Differential expression analysis between high-sugar diet vs control in fruit fly

Shuo Zhang

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Load Libraries

```
library(ggplot2)
library(DESeq2)
## Warning: package 'DESeq2' was built under R version 4.3.1
## Loading required package: S4Vectors
## Loading required package: stats4
## Loading required package: BiocGenerics
##
## Attaching package: 'BiocGenerics'
  The following objects are masked from 'package:stats':
##
##
       IQR, mad, sd, var, xtabs
  The following objects are masked from 'package:base':
##
##
##
       anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##
       colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##
       get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##
       match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
##
       Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,
##
       table, tapply, union, unique, unsplit, which.max, which.min
## Attaching package: 'S4Vectors'
## The following object is masked from 'package:utils':
##
       findMatches
##
## The following objects are masked from 'package:base':
##
##
       expand.grid, I, unname
## Loading required package: IRanges
## Warning: package 'IRanges' was built under R version 4.3.1
## Loading required package: GenomicRanges
## Loading required package: GenomeInfoDb
```

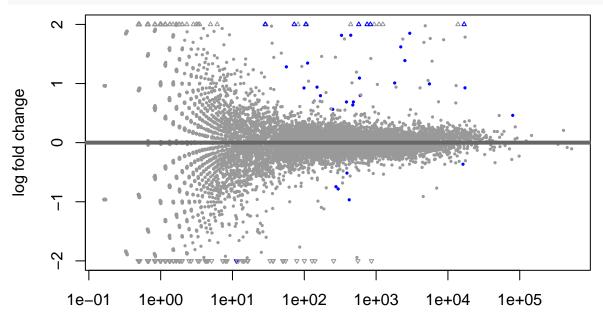
```
## Warning: package 'GenomeInfoDb' was built under R version 4.3.1
## Loading required package: SummarizedExperiment
## Loading required package: MatrixGenerics
## Warning: package 'MatrixGenerics' was built under R version 4.3.1
## Loading required package: matrixStats
## Attaching package: 'MatrixGenerics'
## The following objects are masked from 'package:matrixStats':
##
##
       colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse,
##
       colCounts, colCummaxs, colCummins, colCumprods, colCumsums,
##
       colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs,
##
       colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats,
##
       colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds,
##
       colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads,
##
       colWeightedMeans, colWeightedMedians, colWeightedSds,
##
       colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet,
##
       rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods,
##
       rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps,
##
       rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins,
       rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks,
##
##
       rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars,
       rowWeightedMads, rowWeightedMeans, rowWeightedMedians,
##
##
       rowWeightedSds, rowWeightedVars
## Loading required package: Biobase
## Welcome to Bioconductor
##
##
       Vignettes contain introductory material; view with
##
       'browseVignettes()'. To cite Bioconductor, see
##
       'citation("Biobase")', and for packages 'citation("pkgname")'.
## Attaching package: 'Biobase'
## The following object is masked from 'package:MatrixGenerics':
##
##
       rowMedians
## The following objects are masked from 'package:matrixStats':
##
       anyMissing, rowMedians
library(EnhancedVolcano)
## Loading required package: ggrepel
```

read count table and sample information

```
cts <- read.table('../formatted_HSD_featureCounts.txt', header = TRUE, row.names = 1)
#keep <- rowSums(cts >= 10) >= 3
```

```
#cts <- cts[keep,]</pre>
head(cts)
                  F1_1 F1_2 F1_3 ND_1 ND_2 ND_3
## cac
                  5002 5055 5098 4598 5077 4850
                  4590 4478 4353 4464 4294 4531
## Cngl
## CG11836
                    82
                         82
                              81
                                    68
                                         79
                                              70
                               27
                                              28
## CG33096
                    25
                         22
                                    17
                                         32
## bam
                    0
                         0
                              0
                                   0
                                        0
                                             0
## sisRNA:CR46364
                    15
                         21
                              18
                                  16
                                         20
                                              14
read sample information
coldata <- read.table('../coldata.txt', header = TRUE, row.names =1)</pre>
coldata$condition <- factor(coldata$condition, levels = c("ND", "F1"))</pre>
coldata
        condition
##
## F1_1
               F1
## F1 2
               F1
## F1 3
               F1
## ND_1
               ND
## ND 2
               ND
## ND_3
               ND
quick start
dds <- DESeqDataSetFromMatrix(countData = cts,</pre>
                               colData = coldata,
                               design= ~ condition)
dds <- DESeq(dds)</pre>
## estimating size factors
## estimating dispersions
## gene-wise dispersion estimates
## mean-dispersion relationship
## final dispersion estimates
## fitting model and testing
resultsNames(dds) # lists the coefficients
## [1] "Intercept"
                             "condition_F1_vs_ND"
# or to shrink log fold changes association with condition:
#res <- lfcShrink(dds, coef="condition_trt_vs_untrt", type="apeglm")</pre>
MA-plot
res <- results(dds)
```

plotMA(res, ylim=c(-2,2))

