

QC Tutorial

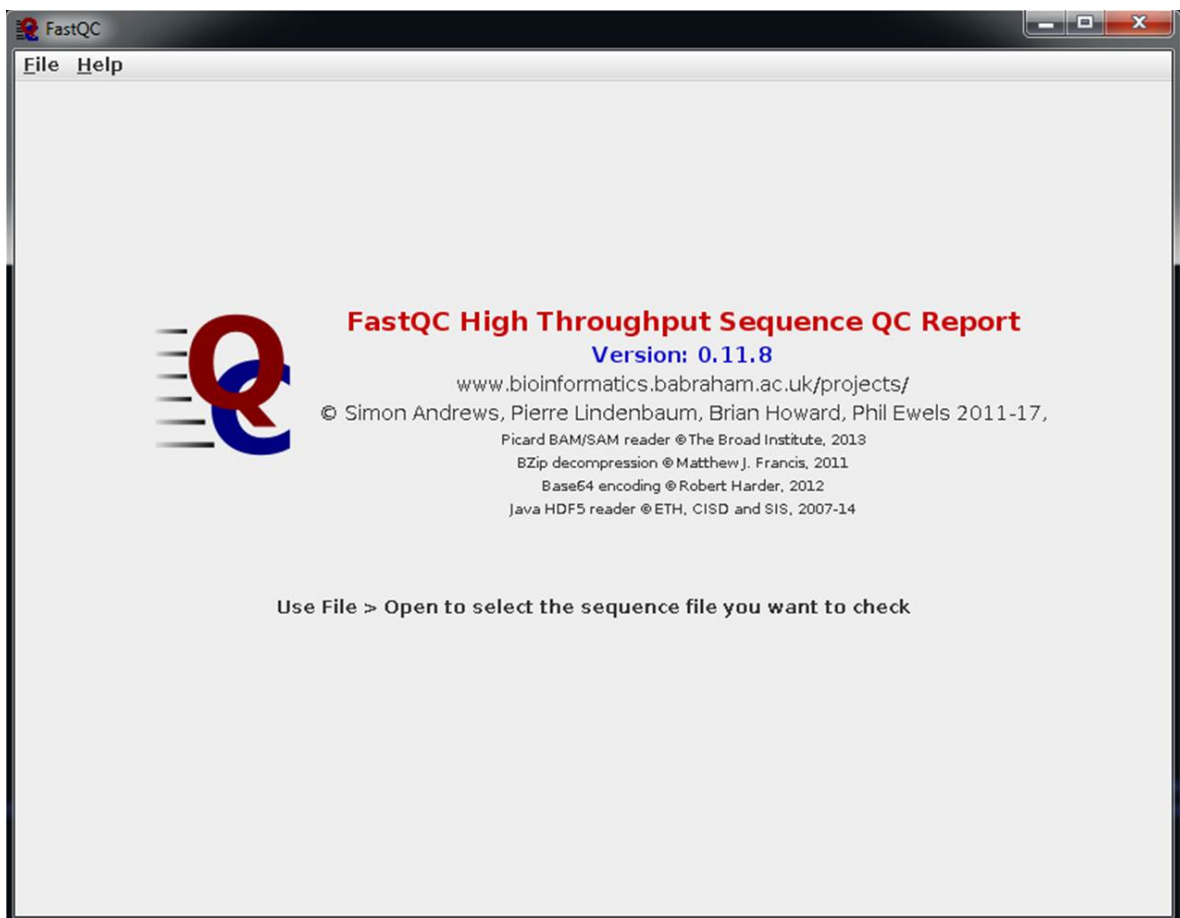
For this tutorial we will work in the folder called **qc_tutorial**

```
$ cd
$ mkdir qc_tutorial
$ cp ~/Course_Data/{PG15-BW001432.R1.fastq.gz,PG15-BW001432.R2.fastq.gz}
~/qc_tutorial
$ cd qc_tutorial
$ ls -lh
```

