

Ribo-seq quality check by riboWaltz

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General directory setting

```
wd <- here::here()
shared <- fs::path(fs::path_dir(wd), "shared")
```

Loading packages

```
library(magrittr)
library(ggplot2)
```

Load common R scripts

```
#source(fs::path(wd, "script_r", "MISC.R"))
#source(fs::path(here::here(), "script_r", "MISC_PALETTE.R"))
```

Load custom scripts

```
# Create annotation data table
source(fs::path(wd, "script_r", "create_annotation_mod.R"))
```

```
# Original code are from riboWaltz@v1.1.0 and slightly modified.
# Instead of specifying the package name in the `txdb` argument,
# we changed it so that we can pass our own txdb object directly to the `txdbanno` argument.
create_annotation_mod <- function(txdbanno = NULL) {
  exon <-
    suppressWarnings(GenomicFeatures::exonsBy(txdbanno, by = "tx", use.names = T))
  utr5 <-
    suppressWarnings(GenomicFeatures::fiveUTRsByTranscript(txdbanno, use.names = T))
  cds <-
    suppressWarnings(GenomicFeatures::cdsBy(txdbanno, by = "tx", use.names = T))
  utr3 <-
    suppressWarnings(GenomicFeatures::threeUTRsByTranscript(txdbanno, use.names = T))
  exon <- data.table::as.data.table(exon[unique(names(exon))])
  utr5 <- data.table::as.data.table(utr5[unique(names(utr5))])
  cds <- data.table::as.data.table(cds[unique(names(cds))])
  utr3 <- data.table::as.data.table(utr3[unique(names(utr3))])
  anno_df <- exon[, list(l_tr = sum(width)), by = list(transcript = group_name)]
  l_utr5 <- utr5[, list(l_utr5 = sum(width)), by = list(transcript = group_name)]
  l_cds <- cds[, list(l_cds = sum(width)), by = list(transcript = group_name)]
  l_utr3 <- utr3[, list(l_utr3 = sum(width)), by = list(transcript = group_name)]
  merge_allx <- function(x, y) merge(x, y, all.x = TRUE)
  anno_df <- BiocGenerics::Reduce(merge_allx, list(anno_df, l_utr5, l_cds, l_utr3))
  anno_df[is.na(anno_df)] <- 0
  return(anno_df)
}
```

Import genomic sequences and annotation data

```

bsg_tair <- BSgenome::getBSgenome("BSgenome.Athaliana.TAIR.TAIR9")
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:stats':
##
##     IQR, mad, sd, var, xtabs
## The following objects are masked from 'package:base':
##
##     anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##     dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##     grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##     order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##     rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##     union, unique, unsplit, which.max, which.min
##
## Attaching package: 'S4Vectors'
## The following objects are masked from 'package:base':
##
##     expand.grid, I, unname
##
## Attaching package: 'GenomicRanges'
## The following object is masked from 'package:magrittr':
##
##     subtract
##
## Attaching package: 'Biostrings'
## The following object is masked from 'package:base':
##
##     strsplit
path_gff <- fs::path(wd, "misc", "gff_gtf",
                    "Araport11_GFF3_genes_transposons.201606_mod.gff")

txdb_araport <-
  GenomicFeatures::makeTxDbFromGFF(
    file = path_gff,
    format = "gff3",
    dataSource = "Araport11",
    organism = "Arabidopsis thaliana",
    circ_seqs = c("ChrC", "ChrM"),
    chrominfo = GenomeInfoDb::seqinfo(bsg_tair)
  )
## Import genomic features from the file as a GRanges object ...
## OK
## Prepare the 'metadata' data frame ... OK
## Make the TxDb object ...
## Warning in .get_cds_IDX(mcols$type, mcols$phase): The "phase" metadata column contains non-NA
## values for features of type
##     exon. This information was ignored.
## OK

annot_dt <- create_annotation_mod(txdb = txdb_araport)

