

Electrical Impedance Tomography for Cardiovascular Imaging and Monitoring

by

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To ...

Abstract

This thesis serves to show the background, methods, planned methods and timeline for the final thesis. The thesis will consist of 6 chapters, in introduction to give background and motivation for the research

Acknowledgements

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Chapter 1

Introduction

Electrical impedance tomography (EIT) is an imaging modality that uses an arrangement of electrodes to simultaneously apply stimulation currents and measure the resulting electric potentials. Measurements of potential at the electrodes are used in conjunction with prior information to reconstruct images of the internal conductivity. In biomedical applications, the variance in conductivity between different tissue types and fluids enables non-invasive imaging of functional activity using electrodes on the body surface.

Thoracic EIT focuses on imaging cardiopulmonary activity using a belt of electrodes placed around the ribcage. In images of the chest, the signal is often dominated by impedance changes due to air movement in the lungs. The much smaller signals due to blood flow are of great interest for continuous monitoring and diagnostic applications, but are challenging to identify.

This thesis explores techniques to improve sensitivity to blood movement, or

perfusion, in EIT through the development of novel meshing techniques and electrode configurations incorporating internal electrodes. We investigate the current limitations of perfusion monitoring in 2D, and analyze novel meshing techniques and electrode placements for their ability to improve thoracic imaging and increase sensitivity to blood flow.

1.1 Motivation

TODO: Briefly

Why should we care?

Why do we measure perfusion?

What are the benefits if this technology works well? (increased sensitivity?)

What are the benefits of using EIT for this?

1.2 Problem

EIT can be used to image blood movement in two ways. First, it is possible to track the flow of blood using a conductivity-contrasting agent injected into a vein or artery and second, filtering the signal to isolate cardiac-frequency impedance changes. Using a conductive bolus injection to image perfusion has been well established and can give an easily detectable signal when using large volumes or high concentrations of the contrasting agent, but they do not allow continuous monitoring and frequent injections may pose risks to the patient. In thoracic EIT bolus injections also typically occur during apnea to facilitate removal from the respiratory component. Filtering

techniques for perfusion imaging are much more appealing for monitoring applications as they are less invasive and could be used continuously, but they present several challenges.

The primary challenge is the small amplitude of the signal. Impedance changes related to cardiac activity are often an order of magnitude smaller than signals related to respiration and can be challenging to identify without additional signals like an electrocardiogram (ECG), and averaging many heartbeats together. When averaging over several heartbeats, the ability of the system to monitor in real time is greatly diminished. Additionally there is some uncertainty surrounding the source of cardiac-frequency impedance changes. It is unclear to what extent the impedance changes stem from pulsatile motion in the thorax compared to the movement of blood, and if they can be used as a true measure of perfusion.

EIT is minimally invasive, requiring only the application of electrodes to the body surface, but does not have high sensitivity in the central regions of the chest where perfusion changes are likely to occur. As electrical current travels through the body it diffuses away from the electrodes, travelling in three dimensions along the paths of least resistance. EIT is most sensitive to impedance changes close to the electrodes where the current density is highest, and along the path of the injected current.

The placement of electrodes internally has the potential to increase current density and sensitivity in the center of the thorax. The benefits of internal electrodes have been simulated in 2D showing great improvements in reconstruction accuracy and sensitivity (Tehrani *et al.*, 2012), but in practice there are several challenges to overcome. Due to differences in physiology it is challenging to model the correct

location of the internal electrodes between subjects and electrodes placed internally may move with relation to the external electrodes during different physiological processes. To accurately reconstruct images with internal electrodes an accurate model is required with precise placement of electrodes and a model that matches the subject and is refined to meet the accuracy requirements. It is in part due to these challenges that internal electrodes have not been widely used in real-world situations, and to our knowledge no implementation of internal electrodes in conjunction with a 3D arrangement of external electrodes has been used for *in-vivo* imaging.

The applications of an EIT system with increased sensitivity to perfusion changes in the thorax are extensive. Methods to monitor blood pressure non-invasively are currently under development and could be greatly improved with increased sensitivity near the region of interest. Increased sensitivity near the heart could enable EIT to be realized as a low-cost solution to monitor and image hemodynamic activity more accurately.

1.3 Thesis objectives

The goal of this project is to improve sensitivity of EIT to perfusion using improved mesh accuracy and custom electrode locations. This thesis approaches problem from two angles (figure 1.1). We investigate both advanced meshing techniques, and novel electrode configurations with internal electrodes and their applications to improve thoracic EIT.

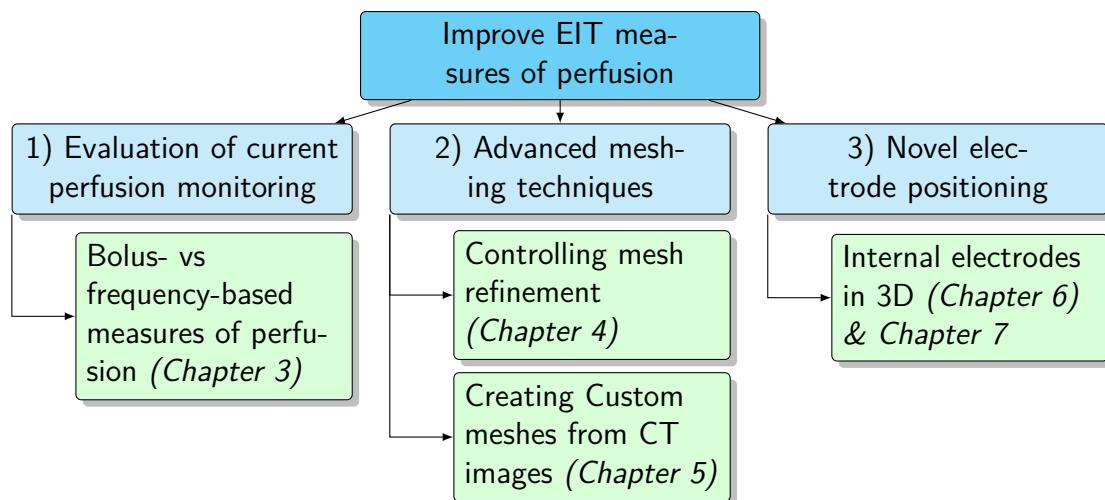


Figure 1.1: We aim to improve EIT measures of perfusion through 3 avenues. 1) Comparing and investigating the limitations of existing perfusion imaging methods; 2) Advancing meshing techniques; and 3) Using novel electrode locations consisting of a 3D external configuration with internal electrodes.

Chapter 2

Background

This section presents the this is short and needs a lot more detail and also figures.

This section briefly reviews the current techniques for lung perfusion and hemodynamic monitoring, and provides a general overview of the state of 3D EIT as used for thoracic imaging and monitoring.

2.1 Impedance Imaging

Impedance imaging has been in use since the early 1900s for geophysical applications. Originally introduced as a technique to image below the earth's surface, current was transmitted between two electrodes placed into the ground and any anomalies in subsurface conductivity produced deviation in the equipotential lines. Including current injections and measurements from multiple locations and using known electrical properties of geological structures Conrad Schlumberger identified features of underground geological structures (Allaud and Martin, 1977).

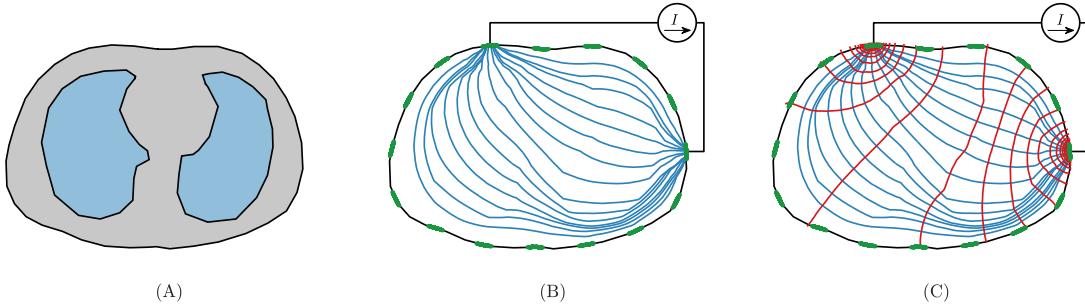


Figure 2.1: (A) A body comprising tissues of different conductivity, (B) Electrodes are placed on the surface and current is injected between a pair or electrodes. The current pathways are indicated by the blue lines. (C) The resulting equipotential lines within the body.

These same techniques can be applied in biomedical applications where voltage is measured on an array of body surface electrodes while current is applied between select electrode pairs (figure 2.1). Due to impedance differences associated with biological tissues and their physiological function (**Geddes1967; McAdams1995**), EIT has been proposed for a wide range of applications from thoracic monitoring to neuronal and brain imaging (**Holder1992; Frerichs2016**).

2.2 Bioimpedance

In thoracic imaging the most commonly measured impedance changes occur due to movement of air in the lungs, the flow of blood, and the motion of organs (Adler and Boyle, 2017). During inhalation, the volume of air in the lungs increases, lowering the conductivity of the lung tissue. The resistivity of lung tissue varies significantly between expiration and inspiration giving a value of $7 \Omega\text{m}$ during expiration and 23

Ωm during inspiration at 100 kHz (Witsoe and Kinnen, 1967), resulting in a measurable variation in impedance during respiration (Eyüboğlu *et al.*, 1989). There are also other significant sources of impedance change that make EIT signal interpretation challenging. Simulations have attributed up to 20 percent of the respiratory signal to the effect of chest expansion and movement of the chest wall (Adler *et al.*, 1994).

The source of impedance changes due to the flow of blood is even more complex. Since the resistivity of blood is so much lower than other tissues ($1.5 \Omega\text{m}$), the increase of blood due to pulsatile flow should decrease the impedance of structures it passes through by a detectable amount (Eyüboğlu *et al.*, 1989). It is often assumed that the component of EIT images at the cardiac frequency is related to the perfusion of blood, but the exact source of cardiosynchronous EIT signals is unclear (Nguyen *et al.*, 2012; Patterson, 2010). A continuous flow of blood alone is insufficient to induce a significant impedance change, as the volume and concentration of the conductive medium is unchanged. Any impedance-based measure of perfusion relies on the cardiosynchronous EIT signals which have numerous possible sources (Adler *et al.*, 2017b).

2.2.1 The Cardiac Cycle

The cardiac cycle consists of the activity in the heart between the beginning of one heart beat, and the next. There are two main stages of the cardiac cycle: the diastole, when the heart relaxes and is filled with blood, and the systole, when contraction of the heart pumps blood to lungs and all other body systems (Pappano and Wier,

2019). A simplified anatomy of the heart is presented in figure 2.2. Since ECG recordings are frequently used to synchronize EIT data, it is helpful to look at the timing of the cardiac cycle as it relates to features of ECG traces. An example ECG waveform is pictured in figure 2.3.

During the first stage of the cardiac cycle is the ventricular diastole, indicated by the P wave on an ECG trace, during which the heart relaxes and expands pulling blood into the ventricles from the atria (Pappano and Wier, 2019). Blood enters the atrium on the right side of the heart through the superior and inferior vena cava and on the left side of the heart, the atrium is filled by oxygenated blood from the lungs through the pulmonary veins (Pappano and Wier, 2019). Next is the atrial contraction during which time the atria pump additional blood into the ventricles, and the ventricular volume and pressure is maximized. At the peak of the ventricular volume, the ventricles contract and depolarize which corresponds to the QRS complex (Pollock and Makaryus, 2021). On the right side of the heart, deoxygenated blood is pumped to the lungs where it perfuses into the lung tissue, and on the left side of the heart oxygenated blood is pumped to the rest of the body through the aorta (Pappano and Wier, 2019). The smallest volume in the ventricles occurs after the ventricular repolarization represented in the ECG by T wave (Pollock and Makaryus, 2021).

2.2.2 Bioimpedance of Perfusion

There are several factors that may contribute some part of the impedance change during the cardiac cycle. Some of these factors include:

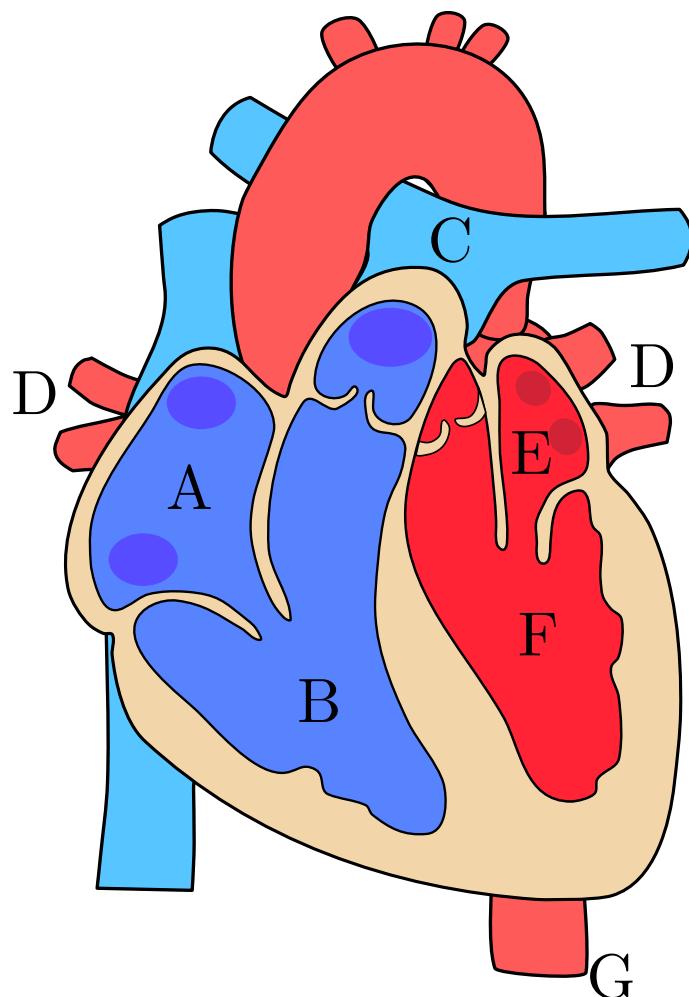


Figure 2.2: The pathway of the blood from entry to the heart to the descending aorta is indicated by the letters. Deoxygenated blood enters the heart through the right atrium (A) where it is moved to the right ventricle (B). From there it passes through the pulmonary artery (C) into the lungs where it perfuses and is oxygenated. Blood return from the lungs through the pulmonary veins (D) then enters the left atrium (E). Finally, the blood is pumped from the left ventricle (F) to the rest of the body through the aorta, and the descending aorta (G).

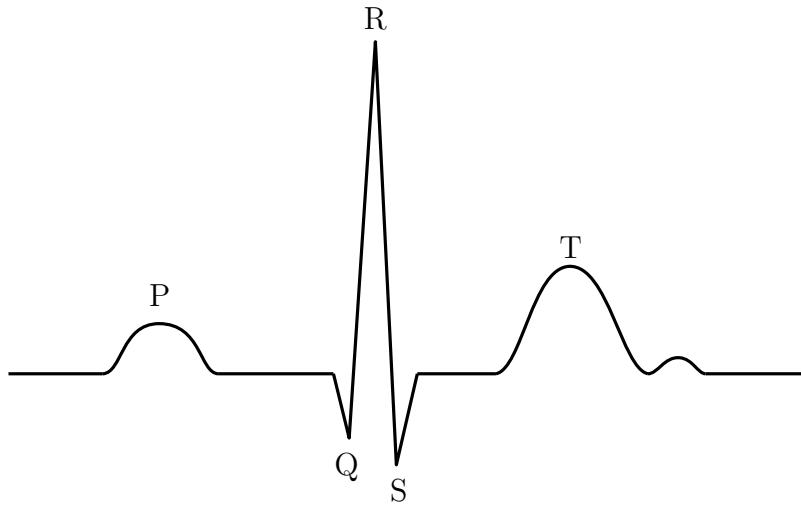


Figure 2.3: An example ECG waveform to compare electrical signals in the heart to blood volume changes. The P wave represents the beginning of the cardiac cycle with the ventricles begin to fill. The QRS complex corresponds with ventricular depolarization and occurs as the ventricles contract they contract. The beginning of the QRS complex corresponds with the maximum ventricular volume. The minimum volume in the ventricles occurs after ventricular repolarization represented by the T wave.

- Changes in blood volume within the heart as blood is pumped. As the ventricles fill with blood the volume increases and results in a more conductive heart (Nyboer *et al.*, 1970).
- Variations in arterial and blood vessel volume. Due to the elasticity of arteries and blood vessels, the pulsatile flow of blood passing through results in variation of vessel diameter, affecting the impedance (Eyüboğlu *et al.*, 1987).
- Physical deformation of structures due to motion of the heart. The motion of the heart can have significant contribution to cardiosynchronous EIT images (Adler *et al.*, 2017b; Proença *et al.*, 2015), with simulations showing that heart motion was the main contributor to impedance change due to the ventricle (Proença *et al.*, 2015).

- The orientation of red blood cells. During pulsatile flow the orientation of red blood cells changes, which had been shown to affect the impedance of the blood (Gaw, 2010).
- Ballistic forces in the body generated by the heart. During each heartbeat blood is pumped downwards through the descending aorta with a large force pushing the rest of the body upwards (Gordon, 1877). Different directions of flow in the aorta result in a repeating ballistic signal on the rest of the body (Kim *et al.*, 2016). This results in motion on the electrodes and body which can introduce significant artefacts in EIT signals (Adler *et al.*, 1994).

The contribution of each of these factors will ultimately depend on the placement of the electrodes and the specific geometry and physiology of a patient. When imaging changes in stroke volume, changes relating to posture, breathing and changes in belt position resulted in changes that overpowers the perfusion signals (Patterson *et al.*, 2001).

Despite the challenges of isolating cardiosynchronous EIT signals related to perfusion there is still a great interest in improving accuracy and stability due to the unique advantages offered by EIT over current state-of-the-art methods.

2.3 Perfusion Imaging

TODO: Discuss the reason for perfusion imaging here There is great interest in monitoring and imaging perfusion in the thorax. Measures and images describing perfusion in the cardiovascular system is widely used to diagnose diseases such as pulmonary

edema. There are several current techniques to image this **TODO: What methods should be discussed here? Initially thinking strictly perfusion and discuss ability to image flow?**

Additional informaion on the function of the cardiovascular system can improve diagnostic accuracy

Several techniques are used to monitor and image perfusion in the heart and lungs.

There are three main and are presented in the following section. **TODO: Additional places for inforation for more indepth stuff?**

2.3.1 Perfusion Imaging Techniques

2.3.1.1 Microspheres

2.3.1.2 Nuclear medicine

2.3.1.3 MRI

2.3.1.4 CT

2.3.1.5 Thermal diffusion

2.3.1.6 contrast agent

2.3.1.7 Ultrasound

2.3.2 Electrical Impedance Tomography for Perfusion Monitoring

2.4 Electrical Impedance Tomography

As described in section 2.1

2.4.1 Imaging Techniques

There are several challenges with EIT that make absolute EIT imaging difficult. The subtle difference in tissue impedance are small relative to artefact introduced by unknown boundary locations and electrode positions (Adler and Boyle, 2017; Adler *et al.*, 2015; Nissinen *et al.*, 2009). EIT measurements are typically used by reconstructing impedance differences between two time points. Time difference EIT

uses a reference frame to image the change in conductivity between two points in time and allows for imaging of functional activity such as the inflation of the lungs and the flow of blood. Time-difference EIT is also much more stable in the presence of errors that remain constant (Adler and Boyle, 2017; Brown, 2003). Frequency difference EIT is also possible based on the different impedance response of tissue types to changing frequencies. Frequency difference uses two or more different frequencies and calculates an image based on the change in electrical properties. Most frequencies used to differentiate between different tissue types are at high frequencies where current starts to flow across the cell membranes. These high frequencies are out of the range of most current EIT systems and frequency changes due to different lower frequencies are limited (Adler and Boyle, 2017). This thesis uses time-difference EIT to images changes in movement and fluid volumes in the thorax.

2.4.2 EIT measurements

EIT measurements are made on a number of external electrodes placed on the body surface around a region of interest. In this thesis we focus on measurements of the thorax. The main aspects affecting EIT measurements are the electrodes places on the body surface and technical details of the current and voltage that are required.

2.4.2.1 Electrodes

EIT is possible using **TODO: How much here? If any? polarizing vs non polarizing?** **brief overview of CEM?** **TODO: electrode impedance on injection vs non-injection measurements** **TODO: Discuss how and why measurement on injecting electrodes are**

typically discarded also talk about this in electrode section

2.4.2.2 Current injection and voltage measurement

To calculate impedance three main considerations must be made with regard to the injected current: the frequency, amplitude and pattern.

Low frequency currents will preferentially pass around cells through the extracellular fluid due to the high capacitive component of the cell membrane (Foster and Lukaski, 1996), whereas high frequency currents will be more affected by the capacitive component (Holder, 2004, December 31). Since EIT requires a linear relationship between the measured voltages and tissue impedance to produce valid reconstructions, it is important to select frequencies of less than 1 MHz to ensure that cell and neural membrane is not the dominant factor in the impedance measurements (Barber and Brown, 1984). Impedance measurements below 100 KHz have shown to be primarily resistive on lung tissue (Witsoe and Kinnen, 1967), and most EIT systems use frequencies between 10 kHz and 1 Mhz (Holder, 2004, December 31).

The selection of current amplitudes is selected based on the **TODO: Ask Andy**.

