The motor programs underlying navigation in *Drosophila* larva

based on *PLoS ONE*, 6:e23180 (2011), with K. Shen, M. Klein, A. Tang, E. Kane, M. Gershow, P. Garrity, and A.D.T. Samuel

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January 9, 2013



Motor programs in Drosophila Iarvae

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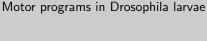
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Introduction

We will look at the motor behaviour of the *Drosophila* larva during navigational motion, paying attention to which segments are used, in which order, etc.

We want to get some insight into the circuits that control this behaviour and the role of sensory feedback by quantifying the motor output at high resolution.





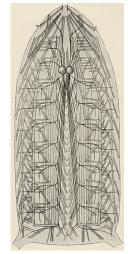
We will look at the motor behaviour of the Drosophila larva during

which order, etc.

navigational motion, paving attention to which segments are used, in We want to get some insight into the circuits that control this behaviour and the role of sensory feedback by quantifying the motor output at high

- 1. Ultimately: trace out full pathway sensory to decision to motor
- 2. future: interfere, now: just look at normal behaviour

Drosophila larva



[Hertweck (1931)]

 $\sim 10^4 \ \text{neurons}.$

Has CNS, spiking neurons,...

Many genetic tools.

Transparent \implies optogenetics.

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Motor programs in Drosophila larvae

└─Drosophila larva



- 1. factor of 10 < adult
- 2. unlike c. elegans
- 3. sequenced genome, $\mathsf{GAL4}/\mathsf{UAS}$ system target cell types

Outline

- Navigation and locomotion
- 2 Imaging and analysis of fluorescent muscles
- Results
- Conclusions and future directions

Imaging and analysis of fluorescent muscles
 Tourists
 Conclusions and future directions

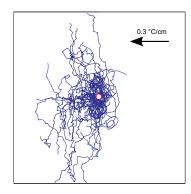
- 1. review how D.larvae navigate, what's known about locomotion circuits
- 2. how larvae with fluorescent muscles will help us, how we use them
- 3. results of this analysis

-Outline

4. conclusions and future directions

Navigation and locomotion

Biased random walks



Alternating runs and reorientations.

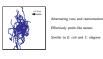
Effectively point-like sensor.

Similar to *E. coli* and *C. elegans*.

Motor programs in Drosophila larvae

Navigation and locomotion

☐Biased random walks



- 1. longer runs in good directions
- 2. has to move sensor to measure gradients
- 3. can do more
- 4. the thing that allows D.larvae to do more...

Head-sweeps



Moves head from side-to-side to sample environment and pick a direction to travel.

Motor programs in Drosophila larvae
Navigation and locomotion
Head-sweeps



- 1. accepted
- 2. rejected

Navigation strategy

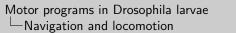
For thermo-/chemo-/photo-taxis, larva modulates:

- head-sweep frequency
- head-sweep size
- head-sweep acceptance probability

Depending on whether conditions are improving/worsening.

[Luo et al. (2010)]





☐Navigation strategy



Navigation strategy

- 1. turns more when things are getting worse
- 2. larger turns when things are getting worse
- 3. more likely to accept when better

Questions

Different types of head-sweep:

- Different circuits?
- When is decision made? With what info?
- Mechano-sensory feedback?

Look for differences in mechanics of different types of head-sweep.



1. difference in initiation \rightarrow makes decision before

Locomotion and sensory feedback

Crawl using peristaltic waves from posterior to anterior that lift and push the body forwards.

Several types of Multidendritic (md) sensory neurons. Repeated in each segment. Possibly used for proprioception.

[Bodmer and Jan (1987), Grueber et al. (2002)]

md neurons are used for locomotion:

ullet Turn off all types o no locomotion

- [Song et al. (2007)]
- Turn off certain subsets → disrupt pattern (toothpasting)

[Hughes and Thomas (2007)]



Motor programs in Drosophila larvae

Navigation and locomotion

Locomotion and sensory feedback

Several types of Multidendritic (md) sensory neurons.

Repeated in each segment. Possibly used for proprioception.

[Biodeser and Jan (1987), Granber et al. (2007)]

md neurons are used for locomotion:

§ Turn off all types. — no locomotion

[Song et al. (2007)]

- 1. We'll see lots of videos of this later.
- 2. each segments waits for posterior segment to contract.
- 3. in future: interfere

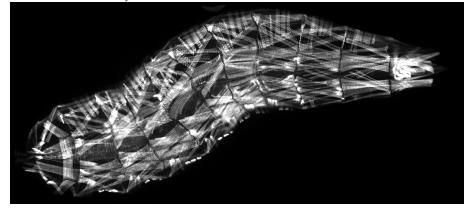
Section 2

Imaging and analysis of fluorescent muscles

Fluorescent muscles

Mutant: w^- ; $\frac{mhc-GFP^{0110}}{CyO}$

[Hughes and Thomas (2007)]



Can see segment boundaries \rightarrow measure length \rightarrow which segment contracts.

