Quake

Main

#### **Overview**

Quake is a package to correct substitution sequencing errors in experiments with deep coverage (e.g. >15X), specifically intended for Illumina sequencing reads. Quake adopts the k-mer error correction framework, first introduced by the EULER genome assembly package. Unlike EULER and similar progams, Quake utilizes a robust mixture model of erroneous and genuine k-mer distributions to determine where errors are located. Then Quake uses read quality values and learns the nucleotide to nucleotide error rates to determine what types of errors are most likely. This leads to more corrections and greater accuracy, especially with respect to avoiding mis-corrections, which create false sequence unsimilar to anything in the original genome sequence from which the read was taken.

http://www.cbcb.umd.edu/software/jellyfish/

Guillaume Marcais and Carl Kingsford, A fast, lock-free approach for efficient parallel counting of occurrences of k-mers. Bioinformatics (2011) 27(6): 764-770

http://www.cbcb.umd.edu/software/quake/

Kelley DR, Schatz MC, Salzberg SL. Quake: quality-aware detection and correction of sequencing errors. Genome Biology 11:R116 2010.



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https://cutadapt.readthedocs.io/en/stable/

### **Cutadapt Removes Adapter Sequences From High-throughput Sequencing Reads**

Marcel Martin

#### Abstract

When small RNA is sequenced on current sequencing machines, the resulting reads are usually longer than the RNA and therefore contain parts of the 3' adapter. That adapter must be found and removed error-tolerantly from each read before read mapping. Previous solutions are either hard to use or do not offer required features, in particular support for color space data. As an easy to use alternative, we developed the command-line tool cutadapt, which supports 454, Illumina and SOLiD (color space) data, offers two adapter trimming algorithms, and has other useful features.

Cutadapt, including its MIT-licensed source code, is available for download at http://code.google.com/p/cutadapt/

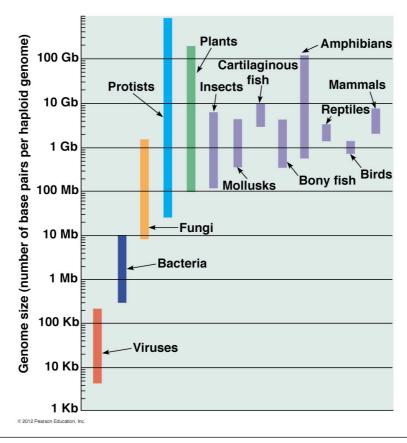
# What's in a genome?

- Genome size
- Genome structure

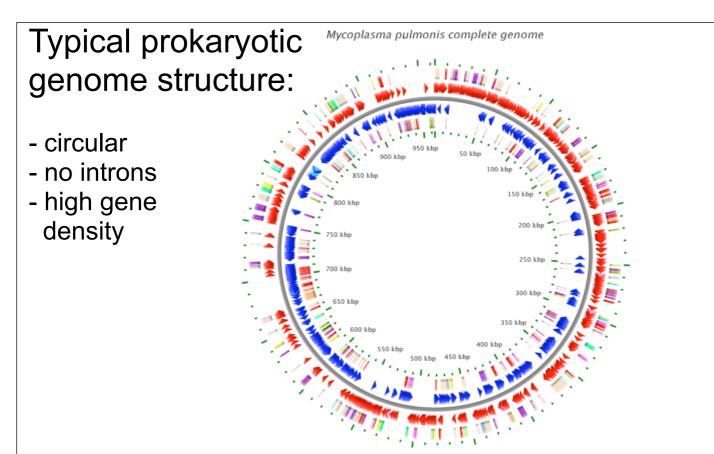
## **Annotation**

Repeat Finding

# Comparative Genome Size



Organism	Genome size (Kb)	Approx no. genes	Notes
Human mitochondrion	16.5	37	
Epstein-Barr virus	172	80	Causes mononucleosis
Nanoarchaeum equitans	401	552	Parasitic Archaea, smallest known genome of a 'true' organism
Encephalitozoon cuniculi	2,508	1,997	Parastic eukaryote
Deinococcus radiodurans	3,254	3,157	2 chromosome, 2 plasmids; highly radiation resistant
Vibrio cholerae	4,033	3,800	2 chromosomes; causes cholera
Escherichia coli	4,639	4,377	4,290 of these are protein-coding
Saccharomyces cerevisiae	12,496	5,770	Budding yeast; eukaryote
Caenorhabditis elegans	100,258	20,532	First multicellular eukaryote to be sequenced
Arabidopsis thaliana	115,410	~25,000	Flowering plant
Drosophila melanogaster	122,654	13,927	Fruit fly
Tetraodon nigroviridis	342,420	27,918	Pufferfish; very compact genome
Rice	390,000	37,544	
Dog	2,400,000	19,300	
Human	3,300,000	20,769	