The pattern of current injection and voltage measurement is commonly referred to in EIT as the stimulation and measurement pattern. Typically a current is injected between two electrodes while the resulting voltages are measured between adjacent pairs. The two most common injection patterns are adjacent, and “skip 4”. Examples of these measurement patterns are shown in figure 2.4 on a 16 electrode system. The terms adjacent and skip 4 describe the space between pairs of injecting or mea-

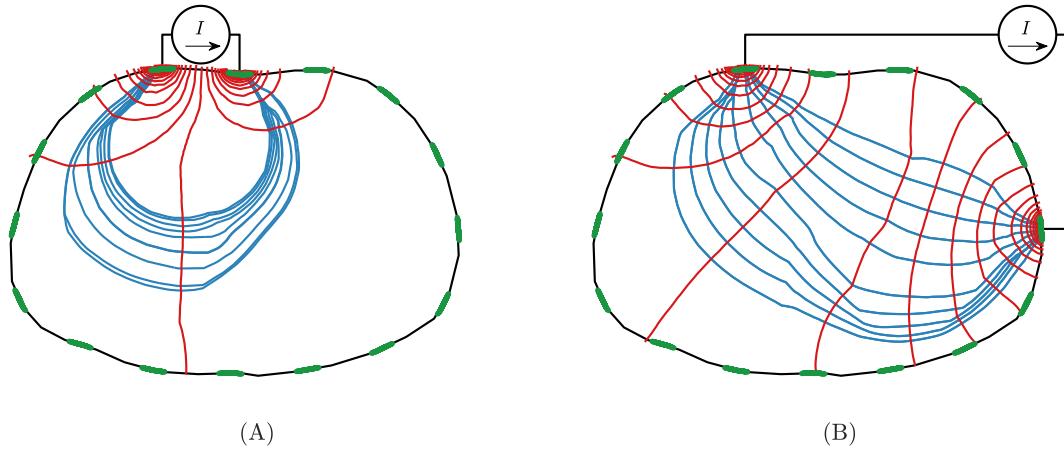


Figure 2.4: Two common stimulation patterns are simulated on a simple human model shown in figure 2.1. Injected current is shown in blue and the resulting equipotential lines are shown in red. (A) An example of an adjacent stimulation pattern. Current is injected between pairs of adjacent electrodes. For each current injection voltage is measured between all sequential pairs. (B) An example of the “skip 4” injection pattern. Current is injected between every 5th electrode skipping 4 between the injecting electrodes. during each current injection, voltage measurements are made using the same skip 4 pattern between each of the 16 electrodes and their corresponding pair. A single frame of data is generated by all measurements made for each possible injection pattern. For 16 electrodes this results in 256 meausurements.

surement electrodes. Regardless of the pattern, a frame of EIT data is generated in the same way. Current is injected between a pair of electrodes as voltage is measured between all remaining pairs, then current is injected between the next pair of electrodes and voltages are measured again. This results in voltage measurements on each electrode pair for every current injection. For a 16 electrode system this results in 256 measurements in each frame. For a 32 electrode system there are 1024. As mentioned in section 2.4.2.1, voltage measurements made on the stimulating electrodes are often removed resulting in 208 measurements per frame on a 16 electrode system, and 928 measurements per frame on a 32 electrode system.

2.4.3 Image Reconstruction

2.4.4 Forward Problem

Using the static formulations of Maxwell's equations, the potential distribution u can be used compute the conductivity ρ and generate images. We are able to use the static form of Maxwell's equations

2.4.5 Discretization and the Finite Element Method

2.4.5.1 2D reconstruction

2.4.5.2 3D Reconstruction

The majority of EIT measurements are done with a single ring of external electrodes but in practice, electrical current cannot be confined to a single plane and using a two dimensional electrode configuration can significantly impact the capabilities of EIT (**Rabbani1991**). The use of 3D electrode configurations in EIT was introduced in 1996 (**Metherall1996**) to overcome the inherent limitations of 2D measurements, but it is still not widely used today. It is thought this is due to the increased complexity of 3D images and the subsequent analysis (**Grychtol2019**).

It has been shown that externally placed 3D electrode configurations consisting of two electrode planes can improve the sensitivity distribution and image quality (**Grychtol2016**), but there is still limited sensitivity in the central-most regions of the chest. The concept of using internal esophageal electrodes has been presented previously (**Pilkington1989; Schuessler1995**) as a method to improve internal

sensitivity and reconstruction quality, but has not been widely used or simulated. Several studies have shown that there may be several advantages to using an internal electrode in EIT recordings; Measurements with an internal electrode have been shown to reconstruct images equally as well as configurations with twice as many external electrodes (**Schuessler1995**), and have shown an increase in sensitivity in a central region of interest (**Kwon2013**; **Czaplik2014**; **Farooq2014**).

2.4.5.3 GREIT

2.4.5.4 Image Regularization

2.4.6 Internal Electrodes

2.4.6.1 Motion Correction

2.4.6.2 Internal Reference Electrodes

2.4.6.3 Inverse Source localization (time permitting)

2.5 Perfusion monitoring

2.5.1 Contrast agent injection

2.5.2 Frequency Filtering

Monitoring lung ventilation is one of the most established clinical uses of EIT, presented initially by Barber and Brown (**Barber1984**). EIT has also been used as a tool to monitor blood perfusion (**Brown1992**) and hemodynamic parameters such

as cardiac output (**Braun2018**) and blood pressure (**Sola2011; Proenca2017**). While the spatial resolution of EIT is much lower than intermittent imaging techniques such as computed tomography (CT) or magnetic resonance imaging (MRI), EIT can have a high temporal resolution enabling continuous or frequent monitoring without concerns regarding radiation exposure. This thesis focuses on time difference EIT for thoracic hemodynamic imaging and monitoring applications.

EIT is sensitive to the movement of blood in two main ways. First, a conductivity-contrasting bolus solution injected into a vein or artery can be used to image the transit of blood through the body and second, the pulsatile changes in conductivity at the cardiac frequency can be isolated through digital filtering (**Leathard1994**). This document proposes methods and techniques to identify and isolate these impedance variations to monitor lung perfusion and aortic flow.

A perfusion scan is a technique for imaging the blood flow through the lungs and when compared with ventilation images can be used to detect pulmonary embolisms when a mismatch is identified. Clinically pulmonary perfusion is done using lung nuclear medical imaging such as single photon emission computed tomography (SPECT) (**Parker2012**). Radio-isotopes are inhaled through a mask for ventilation imaging and injected into the blood to image pulmonary perfusion. Images taken on a gamma camera are compared to look for a mismatch between the ventilation and perfusion distribution. This method of measuring lung perfusion is slow and exposes the subject to low-dose radiation.

EIT has been evaluated for its ability to measure cardiac output and lung perfusion since the late 1980s (**Eyuboglu1989; Blottt1992; Brown1992**;

Frerichs2002). Since then, various configurations of EIT have been evaluated (**Borges2012; Nguyen2015**). Due to the speed and safety of measurement acquisition, EIT might be used to continuously monitor perfusion in subjects.

There are two main challenges with perfusion monitoring using EIT. First, impedance change due to ventilation is 10 times larger than the impedance change due to cardiac-frequency pulsatile activity (**Deibebe2008**) and second, the pulsatile activity outside the lung region can overwhelm the lung perfusion signal (**Stowe2019**). There are several techniques available to mitigate the difference in magnitude such as: pausing ventilation; administering a conductivity-contrasting bolus through the heart and lungs via the jugular (**Frerichs2002**); and digital filtering to isolate activity at the cardiac frequency (**Leathard1994**).

When breathing is paused, the signals based only on the cardiac activity can be more easily extracted. It was found that during apnoea the global impedance recorded with EIT measurements corresponded with stroke volume measured using the thermodilution method with a pulmonary arterial catheter (**Fagerberg2009**). Ventilation perfusion ratios have been calculated during apnoea by comparing ventilation and perfusion signal amplitude with a specified region of interest (**Fagerberg2009a**). There is some concern that the perfusion measured during apnoea may not accurately represent true perfusion during regular respiration as the apnoea impacts the regular respiratory cycle (**Leonhardt2012**).

Using the conductivity-contrasting bolus injection EIT perfusion imaging has been compared to SPECT measurements, and blood flow has been imaged from the right heart into the lungs and back into the left heart using 5-10% hypertonic

saline (**Frerichs2002; Borges2012**). This technique is promising for imaging lung perfusion, but is slow and requires the placement of a venous catheter and repeated saline injections to obtain perfusion measures.

The final method to calculate perfusion using EIT is through filtering to isolate the cardiac related signal. Previous work has shown that principal component analysis (PCA) can be used to separate ventilation and cardiac frequency signals and identify the component related to the heart (**Deibebe2008**). Once the cardiac-frequency component of the EIT signal is identified the pulmonary component must also be isolated. Other than visual identification of the lung region or manually selecting a region of interest, there are few good solutions for isolating pulsatile activity within the lung region.

The use of 3D configurations to differentiation between pulsatile activity in the heart and lungs could allow for an improved perfusion measure using EIT, and a means of continuously monitoring perfusion during ventilation.

EIT has been used clinically to monitor lung perfusion in an animal model (**Leonhardt2012; Nguyen2012**), and it is theorized that the use of an internal electrode for increased sensitivity may allow for imaging of blood flow in the aorta. There is great interest in monitoring cardiac parameters using EIT to determine stroke volume (SV) (**Proenca2017; Braun2018**), and increased sensitivity close to the heart also has the potential to improve these measures.

While there have been some studies researching electrode placement for cardiac imaging in 2D (**Noordegraaf1996**) and 3D electrode configurations (**Graham2007**), there has been little research into determining the optimal

3D external electrode configurations for imaging the heart and aorta.

Additionally when using alternate electrode configurations the current injection and measurement patterns must also be investigated. It has been suggested that an internal electrode in 2D should not be used for current injection in asymmetrical models as the reconstruction performance deteriorates (**NasehiTehrani2012**). It is unclear to what degree injection patterns affect the resulting sensitivity when internal electrodes and alternate electrode arrangements are used in 3D.

This work aims to investigate internal and external electrode configurations for use in imaging blood movement in the thorax, and develop techniques to extract measures of aortic flow and lung perfusion from these reconstructions.

Chapter 3

Comparing bolus- and frequency-based perfusion

3.1 Summary

Objective: Two main functional imaging approaches have been used to measure regional lung perfusion using Electrical Impedance Tomography (EIT): venous injection of a hypertonic saline contrast agent and imaging of its passage through the heart and lungs, and digital filtering of heart-frequency impedance changes over sequences of EIT images. This paper systematically compares filtering-based perfusion estimates and bolus injection methods to determine to which degree they are related.

Approach: EIT data was recorded on 7 mechanically ventilated newborn lambs in which ventilation distribution was varied through changes in posture between prone, supine, left- and right-lateral positions. Perfusion images were calculated using fre-

quency filtering and ensemble averaging during both ventilation and apnoea time segments for each posture to compare against contrast agent-based methods using Jaccard distance score. *Main Results:* Using bolus-based EIT measures of lung perfusion as the reference frequency filtering techniques performed better than ensemble averaging and both techniques performed equally well across apnoea and ventilation data segments. *Significance:* Our results indicate the potential for use of filtering-based EIT measures of heart-frequency activity as a non-invasive proxy for contrast agent injection-based measures of lung perfusion.

3.2 Introduction

Electrical Impedance Tomography (EIT) uses electrical stimulation and measurements at electrodes on the body surface to reconstruct images of internal conductivity distribution and its changes. The most common application of EIT, experimentally and clinically, has been for imaging of the thorax (Frerichs *et al.*, 2017). Using a ring of electrodes around the chest, EIT is able to calculate images of impedance changes in the abdomen. Although most research has focused on imaging of ventilation, there is significant interest in imaging cardiovascular phenomena with EIT (Adler *et al.*, 2012; Leonhardt and Lachmann, 2012).

EIT has been evaluated for its ability to measure cardiac output and lung perfusion since the early 90s (Eyüboğlu *et al.*, 1989; Frerichs *et al.*, 2002; Zadehkoochak *et al.*, 1992). Since then, various configurations of EIT have been evaluated (Borges *et al.*, 2012; Nguyen *et al.*, 2015). The effect of posture on EIT images was evaluated

by Reifferscheid *et al.* (2011), who showed that changing posture introduces a large and reproducible variability into ventilation distribution as imaged by EIT. Based on results showing a common relationship between the effect of gravity and perfusion in both children and adults (Bhuyan *et al.*, 1989), in newborns we expect to see a comparable directional change in perfusion due to the changes in posture. Recently, Braun *et al.* (2018) evaluated EIT’s ability to monitor cardiac output, showing that EIT is more reliable for monitoring cardiac output trends than absolute cardiac output. EIT has also been investigated for monitoring of systemic blood pressure (Solà *et al.*, 2011), and for monitoring of pulmonary arterial pressure (Proença *et al.*, 2017).

EIT measurements are sensitive to blood movement in two main ways. First, it is possible to image the transit of the contrast agent through the heart and lungs via a conductivity-contrasting bolus into the veins and second, through digital filtering of the time series of EIT images at the heart frequency (Leathard *et al.*, 1994). While multiple EIT measures of perfusion are used, their relationship is not well understood. It is currently unclear to what degree pulsatile impedance changes represent blood flow, and how they limit the potential for heart-frequency filtering to correctly estimate the true perfusion (Nguyen *et al.*, 2012).

Injection of a contrast agent to measure regional lung perfusion has been compared with electron beam computed tomography (EBCT) and determined to be feasible for measuring perfusion across different animals (Frerichs *et al.*, 2002). Perfusion measurement via conductivity contrasts has the advantage of measuring the true perfusion, but requires placement of a catheter to introduce the contrast agent. Bolus-derived measurements cannot be made continuously because they rely upon

the circulation of a contrast agent. In addition, the accumulation of NaCl (the main conductivity contrast used) over multiple injections can lead to hypernatremia which limits the rate at which bolus injections can be made.

Calculating the heart-frequency conductivity changes in the thorax offers the benefit of a continuous functional measure calculated directly from EIT signals (possibly in conjunction with a synchronization signal such as the ECG). Heart-frequency EIT signals are typically an order of magnitude smaller than ventilation signals; thus, when measurements are made during tidal ventilation, a large period of data must be used in order to reduce the ventilation signal. On the other hand, measurements during apnoea can be used to eliminate the ventilation signal, but for the safety of the patient the apnoea was limited to 30 s. In healthy human subject of less than one year old it takes a mean of 118 s for the blood oxygen saturation levels to drop below 90% (Fu *et al.*, 1996), however the length of safe apnoea is much shorter for the sick preterm infant. The time period was chosen based on experience in the lab showing that 30 s seconds was not associated with bradycardia or desaturation to less than 90% blood oxygen saturation.

There is a debate within the EIT community about the meaning of heart-frequency EIT signals (Adler and Boyle, 2017; Frerichs *et al.*, 2017). Not all perfusion results in a cardiac-frequency change (for example, continuous blood flow in capillaries), and non-perfusion effects (for example, heart movement in the thoracic cavity) can result in heart-frequency EIT signals. This debate is reflected by the terminology – perfusion vs. pulsatility. Those who prefer “pulsatility” or “heart-frequency fEIT image” seek to emphasise that frequency filtered signals are not “perfusion” (al-

though they may be related). While these pulsatility based EIT images are clearly not a direct measure of perfusion, the signals appear to be useful and are often measured and reported (Bartocci *et al.*, 1999; Ericsson *et al.*, 2016; Halter *et al.*, 2008; Moens *et al.*, 2014). To the authors' knowledge, no systematic comparison of frequency-based perfusion measures has been published.

The heart-frequency signal can be derived from frequency filtering or ensemble averaging. Frequency-filtering uses a filter to isolate the frequency of heart-frequency conductivity changes, and was introduced by Zadehkoochak *et al.* (1992) and Leathard *et al.* (1994). Frequency filtering is susceptible to interference from ventilation when the heart rate is at a harmonic of the breathing rate. Ensemble averaging is another filtering approach which averages signals at a synchronized time, for example at the QRS peak (Bartocci *et al.*, 1999; Deibebe *et al.*, 2008). The impedance change due to each heart beat is aligned and averaged to give a single heart-related impedance change, representative of all heart-beats in the segment.

In this paper, we are motivated to better understand the relationship between lung perfusion and heart-frequency filtering measures, and between the various filtering approaches used to determine heart-frequency components. Our questions are: 1) to what extent do heart-frequency filtering-based measures correspond to perfusion, 2) what are the advantages and disadvantages of different approaches to heart-frequency filtering of EIT data, and 3) which techniques are recommended. In our experimental protocol, we have selected posture-change to introduce changes in the regional distribution of lung perfusion. These changes are then compared using bolus- and filtering-based EIT measures.

3.3 Methods

3.3.1 Overview

Data were acquired as an additional protocol within a study to determine a baseline for lung damage due to gas ventilation in neonatal lambs. This is part of an effort to establish total liquid ventilation (TLV) as a less-injurious ventilation strategy for the delicate lungs of neonatal subjects (Sage *et al.*, 2018). In order to induce changes in ventilation and perfusion patterns, posture changes were made between supine, prone, left and right lateral positions.

3.3.2 Animals

The study was conducted in accordance with the Canadian Council on Animal Care guidelines upon approval by the animal research ethics board of Université de Sherbrooke (protocol 417-17BR).

Seven healthy neonatal lambs (2–4 days old and 2.95 ± 0.27 kg) were used. Animals were anaesthetised (ketamin 10 mg/kg IM at induction followed by propofol 100 mcg/kg/min and ketamin 2 mg/kg/h IV) and placed under mechanical gas ventilation with: peak inspiratory pressure (PIP) 15 cmH₂O, positive end-expiratory pressure (PEEP) 5 cmH₂O, respiratory rate (RR) of 60/min, and fractional concentration of O₂ in inspired gas (FiO₂) of 30%.

A catheter was inserted into the carotid artery for blood gas and continuous blood pressure monitoring. A jugular venous access was inserted to inject the saline bolus for generating perfusion images. Each animal was shaved for placement of a custom

EIT belt around the lower third of the sternum in the transverse plane.

For each animal a bolus injection protocol was used: 1.5 mL of 7.5% saline was injected into the jugular vein at a constant rate over approximately 2s. Before each bolus, ventilation was stopped for ten seconds, and a further twenty seconds of apnoea was maintained before restarting ventilation.

After one hour of ventilation (for stabilization) EIT recordings were made during the position change procedure. Each lamb was rotated onto its right side. Five minutes after turning the subject, the bolus injection protocol was implemented. The animal was then ventilated normally, remaining on the right side for an additional five minutes, before being positioned on the left side for 5 minutes of regular ventilation, followed by the bolus injection protocol.

At 2 hours of ventilation, the position change procedure was repeated, changing the positioning of the lamb from prone to supine as the bolus injection protocol was repeated and EIT recordings were captured.

3.3.3 Data Acquisition and Image Reconstruction

EIT data was acquired with the Pioneer Set (Swisstom, Landquart, Switzerland) using a custom electrode belt (at an acquisition rate of 20 frames/s). The belt uses 32 brass electrodes equally spaced around the thorax, using an ultrasound gel to ensure good contact and minimise the contact impedance. The selected data in this study comes from lateral positioning changes recorded after 1.5 hours of ventilation and prone to supine positioning changes after 2 hours.

EIT images were reconstructed using GREIT (Adler *et al.*, 2009), which calculates

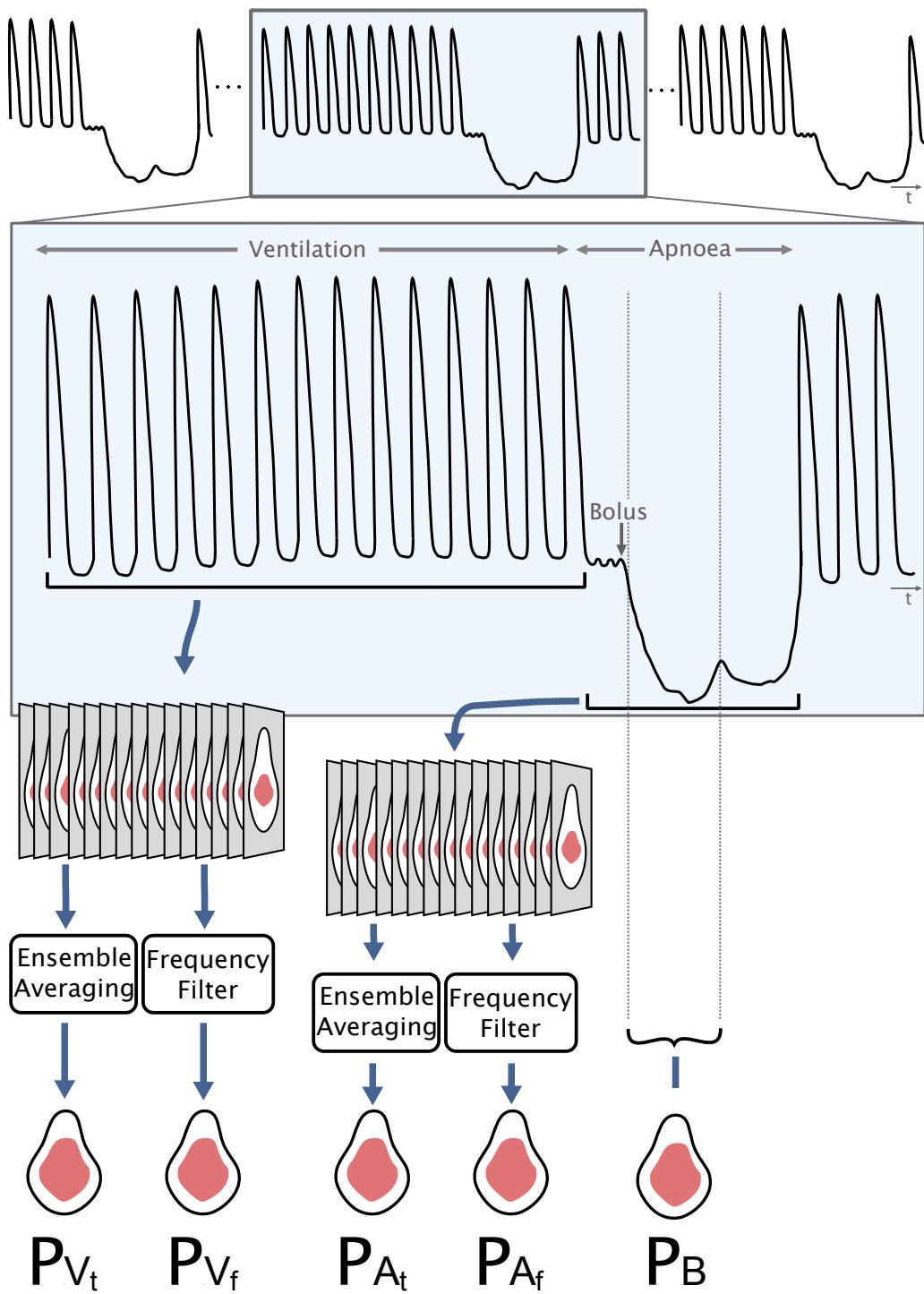


Figure 3.1: This figure is a schematic overview of analysis methods for EIT perfusion. The upper curve illustrates the global EIT signal during a period of ventilation followed by apnoea and renewed ventilation. During apnoea a bolus of conductivity contrasting saline is introduced. From these data 5 fEIT images are calculated: P_{V_t} : pulsatility (perfusion) image during ventilation, calculated by ensemble averaging EIT data during ventilation; P_{V_f} : pulsatility (perfusion) image during ventilation, calculated by frequency filtering EIT data during ventilation; P_{A_t} : pulsatility (perfusion) image during apnoea, calculated by ensemble averaging EIT data during apnoea; P_{A_f} : pulsatility (perfusion) image during apnoea, calculated by frequency filtering EIT data during apnoea; P_B : perfusion image from bolus, calculated between a reference measure during apnoea and one during the bolus

a reconstruction matrix \mathbf{R} from which the reconstructed image is calculated as $\hat{\mathbf{x}} = \mathbf{R}\mathbf{y}$, where \mathbf{y} are the time-difference measurements, $\mathbf{y}(t) = \mathbf{v}(t) - \mathbf{v}(t_r)$, where $\mathbf{v}(t)$ represents the data frame acquired at time, t , and $\mathbf{v}(t_r)$ measurements acquired at a “reference” time, t_r in the case of this experiment the reference was a mean of 10 images preceding the bolus injection.

