

# Prepare MA plots for the E(z) mutant experiment

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Input the data and metadata:

```
inpath <- "~/Desktop/brain"
outpath <- "~/Desktop/brain/_7B_Ez_plotMA"
setwd(inpath)
files <- list.files()
htseq_files <- files[grepl("^JKL.*txt$", files)]
sampleNames <- read.csv("EzSampleNames.csv")
sampleTable <- data.frame(fileName = htseq_files,
                           stringsAsFactors=FALSE)
sampleTable$Library <- gsub("-counts.txt", "", sampleTable$fileName)
sampleTable$Library <- gsub("b", "", sampleTable$Library)
sampleTable$seq.batch <- ifelse(grepl("b", sampleTable$fileName), "B", "A")
sampleTable$seq.batch <- paste(sampleTable$Library, sampleTable$seq.batch)
sampleTable <- merge(sampleTable[,c(1,3)], sampleNames, by = "seq.batch")
sampleTable$genotype <- gsub("-.*$", "", sampleTable$Sample)
sampleTable$genotype <- factor(sampleTable$genotype, levels = c("w", "Ez"))
sampleTable$Temp <- gsub("[A-Z, a-z, -]", "", sampleTable$Sample)
sampleTable$Temp <- factor(sampleTable$Temp, levels = c("25", "29"))
sampleTable$rep <- gsub("^.*-", "", sampleTable$Sample)
sampleTable$condition <- paste(sampleTable$genotype, sampleTable$Temp, sep = "-")
sampleTable
```

##	seq.batch	fileName	Sample	batch	Library	genotype	Temp	rep
## 1	JKL10 A	JKL10-counts.txt	w-25-II	A	JKL10	w	25	II
## 2	JKL10 B	JKL10b-counts.txt	w-25-II	B	JKL10	w	25	II
## 3	JKL11 A	JKL11-counts.txt	w-25-III	A	JKL11	w	25	III
## 4	JKL11 B	JKL11b-counts.txt	w-25-III	B	JKL11	w	25	III
## 5	JKL12 A	JKL12-counts.txt	Ez-25-III	A	JKL12	Ez	25	III
## 6	JKL12 B	JKL12b-counts.txt	Ez-25-III	B	JKL12	Ez	25	III
## 7	JKL13 A	JKL13-counts.txt	w-29-I	A	JKL13	w	29	I
## 8	JKL13 B	JKL13b-counts.txt	w-29-I	B	JKL13	w	29	I
## 9	JKL14 A	JKL14-counts.txt	Ez-29-I	A	JKL14	Ez	29	I
## 10	JKL14 B	JKL14b-counts.txt	Ez-29-I	B	JKL14	Ez	29	I
## 11	JKL15 A	JKL15-counts.txt	w-29-II	A	JKL15	w	29	II
## 12	JKL15 B	JKL15b-counts.txt	w-29-II	B	JKL15	w	29	II
## 13	JKL16 A	JKL16-counts.txt	Ez-29-II	A	JKL16	Ez	29	II
## 14	JKL16 B	JKL16b-counts.txt	Ez-29-II	B	JKL16	Ez	29	II
## 15	JKL17 A	JKL17-counts.txt	w-29-III	A	JKL17	w	29	III
## 16	JKL17 B	JKL17b-counts.txt	w-29-III	B	JKL17	w	29	III
## 17	JKL18 A	JKL18-counts.txt	Ez-29-III	A	JKL18	Ez	29	III
## 18	JKL18 B	JKL18b-counts.txt	Ez-29-III	B	JKL18	Ez	29	III
## 19	JKL19 A	JKL19-counts.txt	w-29-IV	A	JKL19	w	29	IV
## 20	JKL19 B	JKL19b-counts.txt	w-29-IV	B	JKL19	w	29	IV
## 21	JKL20 A	JKL20-counts.txt	Ez-29-IV	A	JKL20	Ez	29	IV
## 22	JKL20 B	JKL20b-counts.txt	Ez-29-IV	B	JKL20	Ez	29	IV
## 23	JKL21 A	JKL21-counts.txt	w-29-V	A	JKL21	w	29	V

```
## 24   JKL21 B   JKL21b-counts.txt    w-29-V    B   JKL21      w   29   V
## 25   JKL22 A   JKL22-counts.txt    Ez-29-V    A   JKL22      Ez  29   V
## 26   JKL22 B   JKL22b-counts.txt    Ez-29-V    B   JKL22      Ez  29   V
## 27     JKL7 A     JKL7-counts.txt    w-25-I    A     JKL7      w   25   I
## 28     JKL7 B     JKL7b-counts.txt    w-25-I    B     JKL7      w   25   I
## 29     JKL8 A     JKL8-counts.txt    Ez-25-I    A     JKL8      Ez  25   I
## 30     JKL8 B     JKL8b-counts.txt    Ez-25-I    B     JKL8      Ez  25   I
## 31     JKL9 A     JKL9-counts.txt    Ez-25-II   A     JKL9      Ez  25  II
## 32     JKL9 B     JKL9b-counts.txt    Ez-25-II   B     JKL9      Ez  25  II
##      condition
## 1      w-25
## 2      w-25
## 3      w-25
## 4      w-25
## 5      Ez-25
## 6      Ez-25
## 7      w-29
## 8      w-29
## 9      Ez-29
## 10     Ez-29
## 11     w-29
## 12     w-29
## 13     Ez-29
## 14     Ez-29
## 15     w-29
## 16     w-29
## 17     Ez-29
## 18     Ez-29
## 19     w-29
## 20     w-29
## 21     Ez-29
## 22     Ez-29
## 23     w-29
## 24     w-29
## 25     Ez-29
## 26     Ez-29
## 27     w-25
## 28     w-25
## 29     Ez-25
## 30     Ez-25
## 31     Ez-25
## 32     Ez-25
```

