DNAPLplume: Spreadsheet Model for Dissolved Plume Attenuation with DNAPL Source Remediation, Aqueous Decay and Volatilization – Model Calibration and First-Order Error Analysis

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1. Introduction

Dense nonaqueous phase liquids (DNAPLs) pose a difficult groundwater remediation challenge due to the impracticability of complete DNAPL source removal and to the long-term persistence of contaminant fluxes when even small DNAPL amounts remain (e.g., Soga et al., 2004). The effectiveness of natural attenuation and the feasibility of engineered remediation strategies are contingent on a number of physical and biological processes that control net source zone mass flux and attenuation within the dissolved phase plume.

A number of recent studies have focused on relationships between contaminant mass flux from DNAPL sources and the amount and distribution of DNAPL remaining in the source. Sale and McWhorter (2000) presented a semi-analytical model for dissolution rates in sources with spatially distributed DNAPL within uniform velocity fields. Their results indicated that while near-equilibrium mass transfer may occur at the local-scale, field-scale mass transfer is primarily controlled by advective-dispersive transport and the geometry of the DNAPL zones. The authors concluded that field-scale dissolution rates will diminish little over time as a function of source mass depletion. Rao and Jawitz (2003) noted that this conclusion is conditioned on assumptions of uniform flow and spatially distributed DNAPL subzones that are uniform in terms of their size, geometry and mass. When these assumptions are not met, greater reductions in contaminant fluxes over time may occur as DNAPL is more quickly depleted in regions with higher velocities and/or smaller initial masses.

Parker and Park (2004) and Park and Parker (2005) presented a field-scale mass transfer function (Parker-Park model) for DNAPL dissolution kinetics and demonstrated its ability to quantify effects of DNAPL mass depletion over time, groundwater velocity within the source zone, and variations in source zone "architecture" based on high resolution numerical simulation results. The foregoing or similar mass flux versus mass depletion relationships have been utilized by a number of authors (Rao et al. 2001, Zhu and Sykes 2004, Jawitz et al. 2005, Falta et al. 2005ab, Christ et al. 2006, Fure et al. 2006, Basu et al. 2007, Saenton and Illangasekare 2007).

While many studies have addressed the biotransformation of chlorinated solvents within dissolved phase plumes, relatively few studies have considered effects of biodecay within DNAPL source areas. Semprini et al. (1992), Mravik et al. (2003) and Ramsburg et al. (2004) have discussed various methods to enhance source zone biodecay. Mass losses due to

volatilization of organic chemicals from groundwater under natural or engineered conditions have also been studied (Jury et al., 1990; Conant et al., 1996; Auer et al., 1996; Parker, 2003). To our knowledge, no models have been presented that incorporate effects of both source zone biodecay and plume-wide volatilization losses on dissolved plume attenuation.

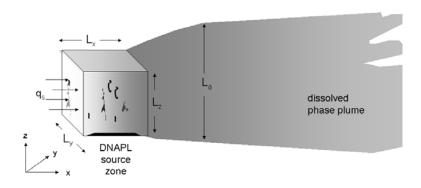


Figure 1. Source zone geometry.

2. Model Description

2.1 DNAPL Dissolution and Source Zone Biodecay

We consider a DNAPL source zone region of height L_z and width L_y with length L_x in the direction of groundwater flow in an aquifer of saturated thickness L_a (Figure 1). DNAPL, which is nonuniformly distributed within this region, undergoes mass transfer-limited dissolution to groundwater, and some of the dissolved mass biodegrades within the source zone before reaching the downgradient source zone boundary. Assuming linear field-scale mass transfer kinetics, first-order dissolved phase biodecay, and pseudo-steady-state advective transport, the areally-averaged source zone concentration distribution along the flow path may be approximated by

$$q_{s} \frac{dC}{dx} = \kappa_{eff} (C_{eq} - C) - \gamma_{s} \phi_{s} C \tag{1}$$

where q_s is the source zone darcy velocity [L T⁻¹], C is aqueous phase concentration [M L⁻³], x is distance in the direction of flow [L], C_{eq} is the equilibrium dissolved phase concentration [M L⁻³],