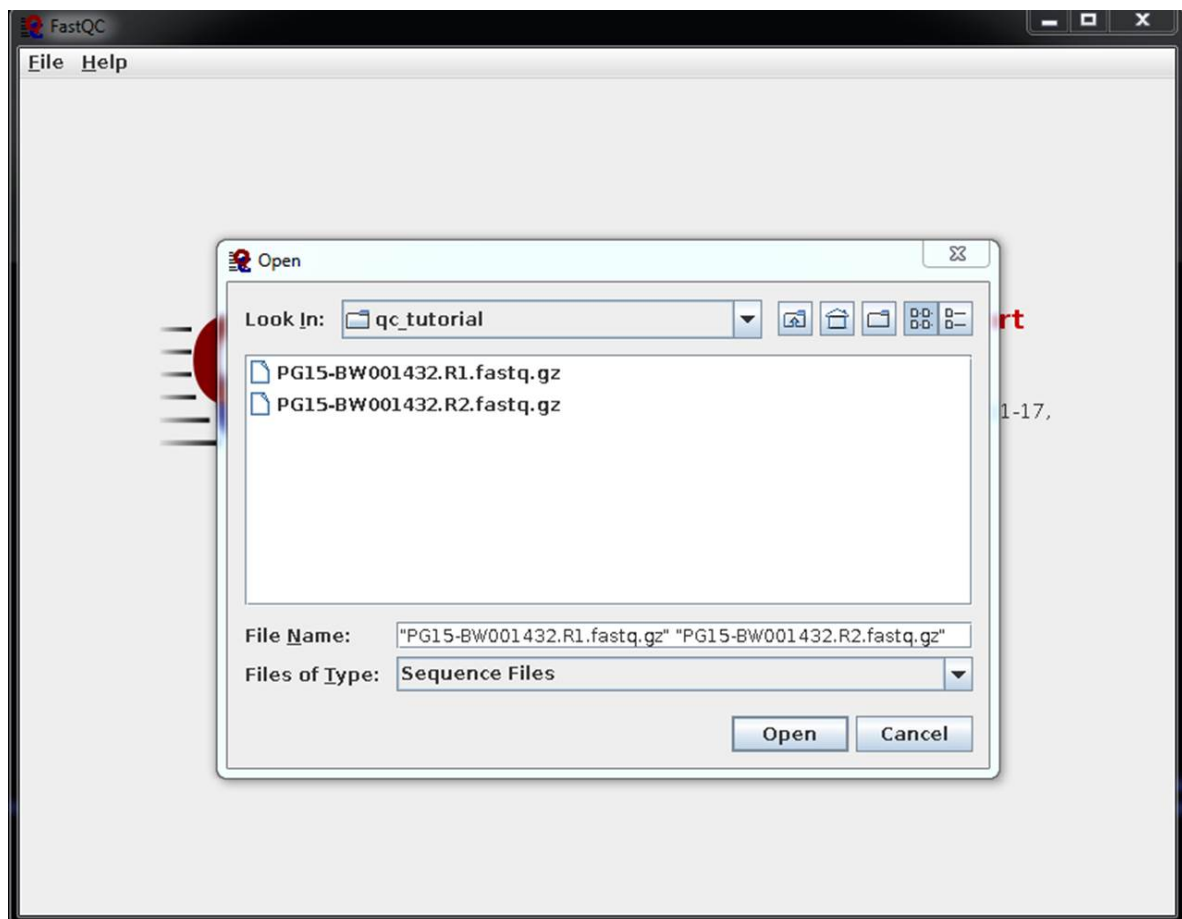
FASTQ Metrics

Opening FastQC

```
$ ~/software/fastqc_v0.11.8/FastQC/fastqc
or
$ fastqc
```



Open both the read files in **qc_tutorial**



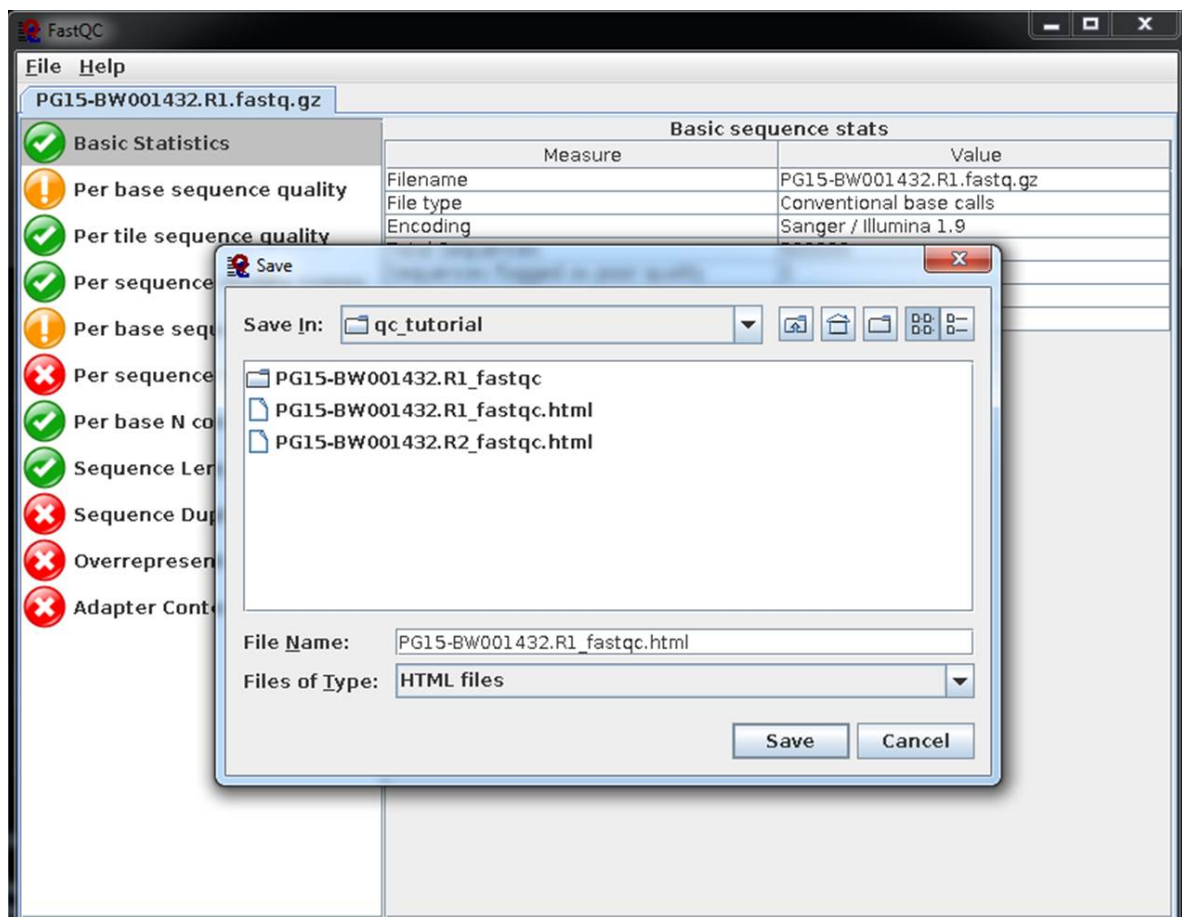
The image shows the FastQC main window with the 'PG15-BW001432.R1.fastq.gz' and 'PG15-BW001432.R2.fastq.gz' tabs selected. The 'Basic Statistics' section is expanded, showing a table of sequence statistics.