Set up the statistical model:

```
design <- formula(~ Temp + genotype)

dds <- DESeqDataSetFromHTSeqCount(sampleTable = sampleTable, directory = inpath, design = design)
# Combine the technical replicates (different runs) by adding the count
# totals for each gene across the two runs:
dds <- collapseReplicates(dds, groupby=dds$Library, run = dds$batch)
dds <- DESeq(dds)
# What does the data look like?
```

```
head(assay(dds)) # This is the sum of the two runs HTSeq-count output!
```

```
##           JKL10 JKL11 JKL12 JKL13 JKL14 JKL15 JKL16 JKL17 JKL18 JKL19
## FBgn0000003      0      0      0      0      0      0      0      0      0      0
## FBgn0000008  1444  1874  1687  1305  1453  1725  1529  1856  1500  1558
## FBgn0000014      0      0      0      0      0      0      1      0      1      0
## FBgn0000015      1      6      1      3      0      1      1      1      2      0
## FBgn0000017  9186 10798  9189  6877  8808  9121  9834 11313  9272  8564
## FBgn0000018   262   306   286   336   288   317   272   346   274   285
##           JKL20 JKL21 JKL22 JKL7  JKL8  JKL9
## FBgn0000003      1      0      0      0      0      1
## FBgn0000008  1353  1589  1367  1980  1861  1884
## FBgn0000014      1      1      1      0      3      0
## FBgn0000015      0      1      0      0      0      2
## FBgn0000017  8697  8196  8612 11453 11360 11139
## FBgn0000018   285   238   284   286   333   333
```