The linear reconstruction matrix $\mathbf{R} = \mathbf{D}\boldsymbol{\Sigma}_t\mathbf{J}^T (\mathbf{J}\boldsymbol{\Sigma}_t\mathbf{J} + \boldsymbol{\Sigma}_n)^{-1}$ is calculated from a finite element model of the body and electrode geometry $F(\cdot)$ and covariance estimates of the image, $\boldsymbol{\Sigma}_t$, noise, $\boldsymbol{\Sigma}_n$ (Grychtol *et al.*, 2016), and a spatial filtering matrix, \mathbf{D} .

EIT data from this experiment was prone to errors consisting of brief periods of zeroed measurements stemming from the synchronisation equipment. Measurements that were zeroed by the device were removed and replaced with linearly extrapolated data to allow for frequency-based analysis over all selected segments of data. A moving median filter with a width of 3 was used to further remove the noise caused by single measurement errors in the signal.

3.3.4 Functional EIT Images

In each animal 4 episodes were recorded — one in each posture — to generate 5 different functional EIT images.

The images Bolus-based measures of lung perfusion (P_B) were calculated using time-difference reconstructions. Heart-frequency filtering during ventilation (P_{Vf}) and apnoea (P_{Af}) used frequency analysis of EIT image sequences, as illustrated in figure 3.3, and ensemble averaging-based methods during ventilation P_{Vt} and apnoea

P_{At} are calculated using ensemble averaging of identified pulsatile components figure 3.4.

The following methods were conducted on segments of data collected both during apnoea and ventilation. Apnoea regions were selected as the total time that ventilation was arrested, including the bolus section and had a duration of 30s. The ventilation data was selected as 30s of data immediately preceding the induction of apnoea. Regions of interest including lung, and heart areas in the images were defined by the lamb model provided in EIDORS (Adler *et al.*, 2017a).

3.3.4.1 Bolus injection image (P_B)

The beginning of the saline bolus injection was determined as the point immediately preceding the drop in impedance from the conductive agent, and is shown in figure 3.2 at the point marked “injection”. The mean of 10 images including and immediately preceding the bolus injection were used as the reference to which all bolus images were reconstructed from. To image perfusion, the point with maximum decline in impedance over the sum of the pixels in the lung region relative to the reference was selected based on the methods presented by Frerichs *et al.* (2002). In figure 3.2 this was found at the point marked “perfusion”. This method was used as the standard perfusion measuring technique against which the other methods were compared.

3.3.4.2 Frequency-Filtering

Heart-frequency EIT images during the selected events were calculated by taking the FFT of the time-series image data after first applying a Blackman window:

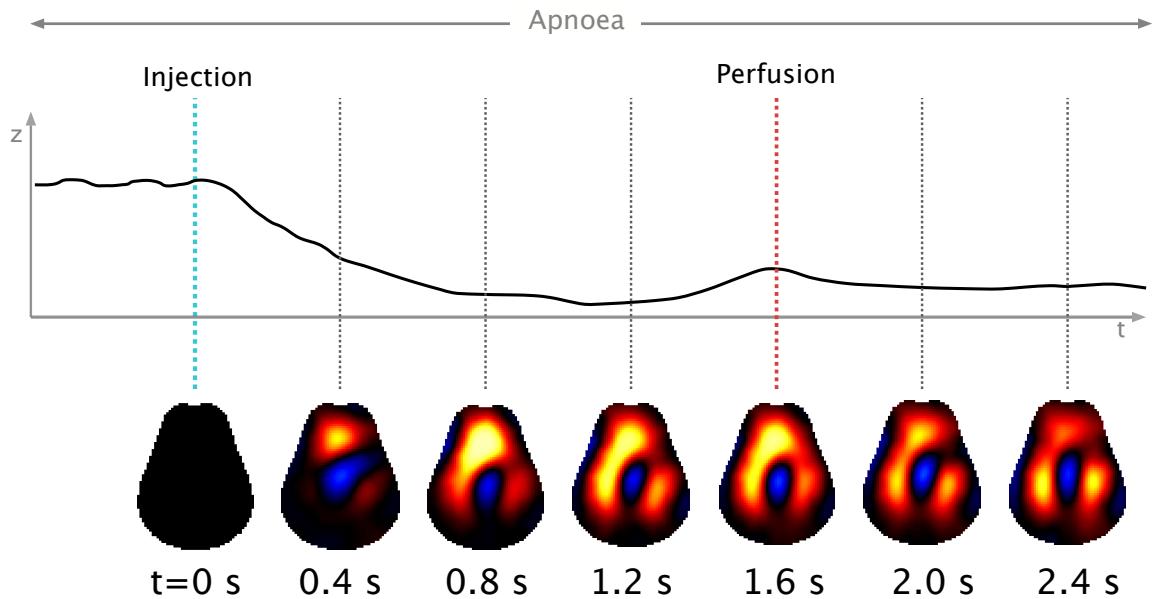


Figure 3.2: The method used to select the perfusion point from the bolus injection is shown in the figure above. The point with the widest spread of high conductivity was selected as the point of perfusion, shown here at 1.6 seconds after the contrast agent injection. The image series shows the conductivity contrast as the bolus injection travels through the thorax.

$w(n) = a_0 - a_1 \cos\left(\frac{2\pi n}{N-1}\right) + a_2 \cos\left(\frac{4\pi n}{N-1}\right)$ with $a_0 = 0.42$, $a_1 = 0.5$ and $a_2 = 0.08$, where N is the number of time-series EIT images in the selected event.

An FFT was calculated from a series of images restricted to pixels in the heart region. From the FFT of all pixels the heart region, the heart frequency was selected as the largest peak between 3 and 4.5 Hz, representing a heart rate between 180 and 240 bpm (typical for a newborn lamb).

The identified heart rate was used to select changes at the heart-frequency in the frequency domain images of the entire thorax. Images at 3 frequencies on either side of the heart rate were also reconstructed to account for changed in heart rate over the course of the data collection. A Blackman window with a length of 7 was applied surrounding the heart frequency to generate a weighted mean of the images, resulting in a single perfusion image from the heart-frequency data.

The output of the frequency filtering method is an image with complex values assigned to each pixel.

Depending on the timing of the pulsatility-based changes within the selected signal the real component of frequency analysed image did not correspond to the maximum conductivity change in the lungs in every event. In order to correct this, each image was displayed along the axis that gave the maximum real component contained within the lung region to ensure the maximum change in impedance related to pulsatile activity in the lungs was calculated.

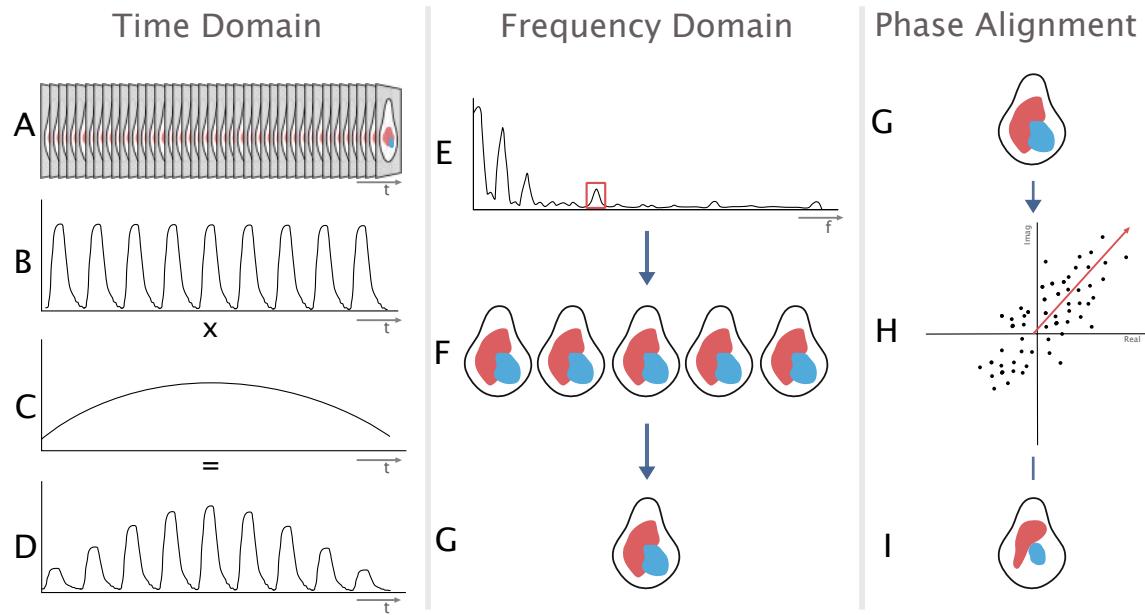


Figure 3.3: Frequency analysis methodology used for obtaining a perfusion image from the time series data. Steps are: A) to reconstruct the images from time series measurements; B) - D) window the time series data before performing a FFT on the data for each element; E) Select the dominant frequency between 3 and 4.5 Hz as the heart frequency; F) reconstruct the image at the heart frequency and selected nearby frequencies; G) take the mean of the images at the heart frequency using a Blackman window to give greater weight to those closer to the center; H) I) select the image that will give the maximum real component contained in the lung region.

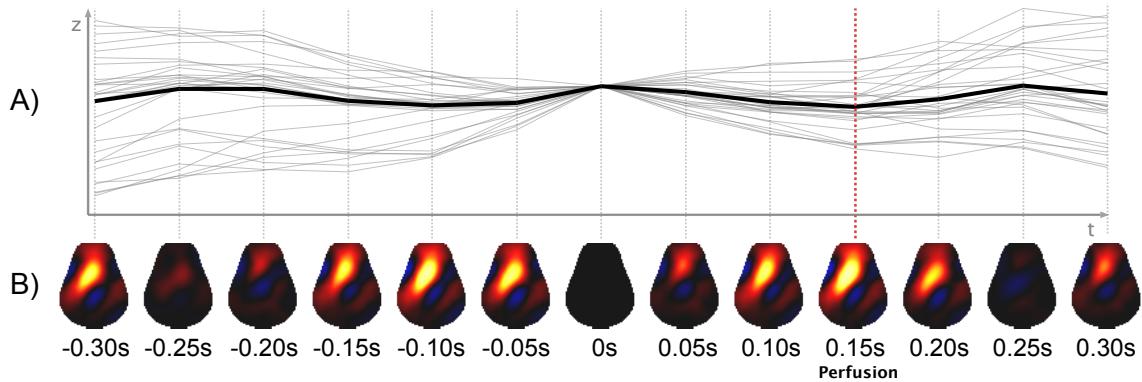


Figure 3.4: Illustration of the stages of the ensemble averaging process: A) an ensemble average of all heartbeats over the time frame is taken from the summed global signal; and B) shows reconstructed images corresponding to each time point in the global ensemble averaged signal above. The selected perfusion image is the image with the maximum impedance increase in the lung region.

3.3.4.3 Ensemble Averaging

Time series data of the total impedance signal for each pixel in the heart region was filtered using a bandpass filter to eliminate noise and breathing changes, and allow the heartbeat to be seen clearly in the signal. Peak detection was used on this heart-region data to select the amplitude peaks in impedance change signal at the heart frequency.

Using the identified time points, the global impedance change signal was ensemble averaged by overlaying all identified peaks to give an averaged heartbeat. 13 images were reconstructed over the course of the heart beat to select the image that resulted in the maximum positive increase impedance within the lung region. This process is outlined in figure 3.4.

3.3.5 Image Comparison

To compare the images the Jaccard distance between functional EIT images was calculated. Negative impedance changes were removed from the images and the images were normalized.

The Jaccard distance was calculated between the reference image calculated using the maximum increase in lung-region conductivity during bolus injection (b), and the frequency-based method (f): $J(x, y) = \sum_i \frac{\min(b_i, f_i)}{\max(b_i, f_i)}$ representing the distance between the two images.

3.3.6 Statistical Analysis

To determine the significance of the change in bolus between postures and methods, the Cohen's d score was calculated to quantify the effect size of the change in the centre of mass of the perfusion image (Cohen, 1977). This was calculated as the difference between two means over the pooled standard deviation. Where the difference between the two means is: $\mu_1 - \mu_2$, and the pooled standard deviation is:

$$\sqrt{\frac{(n_1-1)s_1^2 + (n_2-1)s_2^2}{n_1+n_2-2}}.$$

3.4 Results

The Jaccard scores for each method were compared between ensemble averaging and frequency filtering methods to determine the regions where performance was best for each method. Figure 3.5 shows a comparison between Jaccard distance for each animal, connecting lines indicate different methods performed on the same data

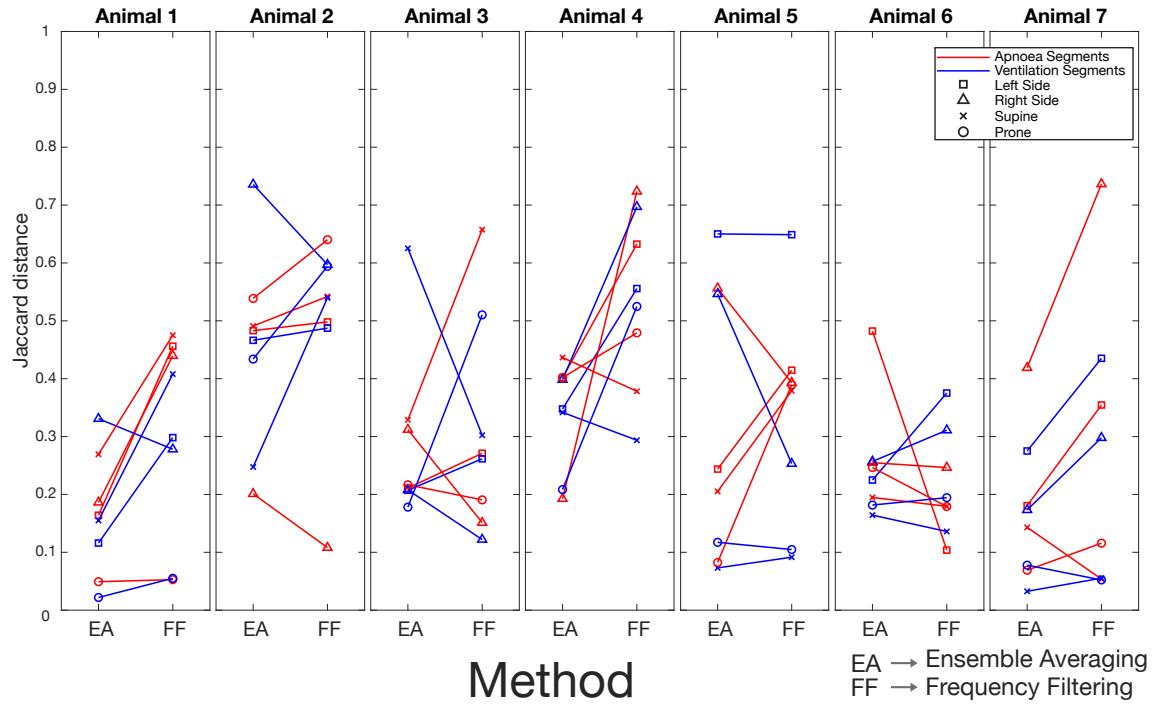


Figure 3.5: Jaccard scores for each method and animal in the comparison. Frequency filtering and ensemble averaging methods performed on the same data segment are connected by solid lines. Red lines and markers indicate apnoea data sections, while blue indicates ventilation data sections. Each posture is denoted by a different shaped marker in the figure.

segment, while each marker shape denotes a separate posture.

On average frequency filtering outperforms ensemble averaging based methods of perfusion calculation ($p=0.04$), and there is no significant difference in performance of the heart-frequency based filtering techniques during periods of apnoea relative to periods of ventilation.

Of the 56 data regions that were analysed, the ensemble averaging performed better in 12 cases and the frequency filtering achieved the best performance in 28 cases, there, were 16 additional cases where the difference in performance was negligible at less than 5%. On average, across all images, frequency filtering based methods

scored 7% higher than ensemble averaging.

The center of mass of the perfusion measure images using the bolus injection method had a Cohen's d score of less than 0.1 between posture changes indicating that there is an insignificant or trivial difference in the means relative to the standard deviation (Cohen, 1977). To demonstrate the visually observable changes due to posture change and the high similarities that can be observed between filtering- and bolus-based perfusion estimates, frequency filtered images from animal 4 are compared to bolus based methods in figure 3.6.

3.5 Discussion

Two primary approaches of EIT perfusion calculation have been compared in this paper: injection of a bolus of contrast-agent resulting in EIT image changes which produce perfusion measures, and digital filtering of EIT image sequences to extract the heart-frequency components. Additionally, various algorithms have been evaluated for digital filtering-base approaches during mechanical ventilation and short apnoea sequences, using both frequency- and ensemble averaging-based techniques. There have been few comparisons of these techniques, and we set out to better understand the relationship between perfusion and heart-frequency measures, and between the various filtering approaches used to determine heart-frequency cardiac changes. We selected an experimental protocol using posture-change to alter the regional distribution of lung ventilation and perfusion in newborn lambs.

Our first question was “to what extent do heart-frequency filtering-based mea-

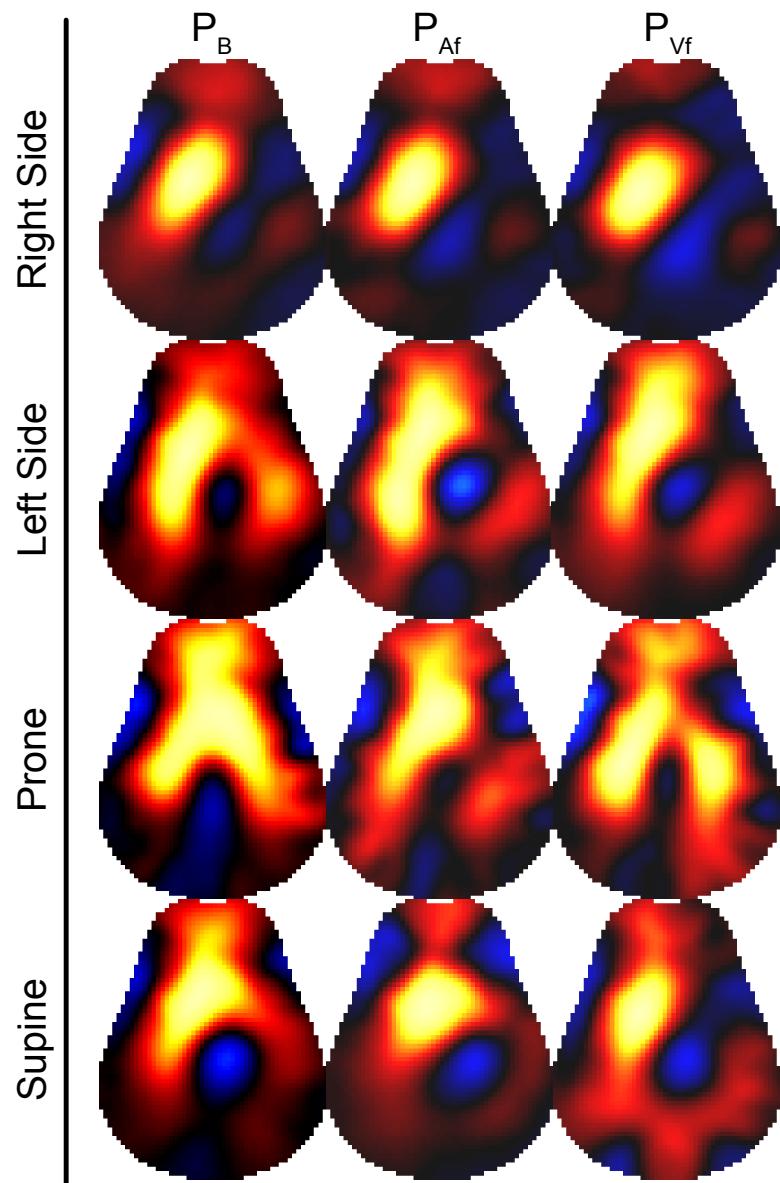


Figure 3.6: This figure shows the tracking of perfusion for frequency filtering measures of perfusion during apnoea and ventilation sections compared to bolus injection for animal 4. P_B is the bolus injection image, P_{Af} uses the frequency filtering method during apnoea and P_{Vf} is the frequency filtering method during ventilation.

sures correspond to perfusion?"

