**Team3ispA: Miniprep**

**source**: mini shaker

**samples**

*culture label side\_label destination plasmid*

A1 pLYC73S-ispA1 A1 Box\_ ispA3/F1 pLYC73S-ispA1

A2 pLYC73S-ispA2 A2 Box\_ ispA3/F2 pLYC73S-ispA2

A3 pLYC73S-ispA3 A3 Box\_ ispA3/F3 pLYC73S-ispA3

A4 pLYC73S-ispA4 A4 Box\_ ispA3/F4 pLYC73S-ispA4

A5 pLYC73S-ispA5 A5 Box\_ ispA3/F5 pLYC73S-ispA5

**Protocol**

* Take an image of the culture block, upload it later to the Github issue
* For each sample, perform 2 minipreps (each on 2mL culture)
* Combine the two identical eluted minipreps in a regular zymo cleanup;

elute with **20uL** ddH2O

**Team3ispA: Sequence**

**source**:

sLYC10 benchtop/lyophilized

**samples**

*label location plasmid oligo*

K1-A Box\_Lyc6/A7 pLYC33K-A sLYC10

K1-B Box\_Lyc6/B7 pLYC33K-B sLYC10

**dilutions**

*label concentration destination*

sLYC10 100uM TPcon4\_stocks/G8

2.66uM sLYC10 2.66uM Box\_Lyc6/F5

**protocol**

* Make 100uM stock of sLYC10
* Make a 2.66uM stock of sLYC10:
  + 97.3 uL ddH2O
  + 2.7 uL oligo
* For each plasmid listed, mix the following sequencing reactions in an eppendorf tube:
  + 4 uL ddH2O
  + 6 uL miniprep DNA (undiluted)
  + 3 uL oligo (2.66 uM)
* Clearly label the tops of the tubes with the “label”
* Take the sequencing reactions and order form to:
  + 237 Stanley Hall (second floor cold room)