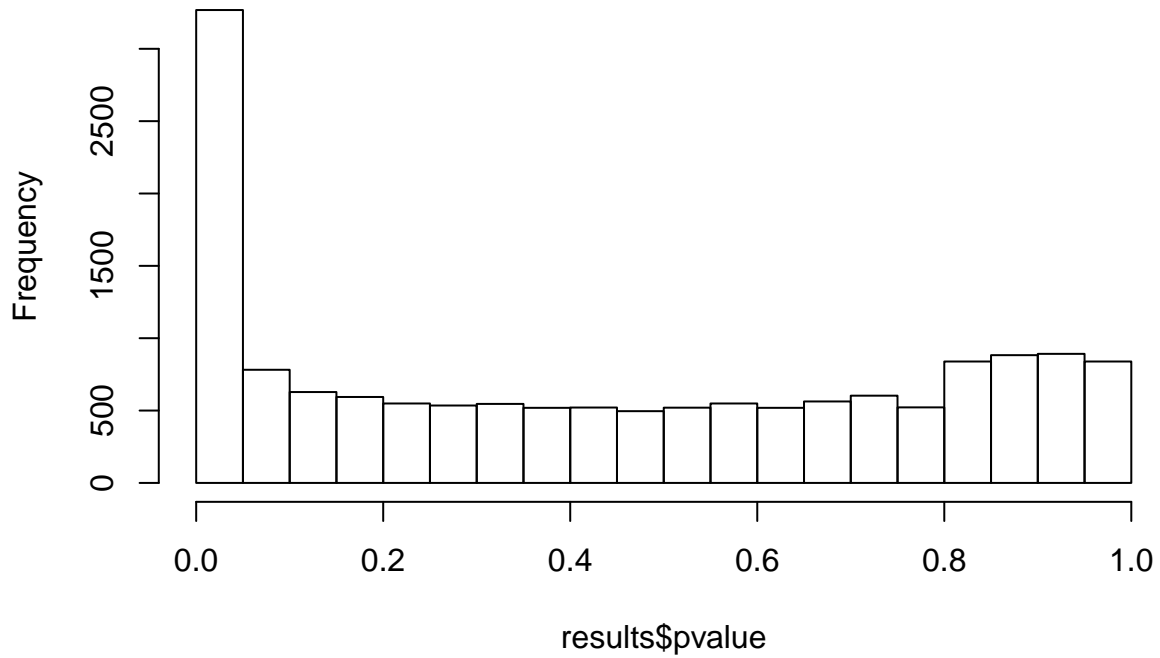
```
# What are the columns?
```

```
colData(dds)
```