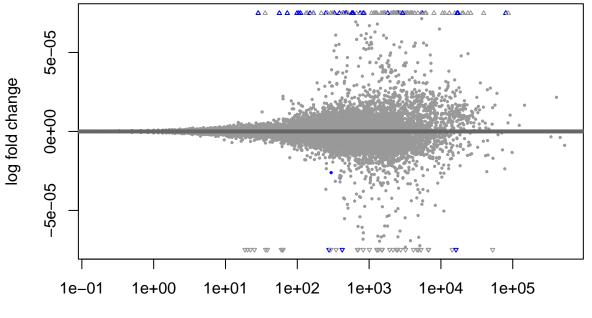


mean of normalized counts

```
resLFC <- lfcShrink(dds, coef="condition_F1_vs_ND", type="apeglm")</pre>
```

using 'apeglm' for LFC shrinkage. If used in published research, please cite:
Zhu, A., Ibrahim, J.G., Love, M.I. (2018) Heavy-tailed prior distributions for
sequence count data: removing the noise and preserving large differences.
Bioinformatics. https://doi.org/10.1093/bioinformatics/bty895

resLFC.Ordered <- resLFC[order(resLFC\$pvalue),]
plotMA(resLFC)</pre>



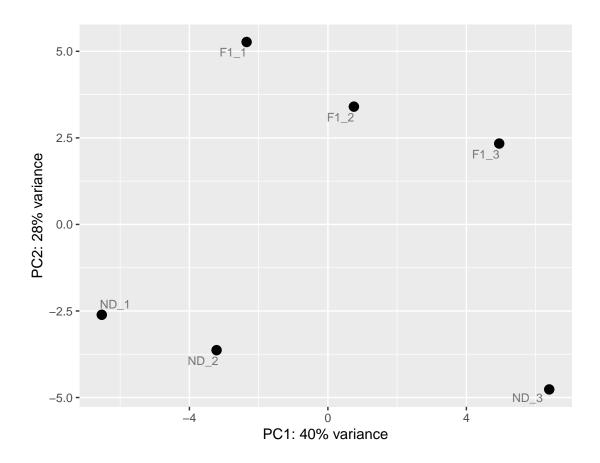
mean of normalized counts

#

PCA

```
rld <- rlog(dds, blind = FALSE)</pre>
plotPCA(rld, intgroup=c("condition"))
    5.0 -
    2.5 -
PC2: 28% variance
                                                                                            group
                                                                                                ND
    0.0 -
                                                                                                F1
   -2.5 -
   -5.0 -
                                                Ö
                         -4
                                                                      4
                                      PC1: 40% variance
pcaData <- plotPCA(rld, intgroup = c("condition"), returnData=TRUE)</pre>
percentVar <- round(100 * attr(pcaData, "percentVar"))</pre>
ggplot(pcaData, aes(PC1, PC2, label = name)) +
```

```
ggplot(pcaData, aes(PC1, PC2, label = name)) +
  geom_point(size=3) +
  xlab(paste0("PC1: ",percentVar[1],"% variance")) +
  ylab(paste0("PC2: ",percentVar[2],"% variance")) +
  coord_fixed() + geom_text_repel(alpha=0.5, size=3)
```

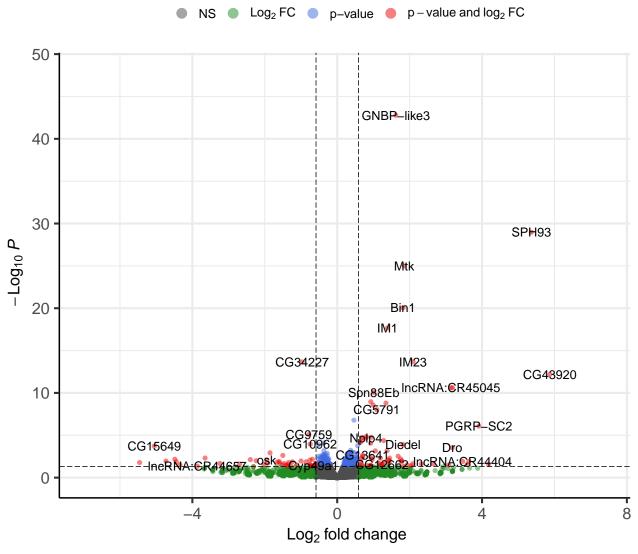


volcano plot

```
EnhancedVolcano(res,
    lab = rownames(res),
    x = 'log2FoldChange',
    y = 'pvalue',
    pCutoff = 0.05,
    FCcutoff = 0.5849625
)
```

Volcano plot

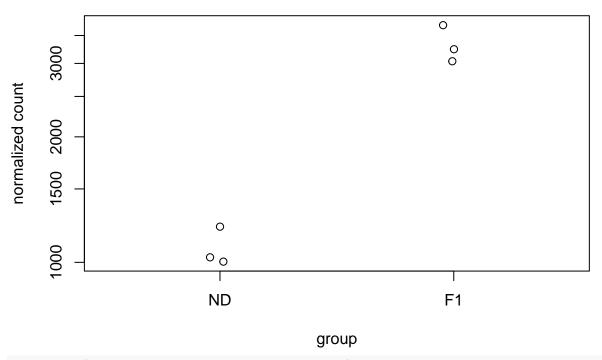
EnhancedVolcano



total = 17137 variables

plotCounts(dds, gene=which.min(res\$padj), intgroup="condition")

GNBP-like3



plotCounts(dds, gene='cad', intgroup="condition")



