Multivariate Statistical Analysis

Homework 4

Lucas Fellmeth, Helen Kafka, Sven Bergmann

03/13/24

```
set.seed(42)
```

Problem 1

Consider the bupa_liver_disorder data set, where a number of individuals are classified into two classes according to blood test results (see full description on p. 258 of the book).

```
bupa_liver_disorder <- read.csv(file = "../Data_csv/bupa_liver_disorder.csv")
bupa_liver_disorder$class <- factor(bupa_liver_disorder$class)</pre>
```

a)

Split the data set into training and test sets (roughly a 70/30 split). Compute the logistic classifier using the training set.

```
out <- glm(formula = class ~ mcv + alkphos + sgpt + sgot + gammagt +
    drinks, data = bupa_liver_disorder_train, family = binomial)
out$coefficients</pre>
```

```
## (Intercept) mcv alkphos sgpt sgot gammagt

## 5.01564297 -0.05348287 -0.01975076 -0.06741025 0.13150896 0.01971347

## drinks

## -0.08709490
```

b)

Construct the misclassification table and compute the misclassification rate for the test set.

```
pi1_hat <- predict(out, type = "response", newdata = bupa_liver_disorder_test)
gr_hat <- ifelse(pi1_hat > 0.5, 1, 2)
mctable <- table(gr_hat, bupa_liver_disorder_test$class)
mctable</pre>
```

```
##
## gr_hat 1 2
## 1 19 44
## 2 28 12

1 - sum(diag(mctable))/length(bupa_liver_disorder$class)
## [1] 0.9101449
```

Problem 2

Consider the ecoli data set. These data were obtained in a study of protein localization sites for 336 examples of E. coli. There are 7 predictor variables and a class variable, localization_site, which indicates the protein localization.

```
ecoli <- read.csv(file = "../Data_csv/ecoli.csv")
ecoli$localization_site <- factor(ecoli$localization_site)</pre>
```

a)

Split the data set into training and test sets (roughly a 80/20 split). Compute the multinomial logistic classifier using the training set.

```
## # weights: 72 (56 variable)
## initial value 698.692358
## iter 10 value 175.691378
## iter 20 value 112.464952
## iter 30 value 107.295386
## iter 40 value 105.913978
## iter 50 value 105.487896
## iter 60 value 105.353525
## iter 70 value 105.242112
## iter 80 value 105.109689
## iter 90 value 105.014041
## iter 100 value 104.969071
## final value 104.969071
## stopped after 100 iterations
```

out

```
## Call:
## nnet::multinom(formula = localization_site ~ mvg + gvh + lip +
      chg + aac + alm1 + alm2, data = ecoli, maxit = 100)
##
## Coefficients:
##
      (Intercept)
                                    gvh
                                              lip
                                                         chg
                       mvg
                                                                     aac
       -14.179741 -1.198919 5.8227058 -1.671139 -7.791876 -0.7240761
## im
## imL -42.802584 37.770240 -49.7078462 33.274120 18.860327 -23.7806465
## imS -15.930816 14.647235
                             3.9962416 -7.935507 -7.823256
                                                               3.0728426
## imU -17.740856 10.842884
                             0.8374873 2.285835 -9.110530 -1.1427249
## om
       -29.106326 7.400551 22.5668026
                                        1.679383 -14.606608 39.1260334
## omL -14.537074 13.877294 -14.1367197 28.299112 -21.495756
                                                              8.0589385
        -4.044233 6.299159 15.6413210 -20.001416 -4.885281
                                                              0.4754381
##
          alm1
                     alm2
## im 36.37688 -4.653052
## imL 18.57044
                4.957740
## imS 26.74233 -9.901364
## imU 29.40923 -2.259731
## om 11.01966 -27.030325
## omL 21.53240 -31.368635
## pp 20.56965 -12.007667
## Residual Deviance: 209.9381
## AIC: 321.9381
mctable <- table(predict(out, newdata = ecoli_test), ecoli_test$localization_site)</pre>
1 - sum(diag(mctable))/length(ecoli$localization site)
```

[1] 0.8095238