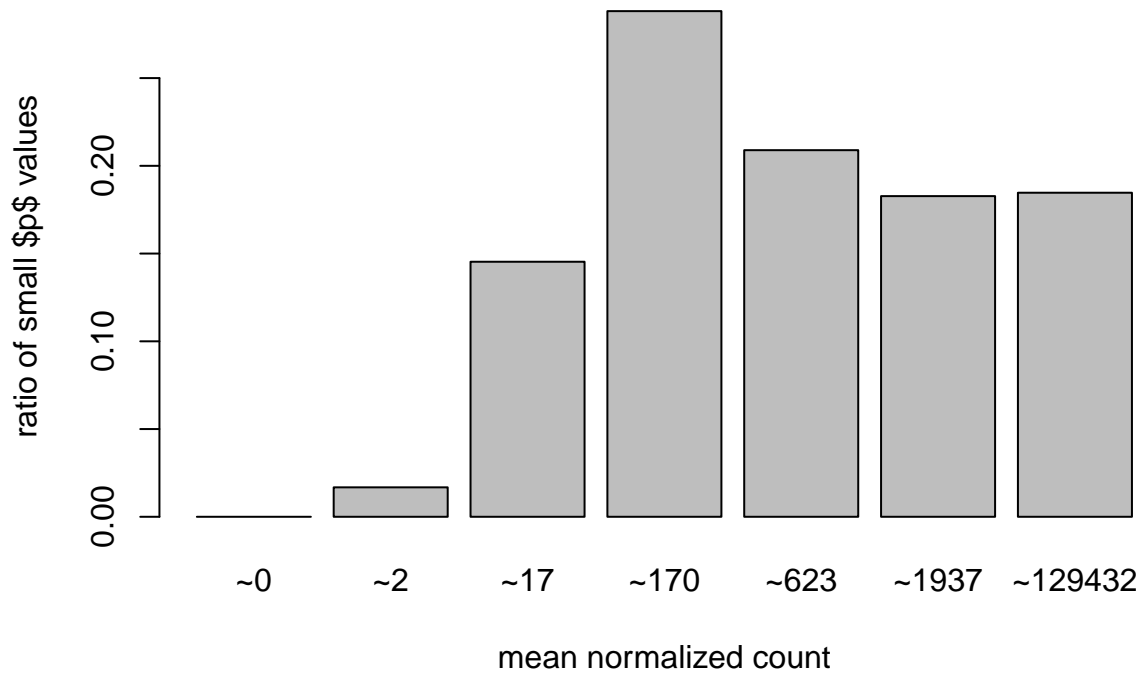
```
## DataFrame with 16 rows and 9 columns
##           Sample  batch  Library genotype  Temp  rep
##           <factor> <factor> <factor> <factor> <factor> <character>
## JKL10    w-25-II      A    JKL10      w      25      II
## JKL11    w-25-III     A    JKL11      w      25      III
## JKL12    Ez-25-III     A    JKL12      Ez      25      III
## JKL13    w-29-I       A    JKL13      w      29      I
## JKL14    Ez-29-I       A    JKL14      Ez      29      I
## ...      ...      ...      ...      ...      ...      ...
## JKL21    w-29-V       A    JKL21      w      29      V
## JKL22    Ez-29-V       A    JKL22      Ez      29      V
## JKL7     w-25-I       A    JKL7       w      25      I
## JKL8     Ez-25-I       A    JKL8      Ez      25      I
## JKL9     Ez-25-II      A    JKL9      Ez      25      II
##           condition runsCollapsed sizeFactor
##           <character> <character> <numeric>
