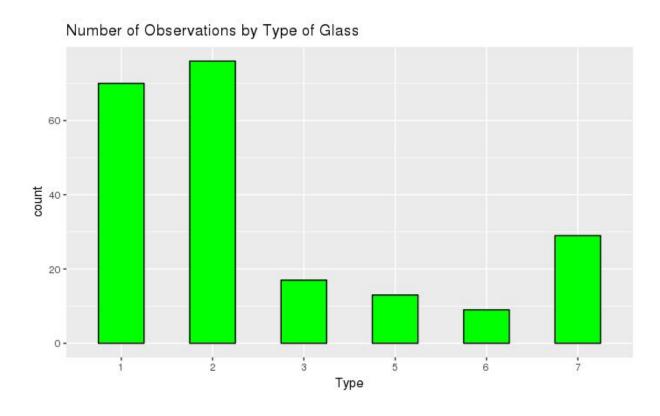
Assignment No:1

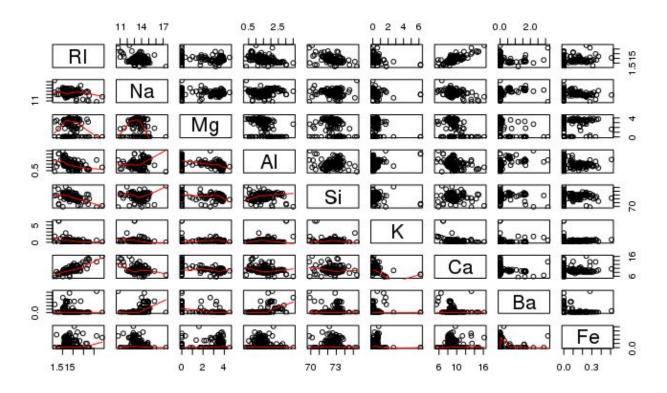
Supriya Bachal

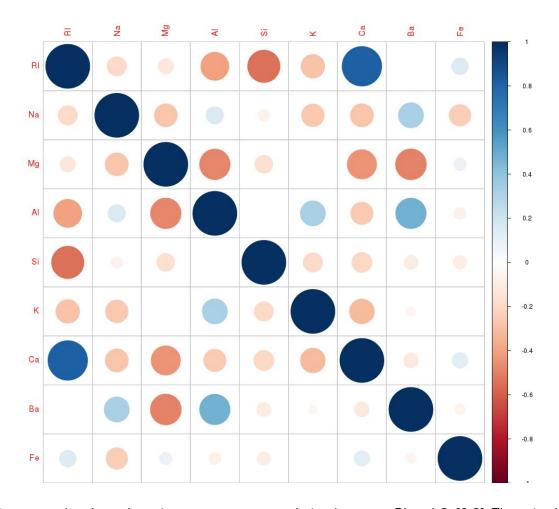
Question 3.1

- (a) Using visualizations, explore the predictor variables to understand their distributions as well as the relationships between predictors.
- (b) Do there appear to be any outliers in the data? Are any predictors skewed?
- (c) Are there any relevant transformations of one or more predictors that

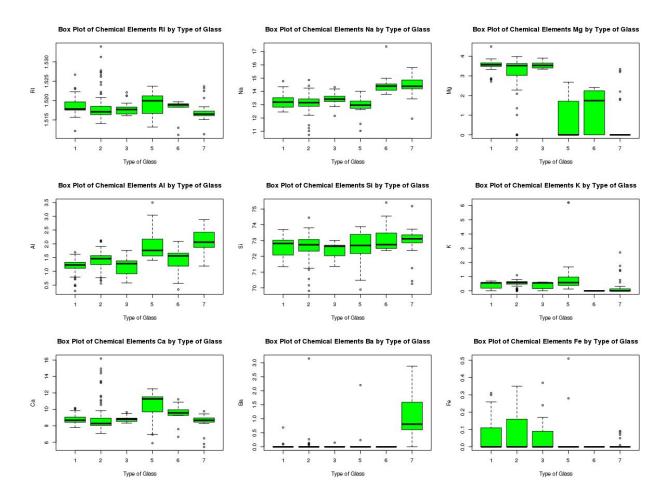


In the first step i identified the number of observations that belonged to each of the type of glass. The above histogram reveals that most of the observations are of glass Type 1 or 2. Least no of observations are of Type 6. Types 1 and 2 represent 68% of the total observations.