The primary results (figure 3.5) use a Jaccard index of the similarity between functional images. Overall it was found that in healthy animals the Jaccard index indicated good agreement with our gold standard. While highly dependant on the data, it was found that there was a high degree of similarity between methods with respect to the overall shape of the perfusion. In both animals 2 and 4, where the signal required little preprocessing before analysis there is a higher Jaccard score across all cases.

The synchronisation box was attached to the EIT system but was not used for this experiment, an error in the connection caused brief periods of the signal (less than 1 s) in some animals to be zeroed. Through careful processing of this signal only brief sections of data were lost and we do not feel this impacts the results.

During the experiment the order of posture change was not randomised. While changes in ventilation due to posture change are not understood to have long term physiological effects, if there is a longer term effect of change in posture the lack of randomisation will impact the results. Nguyen *et al.* (2015) were able to image perfusion changes due to induced pulmonary embolisms and using the peak impedance change on dilution curves, however our data presented insufficient variance in perfusion induced by posture change to complete a center of mass analysis. A higher statistical power could potentially be achieved through initiating posture changes with more dramatic results in perfusion, such as upright to supine (Nakazato *et al.*, 2010).

Throughout the experiment, the perfusion image was selected as the image con-

taining the largest increase in conductivity in the sum of pixels in the lung region, which occurred at different relative times across animals and events. Many factors could affect this including belt positioning changes, and it could be a contributing factor to the inconsistent trends in amplitude changes in the global image across methods. Borges *et al.* (2012) compared EIT perfusion images using first-pass kinetics and heart-frequency filtering based methods to perfusion measures using SPECT, finding that heart-frequency filtering techniques made systemic errors when used to estimate the perfusion. They also determined that there was no discernible relationship between the magnitude of the SPECT images and the heart-frequency images. This was consistent with the findings of this study that image amplitude of the bolus injection and heart-frequency filtering-based methods was not consistent in all animals. This methodology presented by Borges *et al.* (2012) was not part of the comparison in this study as the edentification of the perfusion signal due to the heart could not be consistently identified and removed across all animals. In two dimensions, heart-frequency and ventilation signals have been used to identify the location of the heart and lungs within the EIT electrode plane with known electrode locations and anatomy (Ferrario *et al.*, 2012), but in situations where the electrode location and anatomy is not precisely known EIT tends to perform poorly as a structural imaging modality (Adler and Boyle, 2017). These challenges suggest that configurations with multiple planes of electrodes may be better able to isolate and remove off-plane pulsatility signals related to the heart.

It was observed that the general shape of the perfusion was consistent across all methods despite amplitude variations. One reason for the difference in amplitude

change across animals may be due to slight variations in the belt placement and electrode positioning on the animals. If the belt is closer to the heart, there will be a larger heart-frequency component to the signal and there may be a variance in the impedance change due to bolus injection.

Next, we asked “what are the advantages and disadvantages of different approaches to heart-frequency filtering of EIT data, and which techniques are recommended under which circumstances?”

Our overall recommendation is that, whenever possible, frequency filtering techniques should be used. This is largely because frequency filtering methods tend to be more stable in the presence of noise on the signal. Ensemble techniques are advantageous in some circumstances, because they better use the heart-frequency variability to avoid interference from harmonics of the ventilation at the heart rate. For frequency-filtering techniques, it is necessary to widen the heart-frequency filters to account for such variability. On the other hand, it is sometimes not possible to accurately synchronize heartbeats, due to noise corruption in the signals or the very low amplitude of the heart-frequency signals relative to the ventilation signal. In cases where the signal of the heartbeat was not clearly identifiable through visual inspection of the signal, neither ensemble averaging nor frequency filtering was able to achieve good estimates of perfusion relative to the bolus injection event.

In summary, our goal was to understand the relationship between bolus- and filtering-based EIT measurements of lung perfusion, as well as the relationship between different filtering-based measures of perfusion. Our results indicate there is a common trend between the shape and perfusion estimates of both heart-frequency

and bolus injection images despite the difference in physiological events behind each measure. Amongst filtering techniques, frequency filtering outperforms ensemble averaging across regions of data where there is noise present and the heart signal cannot be readily identified, and both methods were able to approximate the bolus injection measures equally well when applied to apnoea and ventilation regions of data.

Chapter 4

FEM mesh refinement for 3D Electrical Impedance Tomography

4.1 Summary

In this paper we examine the requirement for mesh refinement around electrodes in Electrical Impedance Tomography (EIT). While it has been recommended that models be refined around the electrodes, where current density and sensitivity are highest, the level of refinement required is poorly understood. Using a set number of nodes, we investigate the optimal distribution between the electrodes and the volume of a model. A balance point is used to measure the difference in distribution between the electrode and the centre of the model. To calculate this, all nodes contained between the surface of a selected electrode and the centre of the model were identified and the mean position of nodes along the container axis was computed.

We compare refinement strategies across commonly used meshing software in EIT and compare the model sensitivity error to an ultra-fine reference mesh. In a tank model, for a fixed number of nodes, error in the sensitivity calculation is minimized when the balance point of the nodes is at 85% of the tank radius and the node density dissipates evenly from the electrode surface to the centre of the model. Using this method sensitivity error was decreased in all regions with high sensitivity. This node distribution technique enables the generation of accurate meshes with fewer nodes that can reduce measurement error and computing time.

4.2 Introduction

Electrical Impedance Tomography (EIT) reconstructs images of electrical tissue properties within a body from electrical transfer impedance measurements at surface electrodes. For biomedical imaging applications, it is being actively studied for monitoring the movement of air and blood in the thorax, and for imaging the head and breast. Reconstruction of EIT images requires the solution of an inverse problem in soft field tomography. EIT image reconstruction requires calculation of a sensitivity matrix, \mathbf{J} , representing the relationship between internal changes and measurements. A pseudo-inverse of \mathbf{J} is used to update the image estimate over several iterations. EIT image reconstruction is ill-posed, since the physics of current propagation implies that sensitivity is largest near the electrodes and smallest in the body centre.

It is therefore clear that a precise calculation of \mathbf{J} is required for solution accuracy. Since it is generally not possible to use analytic solutions, because of the non-regular

shapes of biological bodies and the boundary conditions on a conductive electrode, the finite element method (FEM) is typically used. One key advantage of the FEM is that element size can be selectively refined in regions to meet solution accuracy. The accuracy of the FEM solution will increase as more elements are added, so a high mesh density is often desired to achieve an accurate solution. In this paper we will use the term mesh to refer to a specific combination of nodes and elements in a finite element model. In EIT the sensitivity is nonuniform across the entire model. Thus it has generally been recommended in the EIT literature that meshes be refined near electrodes, where the electric field and sensitivity are largest (Adler and Boyle, 2017). This recommendation gives rise to two questions: 1) No thorough analysis has been made to determine how much refinement is required. Given a “mesh element budget”, what should balance of nodes be between the centre of the model and the electrodes? And 2) How do different freely available meshing tools that are commonly used with EIT compare when used to refine 3D meshes?

Previously with EIT, mesh refinement has primarily been either constant, or based on the complexity of geometric surfaces and lines within a model (Grychtol and Adler, 2013). In EIDORS (Adler and Lionheart, 2006) meshes are generated using both Netgen (Schöberl, 1997) and Gmsh (Geuzaine and Remacle, 2009) for 2D and 3D models. Refinement around electrodes is commonly performed by setting a mesh density for the electrodes and allowing the mesh density to decay towards the maximum mesh size. This does not allow the user to specify the rate of decay or precisely control the mesh size.

A model that accurately represents the anatomy of the imaged region can greatly

increase the quality of the reconstructed image (Grychtol *et al.*, 2012), but increasing the complexity of mesh surfaces presents additional challenges for mesh refinement. EIT reconstruction software EIDORS enables users to place electrodes on the surface of complex boundaries (Grychtol and Adler, 2013), but the current functionality does not enable control of the refinement around the electrodes or internal structures. Most commercially available FEM packages do not conveniently provide such capability either.

In this paper we investigate approaches to manage the tradeoff between refinement of the electrode regions versus the bulk volume. We present a comparison between Gmsh and Netgen based mesh refinement around electrodes, and evaluate the effect of mesh refinement techniques on error in the sensitivity matrix, \mathbf{J} .

4.3 METHODS

4.3.1 Overview

We built a cylindrical model in Gmsh and Netgen which was parameterized so that multiple different combinations of mesh refinement were possible. These results were compared to a very high density meshes which was considered the gold standard.

4.3.2 Mesh Generation

A cylinder ($\varnothing = 0.5$ m, height $h = 0.25$ m) with four square electrodes (5 cm edge length) placed equidistantly around the perimeter at mid-height was meshed with Netgen (version 5.3.1) (Schöberl, 1997) and Gmsh (version 4.7.0) (Geuzaine and

Remacle, 2009) meshing software. Current was injected between adjacent electrodes and the voltage was measured between the remaining two electrodes. For 3D meshes an initial analysis was done building on work from Grychtol and Adler (Grychtol and Adler, 2013) where mesh density was set by specifying the maximum edge lengths permitted on electrode surfaces and in the volume of the FEM. Results were compared against those generated using ultra-fine meshes. Calculations were performed with EIDORS (version 3.10) (Adler and Lionheart, 2006) in Matlab 2019b (The Mathworks, Natick, MA, USA).

Meshes of different sizes were generated with Netgen and Gmsh by manipulating the desired maximum edge length (`maxh` parameter) for the entire domain and the electrodes. Two mesh analyses were performed. For the first mesh maximum element lengths were chosen such as to divide the electrode side of 5 cm into an integer number of segments of equal size. The maximum mesh element length ranged from 1 to 7 subdivisions of the electrode edge, while the maximum mesh element length in the ultra-fine reference mesh was 15 subdivisions per electrode edge. Independent reference meshes were generated for each software. Two types of models were generated this way. Constant models C1–C7, where the mesh size was constant, and refined models R1–R7 where the electrode mesh size was specified and dissipated towards an internal mesh element size of 5 cm. Additional refined meshes were generated with reduced mesh size in the internal mesh regions. R1-R7 is referred to as refinement A where the internal element max size was 5 cm. Refinement B had an internal mesh size of 4 cm, C was 3 cm and D was 2 cm. The numeric value in the mesh ID indicated the number of subdivisions per electrode edge. In Netgen the mesh

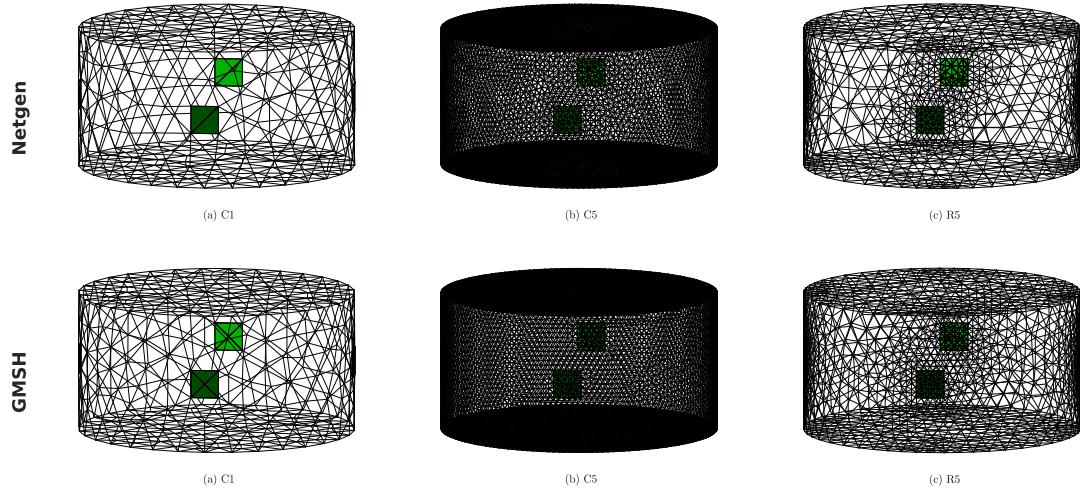


Figure 4.1: Sample meshes generated with Netgen (top row) and Gmsh (bottom row). From left to right: (C1) the coarsest constant mesh; (C5) a refined constant mesh; and (R5) a refined mesh with the same electrode mesh density as C5 but lower internal mesh density.

decay was not controllable, but in Gmsh the size was set to increase evenly from the surface of the electrode to the centre of the model. figure 4.1 shows example meshes of coarse, fine and refined meshes. figure 4.2 shows the generated mesh structure for constant refinement meshes around the electrode for both Netgen and Gmsh.

For the second analysis, the distribution of nodes within the model was changed without altering the total number of nodes to give M1 – M17. Starting with the constant mesh C3 as M1, the maximum mesh element length on the electrode was decreased by 10% and the maximum mesh size in the centre was increased so that the total number of elements in the mesh was within 10% of the original mesh. C3 was chosen as the starting point because several steps of mesh refinement could be generated before the electrode mesh density surpassed the reference meshes. For mesh M17 the specified electrode refinement was equal to the reference mesh. In

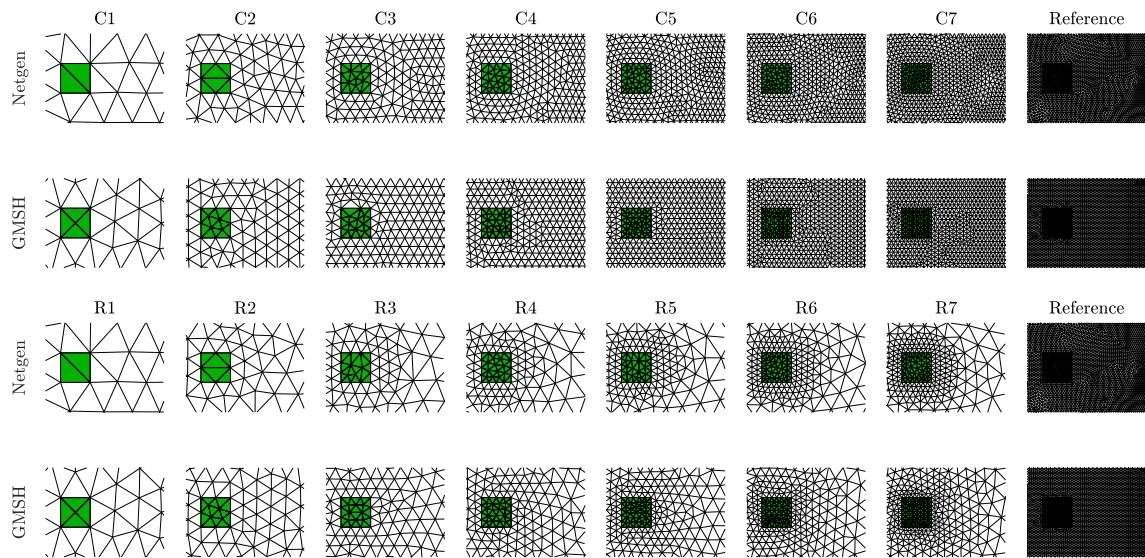


Figure 4.2: A view of the electrode meshing for all constant-density meshes in Netgen and Gmsh. The top two rows show all electrode faces and immediate surroundings from coarsest (C1) to finest (C7). C represents the constant mesh refinement and the number represents the specified mesh subdivisions per electrode edge. The reference mesh is equivalent to C15. The bottom two rows show refined meshes R1 to R7 with both Netgen and Gmsh and shows the rate of mesh dissipation away from the electrodes.

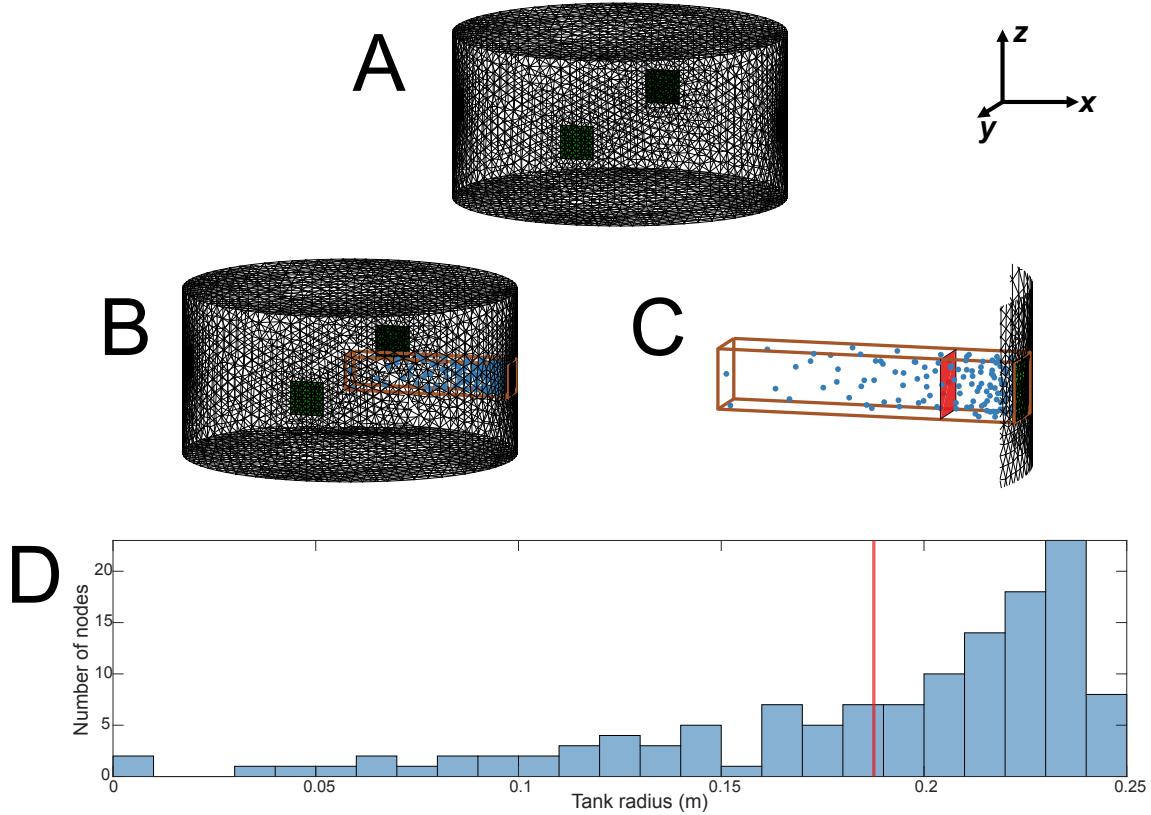


Figure 4.3: A sketch of the process to determine the balance point of generated meshes. A) the starting mesh; B) nodes between the electrode surface and the centre of the model are identified; C) the balance point of the nodes along the x-axis is calculated and indicated by the red plane; D) a histogram showing an example distribution and balance point (red) for the selected model.

Netgen the mesh dissipation rate was not further controlled, and in Gmsh the mesh density decreased evenly from the electrode surface to the centre of the model. To compare these meshes a section of the model was selected encompassing all points between the centre of the model and a selected electrode face. The average distance, or balance point, along the x-axis of the selected points was expressed as a percentage of the tank radius. This process is illustrated in figure 4.3.

When generating meshes to compare across several mesh density profiles as the

balance of the nodes was shifted towards the electrodes, 19 meshes were generated for each software including 2 reference meshes. table 4.1 shows the parameters of the resulting odd numbered meshes.

4.3.3 Simulation

The potential at each node \mathbf{V} of the mesh was calculated using the finite element method (FEM) using the linearization

$$\mathbf{V} = \mathbf{Y}^{-1}\mathbf{C} \quad (4.1)$$

where \mathbf{Y} is the admittance matrix of the FEM (and a function of conductivity distribution) and \mathbf{C} is a matrix representing the current injection pattern, such that \mathbf{C}_{ij} represents the current injected in electrode i during the j -th stimulation. Here, we drive current of 1 A between two adjacent electrodes in a single stimulation, so $\mathbf{C} = [0|0|1|-1]^T$. We pick a node in the centre of the FEM as ground, since it is necessary to assume the potential on one node for \mathbf{Y} to be invertible. We use the complete electrode model and assume contact impedance of 0.01 Ω in the calculation of the admittance matrix (Polydorides and McCann, 2002).

We calculate the sensitivity (or Jacobian) matrix \mathbf{J} of measurements \mathbf{v} to changes in the conductivity σ of individual elements as $\mathbf{J}_{ij} = \frac{\partial v_j}{\partial \sigma_i}$ using the adjoint method (Polydorides and McCann, 2002). Again, since we only have one measurement, \mathbf{J} is in fact a vector. We construct a sensitivity image by assigning each element i of the FEM the value of \mathbf{J}_i divided by the element's volume. Mean sensitivity in

Table 4.1: Mesh parameters for odd numbered meshes generated by Netgen (A) and Gmsh (B) to determine the optimal node balance. Parameters global maxh and electrode maxh refer to the specified input parameters; the remaining columns give parameters from the resulting meshes.