## JKL10      w-25      A,B  0.9090574
## JKL11      w-25      A,B  1.1403424
## JKL12      Ez-25      A,B  1.0027251
## JKL13      w-29      A,B  1.0220507
## JKL14      Ez-29      A,B  0.9570594
## ...      ...      ...      ...
## JKL21      w-29      A,B  0.8750556
## JKL22      Ez-29      A,B  0.9618390
## JKL7       w-25      A,B  1.1341278
## JKL8       Ez-25      A,B  1.1093046
## JKL9       Ez-25      A,B  1.1303920
```

```
results <- results(dds, alpha=0.05)
results$ensembl <- rownames(results)
hist(results$pvalue, breaks=20)
```

## Histogram of results\$pvalue

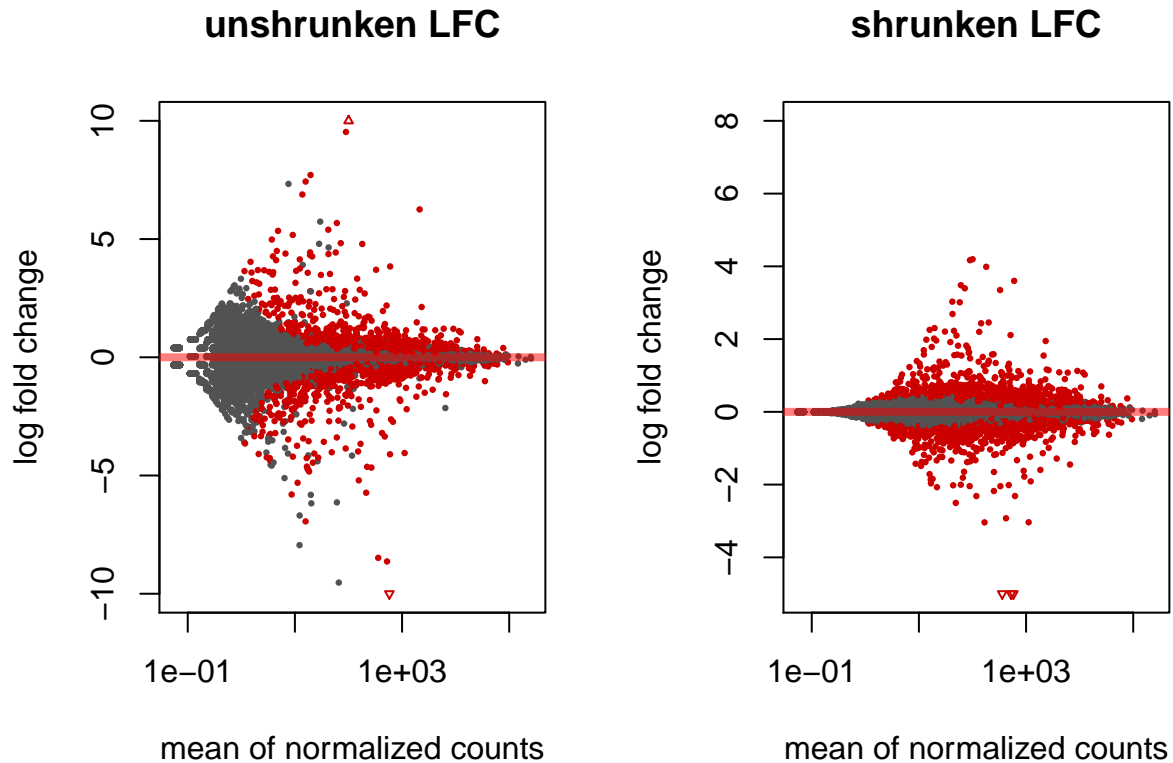


```
qs <- c(0, quantile(results$baseMean[results$baseMean>0], 0:7/7))
bins <- cut(results$baseMean, qs)
levels(bins) <- paste0("~", round(.5*qs[-1] + .5*qs[-length(qs)]))
ratios <- tapply(results$pvalue, bins, function(p) mean(p<.01, na.rm=TRUE))
barplot(ratios, xlab="mean normalized count", ylab="ratio of small $p$ values")
```