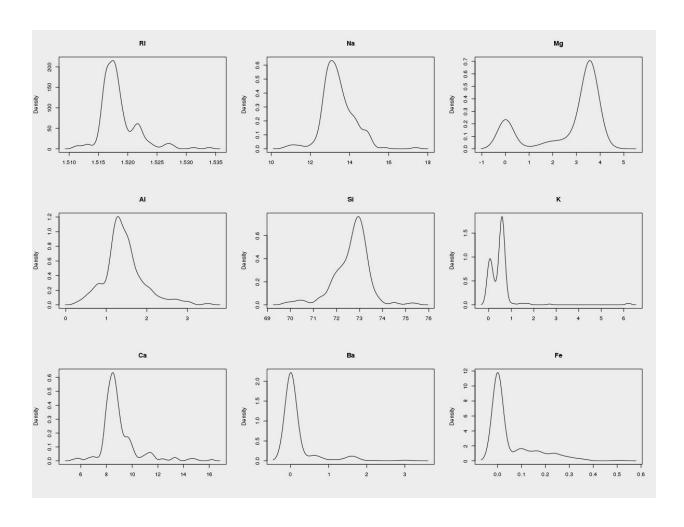


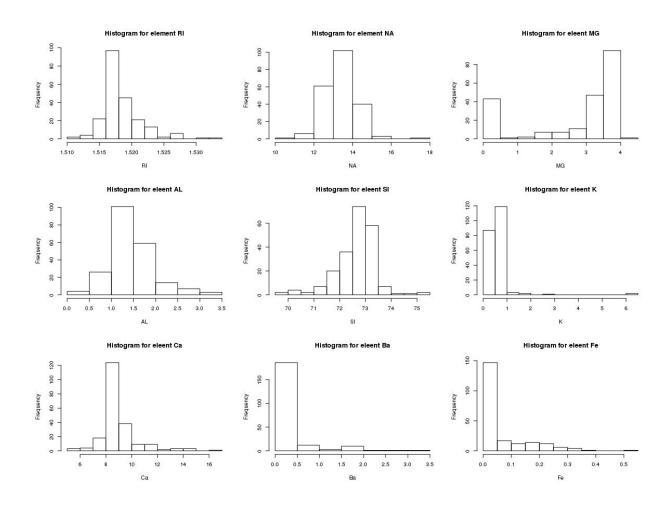
The above graphs show there is a very strong correlation between RI and Ca[0.8]. There is also a correlation between Ai and Ba as also between Ba and Na. There is also a correlation between K and Ai and so on but the strongest one is between RI and ca. The plot also shows that there is a negative correlation between Ri and Si and Ba and Mg.Seeing the high positive Correlation maybe we should drop one of these variables while building a model.



The above box plots show that there are a large no of outliers for Ca for Type 2 and for Mg for Type 2 similarly for Na for and Si. Most of the outliers are for Type 2. Additionally we also can note that for the elements Fe, Ba and k there are instances of 0 observations of a certain Types which implies that we can conclude that if a certain type of glass has traces of fe it cannot be of type 6 etc. The box plots also show that the amount of Fe and Ba in most types of glass is very low. For Mg the concentration is very high for Type 1 and Type 2 and Type 3. If the concentration of Ba is more than 0.1 there is a very high chance that the glass is of Type 7.

To further investigate this we can form histograms and density plots of the elements.





As we saw the distribution from the density plots , we see the histograms as well for the distribution of the elements.the peaks for RI is 1.515 and 1.520, for NA it is 12 to 14 , Mg it is 3 to 4 and so on. To futhur investigate the skewness of the data we use the skewness function which results in the following:

The Skewness of the various elements is as below:

RI 1.6027151	Al 0.8946104	Ca 2.0184463
Ba 3.3686800	Si -0.7202392	Na 0.4478343
Fe 1.7298107	Mg -1.1364523	K 6.4600889

We can conclude that the From the histogram that predictors RI,Mg and Fe are skewed. RI, Na and AI are slightly right skewed while Ca, Ba and Fe are very right skewed .K is double peaked and the density plots show that Mg and K are bimodal.

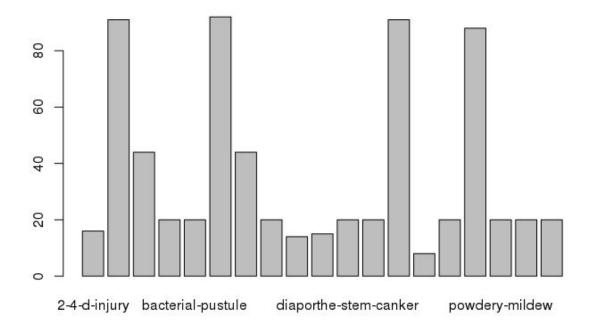
The most common transformations to eliminate skewness are log, square root or inverse transformations and/or outliers are spatial sign transformations. However there are many zero values in some of the elements hence we cannot use log transformations. We can try to find an appropriate transformation using Box -cox method from the Caret package. After scaling, centering and using Box cox transformations the skewness was not reduced to a great extent. However to improve results we could use the Yeo Johnson family of transformations to combat the problem of predictors having zero values.

