

class 10: Structural Bioinformatics pt1

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#PDB statistics

```
stats <- read.csv("Data Export Summary.csv", row.names=1)
head(stats)
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	158,844	11,759	12,296	197	73	32
Protein/Oligosaccharide	9,260	2,054	34	8	1	0
Protein/NA	8,307	3,667	284	7	0	0
Nucleic acid (only)	2,730	113	1,467	13	3	1
Other	164	9	32	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	183,201					
Protein/Oligosaccharide	11,357					
Protein/NA	12,265					
Nucleic acid (only)	4,327					
Other	205					
Oligosaccharide (only)	22					

In uniprot there are 251600,768 proteins.

```
round(183201/251600768*100, 2)
```

```
[1] 0.07
```

```
rm.comma <- function(x){
  as.numeric( gsub(",", "", x))
}
```

```
pdbstats <- apply(stats, 2, rm.comma)
```

Will add the rownames from the original table

```
rownames(pdbstats) <- rownames(stats)
pdbstats
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	158844	11759	12296	197	73	32
Protein/Oligosaccharide	9260	2054	34	8	1	0
Protein/NA	8307	3667	284	7	0	0
Nucleic acid (only)	2730	113	1467	13	3	1
Other	164	9	32	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
	Total					
Protein (only)	183201					
Protein/Oligosaccharide	11357					
Protein/NA	12265					
Nucleic acid (only)	4327					
Other	205					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.
93.16%

```
totals <- apply(pdbstats, 2, sum)
round(totals/totals["Total"] * 100, 2)
```

X.ray	EM	NMR	Multiple.methods
84.83	8.33	6.68	0.11
Neutron	Other	Total	
0.04	0.02	100.00	

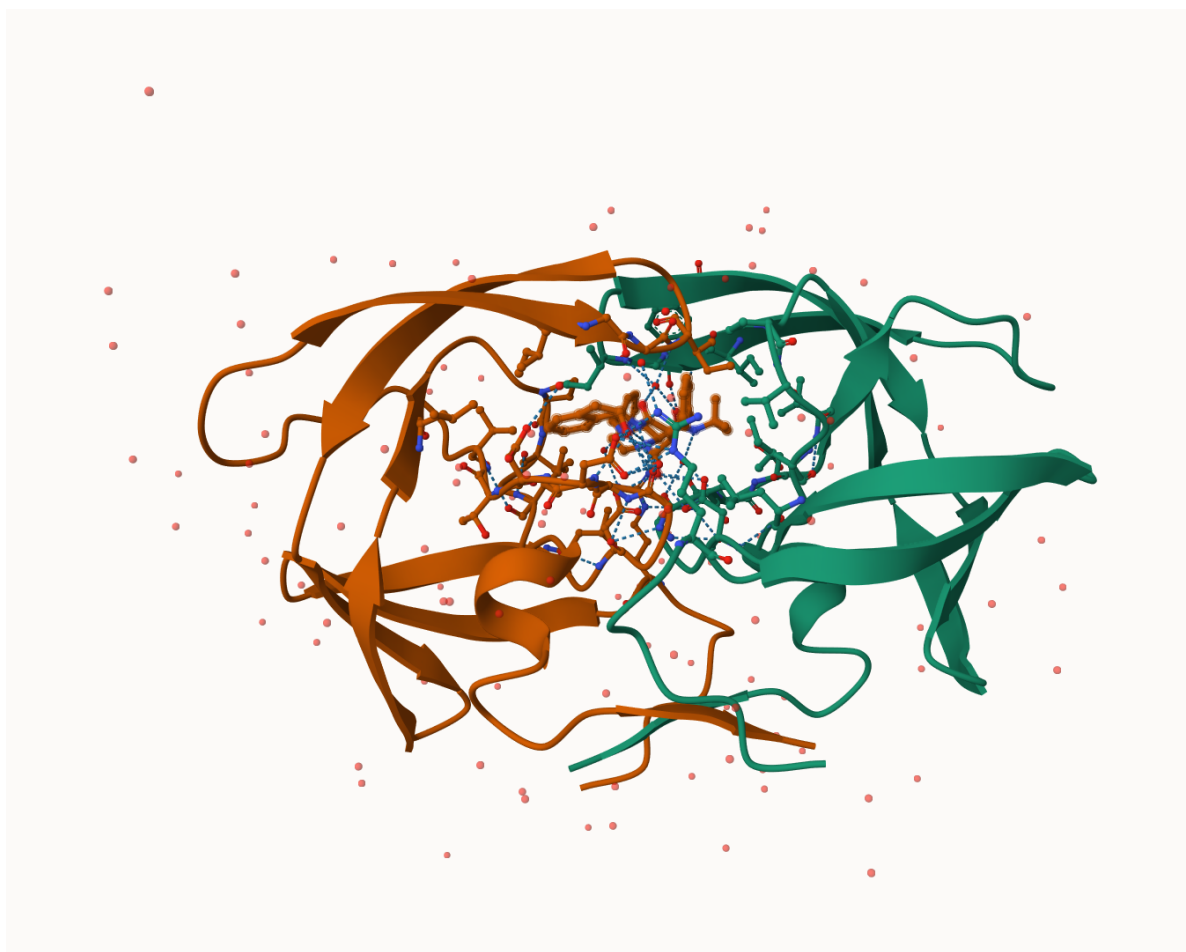
Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure? The resolution is too low 2Å