Mesh ID		lbl.	elec.	# elem.	# nodes	# elec. elem.	minEL ^a [mm]	maxEL ^b [mm]	minEV ^c [mm ³]	maxEV ^d [mm ³]
		maxh [mm]	maxh [mm]							
M-01	A	16.67	16.67	31347	7095	22	9.80	49.45	254.76	6851.01
	B	16.67	16.67	49210	9615	25	10.32	50.00	222.87	2898.59
M-03	A	18.33	15.00	29639	6482	22	10.75	50.41	289.80	5826.14
	B	18.33	15.00	49247	9680	40	7.36	37.11	172.78	2814.55
M-05	A	18.33	13.33	29814	6589	28	9.39	49.91	162.26	5648.41
	B	20.00	13.33	50749	9930	42	7.80	37.93	134.41	3233.59
M-07	A	18.33	11.67	30581	6723	36	8.77	47.88	141.74	6252.36
	B	21.67	11.67	53002	10429	60	6.17	40.84	63.22	4077.18
M-09	A	18.33	10.00	30690	6755	42	7.86	49.18	115.45	5496.39
	B	23.33	10.00	56237	11008	68	5.82	43.81	62.88	4962.89
M-11	A	18.33	8.33	31575	7030	74	6.05	50.99	60.06	6086.88
	B	26.67	8.33	55545	10886	96	5.51	49.72	36.84	7424.70
M-13	A	20.00	6.67	28589	6447	92	4.65	51.85	20.68	6664.11
	B	30.83	6.67	54993	10825	148	4.51	55.36	20.63	10453.90
M-15	A	21.67	5.00	27775	6245	158	3.46	52.60	11.13	9097.30
	B	36.67	5.00	55331	11000	250	3.51	61.66	7.99	15838.09
M-17	A	30.00	3.33	39116	7590	320	1.75	72.83	1.13	23783.17
	B	48.33	3.33	52947	10798	548	2.32	86.60	2.72	32287.34
REF	A	3.33	3.33	6661789	1173243	510	1.75	9.49	1.01	46.09
	B	3.33	3.33	5871464	976558	554	2.10	7.59	1.58	21.65

a: minimum mesh edge length, *b*: maximum mesh edge length

c: minimum mesh element volume, *d*: maximum mesh element volume

the plane of electrodes is then calculated by averaging the sensitivity in fifteen planes parallel to the plane of electrodes and spanning the height of 5 cm. The sensitivity was projected onto a 512×512 array and divided into regions of interest for the centre (C), at the electrode (E) and between the centre and electrode (I). The resulting sensitivity for the reference mesh calculated with Gmsh and the selected regions of interest is presented in figure 4.5.

4.3.4 Electrode refinement for arbitrary FEMs

Our approach for refinement around electrodes in Gmsh with external electrodes also allows for the refinement of arbitrary models with complex structures such as internal electrodes and tissue boundaries. A scenario depicting an approximation of a probe entering a bone with different layers of conductivity. The resulting mesh pictured in figure 4.7 highlights the ability of this technique to be used for refinement around electrodes and the control of mesh density surrounding internal structures which was previously very challenging in EIT software.

4.4 Results

Two analyses of mesh refinement were completed. The first comparing sensitivity error between meshes with constant refinement and refinement only at the electrodes, and the second comparing meshes with different levels of electrode refinement and the same number of nodes.

When comparing constant meshes to meshes with refinement at the electrodes,

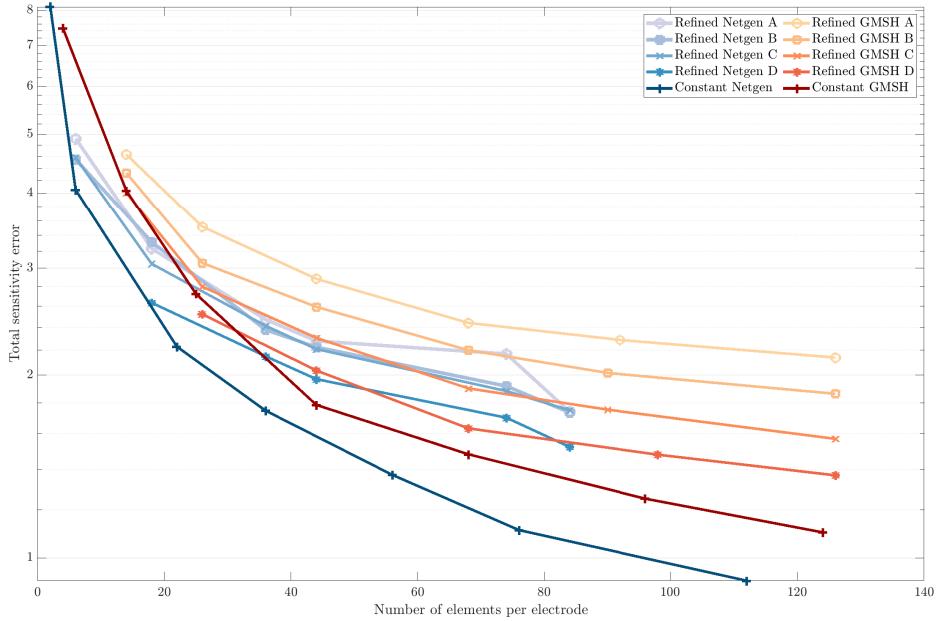


Figure 4.4: Sensitivity error of each mesh as a function of the number of elements per electrode for both Netgen and Gmsh. The darkest lines indicate the constant mesh refinement, lighter lines indicate a larger maximum internal mesh size. The maximum internal mesh sizes are as follows: refinement A - 5 cm; refinement B - 4 cm; refinement C - 3 cm; refinement D - 2 cm.

sensitivity error was decreased as more nodes were added to the mesh and to the electrodes. The sensitivity error was lowest in the constant meshes across both Netgen and Gmsh software. Meshes generated using Netgen provided a slightly lower sensitivity error relative to the respective reference mesh compared to Gmsh, and resulted in meshes with fewer nodes per electrode given the same input parameters. figure 4.4 shows the sensitivity error between constant and refined meshes with respect to the number of nodes per electrode.

Example sensitivity profiles for the M-series meshes are shown in figure 4.5. The

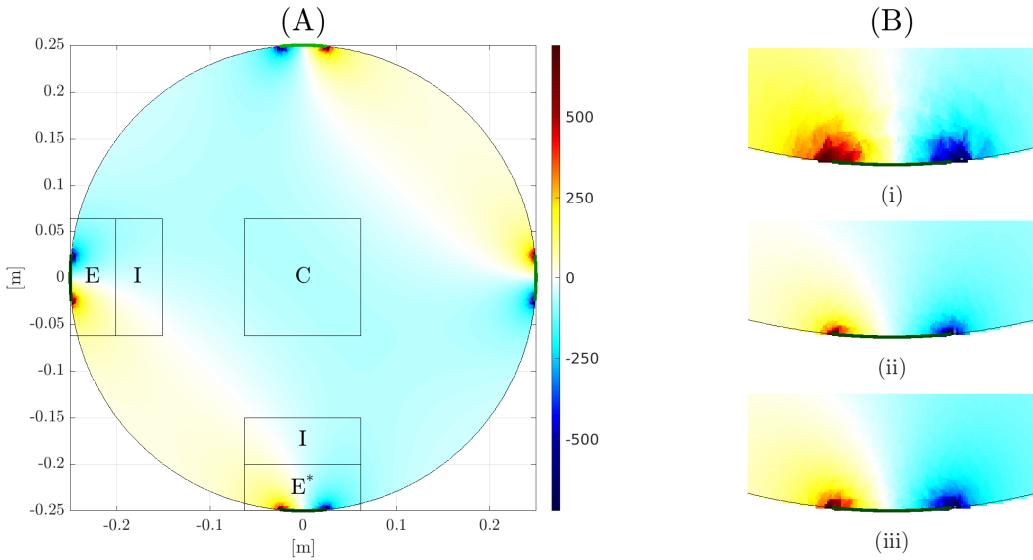


Figure 4.5: (A) Sensitivity distribution for the reference mesh (C15) generated in Gmsh with regions of interest used to compare between models. (B) 3 sensitivity distributions in region E^* next to the electrodes: (i) Constant mesh M1 (ii) refined mesh M15 with a balance point of 82% (iii) reference mesh from (A).

resulting sensitivity profile near the electrodes more closely matched the reference case when refinement at the electrodes was higher.

The total sensitivity error across all meshes is plotted vs. the balance point in figure 4.6. For meshes generated with Gmsh the minimum error was achieved when the node balance point was approximately 85% of the model radius corresponding to model M15-B, and for Netgen generated meshes the minimum sensitivity was achieved in model M13-A at a balance point of approximately 70%. Gmsh achieved a lower sensitivity error measured against the respective reference mesh. For meshes using Netgen refinement, the balance point did not increase evenly as the electrode density was increased and the internal density decreased. To maintain the same number of nodes within the model, Gmsh required a larger internal maxh than

Netgen. Gmsh generated meshes with more nodes for the same input parameters, but generally the resulting mesh sizes were closer to those specified. The resulting mesh parameters for odd numbered meshes can be seen in table 4.1. Across all meshes the measurement error when computing the voltage measurements was insignificant at less than 0.2%.

4.5 Discussion

We consider several questions on the requirement of FEM refinement in the neighbourhood of electrodes and the available tools for mesh refinement in EIT. 1) Given a “FEM element budget”, what should balance of nodes be between the centre of the model and the electrodes? 2) How do different freely available meshing tools that are commonly used with EIT compare when used to refine 3D meshes?

While refining meshes surrounding the electrodes is agreed to be useful, there is a lack of systematic analysis of the required refinement level, and controlling such refinement is difficult. Automatic mesh refinement is an area of active work and there are a number of commercial and free products available. We compare two programs used widely in EIT. Our results show that Netgen and Gmsh control mesh refinement differently and the same input parameters result in meshes with different numbers of nodes. figure 4.2, depicts the difference in mesh dissipation rates between Netgen and Gmsh. The mesh size in Gmsh increased gradually from the surface of the electrode towards the centre of the model, where the mesh size in Netgen increased much more quickly from the edges of the electrode. While we attempted to control

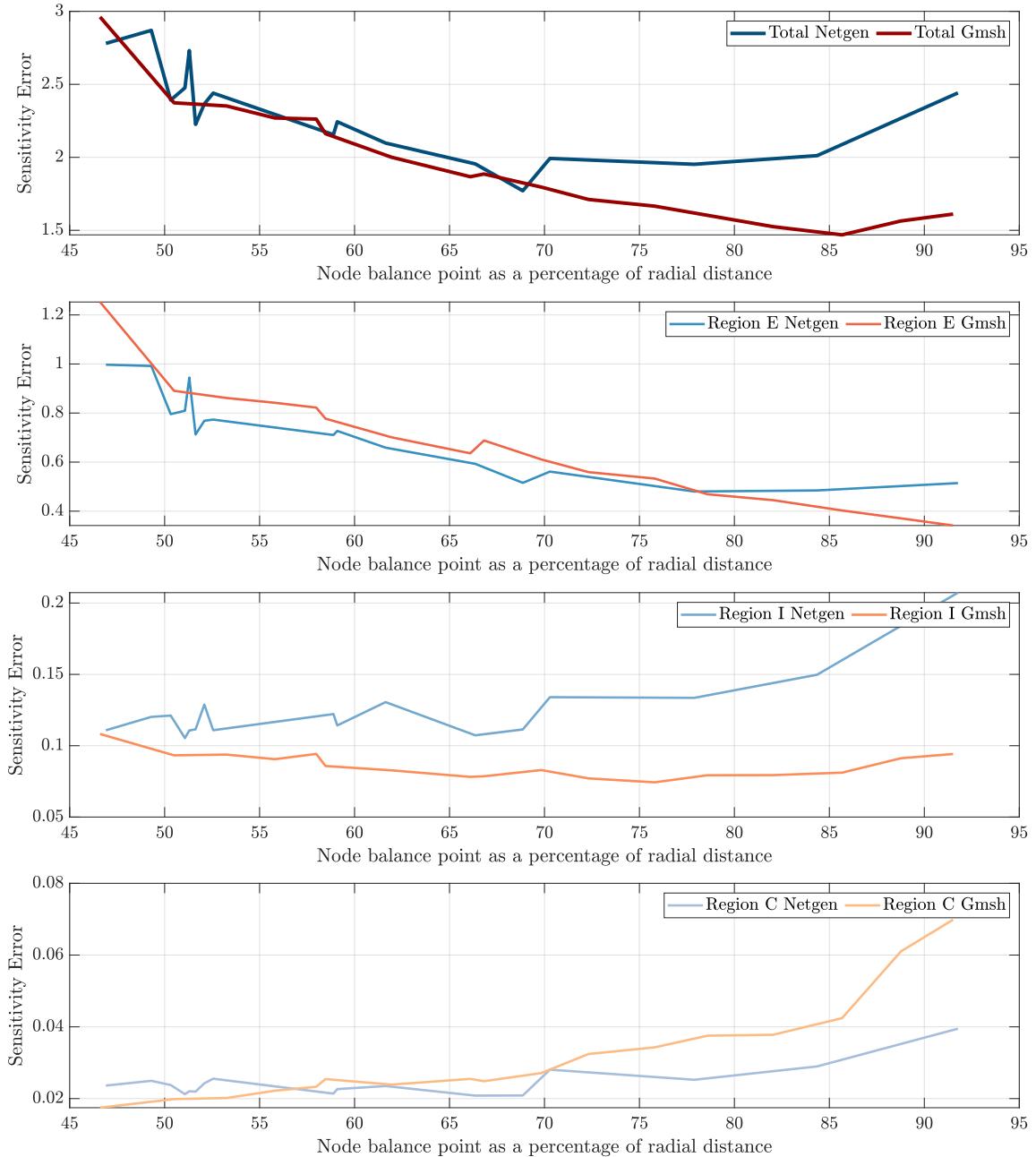


Figure 4.6: Resulting sensitivity error for Netgen (blue) and Gmsh (red) as the balance of the nodes was shifted towards the electrodes. From top to bottom: total sensitivity error, the sum of sensitivity error in region E, the sum of sensitivity error in region I and the sum of sensitivity error in region C. The regions are described in figure 4.5.

the dissipation rate in Netgen by manipulating the mesh density in the centre of the model, we were unable to achieve a smooth transition between the electrode and internal regions of the mesh.

To analyze the benefit of electrode refinement and the difference between Netgen and Gmsh refinement techniques we consider a sequence of refined meshes compared to a “gold standard”, uniformly fine FEM solution. The models were refined either globally or in the electrode neighbourhood, and the error in the sensitivity matrix \mathbf{J} was compared. figure 4.4 displays the difference in sensitivity between constant meshes and meshes with refinement at the electrodes. The sensitivity error was lower in Netgen across all constant refinement meshes, in part because there were more total nodes than Gmsh meshes with the same number of nodes on the electrode. Refinement around the electrode decreased the total sensitivity error, but still had larger error than meshes with constant refinement due to the smaller number of nodes in the model.

Using the balance point analysis, we were able to determine the optimal distribution of nodes to minimize sensitivity in both Gmsh and Netgen. The minimum total sensitivity from figure 4.6 was approximately 70% for Netgen. This was mainly due to the rapid dissipation of node density away from the electrodes. Increasing the refinement near the electrode reduced error in region E, but in region I there were insufficient nodes to reduce the sensitivity error. As the balance of refinement approached 90% the error in region E also started to increase as the node density did not remain fine throughout the entire region. This effect can also be seen in figure 4.2. In Gmsh the optimal balance for refinement was at approximately 85%

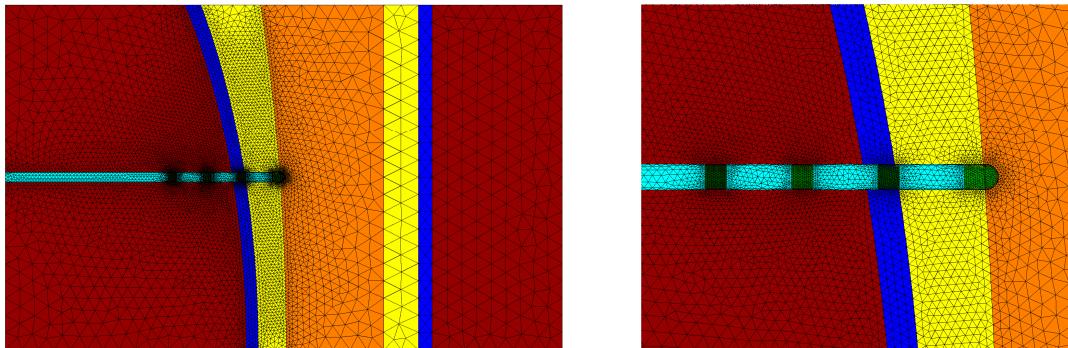


Figure 4.7: Example FEMs of a probe entering a bone from the surrounding tissue. Refinement is specified around the electrodes and tissue interfaces near the probe.

of the radial distance. Since mesh density was set to reduce evenly between the electrode surface and the centre of the model, there was a higher density of nodes that was maintained in regions E and I as the electrode refinement increased. The error in the centre of the model was higher in Gmsh meshes, but since this is where the sensitivity is lowest the total sensitivity error was much lower.

The ability to selectively control the mesh refinement in regions in Gmsh also allows users to generate complex meshes and control mesh density surrounding internal structures and electrodes an example mesh is shown in figure 4.7 where a model was created of a probe entering a bone from the surrounding tissue. These additions fill an important need in EIT to allow for more accurate models of regions surrounding internal structures and electrodes, and

Despite the increased ability to control refinement, the results still show that there are discrepancies between the specified parameters and the resulting meshes. The data in table 4.1 show that the maximum element lengths specified for each mesh were different from the actual maximum edge lengths. As the node density

was increased near the electrodes, in Netgen the balance point did not always shift towards the electrode as the maximum node density was not always higher despite specifying a smaller mesh size. In Gmsh the mesh density was closer to the specified values, typically resulting in more nodes in the refined meshes.

Across both analyses errors away from the refined areas may be higher, but the ability to refine meshes selectively near regions where high sensitivity is required may allow for reduced measurement error while still allowing for quicker meshing times. As more electrodes are added and the model complexity is increased, we expect that refinement around the electrodes will continue to reduce total sensitivity error. However the node balance analysis will not be possible for irregularly shaped models. The node balance analysis provides a straightforward method to determine the ideal placement of a given number of nodes to reduce model errors.

4.6 Conclusion

In summary, as expected, refinement of meshes near electrodes does improve model accuracy in terms of sensitivity. We recommend that, for each EIT imaging case, required model accuracy be determined from an analysis of the system, and to minimize the sensitivity error in the forward solution the balance of the nodes should be approximately 85% towards the radius of the model when the dissipation of mesh refinement is constant.

Chapter 5

Custom EIT Meshes

5.1 Introduction

With EIT it is suggested that more accurate models make a more accurate image...The more closely

5.2 EIT for ARDS Patients

5.3 A Segment Editor Program

5.3.1 Automatic segmentation of the thorax

Algorithm 1: Segment the external body boundary.

Input: image

Output: external boundary

- 1 weiner filter;
 - 2 Set the lung intensity to 0;
 - 3 erode image using disk size 20;
 - 4 reconstruct on image from line 4;
 - 5 dilate with disk of size 20;
 - 6 reconstruct on image from line 6;
 - 7 binarize, thresh = 0.5;
 - 8 fill holes;
 - 9 close - disk of size 2;
 - 10 open using disk size 5;
 - 11 external boundary = largest object;
-

5.3.2 Manual Segmentation Correction

5.3.3 Mesh Generation

5.3.3.1 Extruded Geomerty

5.3.3.2 3D Lung Regions

5.4 Evaluation on ARDS Patients

5.4.1 Methods

5.4.2 Results

Data from 4 ARDS patients with CT and EIT were used to develop a segmentation and correction tool to identify the lungs and boundary of the body. Segmentation was done using the 4th intercostal space, with 10 adjacent CT slices to form an enclosed chest cavity. The lungs and exterior boundary were identified by increasing the contrast and identifying an appropriate threshold. Each segmentation was downsampled to 20 points that could be edited by the user in Matlab. The mesh was generated using `ng_mk_extruded_model Grychtol2012` in EIDORS 3.10 **Adler2019**. Images were reconstructed using GREIT **Adler2009**. The GI index was calculated using the method presented by Zhao et al. **Zhao2009** using the lung regions from the forward model. A ventilated lung estimate was made using the segmentation as $\frac{A_{\text{ventilated lung}}}{A_{\text{total lung}}}$.

Results in figure figure ?? show a reconstructed image with more separable lungs in the enhanced model, and a mean GI index for each breath in the 1 minute recording that follows trend of the ventilated lung percentage in table 5.1.

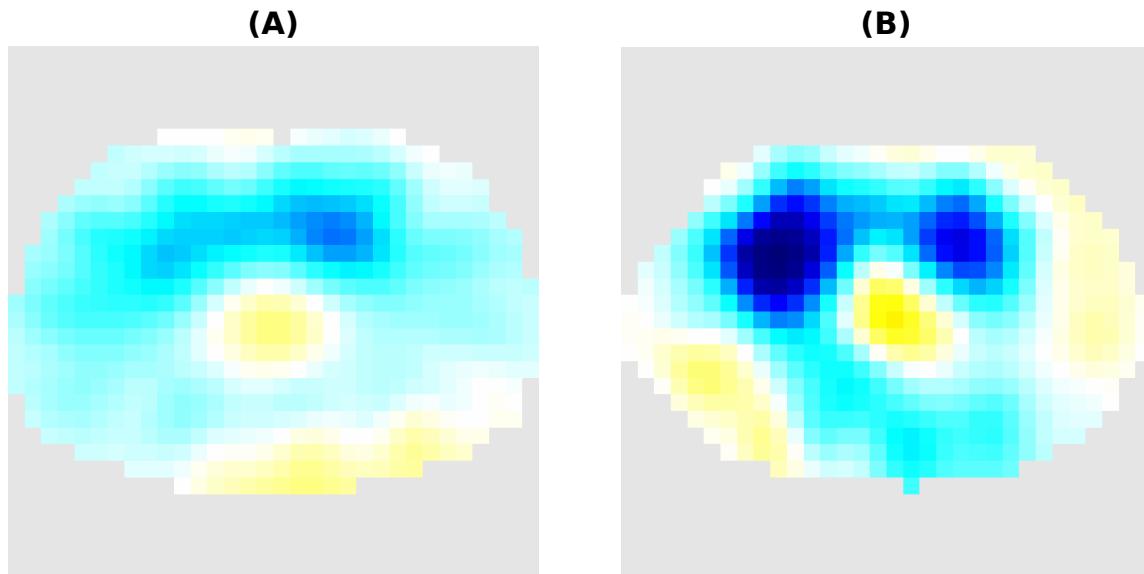


Figure 5.1: Single breath using: A) generic model B) custom model

Table 5.1: Ventilated lung estimate vs. GI index scores.

