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Two tandem and independent sub-activation domains in the amino terminus of p53 require the adaptor complex for activity

Reyes Candau, Daniel M Scolnick, Paula Darpino, Carol Y Ying, Thanos D Halazonetis and Shelley L Berger

The Wistar Institute, Philadelphia, Pennsylvania 19104, USA

The ability of p53 to function as a tumor suppressor is linked to its function as a transcriptional activator, since p53 mutants that do not transactivate are unable to suppress tumor cell growth. Previous studies identified an activation domain in the amino terminal 40 residues of the protein, a region that binds to several general transcription factors and to some oncogene products. For example, mdm-2, a cellular oncoprotein, binds to this region and represses p53 transactivation. Here we describe a new activation domain within the amino terminus of p53 that maps between amino acids 40-83, and whose residues trp-53 and phe-54 are critical for function both in yeast and in mammalian cells. *In vitro* studies in yeast show that the new activation subdomain, unlike the previously described, is mdm-2 independent. Both p53 activation subdomains (1-40 and 40-83) require the yeast adaptor complex ADA2/ADA3/GCN5 for transcriptional activation. Moreover, since activation by p53 requires GCN5's enzymatic histone acetyltransferase domain, p53 may regulate gene expression by influencing chromatin modification.

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Introduction

The tumor suppressor protein p53 (for a review, see Donehower and Bradley, 1993; Ko and Prives, 1996; Levine, 1993 and references therein) is inactivated in more than half of all human tumors (Greenblatt *et al.*, 1994). p53 is a sequence-specific transcription factor that suppresses oncogenic transformation (Eliyahou *et al.*, 1989; Finlay *et al.*, 1989) and induces cell cycle arrest (Leonardo *et al.*, 1994) or programmed cell death (Clarke *et al.*, 1993; Lowe *et al.*, 1993a,b) in response to DNA damage.

Three distinct domains have been identified within p53: the acidic aminoterminal is an activation domain (Fields and Jang, 1990; Funk *et al.*, 1992; Raycroft *et al.*, 1990); the hydrophobic central region contains a sequence-specific DNA-binding domain (Bargonetti *et al.*, 1993; Halazonetis *et al.*, 1993; Wang *et al.*, 1993); and the basic carboxyterminus comprises an oligomerization domain (Clare *et al.*, 1994; Jeffrey *et al.*, 1995; Lee *et al.*, 1994; Sakamoto *et al.*, 1994; Sturzbecher *et al.*, 1992) and a region that regulates DNA binding affinity (Halazonetis and Kandi, 1993; Hupp *et al.*,

1992; Waterman *et al.*, 1995). Hot-spots for p53 mutations in tumors (Hollstein *et al.*, 1991; Levine *et al.*, 1991) are found predominantly in the central DNA-binding domain.

In addition, p53 can act as a transcriptional repressor, to down-regulate promoters lacking wild type p53 binding sites (Ginsberg *et al.*, 1991; Santhanam *et al.*, 1991; Subler *et al.*, 1992). The amino terminal transactivation domain of p53 also is required for its inhibitory effects on transcription (Sang *et al.*, 1994; Subler *et al.*, 1994).

Tumor suppression and transcriptional activation are strongly correlated functions of p53 (Farmer *et al.*, 1992; Fields and Jang, 1990; Funk *et al.*, 1992). Mutations or deletions in the aminoterminal of p53 coordinately abolish transactivation and tumor suppressor functions. Tumor-derived p53 mutants fail to activate transcription of wild type p53 responsive genes. Furthermore, mdm-2, a cellular oncoprotein which is overexpressed in 30-40% of human sarcomas (Oliner *et al.*, 1992), owes its oncogenic ability to its ability to conceal the transactivation domain of p53 (Brown *et al.*, 1993; Momand *et al.*, 1992; Oliner *et al.*, 1992). Biochemical (Lin *et al.*, 1994) and crystallographic analyses (Kussie *et al.*, 1996) of mdm-2 p53 binding show that similar residues in p53 are critical for both mdm-2 binding and transactivation (Lin *et al.*, 1994).

p53 transactivation may operate through direct contacts with components of the general transcription machinery. For example, interactions have been demonstrated between the p53 amino terminus and TATA-binding protein (TBP) (Horikoshi *et al.*, 1995; Liu *et al.*, 1993), TFIIB (the p62) (Xiao *et al.*, 1994), ERCC1 and ERCC3 (Wang *et al.*, 1995) components, as well as the coactivators TBP associated factors (including *Drosophila* TAFII40, TAFII60 (Thut *et al.*, 1995) and human TAFII31 (Lu and Levine, 1995). It is unclear whether these interactions are sufficient for transactivation *in vivo*, or whether additional factors are required.

Although the activation domain of p53 has been mapped to residues 1-40 (Linger *et al.*, 1992), adjacent sequences contribute to overall p53 activity (Chang *et al.*, 1995). The region encompassing residues 1-90 of the protein is remarkably acidic and its primary structure is generally similar to other activation domains. For example, the potent activator of herpes simplex virus, VP16, contains an activation domain between residues 413-490, and, like p53, has overall negative charge with bulky hydrophobic amino acids grouped in clusters. The negative charge contributes to overall potency, but the hydrophobic residues are essential for transactivation (Cress and Triezenberg

Correspondence: S.L. Berger.
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