```

Run the riboWaltz pipeline

```

dir_output <- fs::path("analysis", "out_ribowaltz")
path_out <- function(...) fs::path(wd, dir_output, ...)
fs::dir_create(path_out())

sample_labels <-
  c(
    "zt0_1_ribo", "zt0_2_ribo",
    "zt3_1_ribo", "zt3_2_ribo",

```

```

"zt6_1_ribo", "zt6_2_ribo",
"zt12_1_ribo", "zt12_2_ribo",
"zt18_1_ribo", "zt18_2_ribo",
"zt21_1_ribo", "zt21_2_ribo"
) %>%
paste0(".tr.sort")

```

```

for(bam in sample_labels){
  names(bam) <- bam
  label <- stringr::str_remove(bam, ".tr.sort")

  reads_list <-
    riboWaltz::bamtolist(
      bamfolder = fs::path(wd, "data_preproc", "bam_transcriptome"),
      annotation = annot_dt,
      name_samples = bam
    )

  # read ends heatmaps
  path_out_ <- function(...) path_out("rends_heat", ...)
  fs::dir_create(path_out_())

  gp <- riboWaltz::rends_heat(reads_list, annot_dt, sample = bam)
  ggplot2::ggsave(plot = gp$plot, filename = path_out_(paste0(label, ".png")),
    width = 11, height = 6)
  readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, ".csv")))

  # p-site prediction
  path_out_ <- function(...) path_out("psite_predict", ...)
  fs::dir_create(path_out_())

  psite_offset_start <-
    riboWaltz::psite(reads_list, start = TRUE, plot_dir = path_out_(), plot = TRUE)
  readr::write_csv(
    x = tibble::as_tibble(psite_offset_start),
    file = path_out_(paste0(label, ".csv"))
  )

  reads_psite_list <- riboWaltz::psite_info(reads_list, psite_offset_start)
  rm(reads_list, gp)
  gc();gc()

  # p-site region
  path_out_ <- function(...) path_out("region_psite", ...)
  fs::dir_create(path_out_())

  gp <- riboWaltz::region_psite(reads_psite_list, annot_dt, sample = bam)
  ggsave(plot = gp$plot, filename = path_out_(paste0(label, ".png")),
    width = 8, height = 6)
  readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, ".csv")))

  # frame p-site length
  path_out_ <- function(...) path_out("frame_psite_len", ...)
  fs::dir_create(path_out_())

  gp <-
    riboWaltz::frame_psite_length(
      data = reads_psite_list,
      sample = bam,
      region = "all", cl = 90
    )

```

```

ggsave(plot = gp$plot, filename = path_out_(paste0(label, ".png")),
        width = 10, height = 5)
readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, ".csv")))

# frame p-site
path_out_ <- function(...) path_out("frame_psite", ...)
fs::dir_create(path_out_())

gp <- riboWaltz::frame_psite(data = reads_psite_list, sample = bam,
                             region = "all")
gp$plot <- gp$plot + scale_y_continuous(limits = c(0, 50))
ggsave(plot = gp$plot, filename = path_out_(paste0(label, ".png")),
        width = 10, height = 4)
readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, ".csv")))

# metaprofile p-site
path_out_ <- function(...) path_out("metaplof_psite", ...)
fs::dir_create(path_out_())

gp <-
  riboWaltz::metaprofile_psite(
    data = reads_psite_list,
    annotation = annot_dt,
    sample = bam, plot_title = label,
    utr5l = 20, cdsl = 40, utr3l = 20
  )
ggsave(plot = gp$plot, filename = path_out_(paste0(label, ".png")),
        width = 11, height = 6)
readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, "_all.csv")))

for(nt in 32:35) {
  gp <-
    riboWaltz::metaprofile_psite(
      data = reads_psite_list,
      annotation = annot_dt,
      length_range = nt,
      sample = bam, plot_title = label,
      utr5l = 20, cdsl = 40, utr3l = 20
    )
  ggsave(plot = gp$plot, filename = path_out_(paste0(label, "_", nt, ".png")),
          width = 11, height = 6)
  readr::write_csv(x = tibble::as_tibble(gp$dt), path_out_(paste0(label, "_", nt, ".csv")))
}

rm(reads_psite_list)
gc();gc()
}
Reading zt0_1_ribo.tr.sort.bam
Input reads: 58.905 M
44.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.112 M (1.887 %) reads removed: mapping on negative strand.
Output reads: 57.793 M
Done! zt0_1_ribo.tr.sort.bam has been loaded as zt0_1_ribo.tr.sort
Warning: Removed 6666 rows containing missing values (`geom_tile()`).
processing zt0_1_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting <-- 8% -->

plotting <----- 16% ----->

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plotting          <<----- 20% ----->>
plotting          <<----- 28% ----->>
plotting          <<----- 32% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 44% ----->>
plotting          <<----- 52% ----->>
plotting          <<----- 60% ----->>
plotting          <<----- 64% ----->>
plotting          <<----- 72% ----->>
plotting          <<----- 76% ----->>
plotting          <<----- 84% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt0_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt0_2_ribo.tr.sort.bam
Input reads: 43.505 M
32.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.026 M (2.357 %) reads removed: mapping on negative strand.
Output reads: 42.479 M
Done! zt0_2_ribo.tr.sort.bam has been loaded as zt0_2_ribo.tr.sort
Warning: Removed 6565 rows containing missing values ('geom_tile()').
processing zt0_2_ribo.tr.sort
best offset: 14 nts from the 5' end

plotting          <<-- 8% -->>
plotting          <<---- 16% ---->>
plotting          <<----- 20% ----->>
plotting          <<----- 28% ----->>
plotting          <<----- 32% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 44% ----->>
plotting          <<----- 52% ----->>
plotting          <<----- 60% ----->>
plotting          <<----- 64% ----->>