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

Using Mol* to examine HIV-pr



And a nicer pic colored by secondary structure with catalytic active site ASP 25 shown in each chain

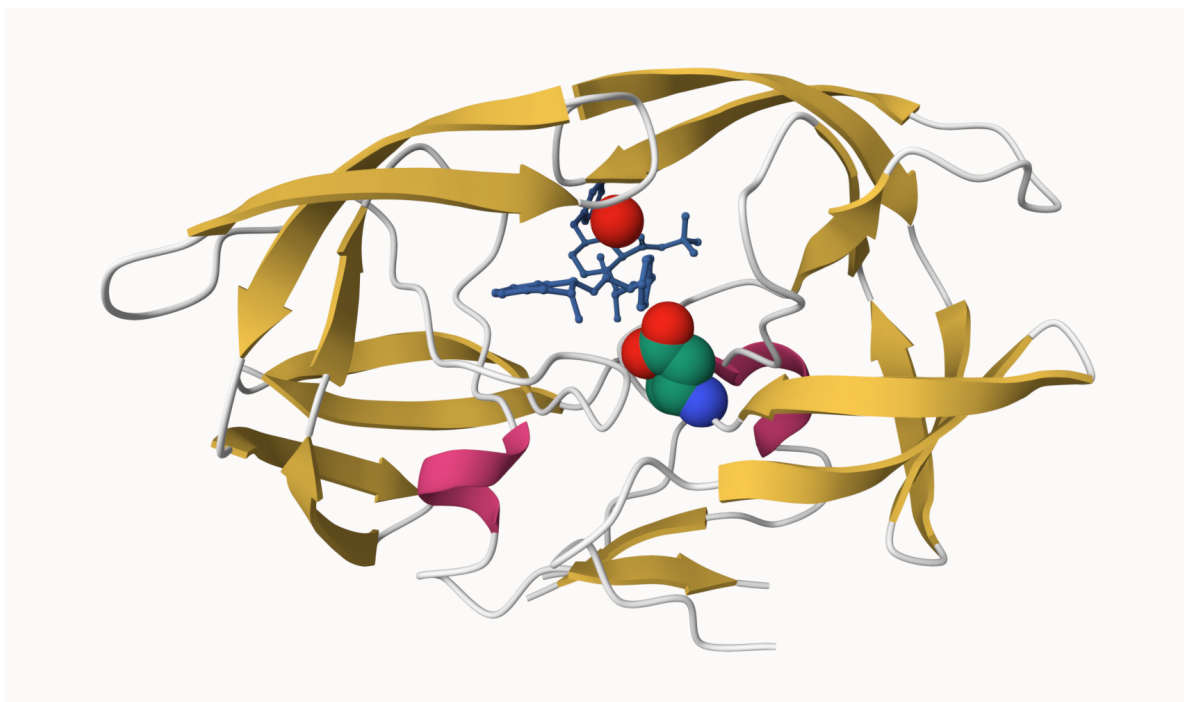


Figure 1: A lovely image

Using the bio3d package

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

`attributes(pdb)`

\$names

[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"

\$class

[1] "pdb" "sse"

`head(pdb$atom)`

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
head(pdb$atom$resid)
```

```
[1] "PRO" "PRO" "PRO" "PRO" "PRO" "PRO"
```

```
aa321( pdb$atom$resid[ pdb$calpha ] )
```

```
[1] "P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q"
[19] "L" "K" "E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M"
[37] "S" "L" "P" "G" "R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I"
[55] "K" "V" "R" "Q" "Y" "D" "Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I"
[73] "G" "T" "V" "L" "V" "G" "P" "T" "P" "V" "N" "I" "I" "G" "R" "N" "L" "L"