Prepare MA plots:

```
# For Maximum likelihood estimates:
resultsMLE <- results(dds, addMLE=TRUE, alpha = 0.05)
par(mfrow=c(1,2))
plotMA(resultsMLE, MLE=TRUE, alpha = 0.05, main="unshrunk LFC", ylim=c(-10,10))
plotMA(results, alpha = 0.05, main="shrunk LFC", ylim=c(-5,8))
```



Add Annotation

```
# Add usefull gene names:
library(biomaRt)

# Get the archived version:
mart = useMart("ENSEMBL_MART_ENSEMBL",
              host="aug2017.archive.ensembl.org")
#x <- listDatasets(mart)
mart = useMart("ENSEMBL_MART_ENSEMBL", host="aug2017.archive.ensembl.org",
              dataset = "dmelanogaster_gene_ensembl")
#listAttributes(mart)
genemap <- getBM(attributes = c("ensembl_gene_id", "entrezgene", "flybasecgid_gene",
                              "external_gene_name"),
                 filters = "ensembl_gene_id",
                 values = results$ensembl,
                 mart = mart)

idx <- match(results$ensembl, genemap$ensembl_gene_id)
results$entrez <- genemap$entrezgene[idx]
```

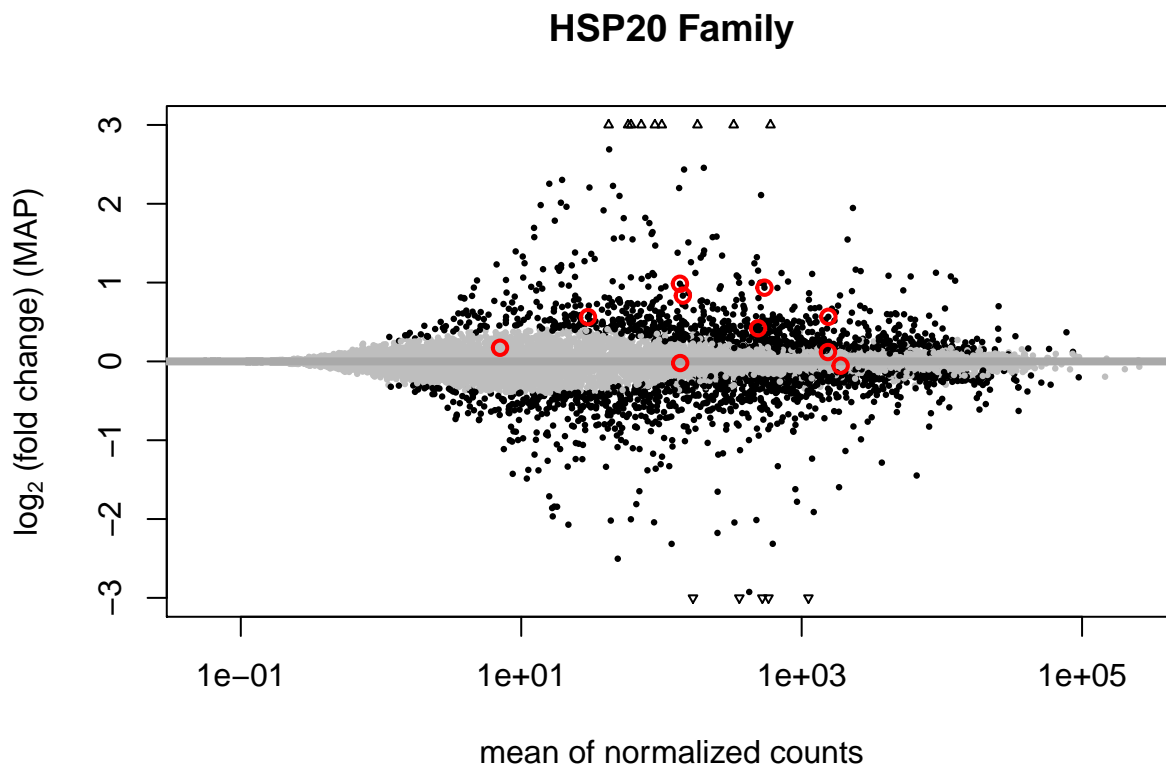
```
results$geneSymbol <- genemap$external_gene_name[idx]
results$cg <- genemap$flybasecgid_gene[idx]
```

## Prepare MA plots of small HSP family

```
small_hsp <- read.delim(file.path(inpath, "FBgg0000507.txt"))
colnames(small_hsp) <- c("FlyBase", "NAME", "SYMBOL")

# Small HSP Family:
jpeg(file=paste(outpath, "Ez_sHSP_MAP.jpg", sep="/"), width=480, height=480)
results$shsp <- results$ensembl %in% small_hsp$FlyBase
plotMA(results, alpha = 0.05, ylim=c(-3,3), colNonSig="gray", colSig="black", colLine="darkgrey",
        ylab=expression("log"[2]*" (fold change) (MAP)"), main="HSP20 Family")
with(results[results$shsp==TRUE,], {points(baseMean, log2FoldChange, col = "red", cex=1, lwd=2)})
dev.off()

## pdf
## 2
plotMA(results, alpha = 0.05, ylim=c(-3,3), colNonSig="gray", colSig="black", colLine="darkgrey",
        ylab=expression("log"[2]*" (fold change) (MAP)"), main="HSP20 Family")
with(results[results$shsp==TRUE,], {points(baseMean, log2FoldChange, col = "red", cex=1, lwd=2)})
```