```

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plotting          <<----- 72% ----->>
plotting          <<----- 76% ----->>
plotting          <<----- 84% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt0_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt3_1_ribo.tr.sort.bam
Input reads: 53.506 M
40.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.160 M (2.168 %) reads removed: mapping on negative strand.
Output reads: 52.346 M
Done! zt3_1_ribo.tr.sort.bam has been loaded as zt3_1_ribo.tr.sort
Warning: Removed 7171 rows containing missing values ('geom_tile()').
processing zt3_1_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting          <<-- 8% --->
plotting          <<-- 12% --->
plotting          <<-- 20% --->
plotting          <<-- 24% --->
plotting          <<-- 32% --->
plotting          <<-- 36% --->
plotting          <<-- 44% --->
plotting          <<-- 48% --->
plotting          <<-- 56% --->
plotting          <<-- 60% --->
plotting          <<-- 68% --->
plotting          <<-- 72% --->
plotting          <<-- 80% --->
plotting          <<-- 84% --->
plotting          <<-- 92% --->
plotting          <<-- 96% --->
plotting          <<-- 100% --->

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plotting      <----- 100% ----->
processing zt3_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt3_2_ribo.tr.sort.bam
Input reads: 39.231 M
17.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.046 M (2.667 %) reads removed: mapping on negative strand.
Output reads: 38.185 M
Done! zt3_2_ribo.tr.sort.bam has been loaded as zt3_2_ribo.tr.sort
Warning: Removed 6161 rows containing missing values ('geom_tile()').
processing zt3_2_ribo.tr.sort
best offset: 14 nts from the 5' end

plotting      <-- 8% -->

plotting      <----- 16% ----->

plotting      <----- 20% ----->

plotting      <----- 28% ----->

plotting      <----- 32% ----->

plotting      <----- 40% ----->

plotting      <----- 44% ----->

plotting      <----- 52% ----->

plotting      <----- 60% ----->

plotting      <----- 64% ----->

plotting      <----- 72% ----->

plotting      <----- 76% ----->

plotting      <----- 84% ----->

plotting      <----- 88% ----->

plotting      <----- 96% ----->

plotting      <----- 100% ----->

plotting      <----- 100% ----->
processing zt3_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt6_1_ribo.tr.sort.bam
Input reads: 37.576 M
78.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.024 M (2.726 %) reads removed: mapping on negative strand.
Output reads: 36.551 M
Done! zt6_1_ribo.tr.sort.bam has been loaded as zt6_1_ribo.tr.sort
Warning: Removed 5252 rows containing missing values ('geom_tile()').
processing zt6_1_ribo.tr.sort
best offset: 13 nts from the 5' end

```