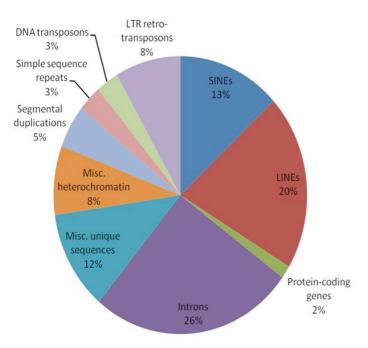


http://microbewiki.kenyon.edu/index.php/File:Genome.png

Length: 963,879 bp; Genes: 814

Accession: NC\_002771

## Human genome: not homogeneous

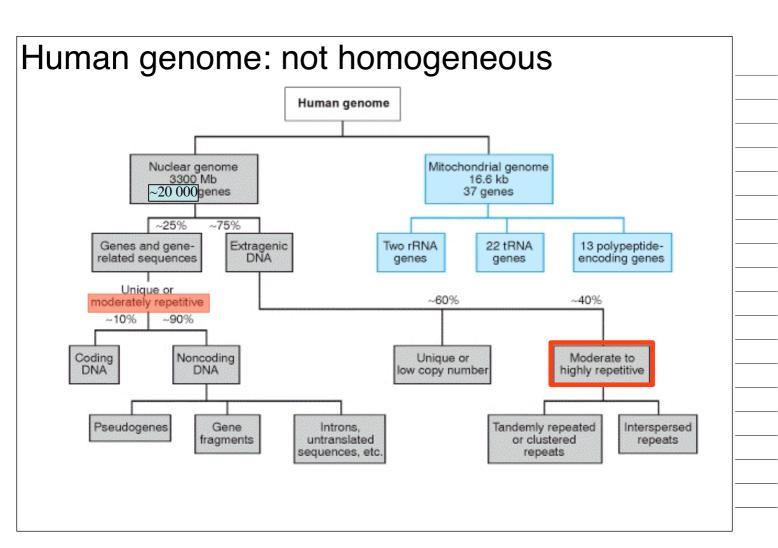


### Two genomes:

- Mitochondrial:16.6 kb 37 genes
- Nuclear:3,300,000 kb -~20,000 genes

Linear
Complex gene structure
Low gene density

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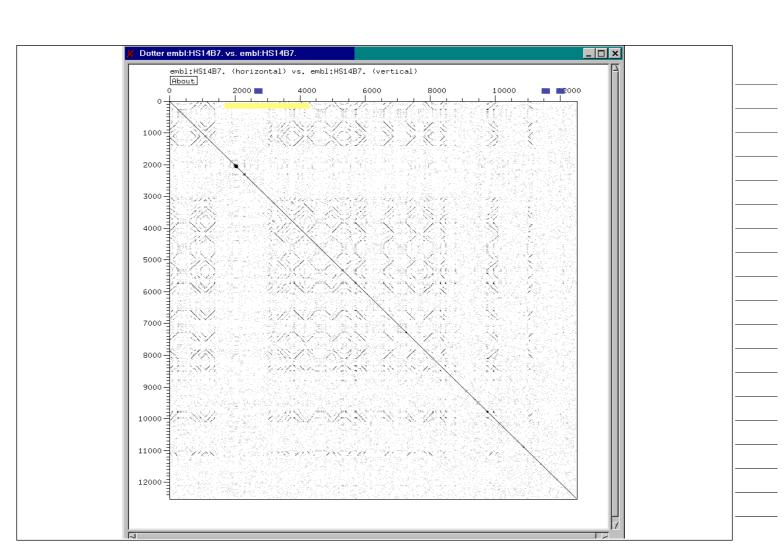