Question 3.2:

- a) Investigate the frequency distributions for the categorical predictors. Are any of the distributions degenerate in the ways discussed earlier in this chapter?
- (b) Roughly 18 % of the data are missing. Are there particular predictors that are more likely to be missing? Is the pattern of missing data related to the classes?
- (c) Develop a strategy for handling missing data, either by eliminating predictors or imputation.

The first observation about this dataset was that it has missing values and the predictor as well as the predicted variables are factors.

Distribution of variables:



Degenerate distribution basically means that there is not enough variability in a predictor to be informative about the prediction.

The ordered descending table the diseases are as follows:

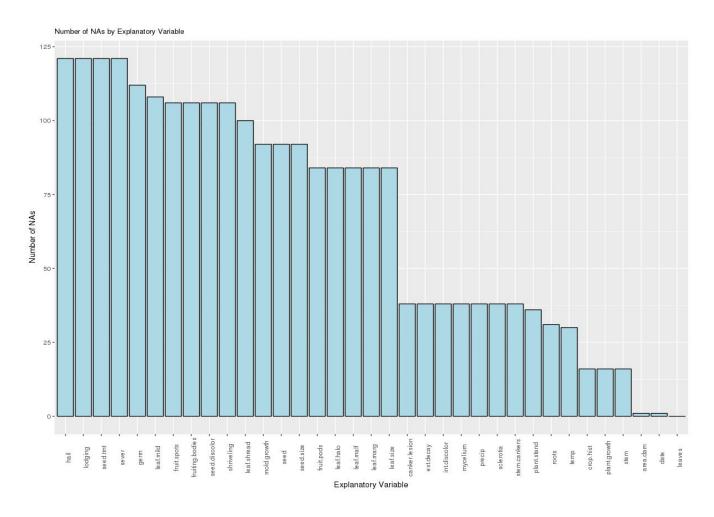
- 1 brown-spot 92
- 2 alternarialeaf-spot 91
- 3 frog-eye-leaf-spot 91
- 4 phytophthora-rot 88
- 5 anthracnose 44
- 6 brown-stem-rot 44

To handle the missing values

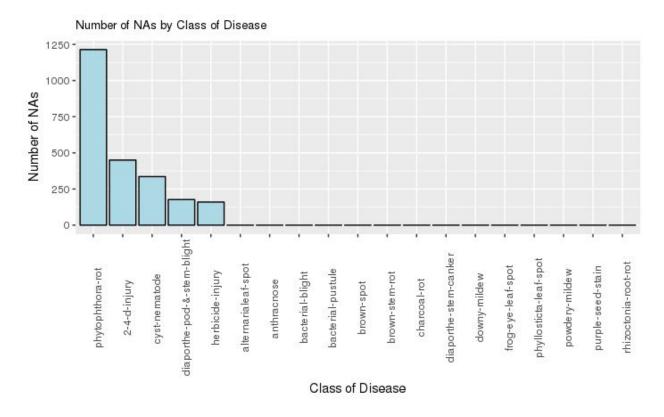
na.count		x.var	
1	1	date	
	•		
2	36	plant.stand	
3	38	precip	
4	30	temp	
5	121	hail	
6	16	crop.hist	
7	1	area.dam	
8	121	sever	
9	121	seed.tmt	
10	112	germ	
11	16	plant.growth	
12	0	leaves	
13	84	leaf.halo	
14	84	leaf.marg	
15	84	leaf.size	
16	100	leaf.shread	
17	84	leaf.malf	
18	108	leaf.mild	
19	16	stem	
20	121	lodging	
21	38	stem.cankers	
22	38 c	38 canker.lesion	

106 fruiting.bodies

- 24 38 ext.decay
- 25 38 mycelium
- 26 38 int.discolor
- 27 38 sclerotia
- 28 84 fruit.pods
- 29 106 fruit.spots
- 30 92 seed
- 31 92 mold.growth
- 32 106 seed.discolor
- 33 92 seed.size
- 34 106 shriveling
- 35 31 roots



We find that the predictor variables have a maximum count of 120. This is about 17 percent of the data.

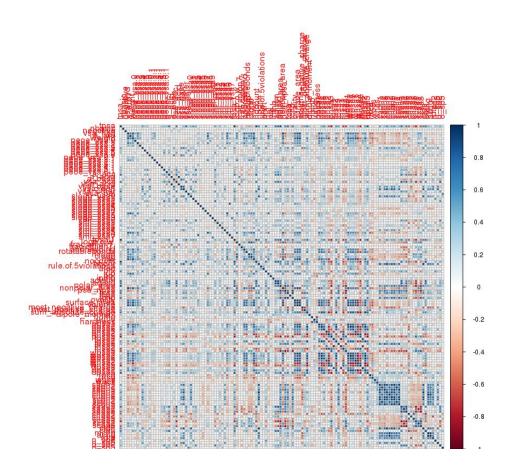


The classes that show maximum number of NA values are Phytophthora-rot 2-4-d-injury and cyst nematode. These classes are likely to have more missing values than the others.