[91] "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P" "Q" "I" "T" "L" "W" "Q" "R" "P"
[109] "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E" "A" "L" "L" "D" "T" "G"
[127] "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R" "W" "K" "P" "K"
[145] "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q" "I" "L"
[163] "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
[181] "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"
```

#Q7: How many amino acid residues are there in this pdb object? 198 #Q8: Name one of the two non-protein residues? HOH, MK-1 #Q9: How many protein chains are in this structure? 2

Predicting functional motions of a single structure

Run a Normal Mode Analysis (NMA)

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

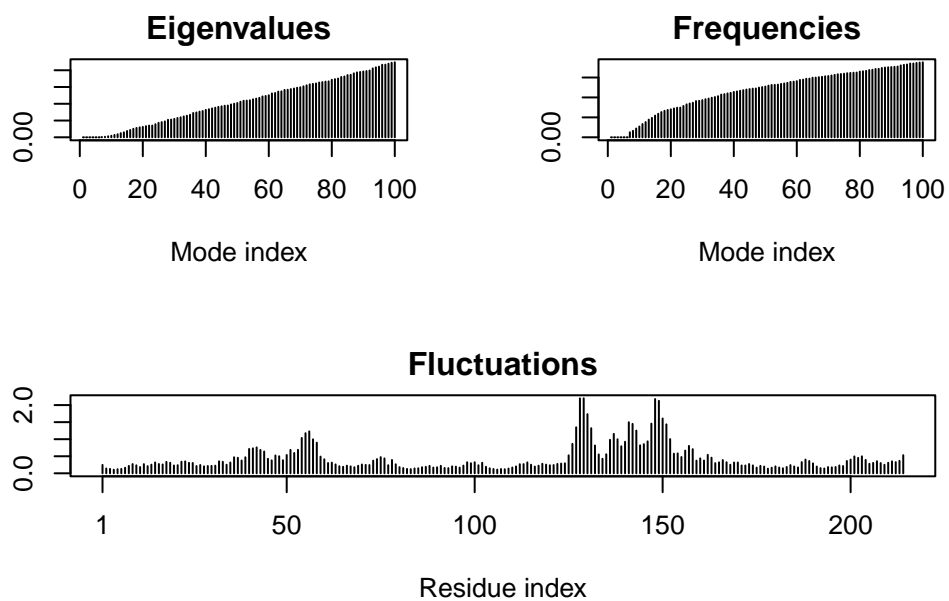
PDB has ALT records, taking A only, rm.alt=TRUE

```
modes <- nma(adk)
```

Building Hessian... Done in 0.014 seconds.

Diagonalizing Hessian... Done in 0.255 seconds.

```
plot(modes)
```



```
mktrj(modes, file="modes.pdb")
```