Sub- ject	Venti- lated lung (%)	GI (basic model)	GI (cus- tom model)
1	99.9	0.353±0.00	40.690±0.005
2	85.5	0.640±0.02	20.771±0.020
3	79.6	0.695±0.00	70.857±0.009
4	27.0	0.614±0.01	11.81±0.053

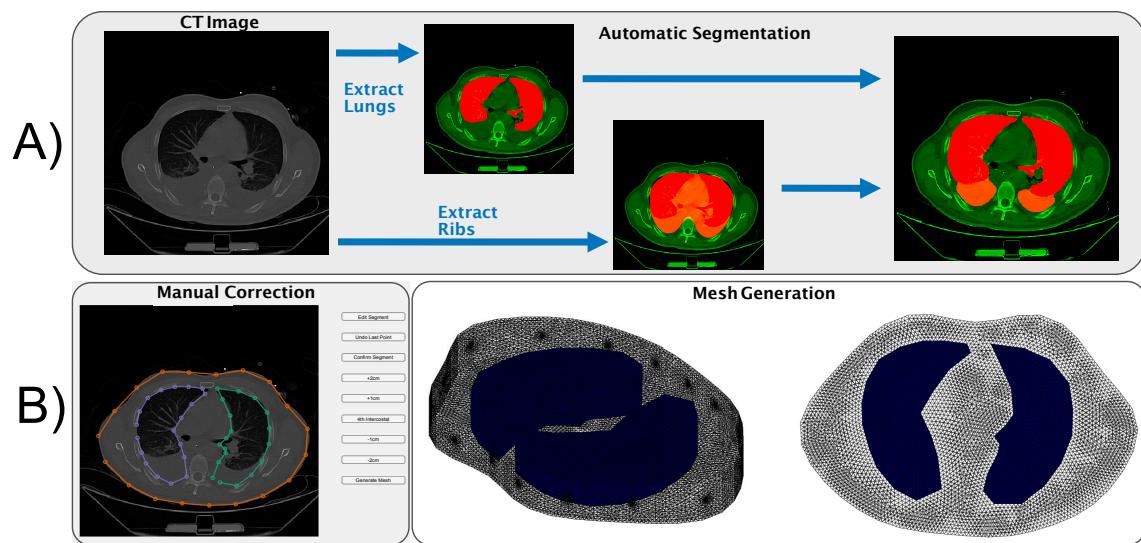


Figure 5.2: An overview of the segmentation and editing process showing: A) A sample raw CT which was thresholded, scaled and adjusted over several adjacent slices to identify the lung regions and an enclosed rib area, and the resulting lung estimate; and B) A screen capture of the manual mesh correction process and 2 views of the generated mesh.

Chapter 6

Internal Electrodes

The sensitivity analysis work and preliminary animal reconstructions have been presented in part at: the 42nd Annual International Conference of the IEEE Engineering in Medicine & Biology Society (EMBC 2020) (Stowe and Adler, 2020), and the 21st International Conference on Biomedical Applications of Electrical Impedance Tomography (EIT 2021) (Stowe et al., 2021).

Currently the most common implementations of EIT in biomedical applications measure voltages and inject currents from the body surface using one or two planes of electrodes. Internal electrodes are not used clinically despite promising simulation studies that have shown large improvements in reconstruction accuracy and internal sensitivity in 2D. In practice, EIT images with internal electrodes are challenging to interpret as the measurements are prone to motion artefacts. A shift in probe location between measurements, or modelling the probe location incorrectly can re-

sult in a large reconstruction error. In this chapter, we assess the sensitivity and accuracy benefits of internal electrode imaging in 3D, and present the effect of probe motion and localization error on reconstructions. We also present a reconstruction technique based on the single-step Gauss–Newton algorithm reconstructing for both motion and electrode locations in 2D (Soleimani *et al.*, 2006), to reduce the impact of motion artefacts on EIT images when using internal electrodes. In simulations on a tank model, **TODO: also have un processed tank data...** motion artefact due to electrode probe movement of up to 10% of the model radius was corrected. This chapter explores the use of internal electrodes in 3D and evaluates the potential to use esophageal electrodes to monitor ventilation and perfusion *in-vivo*.

6.1 Introduction

Currently the most common use of EIT is thoracic imaging, where it is used to image the movement of fluids due to cardiopulmonary activity. In the thorax the impedance signal is often dominated by the lungs and the change in volume due to breathing or ventilation. As discussed in chapter 3, one of the main challenges in perfusion imaging is the large difference in amplitude between the ventilation and cardiac signals. When using external electrodes, the respiratory amplitude is often 10 times larger than the cardiac component [refs]. Techniques such as breath holds or induced apnoea have been used to reduce the amplitude of the respiratory signal [several refs], but they are not feasible for long-term or continuous monitoring.

Internal electrode have been proposed several times as a method to increase sen-

sitivity in the most central regions of the chest.

One of the shortcomings of thoracic EIT imaging is low sensitivity in the especially surrounding the heart where it is challenging to isolate cardiac activity and the signals are often overpowered by the ventilation signals from the boundary regions with higher sensitivity [1].

The low sensitivity in central regions relative to the boundary limits the clinical applications of EIT, reducing the ability to accurately reconstruct and identify impedance changes due to the heart.

External electrodes limit the sensitivity of EIT there is limited sensitivity in the internal regions, making it challenging to identify the perfusion component of a signal dominated by ventilation.

Past research in 2D identified a sixfold increase in the cardiac frequency component of the EIT signal when 2 of 16 total electrodes were placed in the esophagus or trachea (Czaplik *et al.*, 2014). It is not clear whether the identified increase in cardiac-frequency amplitude stems from pulsatile (motion-based), changes or whether these changes represent true perfusion.

Several works have shown that motion **TODO: ref from notes on percent electrode movement**

the difference in amplitude between

There is potential for clinical use of internal electrodes to monitor ventilation, perfusion and hemodynamic changes in the intensive-care unit (ICU), where patients typically have breathing and feeding tubes in place.

EIT applications in development include mechanical ventilation guidance and

monitoring[refs needed] perfusion monitoring [refs], blood pressure monitoring [refs], and cardiovascular output [refs]. One of the main challenges for continuous ventilation and perfusion monitoring has been the small amplitude of the cardiac signal from the surface electrodes (Nguyen *et al.*, 2012).

EIT has low sensitivity away from the electrodes which are typically placed on the body surface. This chapter will investigate the use and application of internal electrodes and quantify the improvement in sensitivity and image reconstruction accuracy in simulation.

6.2 Motivation

One of the shortcomings of EIT is low sensitivity away from the electrodes. For continuous monitoring of cardiac-related events an EIT protocol with an improved sensitivity near the heart and innermost regions of the lungs may improve results and imaging accuracy.

6.3 Summary

This chapter provides proof of principle of internal electrode configurations to improve EIT sensitivity and reconstruction accuracy. The goal of this chapter is to determine the viability of internal electrode use through a series of simulations and phantom measurements then validate the imaging ability using one or two animals.

We expect to show that using internal electrodes increases the sensitivity of the measurements to internal impedance changes and enables a higher resolution of image

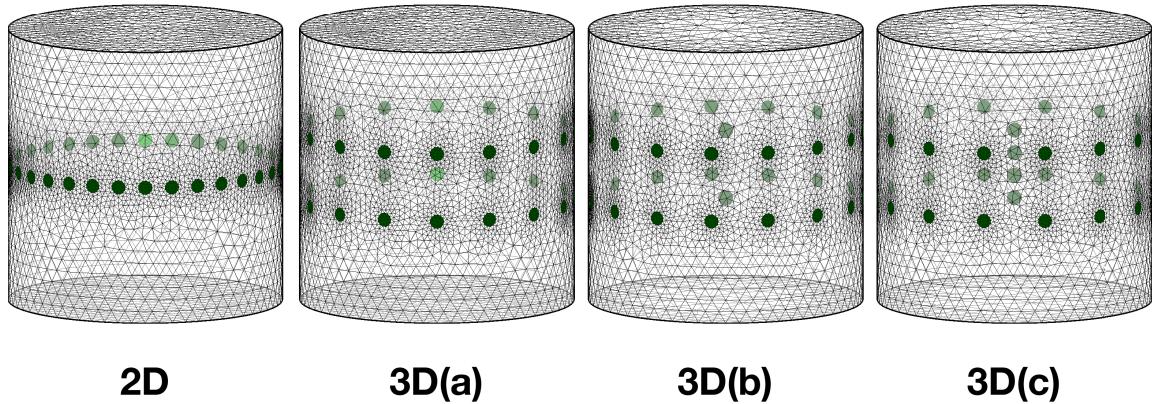


Figure 6.1: 4 configurations of electrodes were tested: 2D) a single ring of 32 electrodes; 3D(a) 2 rows of 16 external electrodes; 3D(b) 2 rows of 15 external electrodes with 2 internal electrodes; and 3D(c) 2 rows of 14 external electrodes and 4 internal electrodes.

reconstructions in these central regions of the thorax.

6.4 Simulations

To analyze the sensitivity changes due to different electrode configurations finite element models (FEMs) of a cylindrical tank were created with each of the tested electrode configurations. Figure 6.1 shows the four different configurations that were tested: a 2D electrode ring of 32 electrodes; a 3D configuration of 2 layers of 16 electrodes (3D(a)); a second 3D configuration of 2 layers of 15 electrodes plus 2 central internal electrodes inline with the electrode planes (3D(b)); and a final 3D configuration of 2 layers of 14 electrodes with 4 central internal electrodes evenly spaced between the electrode planes (3D(c)).

The tank in the simulations has a height of 2 m, radius of 1 m, and the electrode radius is 0.05 m for both the round external electrodes and the spherical internal

electrodes. In the 3D configurations the plane separation is 0.5 m and in all configurations the radial spacing between electrodes is equal. The background conductivity of the tank was 1 S/m and the conductivity of the target was 10 S/m.

When reconstructing images a conductive target was added to the tank centred at a height of 1 m at the midpoint of the tank radius. The target object radius is 0.4 m.

The reconstructions of the conductive object with and without additive noise are shown in Figure 6.2. To generate EIT images from voltage measurements, the 3D GREIT reconstruction algorithm was used (**Adler2009**). A spherical conductive target with a radius of 20% of the tank radius was placed midway between the centre and boundary of the tank, in a region with typically low sensitivity. The inverse problem hyperparameter was selected so that in all instances the amount of measurement noise that propagated from the measurements into the final images was equal.

Preliminary reconstructions on simulated data show that internal electrodes are able to reconstruct a conductive object closer to its true size based on visual inspection, but more simulations are required to quantify the improvements. These preliminary reconstructions are shown in Figure 6.2. The top row shows reconstructions with no additive noise, and the second row shows reconstructions on measurements with 5dB of additive noise.

Preliminary results shown in Figure 6.3 show internal sensitivity distribution changes when using 2 and 4 internal electrodes compared to the typical 2D and 3D configurations with only external electrodes.

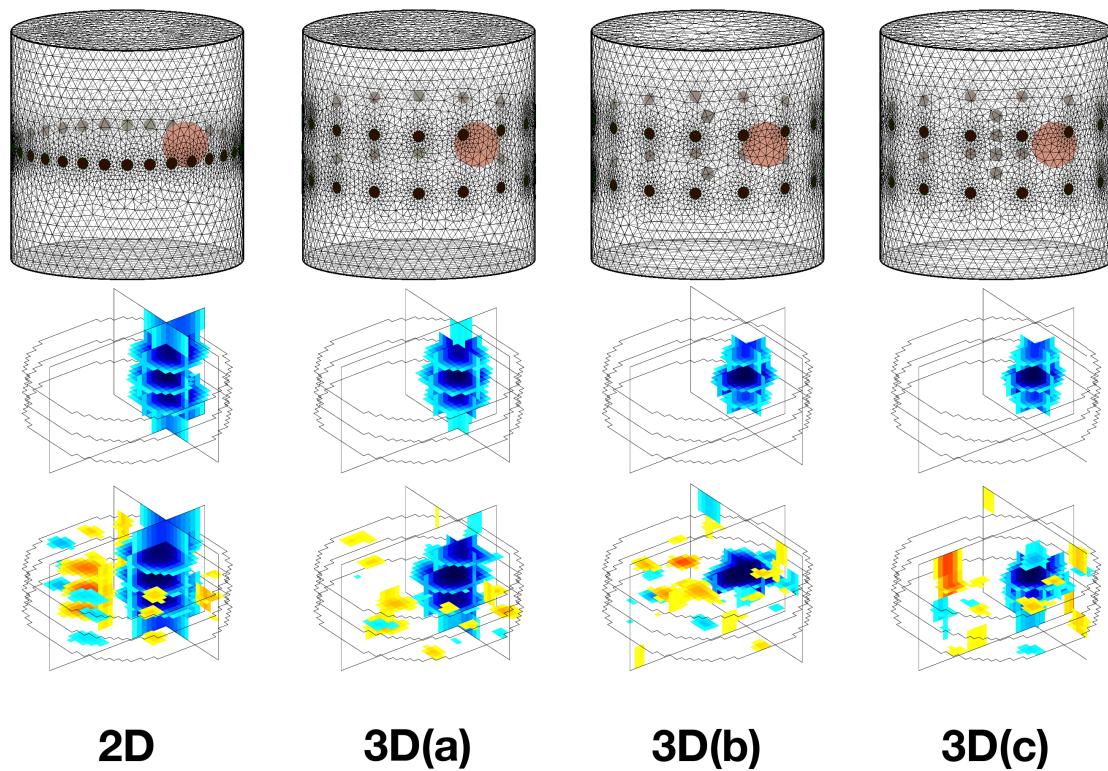


Figure 6.2: The top row shows reconstructions with no additive noise, and the second row shows reconstructions on measurements with 5dB of additive noise.

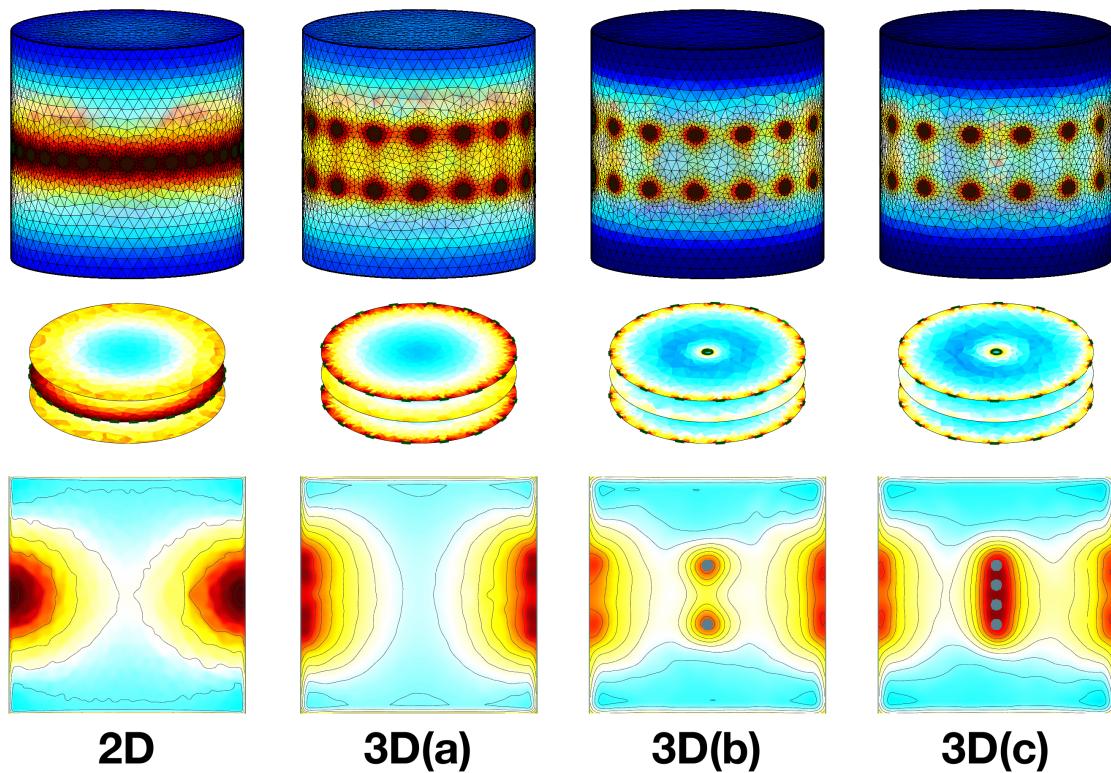


Figure 6.3: Sensitivity distributions for electrode patterns from left to right: A single 2D electrode plane; 2 electrode planes of 16 electrodes each; 2 internal electrodes and 2 external electrode rings of 15 electrodes; 4 internal electrodes arranged between 2 planes of 14 external electrodes

The sensitivity is then calculated from the jacobian (J) of the reconstruction matrix as:

$$S = \frac{\sqrt{\sum_i J_{ij}^2}}{V_i}$$

where V_i is the volume of each respective voxel.

These results show the expected increased sensitivity in the central regions of the model. To further improve internal sensitivity a new measurement pattern is proposed that uses more measurements between the internal probe and peripheral electrodes. The proposed injection and measurement pattern is shown in Figure 6.4. The sensitivity of the proposed pattern was compared to the sensitivity profile of the same configuration using the basic “skip 4” injection and measurement pattern which has been found to give good sensitivity in 2D and 3D external electrode configurations **Grychtol2016**.

Using this injection pattern preliminary results show a further increase in sensitivity in the internal regions without increasing the measurement acquisition time. The sensitivity distribution for the new injection pattern is pictured in Figure 6.5.

To quantify image quality, the same object will be reconstructed in multiple situations and is computed as a combination of the following metrics: mean squared error (MSE) between the reconstructed object and the actual geometry; full width half max (FWHM) of the reconstructed object; and separability of two identical circular objects in the same reconstruction.

Building on this work simulations will be used to answer more questions on the use of internal electrodes:

- What internal electrode configuration most improves the internal sensitivity?

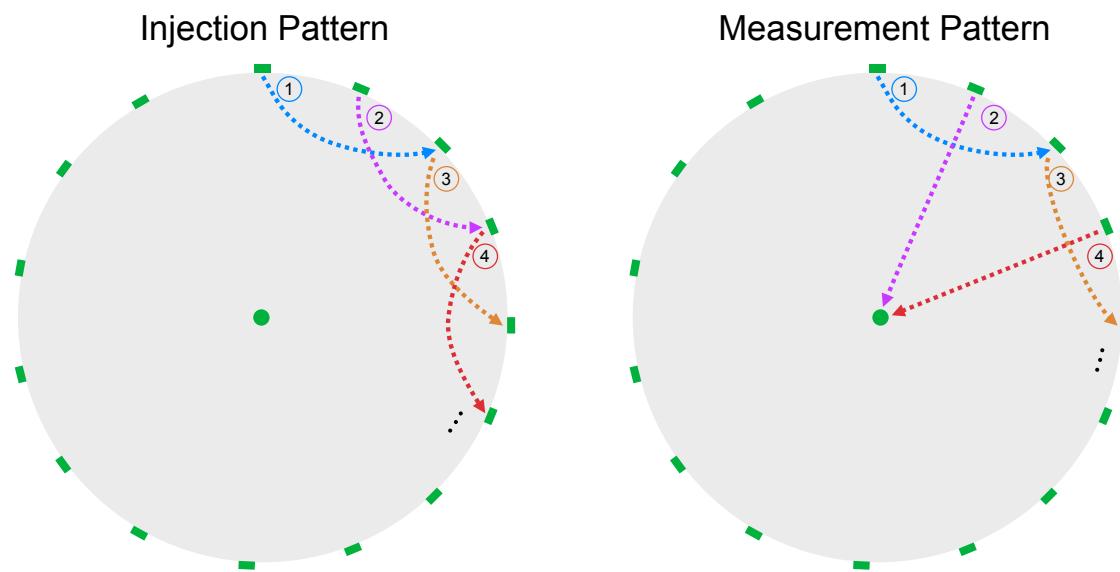


Figure 6.4: A proposed current injection and measurement pattern for EIT imaging with 2 internal electrodes. The injection pattern is a typical “skip 4” pattern injecting between every 5th electrode in a square electrode layout and the measurement pattern replaces every 2nd measurement in the typical method with a measurement between the internal probe and external rings. Note: this figure does not differentiate between upper and lower electrode planes, but all injections and measurements are done between the 2 planes.