- (b) Do any of the individual predictors have degenerate distributions?
- (c) Generally speaking, are there strong relationships between the predic-

tor data? If so, how could correlations in the predictor set be reduced?

Does this have a dramatic effect on the number of predictors available for



There are many positive and many negative correlations in the data, and since most of the variables are related to each other the data will be reduced to a large extent as we may drop many of the variables.

Appendix:

Question 1:

title: "Assignment 1"

author: "Supriya Bachal"

date: "September 30, 2017"

```
output: html_document
```{r setup, include=FALSE}
knitr::opts_chunk$set(echo = FALSE)
```{r}
library(mlbench)
data(Glass)
str(Glass)
```{r}
data(Glass)
summary(Glass$Type)
library(ggplot2)
ggplot(Glass,aes(x=Type))+
stat_count(width = 0.5,fill="Green",colour="black")+
ggtitle("Number of Observations by Type of Glass")

```{r}
par(mfrow=c(3,3))
library(fBasics)
for(i in 1:9) {
d<-density(Glass[,i])
plot(d,main=names(Glass)[i],xlab = "")
```

```
}
```{r}
par(mfrow=c(3,3))
for(i in 1:9){
 plot(Glass$Type,Glass[,i],col="Green",
 xlab="Type
 of
 Glass",ylab=names(Glass)[i],main=paste("Box
 Plot
 Chemical
 of
Elements", names(Glass)[i], "by Type of Glass"))
}
```{r}
pairs(Glass[,1:9],lower.panel=panel.smooth)
m<-cor(Glass[1:9])
cor(Glass[1:9])
corrplot(m, order="hclust")
```{r}
library(e1071)
apply(Glass[,-10], 2, skewness)
par(mfrow=c(3,3))
hist(Glass$RI,main="Histogram for element RI",xlab="RI")
hist(Glass$Na,main="Histogram for element NA",xlab="NA")
hist(Glass$Mg,main="Histogram for eleent MG",xlab="MG")
hist(Glass$Al,main="Histogram for eleent AL",xlab="AL")
hist(Glass$Si,main="Histogram for eleent SI",xlab="SI")
```

```
hist(Glass$K,main="Histogram for eleent K",xlab="K")
hist(Glass$Ca,main="Histogram for eleent Ca",xlab="Ca")
hist(Glass$Ba,main="Histogram for eleent Ba",xlab="Ba")
hist(Glass$Fe,main="Histogram for eleent Fe",xlab="Fe")
```{r}
Sorted<-Glass[order(Glass$Type),]
library(e1071)
for(i in 1:9)
{
a[i]<-skewness(Glass[,i])
}
names(a)<-c("RI","Na","Mg","AI","Si","K","Ca","Ba","Fe")
```{r}
f=Glass[,-10]
trans <- preProcess(f,method = c("BoxCox"))
transformed <- predict(trans, f)</pre>
fulldata.scale = data.frame(predict(pred.scale, as.data.frame(Glass[, -10])),Type = Glass$Type)
par(mfrow=c(3,3))
for (i in names(fulldata.scale)[1:9]) {
```

```
ggplot(fulldata.scale, aes_string(x = i)) + geom_histogram(fill = "SeaGreen") + labs(y = "")
}
par(mfrow=c(3,3))
graphlist
skewness(fulldata.scale[,i])
yjTrans <- preProcess(Glass[, -10], method = "YeoJohnson")
yjData <- predict(yjTrans, newdata= Glass[, -10])</pre>
Q2
title: "Assignment 2"
author: "Supriya Bachal"
date: "October 1, 2017"
output: html_document
```{r setup, include=FALSE}
knitr::opts_chunk$set(echo = TRUE)
```{r}
```

```
d=table(Soybean$Class)
barplot(d)
which.max(d)
d[which.min(d)]
gg=data.frame(d[order(d,decreasing = T)])
head(gg)
```{r}
Missing values
isna.byclass<-Soybean%>%
group_by(Class)%>%
do(data.frame(sum(is.na(.))))
names(isna.byclass)<-c("Class","na.count")</pre>
isna.byclass
isna.byclass<-Soybean%>%
 group_by(Class)%>%
 do(data.frame(sum(is.na(.))))
names(isna.byclass)<-c("Class","na.count")</pre>
Isna.byclass
Question:3
```{r}
```

library(caret)

data(BloodBrain)

correlations<-cor(bbbDescr)

library(corrplot)

corrplot(correlations)

\*\*\*

#### References:

https://rpubs.com/fraki22/122619

https://rpubs.com/chidungkt/221544

https://rpubs.com/chidungkt/221544

Chapter 3 from notes.