 γ_s is a source zone biodecay coefficient [T⁻¹], ϕ_s is source zone porosity [-], and κ_{eff} is an effective field-scale dissolution rate coefficient [T⁻¹] that may be described (Parker and Park, 2004; Park and Parker, 2005) by

$$\kappa_{eff} = \kappa_o q_s \left(\frac{M}{M_o}\right)^{\beta} \tag{2}$$

where κ_0 is a flow-normalized initial dissolution rate coefficient [L⁻¹], M_o is initial DNAPL mass [M], M is the current DNAPL mass [M], and β is a mass depletion exponent [-]. Values of β greater than one reflect dissolution rates diminish more rapidly than relative mass reduction, while values less than one indicate disproportionately slower rate reductions. Studies by Park and Parker (2005) indicate β values greater than 1 for finger-dominated residual DNAPL and less than 1 for DNAPL pools and lenses, while Falta et al. (2005ab) suggest that sites with DNAPL located predominantly in low permeability zones exhibit $\beta > 1$ while sites with DNAPL in higher permeability zones have $\beta < 1$. Integrating (1) along the source zone flow path gives

$$C(x') = \frac{C_{eq} \kappa_{eff}}{\kappa_{eff} + \gamma_s \phi_s} \left(1 - \exp\left(\frac{-(\kappa_{eff} + \gamma_s \phi_s) x'}{q_s}\right) \right)$$
(3)

where C(x') is the mean concentration within the source perpendicular to the flow direction at a distance x' from the upgradient source boundary. The net contaminant flux from the source, J_{net} [M T⁻¹], is

$$J_{net} = A_s q_s C_{out} \tag{4}$$

where $A_s = L_y L_z$ is the gross source area perpendicular to the flow direction and $C_{out} = C(x'=L_x)$, which yields

$$J_{net} = \frac{q_s A_s C_{eq} \kappa_{eff}}{\kappa_{eff} + \gamma_s \phi_s} \left(1 - \exp\left(\frac{-(\kappa_{eff} + \gamma_s \phi_s) L_x}{q_s}\right) \right).$$
 (5)

A mass balance for the source may be written as

$$\frac{dM}{dt} = -J_{tot}
= -(J_{net} + J_{bio})$$
(6)

where J_{tot} is the contaminant dissolution rate [M T⁻¹] and J_{bio} is the biodecay rate within the source zone [M T⁻¹] given by

$$J_{bio} = C_{avg} V_s \phi_s \gamma_s$$

$$= f C_{out} V_s \phi_s \gamma_s$$
(7)

in which $V_s = L_x L_y L_z$ is the gross source zone volume [L³], C_{avg} is the average dissolved phase concentration within the source zone, and $f = C_{avg}/C_{out}$ may be computed as

$$f = \frac{1}{LC_{out}} \int_{0}^{L_{x}} C(x')dx'.$$
 (8)

Integration of (8) with (3) for C(x') gives

$$f = \frac{a - 1 + \exp(-a)}{a(1 - \exp(-a))} \tag{9}$$

where $a = \kappa_{eff} L_x/q_s + \gamma_s \phi_s L_x/q_s$. Eq. (9) indicates that $f \to 0.5$ if $\kappa_{eff} L_x/q_s \ll 1$ and $\gamma_s \phi_s L_x/q_s \ll 1$ and $f \to 1$ if $\kappa_{eff} L_x/q_s \gg 1$ or $\gamma_s \phi_s L_x/q_s \gg 1$. If $\kappa_o L_x \ll 1$, then $\kappa_{eff} L_x/q_s \ll 1$ independent of source depletion and f will be controlled by the magnitude of $\gamma_s \phi_s L_x/q_s$ over time as source depletion proceeds. Therefore, if $\kappa_o L_x < 1$, we may disregard time-dependence of f and employ $a = \kappa_o L_x + \gamma_s \phi_s L_x/q_s$ with little loss of accuracy. Note that if $\kappa_{eff} L_x/q_s > 1$ with low biodecay, eq. (3) predicts the average source zone exit concentration will approach equilibrium ($C/C_{eq} > 0.63$). Since such average concentrations are rarely observed in the field, we infer that the condition $\kappa_{eff} L_x/q_s \ll 1$ is a valid approximation for most practical purposes.