```

plotting          <-- 8% -->
plotting          <-- 16% -->
plotting          <-- 20% -->
plotting          <-- 28% -->
plotting          <-- 32% -->
plotting          <-- 40% -->
plotting          <-- 44% -->
plotting          <-- 52% -->
plotting          <-- 60% -->
plotting          <-- 64% -->
plotting          <-- 72% -->
plotting          <-- 76% -->
plotting          <-- 84% -->
plotting          <-- 88% -->
plotting          <-- 96% -->
plotting          <-- 100% -->

plotting          <-- 100% -->
processing zt6_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt6_2_ribo.tr.sort.bam
Input reads: 38.352 M
37.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.923 M (2.407 %) reads removed: mapping on negative strand.
Output reads: 37.429 M
Done! zt6_2_ribo.tr.sort.bam has been loaded as zt6_2_ribo.tr.sort
Warning: Removed 6060 rows containing missing values ('geom_tile()').
processing zt6_2_ribo.tr.sort
best offset: 14 nts from the 5' end

plotting          <-- 8% -->
plotting          <-- 16% -->
plotting          <-- 20% -->
plotting          <-- 28% -->
plotting          <-- 32% -->
plotting          <-- 40% -->
plotting          <-- 44% -->
plotting          <-- 52% -->

```



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plotting          <<----- 60% ----->>
plotting          <<----- 64% ----->>
plotting          <<----- 72% ----->>
plotting          <<----- 76% ----->>
plotting          <<----- 84% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt6_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt12_1_ribo.tr.sort.bam
Input reads: 49.633 M
58.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.574 M (1.156 %) reads removed: mapping on negative strand.
Output reads: 49.059 M
Done! zt12_1_ribo.tr.sort.bam has been loaded as zt12_1_ribo.tr.sort
Warning: Removed 9292 rows containing missing values ('geom_tile()').
processing zt12_1_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting          <<-- 8% -->>
plotting          <<---- 16% ---->>
plotting          <<----- 24% ----->>
plotting          <<----- 32% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 48% ----->>
plotting          <<----- 56% ----->>
plotting          <<----- 64% ----->>
plotting          <<----- 72% ----->>
plotting          <<----- 80% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt12_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region

```

```

Reading zt12_2_ribo.tr.sort.bam
Input reads: 39.851 M
78.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
1.266 M (3.177 %) reads removed: mapping on negative strand.
Output reads: 38.585 M
Done! zt12_2_ribo.tr.sort.bam has been loaded as zt12_2_ribo.tr.sort
Warning: Removed 4444 rows containing missing values ('geom_tile()').
processing zt12_2_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting <-- 8% -->
plotting <-- 16% -->
plotting <-- 20% -->
plotting <-- 28% -->
plotting <-- 32% -->
plotting <-- 40% -->
plotting <-- 44% -->
plotting <-- 52% -->
plotting <-- 60% -->
plotting <-- 64% -->
plotting <-- 72% -->
plotting <-- 76% -->
plotting <-- 84% -->
plotting <-- 88% -->
plotting <-- 96% -->
plotting <-- 100% -->
plotting <-- 100% -->
processing zt12_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt18_1_ribo.tr.sort.bam
Input reads: 47.327 M
40.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.688 M (1.453 %) reads removed: mapping on negative strand.
Output reads: 46.639 M
Done! zt18_1_ribo.tr.sort.bam has been loaded as zt18_1_ribo.tr.sort
Warning: Removed 7070 rows containing missing values ('geom_tile()').
processing zt18_1_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting <-- 8% -->
plotting <-- 16% -->

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plotting          <<----- 20% ----->>
plotting          <<----- 28% ----->>
plotting          <<----- 32% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 44% ----->>
plotting          <<----- 52% ----->>
plotting          <<----- 60% ----->>
plotting          <<----- 64% ----->>
plotting          <<----- 72% ----->>
plotting          <<----- 76% ----->>
plotting          <<----- 84% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>

plotting          <<----- 100% ----->>
processing ztl8_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading ztl8_2_ribo.tr.sort.bam
Input reads: 45.696 M
28.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.644 M (1.409 %) reads removed: mapping on negative strand.
Output reads: 45.052 M
Done! ztl8_2_ribo.tr.sort.bam has been loaded as ztl8_2_ribo.tr.sort
Warning: Removed 7474 rows containing missing values ('geom_tile()').
processing ztl8_2_ribo.tr.sort
best offset: 14 nts from the 5' end

plotting          <<-- 8% -->>
plotting          <<----- 16% ----->>
plotting          <<----- 20% ----->>
plotting          <<----- 28% ----->>
plotting          <<----- 36% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 48% ----->>
plotting          <<----- 56% ----->>
plotting          <<----- 60% ----->>
plotting          <<----- 68% ----->>