Motor programs in Drosophila larvae Imaging and analysis of fluorescent muscles

Fluorescent muscles



- 1. we see 11 segments, some people talk about A9 (terminal, too small), mouth segment (involute during early development)
- 2. can't automate this yet.

Intensity pattern



Muscles contract \rightarrow same GFP in smaller volume \rightarrow increase concentration \rightarrow increase brightness.



Motor programs in Drosophila larvae Limaging and analysis of fluorescent muscles

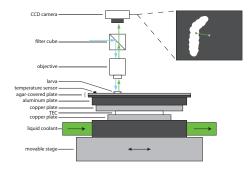
Muscles contact -- same GFP in smaller volume -- increas

Intensity pattern

Intensity pattern

1. another measure of contraction. less noisy

Apparatus



- Temperature varied from $14-16^{\circ}\mathrm{C}$ with period $300\,\mathrm{s}$.
- Movable stage keeps larva in camera frame.

Motor programs in Drosophila larvae

Imaging and analysis of fluorescent muscles

—Apparatus



- 1. triggers many head-sweeps
- 2. allows comparison of head-sweepin warming/cooling



Find boundary, head, tail and bend angle automatically

Motor programs in Drosophila larvae —Imaging and analysis of fluorescent muscles

└─Image analysis



- 1. allows us to flag interesting bits
- 2. slowest part
- 3. automatic again. look for asymmetry
- 4. less noisy



User clicks on segment boundaries

Motor programs in Drosophila larvae Imaging and analysis of fluorescent muscles

☐Image analysis



- 1. allows us to flag interesting bits
- 2. slowest part
- 3. automatic again. look for asymmetry
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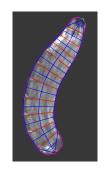


Map to boundary. Find segment lengths.

Motor programs in Drosophila larvae Imaging and analysis of fluorescent muscles

☐Image analysis

- 1. allows us to flag interesting bits
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- 3. automatic again. look for asymmetry
- 4. less noisy



Split segment into quadrants. Mean pixel value \rightarrow intensity.

Motor programs in Drosophila larvae

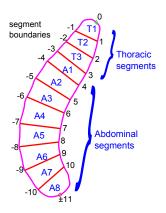
Imaging and analysis of fluorescent muscles

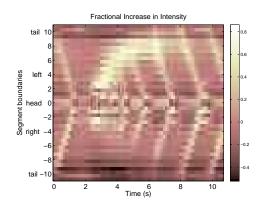
Spite augment into quadrants. Mano pinel value --> intensi

└─Image analysis

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Coordinate system





Motor programs in Drosophila larvae

Imaging and analysis of fluorescent muscles

Coordinate system

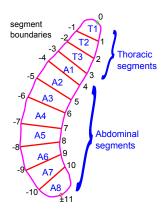


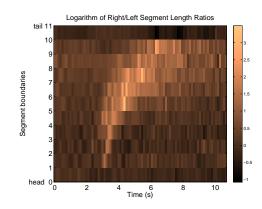
this slide just to explain how to read graphs. Interpret later.

thorax -3 to 3, rest abdomen

- 1. Head in middle, left above, right below. Bright spots: contraction. See peristalsis go from tail to head
- 2. Head at bottom, tail at top. Remove peristalsis, just see bend. Bright: left bend, dark: right bend.

Coordinate system





Motor programs in Drosophila larvae —Imaging and analysis of fluorescent muscles





Coordinate system

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Motor programs in Drosophila Iarvae Results

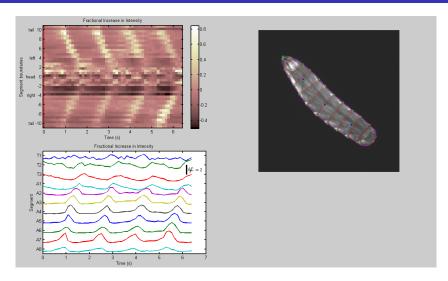


Section 3

Results



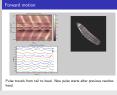
Forward motion



Pulse travels from tail to head. New pulse starts after previous reaches head.

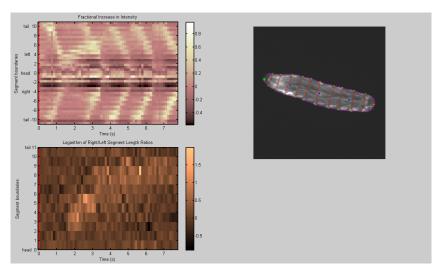
Motor programs in Drosophila larvae Results

Forward motion



- 1. Mouth hooks drown out all else (ratio) in T1,T2.
- 2. If we interfere with sensory feedback, could use this to measure effects.

Small accepted head-sweep

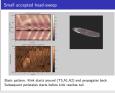


Basic pattern: Kink starts around (T3,A1,A2) and propagates back.

Subsequent peristalsis starts before kink reaches tail.