From (4) and (7), we observe that

$$F_{bio} = \frac{J_{bio}}{J_{tot}} = \frac{f \gamma_s \phi_s L_x}{q_s + f \gamma_s \phi_s L_x}$$
(10)

which may be rearranged to compute γ_s from F_{bio} . The latter may be estimated from the ratio of total daughter product to primary contaminant species molar fluxes near the downgradient edge

of the source. Using F_{bio} we may express J_{net} in terms of J_{tot} as

$$J_{net} = (1 - F_{bio})J_{tot}. (11)$$

If $\kappa_o L_x < 1$ (hence $\kappa_{eff} L_x / q_s < 1$), then $C(L_x) << C_{eq}$ in eq. (3) and the magnitude of source

biodecay (γ_s) will have little effect on the computed rate of source dissolution as represented by the first term on the RHS of (3). Under these conditions, the source dissolution rate can be computed using the model of Parker and Park (2004) given here in the simplified form

$$J_{tot}(t) = J_o \left(\frac{M(t)}{M_o}\right)^{\beta}$$
 (12)

in which M(t) is the source mass at time t, M_o is the source mass at the release date $t = t_o$, and J_o is the corresponding source dissolution rate [M T⁻¹]. Substituting (12) into (11) yields

$$J_{net}(t) = (1 - F_{bio})J_o \left(\frac{M(t)}{M_o}\right)^{\beta}.$$
 (13)

We caution that although field-scale concentration mass transfer gradients may not be significantly affected by source zone biodecay, *pore-scale* concentration gradients may be affected, which, in turn, impact *field-scale* mass transfer coefficients. For convenience in model calibration and to accommodate effects of remedial actions on dissolution kinetics, we recast (13) as

$$J_{net}(t) = (1 - F_{bio\ 0}) J_{cal} \left(\frac{M(t)}{M_{cal}}\right)^{\beta} \qquad t_o < t \le t_{rem}$$

$$= (1 - F_{bio\ f}) f_{mt} J_{cal} \left(\frac{M(t)}{M_{cal}}\right)^{\beta} \qquad t > t_{rem}$$

$$(14)$$

where $J_{cal} = J_{tot}(t=t_{cal})$ and $M_{cal} = M(t=t_{cal})$ in which t_{cal} denotes a reference time used for model calibration (e.g., when reliable field data becomes available), t_{rem} is the remediation date, F_{biof} and $F_{bio 0}$ are values of F_{biof} before and after remediation, and f_{mt} is the ratio of the post- to preremediation mass transfer coefficients κ_{eff} . For example, Sorenson (2006) reported that enhanced source zone biodecay caused dissolution rate coefficients to increase by factors (i.e., f_{mt}) of 2 to 6 in laboratory studies and 3 to 8 in field studies. Also, Parker and Park (2004) have shown that field-scale dissolution rate coefficients vary in direct proportion to changes in source zone darcy flux. Thus, if remediation decreased source zone permeability by a factor of 2, for example, f_{mt} would be 0.5 in the absence of other effects.

Integration of (6) with (14) and (11) as described by Park and Parker (2005) gives source mass remaining versus time following the release date t_o subject to the stipulation that $t_{cal} \le t_{rem}$ as

$$M(t) = \begin{cases} \left[M_{cal}^{1-\beta} - (1-\beta)B(t-t_{cal}) \right]^{1/(1-\beta)} & \text{for } \beta \neq 1 \\ M_{cal} \exp\left(-B(t-t_{cal})\right) & \text{for } \beta = 1 \end{cases}$$

$$for t > t_{rem}$$

$$M(t) = \begin{cases} \left[M_{remf}^{1-\beta} - f_{mt}(1-\beta)B(t-t_{rem}) \right]^{1/(1-\beta)} & \text{for } \beta \neq 1 \\ M_{remf} \exp\left(-f_{mt}B(t-t_{rem})\right) & \text{for } \beta = 1 \end{cases}$$

$$(15)$$

where $B = J_{cal} / M_{cal}^{\beta}$, and M_{remf} is the source mass following remediation computed by