```

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plotting          <<----- 76% ----->>
plotting          <<----- 80% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt18_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt21_1_ribo.tr.sort.bam
Input reads: 41.954 M
48.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.710 M (1.691 %) reads removed: mapping on negative strand.
Output reads: 41.245 M
Done! zt21_1_ribo.tr.sort.bam has been loaded as zt21_1_ribo.tr.sort
Warning: Removed 7070 rows containing missing values (`geom_tile()`).
processing zt21_1_ribo.tr.sort
best offset: 13 nts from the 5' end

plotting          <<-- 8% -->>
plotting          <<---- 16% ---->>
plotting          <<---- 20% ---->>
plotting          <<----- 28% ----->>
plotting          <<----- 36% ----->>
plotting          <<----- 40% ----->>
plotting          <<----- 48% ----->>
plotting          <<----- 56% ----->>
plotting          <<----- 60% ----->>
plotting          <<----- 68% ----->>
plotting          <<----- 76% ----->>
plotting          <<----- 80% ----->>
plotting          <<----- 88% ----->>
plotting          <<----- 96% ----->>
plotting          <<----- 100% ----->>
plotting          <<----- 100% ----->>
processing zt21_1_ribo.tr.sort
1. adding p-site position
2. adding transcript region
Reading zt21_2_ribo.tr.sort.bam
Input reads: 47.392 M

```

```

29.000 (< 0.001 %) reads removed: exceeding indel_threshold.
Great! All reads' reference transcript IDs were found in the annotation table. No reads removed.
0.747 M (1.576 %) reads removed: mapping on negative strand.
Output reads: 46.645 M
Done! zt21_2_ribo.tr.sort.bam has been loaded as zt21_2_ribo.tr.sort
Warning: Removed 7272 rows containing missing values (`geom_tile()`).
processing zt21_2_ribo.tr.sort
best offset: 14 nts from the 5' end

```

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plotting          <-- 8% -->
plotting          <-- 16% -->
plotting          <-- 20% -->
plotting          <-- 28% -->
plotting          <-- 36% -->
plotting          <-- 40% -->
plotting          <-- 48% -->
plotting          <-- 56% -->
plotting          <-- 60% -->
plotting          <-- 68% -->
plotting          <-- 76% -->
plotting          <-- 80% -->
plotting          <-- 88% -->
plotting          <-- 96% -->
plotting          <-- 100% -->
plotting          <-- 100% -->
processing zt21_2_ribo.tr.sort
1. adding p-site position
2. adding transcript region

```

```

rm(annot_dt, bsg_tair, psite_offset_start, txdb_araport)
gc(); gc()

```

	used	(Mb)	gc trigger	(Mb)	limit (Mb)	max used	(Mb)
Ncells	11772353	628.8	24850738	1327.2	NA	24850738	1327.2
Vcells	799173066	6097.3	1960606682	14958.3	16384	2123736709	16202.9

	used	(Mb)	gc trigger	(Mb)	limit (Mb)	max used	(Mb)
Ncells	11772518	628.8	24850738	1327.2	NA	24850738	1327.2
Vcells	799169844	6097.2	1960606682	14958.3	16384	2123736709	16202.9