# What's in a (Human) Genome?

Varying %GC: 1Mb of 16p13.3



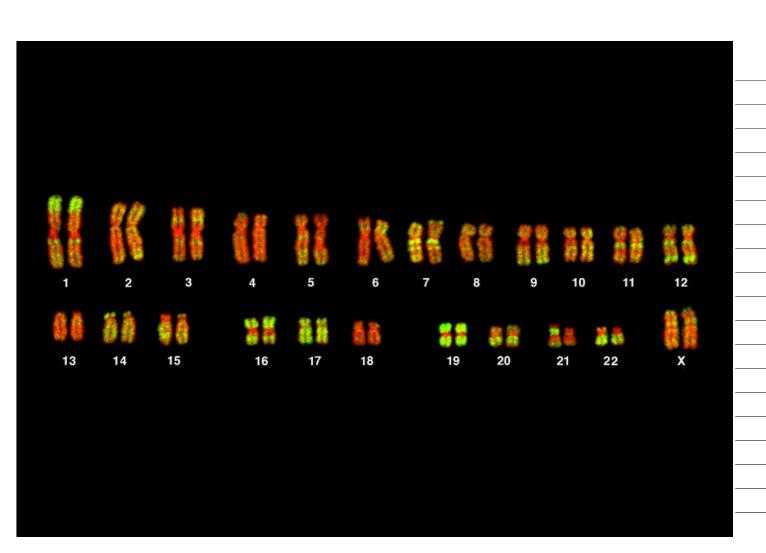
- 95% Intergenic sequence
  - ~50% genome is repetitive
    - Low complexity/ tandem repeats/ centromeric/ telomeric
  - Mobile elements (selfish DNA)
    - Transposons/ retroviruses
      - Long interspersed repeats (LINE)
        - » E.g. 6kb L1 LINE, old, often partial.
      - Short interspersed sequences (SINE)
        - » E.g. ~10e6 copies of 300bp ALU repeat, young, mostly complete.
- %GC; Alu density; gene density all related (LINE inversely)



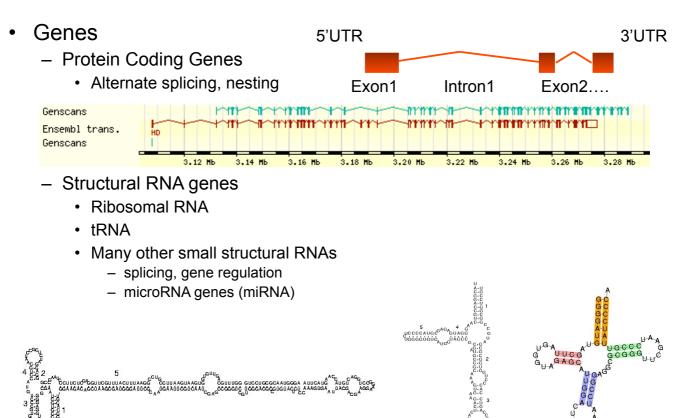
## CpG Islands

- · C modified by methylation to methyl-C
- Methyl-C frequently mutates to T

- CpG is normally ~5 fold under-represented from genomic GC% (in intergenic regions)
- CpG more frequent where methylation is suppressed (e.g. gene promoter regions)



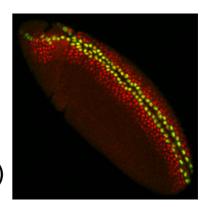
## What's in a Genome? II



# What's in a Genome? III

- Regulatory Sequences
  - Gene transcription
  - DNA replication
  - Gene splicing
  - (Gene re-arrangment: Ig, TCR genes)
  - (Chromatin packing)