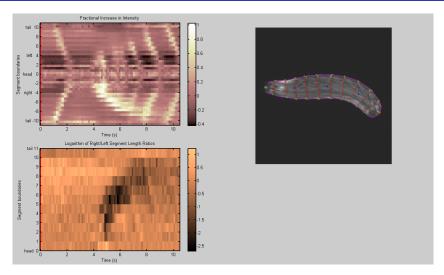
Motor programs in Drosophila larvae
Results

└─Small accepted head-sweep



- 1. Completes head-sweep with peristalsis, not unbending.
- 2. non-overlapping

Large accepted head-sweep



Basic pattern: Kink starts around (T3,A1,A2) and propagates back.

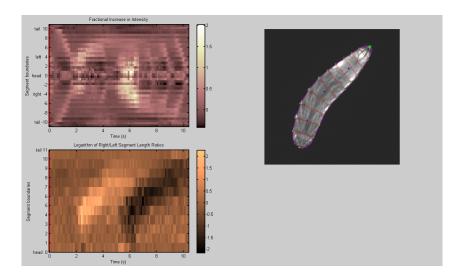
Subsequent peristalsis starts from kink, not tail.

Large accepted head-sweep



1. same as small, statistics later

Rejected head-sweep



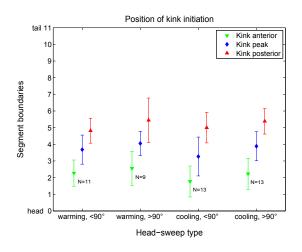
Rejected head-sweep not undone until next one.

Rejected head-sweep



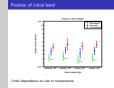
1. no unbending program

Position of initial bend



Little dependence on size or temperature.

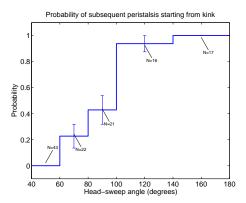




—Position of initial bend

1. error bars ar std dev, not std err.

Position of start of peristalsis



- Transition is around $90 100^{\circ}$.
- Varies from animal to animal.
- Not fully determined by angle.

Subhaneil Lahiri (Harvard)

Motor programs in Drosophila larvae ☐Results





Transition is around 90 - 100^o
Varies from animal to animal.
Not fully determined by angle.

Possible explanations

- Mechanical reason?
 - $> 90^{\circ}$ tail would move wrong way.
 - $< 90^{\circ}$ starting from kink would be slower.
- Neural circuit?
 Stretch-sensors involved in locomotion pattern. If one side is already contracted, segment just anterior to kink might think peristaltic pulse has already reached it
- Central pattern generator?
 Body re-coupling in mid-cycle dependence on head-sweep size?

Motor programs in Drosophila Iarvae
Results

Possible explanations

Possible explanations

Mechanical reason?

- > 90° tail would move wrong way. < 90° starting from kink would be slower.
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Section 4

Conclusions and future directions

Conclusions

All head-sweeps start at the same segments. Same circuits? Decision on size of head-sweep made later?

Navigation results from combining two basic motor programs: peristalsis and asymmetric contraction. Pathway from sensory input \rightarrow motor output simpler than previously thought.

No "unbending" motor program.

Large head-sweeps: subsequent peristalsis starts at kink. Shows that peristalsis can start anywhere. Implications for circuits that control forward motion.

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Motor programs in Drosophila larvae

Conclusions and future directions

└─ Conclusions

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Navigation results from combining two basic motor programs: peristalsis and asymmetric contraction. Pathway from sensory input → motor outp simpler than represently through?

No Sushanding mater program

Large head-sweeps: subsequent peristalsis starts at kink. Shows that peristalsis can start anywhere. Implications for circuits that control

- 1. need to interfere with sensory input during head-sweep optogenetically.
- 2. only need to decide when to switch programs
- 3. can only reject by going other way.
- 4. peristalsis initiator not localised

Future directions

Interfere with motor patterns (optogenetically).

Fully automate image analysis.

Other stimuli.

Reverse crawling, hunching, and rolling.

2013-01-09

Motor programs in Drosophila larvae

Conclusions and future directions

Future directions



- 1. requires next point
- 2. machine learning training data
- 3. we did temperature, could do odour. light difficult. Unlikely to be any difference.
- 4. nociceptive and rapid avoidance responses

Acknowledgements

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Motor programs in Drosophila larvae Conclusions and future directions

Acknowledgements

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And Thank

And Thank

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Anthry Carter

Anthry Carter

Aran Samuel

Gently lab

1. Last slide!

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Subhaneil Lahiri, Konlin Shen, Mason Klein, Anji Tang, Elizabeth Kane, Marc Gershow, Paul Garrity, and Aravinthan D. T. Samuel.

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Motor programs in Drosophila larvae

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Conclusions and future directions

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Motor programs in Drosophila larvae Conclusions and future directions

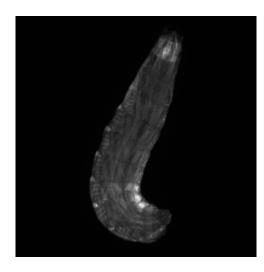
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Toothpasting



Motor programs in Drosophila larvae Conclusions and future directions

Toothpasting

☐ Toothpasting