Sessioninfo

```
sessionInfo()
```

```

R version 4.2.1 (2022-06-23)
Platform: aarch64-apple-darwin20 (64-bit)
Running under: macOS Ventura 13.1

Matrix products: default
BLAS:   /Library/Frameworks/R.framework/Versions/4.2-arm64/Resources/lib/libRblas.0.dylib
LAPACK: /Library/Frameworks/R.framework/Versions/4.2-arm64/Resources/lib/libRlapack.dylib

locale:
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8

attached base packages:
[1] stats4      stats      graphics  grDevices datasets  utils      methods
[8] base

other attached packages:
[1] BSgenome.Athaliana.TAIR.TAIR9_1.3.1000
[2] BSgenome_1.64.0
[3] rtracklayer_1.56.1
[4] Biostrings_2.64.1
[5] XVector_0.36.0
[6] GenomicRanges_1.48.0
[7] GenomeInfoDb_1.32.4
[8] IRanges_2.30.1
[9] S4Vectors_0.34.0
[10] BiocGenerics_0.42.0
[11] ggplot2_3.4.2
[12] magrittr_2.0.3

loaded via a namespace (and not attached):
[1] bitops_1.0-7           matrixStats_0.62.0
[3] fs_1.5.2               bit64_4.0.5
[5] filelock_1.0.2         progress_1.2.2
[7] httr_1.4.5             rprojroot_2.0.3
[9] tools_4.2.1            utf8_1.2.2
[11] R6_2.5.1               DBI_1.1.3
[13] colorspace_2.0-3       withr_2.5.0
[15] tidyselect_1.2.0       prettyunits_1.1.1
[17] bit_4.0.5              curl_4.3.3
[19] compiler_4.2.1         textshaping_0.3.6
[21] cli_3.6.0              Biobase_2.56.0
[23] xml2_1.3.3             DelayedArray_0.22.0
[25] labeling_0.4.2         scales_1.2.1
[27] readr_2.1.4            rappdirs_0.3.3
[29] systemfonts_1.0.4      stringr_1.5.0
[31] digest_0.6.31          Rsamtools_2.12.0
[33] rmarkdown_2.24         pkgconfig_2.0.3
[35] htmltools_0.5.3        riboWaltz_1.2.0
[37] MatrixGenerics_1.8.1   dbplyr_2.3.2
[39] fastmap_1.1.0          rlang_1.1.0
[41] rstudioapi_0.14        RSQLite_2.2.18
[43] BiocIO_1.6.0           generics_0.1.3
[45] farver_2.1.1           jsonlite_1.8.4
[47] vroom_1.6.0            BiocParallel_1.30.4
[49] dplyr_1.1.1            RCurl_1.98-1.9
[51] GenomeInfoDbData_1.2.8 Matrix_1.6-4
[53] Rcpp_1.0.11            munsell_0.5.0
[55] fansi_1.0.3            lifecycle_1.0.3
[57] stringi_1.7.12         yaml_2.3.6
[59] SummarizedExperiment_1.26.1 zlibbioc_1.42.0
[61] BiocFileCache_2.4.0    grid_4.2.1
[63] blob_1.2.3             parallel_4.2.1

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[65] crayon_1.5.2	lattice_0.20-45
[67] GenomicFeatures_1.48.4	hms_1.1.3
[69] KEGGREST_1.36.3	knitr_1.42
[71] pillar_1.9.0	rjson_0.2.21
[73] codetools_0.2-18	biomaRt_2.52.0
[75] XML_3.99-0.11	glue_1.6.2
[77] evaluate_0.20	data.table_1.14.4
[79] renv_1.0.3	BiocManager_1.30.18
[81] tzdb_0.3.0	png_0.1-7
[83] vctrs_0.6.1	gtable_0.3.1
[85] cachem_1.0.6	xfun_0.40
[87] restfulr_0.0.15	ragg_1.2.5
[89] tibble_3.2.1	GenomicAlignments_1.32.1
[91] AnnotationDbi_1.58.0	memoise_2.0.1
[93] here_1.0.1	