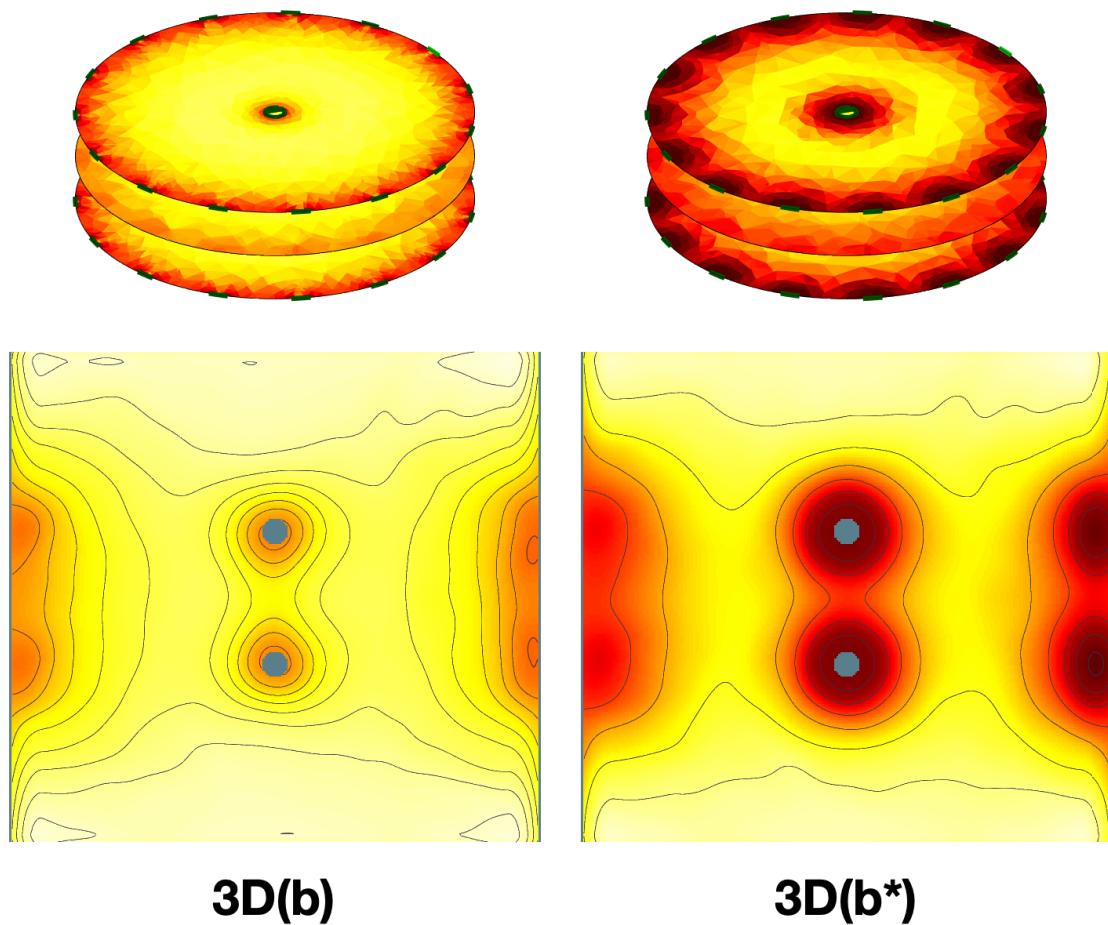


Figure 6.5: A comparison between the sensitivity distributions for a typical “skip 4” injection pattern pictured on the left (3D(b)) and the modified injection and measurement pattern on the right (3D(b^{*})).

- What is the best injection pattern for measuring activity in the heart region?
- What is the maximum number of electrodes that can be placed internally while maintaining image quality?
- How should internal electrodes be used for current injections?
- How sensitive is the resulting image to movement of the internal electrodes?
- In what regions of the FEM is the object reconstruction improved?
- Do internal electrodes pose any risk to patient safety?

6.5 Phantom Measurements

Phantom measurements are used to assess the real-world performance of the measurements and configurations that were identified in simulations. Two non-conductive circular objects are placed and imaged at 4 different separations: 1 cm, 6 cm, 11 cm and 16 cm mid-way between the central probe and the tank wall. These separations are used to determine the change in resolution between the different configurations.

In addition to determining the real-world separability the phantom measurements will be used to image a conductive object in a region representing the heart region and determine the FWHM and MSE based on the measured location of the object within the phantom dimensions. This will be compared to the results obtained through simulations.

6.6 Motion Correction

6.7 Inverse Source Localization

6.8 Animal Data

Internal electrodes present the opportunity to obtain a significantly higher sensitivity and with motion correction may be used to give a more accurate estimate of arterial pressure. This paper presents a method of using internal electrodes while correcting for motion of the internal probe to yield high sensitivity near to the esophageal probe.

6.9 Methods

The forward model was constructed using EIDORS version 3.10 **Adler2019** using `mk_library_model` **Grychtol2012** the internal electrode was added as an extra structure. This model was also used to identify the heart and lung regions. The sensitivity of the lamb model was calculated from the Jacobian using the method from **Stowe2020**. Sensitivity profiles using 32 electrodes with and without 4 internal electrodes are shown in fig. 6.6.

Data were collected in 3 ewes during ventilation under general anesthetic using the SenTec EIT Pioneer Set. 30 second recordings were made during regular ventilation with a volume of 400 ± 50 ml, frequency of 0.2 ± 0.05 Hz, and peep of 6. Recordings were repeated for several ventilation scenarios including: high volume (+100 ml), low volume (-100 ml), high frequency (+0.17 Hz), low frequency (0.07 Hz), high peep

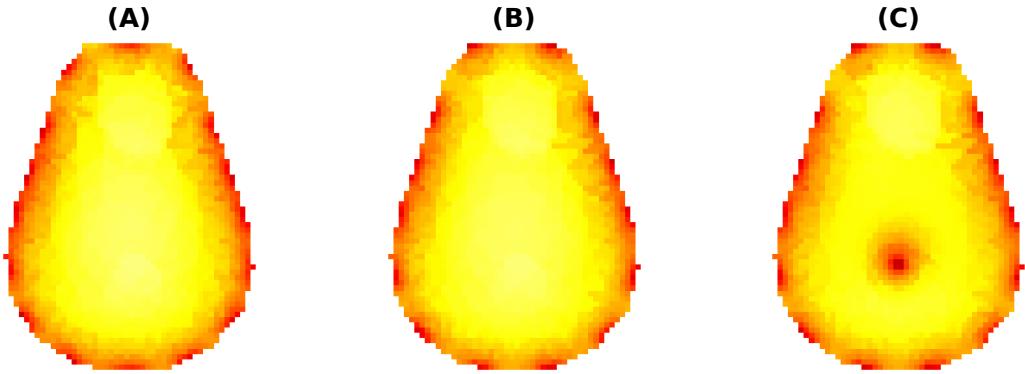


Figure 6.6: Sensitivity distribution averaged across 10 evenly spaced layers between the electrode planes in the lamb model for: A) 32 external electrodes B) 28 external electrodes c) 28 external electrodes and 4 internal electrodes

(10) and low peep (4).

All breaths in each 30 second segment were ensemble averaged to give one representative breath for each scenario.

Images of one averaged breath per recording were reconstructed using the 3D GREIT algorithm **bartek2016** which minimizes the effect of electrode motion on the resulting image. Results were compared for both external only and internal electrode configurations using the same recording. To obtain results with only external electrodes all injections and measurements using internal electrodes were removed prior to reconstruction. Measurements on injecting electrodes were always removed. Images from subject 3 during regular ventilation are shown below in fig. 6.7. With internal electrodes impedance changes at the cardiac frequency had a magnitude of $6.4\% \pm 4.35\%$ of the ventilation frequency and without internal electrodes the amplitude of the cardiac frequency was $0.8\% \pm 0.3\%$ of the ventilation frequency signal.

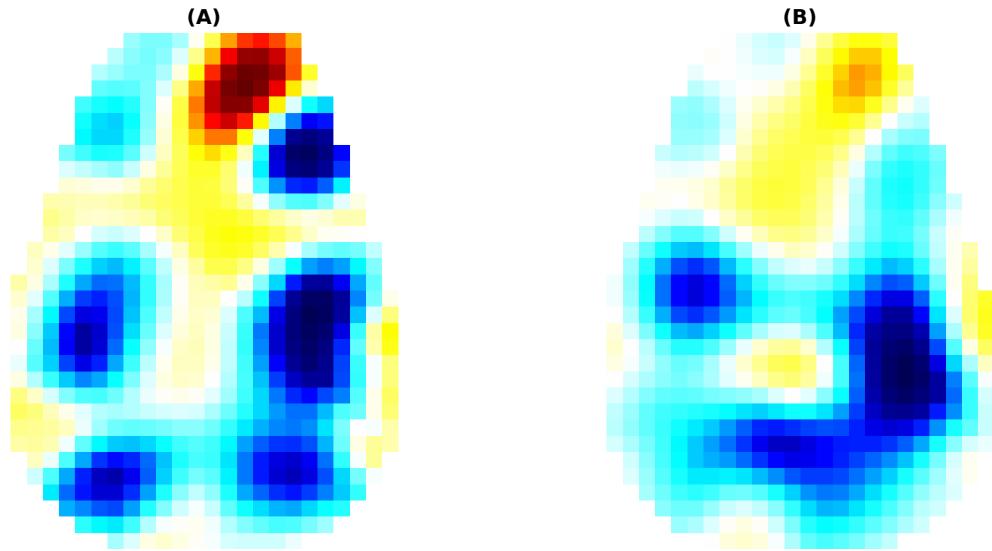


Figure 6.7: A single breath imaged with: A) no current injections or measurements on internal electrodes b) with internal electrodes

6.10 Conclusions

Reconstructions using the GREIT algorithm with internal electrodes on an esophageal probe were able to give increased sensitivity to cardiac-frequency impedance changes and may allow for better measures of blood pressure and pulse wave velocity.

Chapter 7

Internal Electrode Motion

7.1 Introduction

As discussed in section 2.2, there are several factors that contribute towards the cardiosynchronous signal in EIT. One of the largest contributions if movement; both the movement of body structures and the movement of electrodes contribute significantly to the measured impedance and the resulting reconstructed image (Adler *et al.*, 2017b; Proen  a *et al.*, 2015). The previous chapters support that more accurate meshing and internal electrodes may help to identify the movement of structures and organs, reducing their impact on the reconstructed images, but the probelm of electrode motion is amplified when internal electrodes are used. Internal electrodes placed on a probe are challenging to locate within a subject due to difference in individual geometries. Incorrectly modelling the electrode locations can introduce an artefact in the image (Boyle and Adler, 2011), but in time difference EIT these effects

ar minimized (Adler and Boyle, 2017). The larger problem appears to be the movement of the internal probe relative to the external electrodes between measurements. Movement correction algorithms using the movement jacobian have been designed in 2D and 3D (Gómez-Laberge and Adler, 2007; Gómez-Laberge and Adler, 2008; Soleimani *et al.*, 2006) and used to reconstruct electrode movement on 3D models with a 2D arrangement of electrodes (Boyle, 2016). There are several available methods to calculate the movement jacobian (Boyle *et al.*, 2017), the simplest of which, the naïve perturbation method (Gómez-Laberge and Adler, 2008), has been used here as a proof of concept. A more detailed explanation of this jacobian calculation method is presented in section 2.4.6.1 of the background. As an internal probe is moved towards one side of the model the distance between the probe and the external electrodes changes. This results less impedance where the distance has decreased, and a higher impedance where the distance has increased. Without knowing the new probe location we can minimize the effect of this error, but there still remains a mismatch between the modeled and actual probe locations. The artefact that is present can give hints regarding the motion of electrodes although it can be difficult to separate the effect of electrode movement from biological impedance changes of interest (Boyle, 2016). This chapter presents a method that builds on current algorithms available to compensate for electrode movement, to create a new, corrected model that helps to correct for the effect of electrode location. The known shape of the artefact due to movement of the probe is used to derive the probe location, and create a corrected model to improve reconstruction accuracy in the presence of probe motion.

The goal of this chapter is to establish a technique to reduce the impact of probe movement on image reconstructions, and then to use this technique in an animal model. Although simulations in 2D and now in 3D show that internal electrodes can provide a desireable increase in sensitivity, the benefit has not been realized in real-world recordings. This chapter presents a novel method to correct for electrode displacement artefact, and compares established techniques for 3D EIT image reconstruction to reconstructions with the novel internal electrode approach in a limited number of subjects.

7.2 Methods

This chapter consists of two experiments: a simulation study correcting for probe motion in a tank model, and preliminary *in-vivo* recordings that were conducted as an addition to another experimental protocol.

7.2.1 Simulations

Simulations were done using EIDORS (v3.10) (Adler *et al.*, 2017a) meshes were generated using Netgen (version 5.3.1) (Schöberl, 1997) with Matlab 2019b (Matworks, Natick, MA, USA).

7.2.1.1 Tank models

A model of a tank with 28 external electrodes placed in twp planes of 14 evenly around the exterior and 4 internal electrodes was created. The tank radius and

height were 1 m, and the external electrode radius was 5cm. The external electrode planes were placed at a height of 0.3 m and 0.7 m on the tank. The internal electrodes were spaced evenly between the external electrode planes at 0.3 m, 0.433 m, 0.567 m and 0.7m. The internal electrodes were specified in 2 different ways. The first used spheres with a radius of 2.5 cm, and the second used a hollow cylinder with a radius of 2.5 cm containing cylindrical electrodes with a radius of 2.5 cm and a height of 5 cm. Both models are shown in figure 7.1.

These models were compared to confirm that both performed adequately with motion correction, as internal electrodes are challenging to model and many different techniques are used. When using internal electrodes on a complex model, it is currently easiest to add electrodes along a hollow tube cut through the model.

To specify a conductive region elements within a radius were assigned a conductivity of twice the background conductivity of the tank model. The conductive target was placed midway between the center of the tank and the boundary and the probe. The radius of the target was 20 cm. Figure 7.1 shows the conductive target for both internal probe types.

7.2.1.2 Measurements

Despite sensitivity advantages ascribed to custom injection and measurement patterns, the skip 4 pattern was used as it is straight forward to implement with currently available EIT systems.

Reference measurements were made with no conductive object and electrodes in the center of the model. Measurements with a conductive object were made with

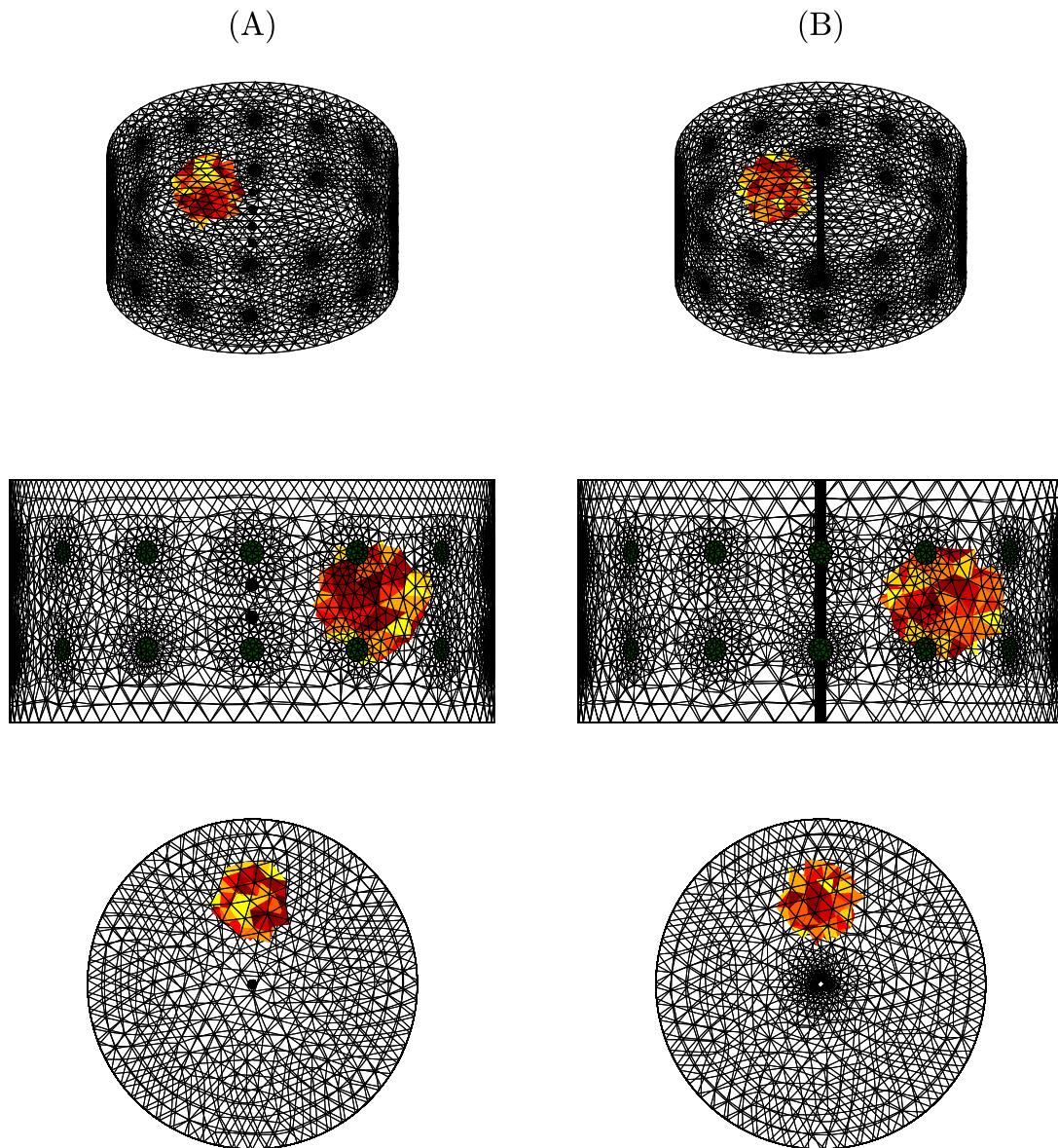


Figure 7.1: On the left (A) internal electrodes were modelled as 4 spherical electrodes between the external electrode planes. On the right (B) the intenal electrodes were created using a hole through the center of the model and using cylindrical electrodes on the inner surface of the model. The conductive region was specified by setting the conductiviy of all elements contained in the target to be twice the background conductivitity.

the probe centered, then with the probe was shifted by 1, 5, and 10 percent of the tank radius in a randomized direction. The direction was randomized between trials but was consistent for each of the 1, 5 and 10 percent probe error models that was compared.

7.2.2 Movement correction

The movement correction jacobian was calculated using the methods presented by Gómez-Laberge and Adler (2008). In a model with a centered electrode probe and uniform conductivity, each of the 4 electrodes on the probe was perturbed by 0.001 m in each of the **x**, **y**, and **z** directions. A measurement was made for each electrode and each of the 3 dimensions of movement. The resulting measurements were divided by the perturbation amount. The measurements on each electrode (V_j) and from each direction of movement (**x**, **y**, and **z**) were combined to form the movement jacobian (J_M) using the following equation from Gómez-Laberge and Adler (2008):

$$J_M = \left[\frac{\partial \mathbf{V}_j}{\partial \mathbf{x}}, \frac{\partial \mathbf{V}_j}{\partial \mathbf{y}}, \frac{\partial \mathbf{V}_j}{\partial \mathbf{z}} \dots \frac{\partial \mathbf{V}_n}{\partial \mathbf{x}}, \frac{\partial \mathbf{V}_n}{\partial \mathbf{y}}, \frac{\partial \mathbf{V}_n}{\partial \mathbf{z}} \right] \quad (7.1)$$

This movement jacobian was used in conjunction with the following single step formulation for a reconstruction matrix presented by Adler *et al.* (1994). The regular impedance based reconstruction matrix is denoted by R_C and the jacobian for impedance based reconstruction is denoted by J_C .

$$R_C = \frac{[J_C]^T [W]}{[J_C][W][J_C]^T + [W]} \quad (7.2)$$

W in the above equation represents the laplace prior (Soleimani *et al.*, 2006). Combining this formulation with the movement jacobian yeilds the following equation for the reconstruction matrix (R_M) (Soleimani *et al.*, 2006) where R_M denotes the reconstruction matrix for motion correction:

$$R_M = \frac{[J_C]^T[W]}{[J_C][W][J_C]^T + \mu[J_M][J_M]^T + [W]} \quad (7.3)$$

In the above equation μ represents the weighting of the motion correction. In this experiment the motion correction weighting was set to 100.

Combining equation (7.2) and equation (7.3) a reconstruction matrix to reconstruct exclusively the noise due to movement (R_N) in an image can be generated.

$$R_N = R_M - R_C \quad (7.4)$$

An image (X) is reconstructed using the measurements (b) and reconstruction matrix (R) as follows:

$$X = bR \quad (7.5)$$

Three images: X_C , X_M and X_N are recontsructed from their respective reconstruction matrix.

A new movment correction strategy had been built around the reconstructed image X_N . The noise due to exclusively motion is reconstructed into a 3D image, and the center of mass of the positive change is located. This is assumed to be the direction of the motion. The amplitude of the electrode motion is estimated as half of

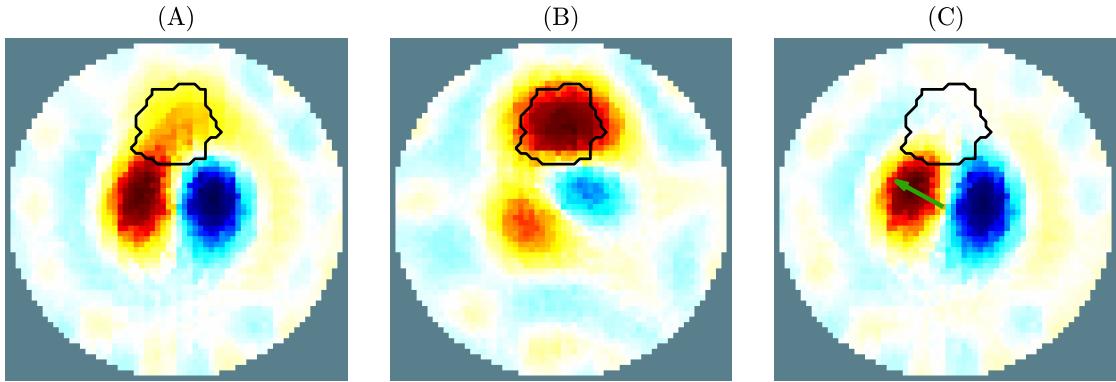


Figure 7.2: Three image reconstruction matrices are generated: one to calculate an image based only on the measurements (A), one that is able to reduce the impact of noise using a movement jacobian (B) and finally a reconstruction that reconstructs only the noise due to motion of the probe (C). The center of mass of the conductive object is used to estimate the probe location and direction of movement. The green arrow indicates the calculated direction of movement (multiplied by 5 for visibility). The black line indicates the outline of the region of conductivity at the imaged plane. The images depict an average of 10 slices between the external electrode planes.

the distance between the conductive object and the probe this method is illustrated in figure 7.2. Images were reconstructed and displayed on a 64 by 64 grid to give a more accurate position of the electrode position between the model and the image.

