diag=F, tl.col =
'black',tl.srt=15)
#We see that there are extremely high correlations between some of the
variables
w.corr<-cor(data[,c(3:32)],method="pearson")</pre>
corrplot(w.corr, order='hclust', method='ellipse',addCoef.col =
'black', type='lower', number.cex = 0.25, tl.cex = 0.25, diag=F,tl.col =
'black',tl.srt=15)
col<-colorRampPalette(c('blue','white','red'))(20)</pre>
heatmap(x=w.corr, col=col,symm=T)
#Training and testing datasets
dataset=data
head(dataset)
#Dataset is divided into two datasets: training (70%) and testing (30%)
set.seed(123)
smp size <- floor(0.70 * nrow(dataset))</pre>
train ind <- sample(seq len(nrow(dataset)), size = smp size)</pre>
train <- dataset[train ind, ]</pre>
test <- dataset[-train ind, ]</pre>
#Let's check the target variable.
prop.table(table(train$diagnosis))*100
prop.table(table(test$diagnosis))*100
#K-nn
control <- trainControl(method='repeatedcv',</pre>
                         number=10,
                         repeats=3)
knnFit <- train(diagnosis ~ ., data = train[,-1], method = "knn", trControl
= control, tuneLength = 20)
plot(knnFit)
#Testing the model
knnPredict <- predict(knnFit,newdata = test )</pre>
cm knn<-confusionMatrix(knnPredict, test$diagnosis )</pre>
cm knn
#Naive Bayes
#Training the model
learn nb <- naiveBayes(train[,-c(1,2)], train$diagnosis)</pre>
#Testing the model
pre_nb <- predict(learn_nb, test[,-c(1,2)])</pre>
cm nb <- confusionMatrix(pre nb, test$diagnosis)</pre>
cm nb
#Classification tree
#Training the model
```

```
learn ct <- ctree(diagnosis~., data=train[,-1],</pre>
controls=ctree control(maxdepth=2))
#Testing the model
pre ct <- predict(learn ct, test[,-c(1,2)])</pre>
        <- confusionMatrix(pre ct, test$diagnosis)</pre>
cm ct
# K-means
#Training the model.
predict.kmeans <- function(newdata, object) {</pre>
  centers <- object$centers
  n centers <- nrow(centers)</pre>
  dist mat <- as.matrix(dist(rbind(centers, newdata)))</pre>
  dist mat <- dist mat[-seq(n centers), seq(n centers)]</pre>
 max.col(-dist mat)
learn kmeans \leftarrow kmeans (train[,-c(1,2)], centers=2)
#Testing the model.
pre kmeans <- predict.kmeans(test[,-c(1,2)],learn kmeans)</pre>
pre_kmeans <- factor(ifelse(pre_kmeans == 1,"B","M"))</pre>
cm kmeans <- confusionMatrix(pre kmeans, test$diagnosis)</pre>
cm kmeans
#Hierarchical
# Dissimilarity matrix
d <- dist(df, method = "euclidean")</pre>
# Hierarchical clustering using Complete Linkage
hc1 <- hclust(d, method = "complete" )
# Plot the obtained dendrogram
plot(hc1, cex = 0.6, hang = -1)
# Cut tree into 3 groups
sub grps \leftarrow cutree (hc1, k = 3)
# Visualize the result in a scatter plot
fviz cluster(list(data = df, cluster = sub grps))
# Plot the obtained dendrogram with
# rectangle borders for k clusters
plot(hcl, cex = 0.6, hang = -1)
rect.hclust(hc1, k = 3, border = 2:4)
col <- c("#ed3b3b", "#0099ff")
par(mfrow=c(2,3))
fourfoldplot(cm ct$table, color = col, conf.level = 0, margin = 1,
main=paste("CTree (",round(cm ct$overall[1]*100),"%)",sep=""))
fourfoldplot(cm nb$table, color = col, conf.level = 0, margin = 1,
main=paste("NaiveBayes (",round(cm_nb$overall[1]*100),"%)",sep=""))
fourfoldplot(cm_knn$table, color = col, conf.level = 0, margin = 1,
main=paste("K-nn (",round(cm ranger$overall[1]*100),"%)",sep=""))
```