Signals are small
Signals are often conserved between organisms

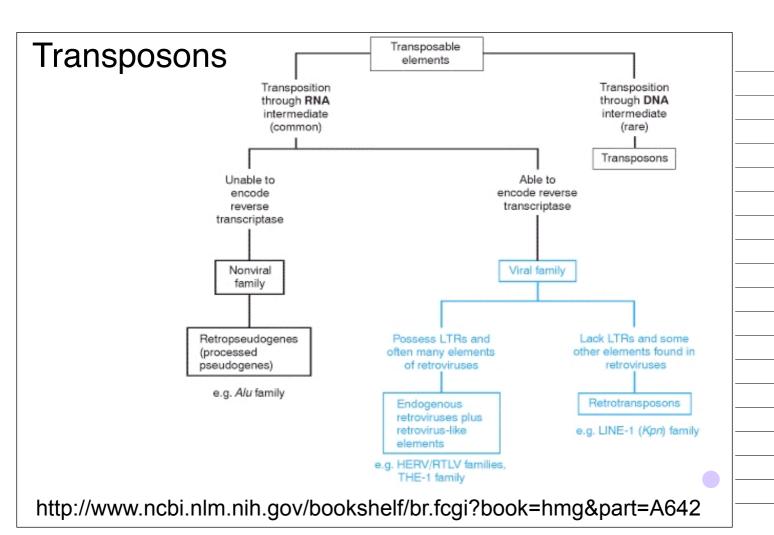


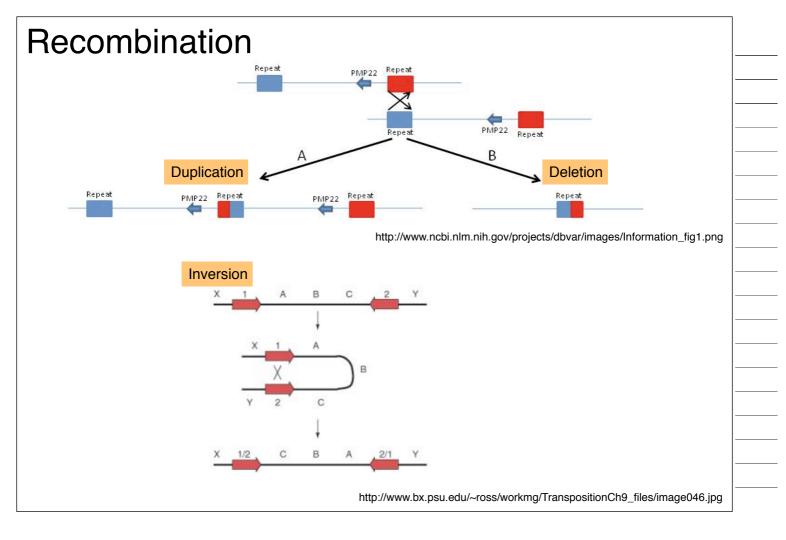
### Indian Muntjac

### Chinese Muntjac









## **Annotation**

Repeat Finding

# **Existing Databases**

- RepBase
  - All types of repeats; actual sequencehttp://www.girinst.org/repbase/index.html
- Dfam
  - Alignments, HMMs and match lists of repeats

http://dfam.org/

# RepeatMasker

- Screens using modified RepBase library
- Alignment with engine similar to blast but optimised for repeats
- Masks ~50% of human sequence

# de novo repeat finding

- Start from assembled genome: High-coverage kmers, align, greedy extension e.g. RepeatScout
- Start from reads: calculate overlaps, generate graph – vertices repeats elements, edges overlaps and cluster e.g. RECON
- Combined in RepeatModeller

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## **Problems**

- Speed
- False positive rate esp. repetitive elements in genes
- Classification
- Saha et al. Empirical comparison of ab initio repeat finding programs – Nucleic Acid Research 2008

### References

Benson - Tandem repeats finder: a program to analyze DNA sequences – Nucleic Acid Research 1999

Smit, AFA, Hubley, R & Green, P. RepeatMasker Open-3.0. 1996-2010

http://www.repeatmasker.org

RepeatScout: Price AL, et al. - De novo identification of repeat families in large genomes. Bioinformatics. 2005

RECON: Bao Z, Eddy SR – Automated de novo identification of repeat sequence families in sequenced genomes. Genome Res. 2002

RepeatModeller: http://www.repeatmasker.org/RepeatModeler.html

Saha *et al.* - Empirical comparison of ab initio repeat finding programs – Nucleic Acid Research 2008

#### Online resources to explore in your own time - I

#### Resource centres:

The EBI: www.ebi.ac.uk

The NCBI: www.ncbi.nlm.nih.gov UCSC: genome.ucsc.edu

#### Model organism databases:

Budding yeast: www.yeastgenome.org
Worm: www.wormbase.org
Fly: www.flybase.org

Mouse: www.informatics.jax.org

Rat: rgd.mcw.edu Zebrafish: zfin.org

### Database building tool: www.intermine.org InterMine databases:

http://yeastmine.yeastgenome.org

http://www.mousemine.org/mousemine

https://phytozome.jgi.doe.gov/phytomine/begin.do

https://apps.araport.org/thalemine

http://www.humanmine.ora

http://targetmine.mizuguchilab.org http://mitominer.mrc-mbu.cam.ac.uk

http://ratmine.mcw.edu

http://intermine.wormbase.org

http://zmine.zfin.org

http://intermine.modencode.org

http://www.flymine.org

Ontologies:

Gene ontology:

www.geneontology.org

Sequence ontology:

www.sequenceontology.org

#### DNA sequence/ genomes:

Ensembl: www.ensembl.org

Short read archive: www.ncbi.nlm.nih.gov/sra

Proteins:

www.uniprot.org

www.ebi.ac.uk/interpro

RNA:

rfam.sanger.ac.uk

#### Pathways:

www.reactome.org www.genome.jp/kegg/

#### Online resources to explore in your own time - II

#### A selection of tools:

GBrowse genome browser: gmod.org/wiki/Gbrowse CGL genome annotation manipulation:

www.yandell-lab.org/software/cgl.html

bio{perl, python, java, ruby}.org: much useful functionality mummer overview genome comparisons: mummer.sourceforge.net

Galaxy: powerful online analysis system: galaxy.psu.edu

For next generation sequencing related questions/discussions: http://seqanswers.com

Pubmed - http://www.ncbi.nlm.nih.gov/pubmed

WoK - http://wok.mimas.ac.uk/

Google - http://scholar.google.co.uk