Using the probe location estimate calculated from X_N a new model was reconstructed with the probe repositioned to the reconstructed location. This model was then used to calculate a new reconstruction matrix using equation (7.3) with an updated jacobian based on the new probe location.

7.3 Image comparison

To simplify processing of the reconstructions, 10 images between the external electrode planes were averaged together to generate a 2D representation of the 3D data.

This did not allow for analysis of movement in the vertical direction, but we did not expect this to be a major contribution in real-world data. The reconstructed images were compared in 2 ways. First the accuracy of the reconstruction was evaluated by computing the Jaccard index between the actual and imaged boundaries of the conductive target. The imaged boundary was drawn at half the maximum value of the brightest object in the image. The second metric computed was a noise estimate. This was calculated as the amplitude of the imaged object relative to the amplitude of the entire image through the following equation:

$$N = 1 - \frac{A_{\text{object}}}{A_{\text{image}}} \quad (7.6)$$

Where A_{object} is the amplitude of the object, A_{image} is the amplitude of the image, and it is subtracted from 1 so that a noise estimate of 0 corresponds with all of the image signal corresponding to the conductive target.

7.3.1 *In-vivo* recordings

Data were collected in 3 ewes during ventilation under general anesthetic using the SenTec EIT Pioneer Set. A skip 4 measurement pattern was used with an injection current of 3 mA at 20 kHz. Electrodes 29 to 32 were connected to the internal electrode probe.

7.3.1.1 Internal electrode probe design

The internal electrode probe consisted of 4 brass electrodes on a flexible tube. Each electrode was 1 cm in length and the spacing between electrodes was 2 cm. The

electrodes were created from a brass sheet with a thickness of 2 mm that was bent to the shape of the flexible flexible tubing and soldered to achieve a smooth surface. The electrodes were glued in place on the tube protruding 2 mm from probe surface. The constructed probe is shown in figure 7.3.

7.3.1.2 Ovine model

An internal probe was added to an ovine model by adapting the `mk_library_model` function in EIDORS (Adler *et al.*, 2017a). A circular region was added to the geometry library that was extruded upwards to make a hole at the probe location. External electrodes were placed in a “square” pattern on the boundary, and internal electrodes were added as cylindrical objects in the central hole of the model. The resulting model is shown in figure 7.4.

7.3.1.3 Reconstruction

Images were reconstructed with 4 methods. The 3 methods discussed in section 7.2.2, and GREIT (Grychtol *et al.*, 2016).

7.3.1.4 Pulsatile amplitude

To measure the advantage provided by internal electrodes with regards to pulsatility detection, a fast fourier transform (FFT) of each signal was calculated. The cardiosynchronous component was selected as the highest the amplitude near the recorded heartrate range of 65-80 bpm. This was divided by the amplitude of the ventilation frequency, which was the largest low frequency amplitude in the FFT.



Figure 7.3: A prototype of an internal electrode probe for esophageal use in an ovin model. Four brass electrodes are bent around the outside of a flexible tube designed for esophagaea use. Edges of the electrodes are soldered and filed to be smooth. The electrodes are glued in place and wires run down the hollow center of the tube to connect to the EIT system.

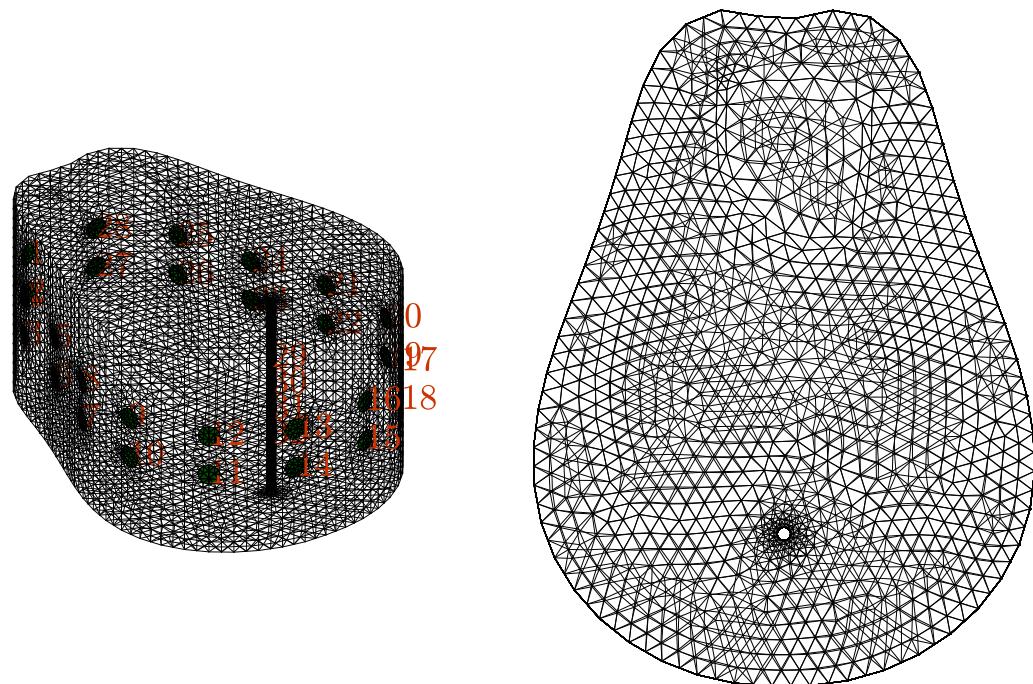


Figure 7.4: An internal electrode was added to an ovine model by creating a hole at desired probe location and added cylindrical electrodes at the desired heights. External electrodes were placed in a “square” electrode configuration on the boundary. Electrodes 1 to 28 were external and 29 to 32 were internal.

Table 7.1: The jaccard indexes were calculated for each of the reconstructions in figure 7.5. Method A does not use any motion correction. Method B incorporates the movement jacobian, and method C uses the new probe location correction technique. For jaccard index closer to 1 is better.

Movement (% of radius)	Method A	Method B	Method C
1	0.732	0.809	0.802
5	0.038	0.654	0.808
10	0.003	0.227	0.577

This was repeated for all 12 recordings across 3 animals.

7.4 Results

The following section presents the results for both simulation and *in-vivo* work.

7.4.1 Simulation

It was found that there was no measureable difference between simulations of models using spherical internal electrodes relative to the cylindrical electrodes. The results are presented using the cylindrical model to be consistent with the lamb models. The probe was moved in the $-y$ direction for figures and calculations in the results.

Figure 7.5 shows the result of the 3 reconstruction methods. The boundaries of the reconstructed and actual targets are outlined to highlight the performance. The new method is able to reconstruct the correct location with motion of up to 10 % of the tank boundary between measurements. The jaccard index and noise estimate for each method are presented below in table 7.1 and table 7.2.

For movement of 1 percent of the tank boundary motion correction using only

Table 7.2: Noise estimate values calculated for each of the reconstructions in figure 7.5. Method A does not use any motion correction. Method B incorporates the movement jacobian, and method C uses the new probe location correction technique. For the noise estimate a lower score is better, a score of 0 indicates all image changes occur within the target boundary, and a core less than 1 indicates most of the changes in the image are due to the identified target.

Movement (% of radius)	Method A	Method B	Method C
1	0.588	0.537	0.529
5	0.781	0.669	0.618
10	0.777	0.747	0.718

the movement jacobian was roughly equivalent to the probe location correction algorithm. Across all other scenarios the probe location correction algorithm achieved a higher jaccard score. Across all scenarios the new tecnique to correct for motion had the lowest noise estimate score.

When looking at the reconstructions using the three methods, the new probe correction technique reconstructed the target as the brightest object in all scenarios. With no motion correction the target was identified correctly only in the first scenario. Using the movement jacobain reconsturciton method, the target was correctly identified when the probe was moved 1 or 5 percent of the tank radius.

7.4.2 *In-vivo*

Recordings in each of the three ewes are presented in this section. Reconstructions with internal electrodes are shown in figure 7.6

Reconstructions with movement correction applied to the internal electrode appear to show a slight improvement in lung distinguishability and a slight reduction in noise surrounding the internal electrode. There is limited difference between the

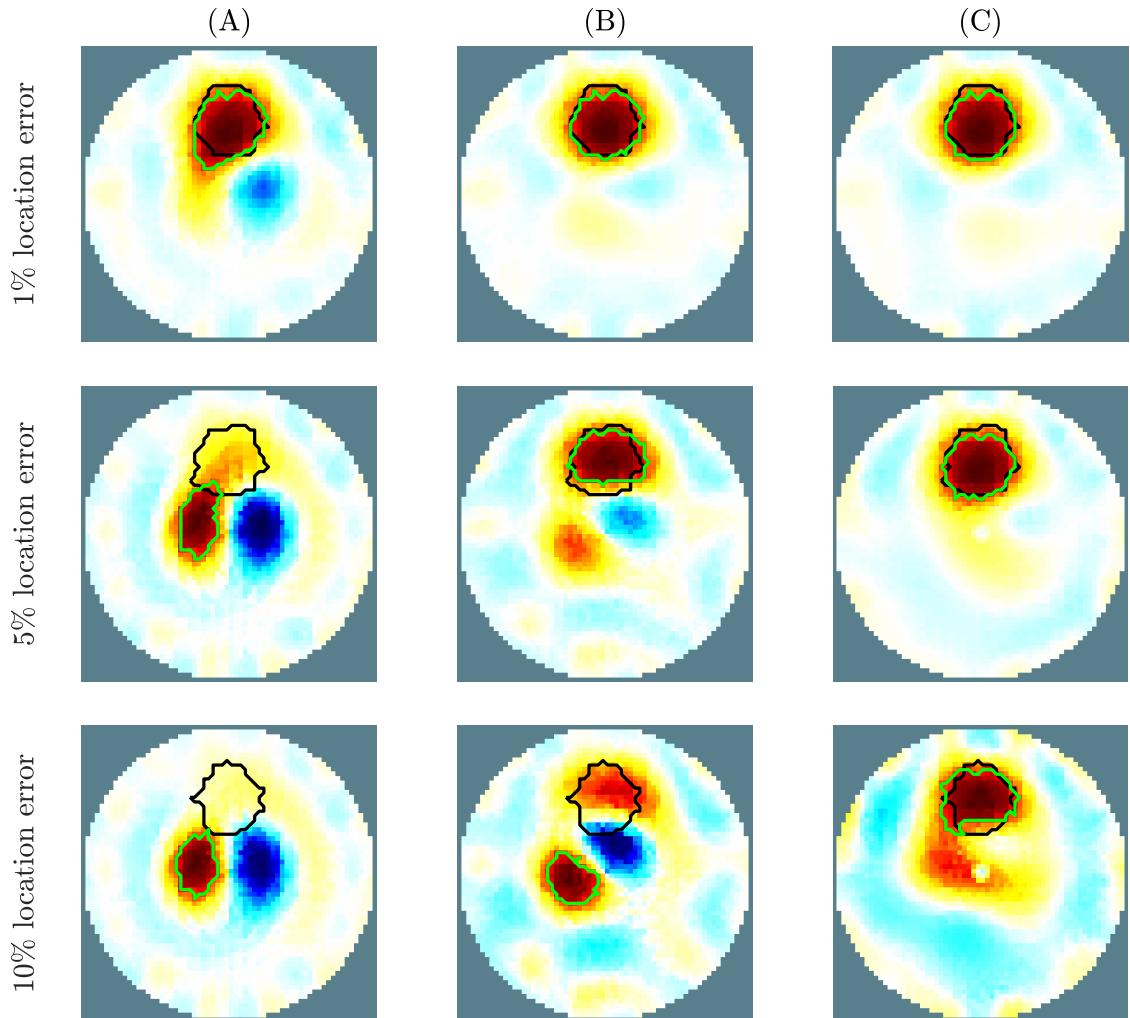


Figure 7.5: The results of the probe location correction are presented. The rows from top to bottom show results with 1, 5, and 10% shifts in probe location relative to the tank radius. Column (A) shows the results of the reconstruction with no motion correction, column (B) shows the method using the movement jacobian, and column (C) shows the results of the new probe location correction method. The green outline indicates the reconstructed boundary of the conductive target, and the black outline is the actual boundary.

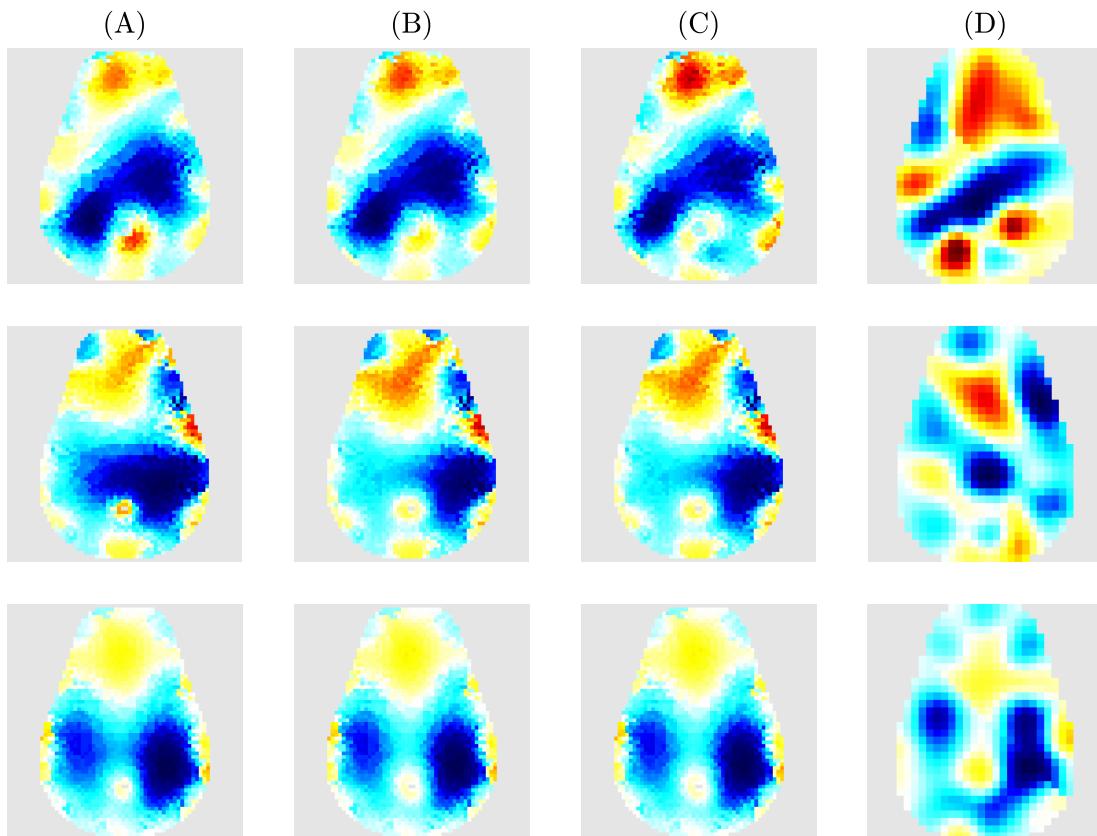


Figure 7.6: preliminary results on 3 ewes. Each column represents a reconstruction method. Column (A) uses no motion correction, column (B) uses the movement jacobian and column (C) uses the new method for electrode location correction. Column (D) contains reconstructions using the GREIT 3D algorithm. The number of pixels is different between images due to the methodology of probe localization and the limitations of GREIT.

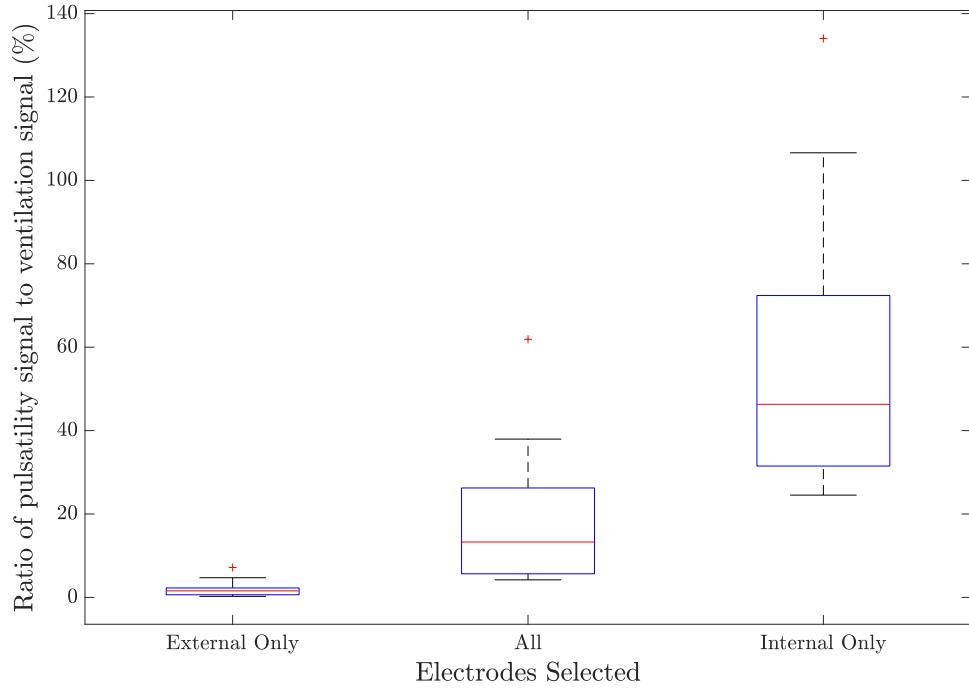


Figure 7.7: The cardiosynchronous signal component was divided by the ventilation frequency component to compare the detectability of the pulsatile changes. The ratio was compared for measurements using only external electrodes, all electrodes, and internal electrodes only.

two methods accounting for motion on the electrode.

The ratio of cardiosynchronous to respiratory rate impedance signals is shown below in figure 7.7. The results show a much higher amplitude of the cardiopulmonary signal relative to ventilation when internal electrodes are used.

7.5 Discussion

This chapter aimed to provide a technique to reduce noise due to motion on internal electrodes when reconstructing images in 3D and demonstrate the noise reduction technique *in-vivo*. The new technique to reconstruct images in the presence of motion on internal electrodes was able to reconstruct an image of a conductive target more accurately and with less noise in the surrounding image. *In-vivo* reconstructions showed a slight improvement in lung distinguishability when correcting for motion artefact on internal electrodes compared to methods without this correction including GREIT.

It was found that GREIT performed poorly when reconstructing data with internal electrodes. The figures of merit for GREIT designed and optimized to work with external electrodes where sensitivity is low in the center of the model and high at the edges (Adler *et al.*, 2009). An adaptation to GREIT that is able to account for internal electrodes and motion of an internal probe is planned as a continuation of this project.

Two different types of internal electrodes were created in this chapter. One with spherical internal electrodes and one with a hollow probe and cylindrical electrodes. Meshing and creating internal electrodes can be challenging due to limited meshing tools for using internal electrodes with EIT and the method of adding internal probes to anatomical models using a hollow region in the center of the model was created for this project. Code to generate this model with internal electrodes is planed to be included in a future release of EIDORS.

The noise estimate score calculations were not perfect representations of the noise

in the images due to the way the reconstructed object was identified. Since the brightest object was always identified as the object, the noise estimate was artificially low in some cases. Comparing the amplitude of changes contained within the actual object boundaries could be a better metric of the noise present in the image, but this would also be dependant on the accuracy of reconstruction and would have favoured the method that reconstructed the object most accurately. The boundary of the actual target also changes due to differences in element locations as the probe is moved, so the comparison would not be consistent across all probe movement states. Despite this limitation, the noise estimate does help to quantify the amount of noise seen in the images for each technique. The new method performed better than existing techniques with regards to the computed noise metrics.

In cases of less than 1 percent error in probe location, there was very little difference between the two motion correction algorithms with regards to reconstruction accuracy of the object. Despite this there was still a slight reduction in background noise in the image.

Despite the success of the probe correction algorithm with regards to location accuracy and noise reduction there are several limitations to its use. The most significant is the increased modelling time required. This method required an extra model, and inverse solution compared to the single step solutions. For simple models such as a tank the added time difference is small, but for complex models this could increase reconstruction time drastically. When several frames are reconstructed each one requires a unique model with a modified probe location increasing the time required significantly. To reduce the overall time, correcting the probe location only

when it is above 1 percent of the model width may be helpful. The benefits of the probe localization technique are not substantial at this level of movement to be required. Additionally in the images presented and recorded there was not seen to be significant probe movement. It is not yet clear what the typical movement of the electrode probe is expected to be in the esophagus.

Previous studies with internal electrodes showed that there were also cases where contact impedance of the probe was not ideal for electrodes placed on the breathing and feeding tubes (Czaplik *et al.*, 2014). The electrode used was shown to have good contact impedance on the SenTec Pioneer Set interface, but the current design protrudes slightly from the tube and may be challenging to integrate into a clinical esophageal or tracheal tube.

TODO: Add as many references as we can find and discuss the amplitudes then
proofread

Chapter 8

Conclusion

8.1 Summary of Findings

8.2 Future Work

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