**Team3ispA: Miniprep**

**source**: mini shaker

**samples**

*culture label side\_label destination plasmid*

A1 pLYC73S-ispA1 A1 Box\_ ispA3/F1 pLYC73S-ispA1

A2 pLYC73S-ispA2 A2 Box\_ ispA3/F2 pLYC73S-ispA2

A3 pLYC73S-ispA3 A3 Box\_ ispA3/F3 pLYC73S-ispA3

A4 pLYC73S-ispA4 A4 Box\_ ispA3/F4 pLYC73S-ispA4

A5 pLYC73S-ispA5 A5 Box\_ ispA3/F5 pLYC73S-ispA5

**Protocol**

* Take an image of the culture block, upload it later to the Github issue
* For each sample, perform 2 minipreps (each on 2mL culture)
* Combine the two identical eluted minipreps in a regular zymo cleanup;

elute with **20uL** ddH2O

**Team3ispA: Sequence**

**source**:

sISPA1 benchtop/lyophilized

sISPA2 benchtop/lyophilized

sISPA3 benchtop/lyophilized

sISPA4 benchtop/lyophilized

sISPA5 benchtop/lyophilized

**samples**

*label location plasmid oligo*

A1 Box\_ ispA3/F1 pLYC73S-ispA1 sISPA1

A2 Box\_ ispA3/F2 pLYC73S-ispA2 sISPA2

A3 Box\_ ispA3/F3 pLYC73S-ispA3 sISPA3

A4 Box\_ ispA3/F4 pLYC73S-ispA4 sISPA4

A5 Box\_ ispA3/F5 pLYC73S-ispA5 sISPA5

**dilutions**

*label concentration destination*

sISPA1 100uM ispA\_stocks/A1

2.66uM sISPA1 2.66uM Box\_ ispA3/G1

sISPA2 100uM ispA\_stocks/A2

2.66uM sISPA2 2.66uM Box\_ ispA3/G2

sISPA3 100uM ispA\_stocks/A3

2.66uM sISPA3 2.66uM Box\_ ispA3/G3

sISPA4 100uM ispA\_stocks/A4

2.66uM sISPA4 2.66uM Box\_ ispA3/G4

sISPA5 100uM ispA\_stocks/A5

2.66uM sISPA5 2.66uM Box\_ ispA3/G5

**protocol**

* Make 100uM oligo stocks:
* Make 2.66uM oligo stocks:
  + 97.3 uL ddH2O
  + 2.7 uL oligo
* For each plasmid listed, mix the following sequencing reactions in an eppendorf tube:
  + 4 uL ddH2O
  + 6 uL miniprep DNA (undiluted)
  + 3 uL oligo (2.66 uM)
* Clearly label the tops of the tubes with the “label”
* Take the sequencing reactions and order form to:
  + 237 Stanley Hall (second floor cold room)