Basic sequence stats		
	Measure	Value
	Filename	PG15-BW001432.R1.fastq.gz
	File type	Conventional base calls
	Encoding	Sanger / Illumina 1.9
	Total Sequences	500000
	Sequences flagged as poor quality	0
	Sequence length	250
	%GC	46

The left sidebar shows a list of analysis modules with status icons:

- Basic Statistics (Green checkmark)
- Per base sequence quality (Yellow warning icon)
- Per tile sequence quality (Green checkmark)
- Per sequence quality scores (Green checkmark)
- Per base sequence content (Yellow warning icon)
- Per sequence GC content (Red X icon)
- Per base N content (Green checkmark)
- Sequence Length Distribution (Green checkmark)
- Sequence Duplication Levels (Red X icon)
- Overrepresented sequences (Red X icon)
- Adapter Content (Red X icon)

Save the report in HTML format



FastQC without GUI

FastQC can be invoked from the command line to directly run on a batch of read files in any folder

```
$ mkdir output
$ fastqc -h
$ fastqc *.gz -o output/ --extract
```

-o : output directory

--extract : Unzips the files after creation

All the output files are now in **~/qc_tutorial/output/**

```
$ cd output
$ ls -lh
$ lynx PG15-BW001432.R1_fastqc.html
$ cd PG15-BW001432.R1_fastqc/Images
$ ls -lh
$ eog adapter_content.png
$ cd ~/qc_tutorial
```

Trimming Reads

Running Trim Galore

```
$ ~/software/TrimGalore-0.6.5/trim_galore --help
or
$ trim_galore --help
$ cd ~/qc_tutorial
$ trim_galore
```

Trimming adapters and Low quality bases

```
$ mkdir output_q30
$ trim_galore *.gz -q 30 --phred33 -o output_q30/ --illumina --max_n 2 --paired
$ cd output_q30
$ nano PG15-BW001432.R1.fastq.gz_trimming_report.txt
```

-q : Quality score to use

-o : Output folder

--phred33 : ASCII code used for quality scores

--illumina : Adapters to be trimmed (can specify using **-a** and **-a2**)

--max_n : Total no of 'N' in a read before it is removed

--paired : Reads are paired

- How many Reads contain adapters?
- How many reads passed?
- How many bases were quality trimmed?
- Open the trimmed files in FastQC (*val_1.fq.gz and *val_2.fq.gz)

Increasing and decreasing the stringency of Trimming

```
$ mkdir output_q40
$ trim_galore *.gz -q 40 --phred33 -o output_q40/ --illumina --max_n 2 --paired
$ cd output_q40
$ nano PG15-BW001432.R1.fastq.gz_trimming_report.txt
$ ls -lh
```

- How many Reads contain adapters?
- How many reads passed?
- How many bases were quality trimmed?

```
$ mkdir output_q10
$ trim_galore *.gz -q 10 --phred33 -o output_q10/ --illumina --max_n 2 --paired
$ cd output_q10
$ nano PG15-BW001432.R1.fastq.gz_trimming_report.txt
```

- How many Reads contain adapters?
- How many reads passed?
- How many bases were quality trimmed?
- Open the trimmed files in FastQC

Using wrong adapters for trimming

```
$ mkdir wrong_adapter
$ trim_galore *.gz -q 30 --phred33 -o wrong_adapter/ --nextera --max_n 2 --
paired
$ cd wrong_adapter
$ nano PG15-BW001432.R1.fastq.gz_trimming_report.txt
```

- How many Reads contain adapters?
- Open the trimmed files in FastQC to check adapter content