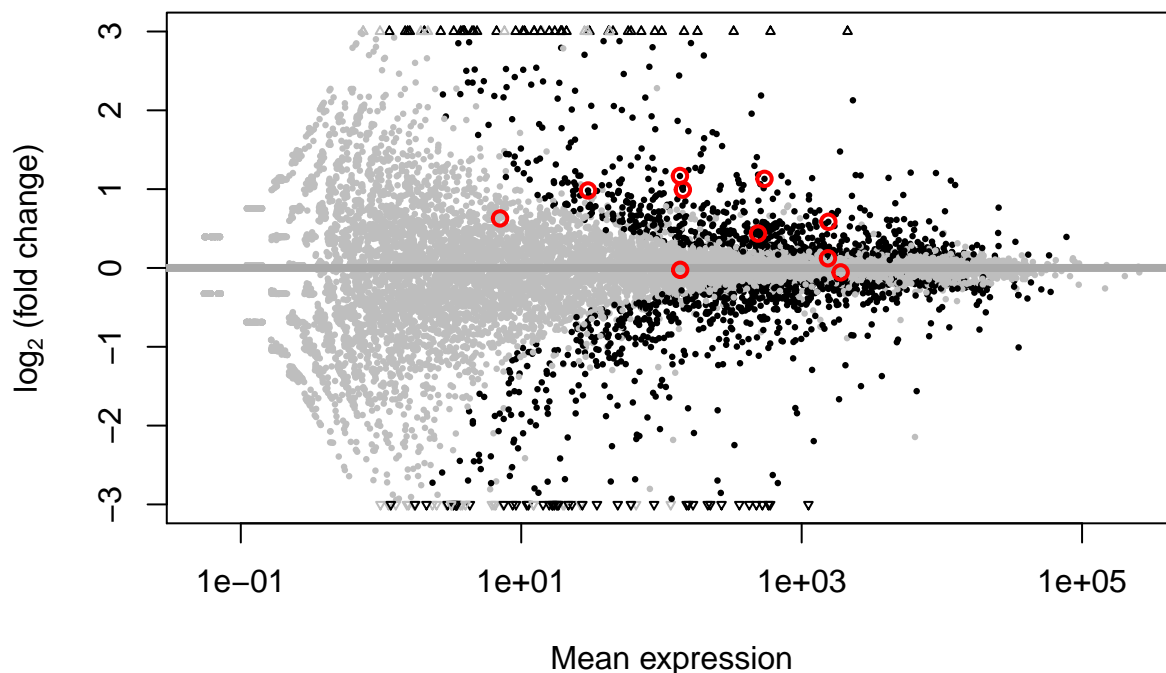
## Prepare MA plots using MLE:

```
# Small HSP Family
#####Prepare Publication Plot#####
jpeg(file=paste(outpath, "Fig7b_Ez_sHSP_MLE.jpg", sep="/"),
      quality=100,
      res=300,
      width=960,
      height=960)
resultsMLE$shsp <- rownames(resultsMLE) %in% small_hsp$FlyBase
par(mai=c(0,0,0,0), mar=c(2,4.5,3,1)+.5)
plotMA(resultsMLE, alpha = 0.05, ylim=c(-3,3), colNonSig = "gray", colSig = "black",
        MLE=TRUE, colLine = "darkgrey",
        las=1, cex.lab=1, cex.axis=1, cex.main=1.5,
        ylab="", xlab = "", xaxp=c(1,2,1), xaxt="n",
        main="")
axis(1, at=c(1,10, 100, 1000, 10000), labels=c(1,10, 100, 1000, 10000), cex.axis=1)
with(resultsMLE[resultsMLE$shsp==TRUE,], {points(baseMean, lfcMLE, col = "red", cex=1, lwd=3)})
dev.off()

## pdf
## 2

#####
plotMA(resultsMLE, alpha = 0.01, ylim=c(-3,3), colNonSig = "gray", colSig = "black",
        MLE=TRUE, colLine = "darkgrey",
        ylab=expression("log"[2]*" (fold change)"), xlab = "Mean expression",
        main="HSP20 Family is \nupregulated in E(z) mutant brains")
with(resultsMLE[resultsMLE$shsp==TRUE,], {points(baseMean, lfcMLE, col = "red", cex=1, lwd=2)})
```

### HSP20 Family is upregulated in E(z) mutant brains



```
Sys.info()
```

```
##                                     sysname
##                                     "Darwin"
##                                     release
##                                     "15.6.0"
##                                     version
## "Darwin Kernel Version 15.6.0: Thu Jun 21 20:07:40 PDT 2018; root:xnu-3248.73.11~1/RELEASE_X86_64"
##                                     nodename
##                                     "Jasons-MacBook-Pro.local"
##                                     machine
##                                     "x86_64"
##                                     login
##                                     "jasonkennerdell"
##                                     user
##                                     "jasonkennerdell"
##                                     effective_user
##                                     "jasonkennerdell"
```