$$M_{rem f} = M_{rem 0} - \Delta M_{rem}$$

$$M_{rem 0} = \begin{cases} \left[M_{cal}^{1-\beta} - (1-\beta)B(t_{rem} - t_{cal}) \right]^{1/(1-\beta)} & \text{for } \beta \neq 1 \\ M_{cal} \exp(-B(t_{rem} - t_{cal})) & \text{for } \beta = 1 \end{cases}$$
(16)

where M_{rem0} is the source mass prior to remediation, and ΔM_{rem} is the source mass eliminated by remediation initiated at $t=t_{rem}$ and assumed to be effective immediately thereafter. The stipulation that $t_{cal} \leq t_{rem}$ is made to avoid numerical problems that can arise if t_{rem} is inadvertently set to a date when the model parameters predict the source is depleted (e.g., during the process of model calibration). The requirement does not limit the model applicability or the use of post-remediation data for calibration.

The net source flux reduction caused by remediation may be computed from (14) as

$$R_{Jrem} = \frac{J_{net f}}{J_{net 0}} = \frac{f_{mt}(1 - F_{bio f})}{(1 - F_{bio 0})} \left(\frac{M_{rem f}}{M_{rem 0}}\right)^{\beta}$$
(17)

where J_{net0} and J_{netf} denote the net source flux before and after remediation.

2.2 Volatilization from Groundwater

Volatilization from groundwater is modeled following the approach of Parker (2003), which takes into consideration dissolved phase vertical dispersion within the aquifer, vadose zone vapor diffusion and dispersion, vapor phase advection driven by cyclic barometric pressure or water table fluctuations, and aqueous phase advection in the unsaturated zone. Resulting volatilization losses from groundwater follow the first-order kinetic expression

$$s_{vol} = \lambda_{vol} \phi_a C_z \tag{18}$$

where s_{vol} is the rate of mass loss per aquifer volume due to volatilization [ML⁻³T⁻¹], λ_{vol} is a volatilization coefficient [T⁻¹], ϕ_a is aquifer porosity [-], and C_z is the dissolved phase concentration [ML⁻³] at depth z below the water table (we disregard capillary fringe thickness here for simplicity). The volatilization coefficient may be formulated as

$$\lambda_{vol} = \frac{q_u \kappa_{sat}}{\phi_a L_a \left(\exp\left(\frac{q_u L_u}{D_{eff} H}\right) - 1 \right)} \quad \text{for } q_u \neq 0$$

$$\lambda_{vol} = \frac{D_{eff} H \kappa_{sat}}{\phi_a L_a L_u} \quad \text{for } q_u = 0$$

$$(19)$$

where q_u is the unsaturated zone darcy velocity (+ downwards) [L T⁻¹], L_u is the unsaturated zone thickness (distance from ground surface to water table) [L], L_a is the saturated zone thickness over which volatilization losses are averaged [L], H is a dimensionless Henry's coefficient for the chemical of concern [-], D_{eff} is an effective diffusion coefficient for vapor transport in the unsaturated zone [L²T⁻¹], and κ_{sat} is a nonequilibrium factor for vertical mass transfer within the aquifer [-]. Note that L_a may represent the entire aquifer thickness for a vertically-integrated model or the saturated thickness of the upper model layer for a vertically-discretized model. The effective vapor phase diffusion coefficient is defined by

$$D_{eff} = D_{diff} + D_{disp} \tag{20}$$

where D_{diff} is the porous media molecular diffusion coefficient [L²T⁻¹] estimated by the Millington-Quirk model as

$$D_{diff} = \phi_{v}^{10/3} \phi_{u}^{-2} D_{v} \tag{21}$$

where ϕ_v and ϕ_u are the air-filled and total porosities, respectively, in the unsaturated zone [-], and D_v is the molecular diffusion coefficient in free air [L²T⁻¹]. Assuming a linear increase in vapor dispersivity with travel distance, vapor phase dispersion is described by

$$D_{disp} = a\varepsilon L_u \tag{22}$$

where a is the effective vapor phase velocity (L T⁻¹], and ε is the ratio of vertical dispersivity to travel distance [-] for vapor transport. For cyclic atmospheric pressure fluctuations, a may be approximated by

$$a = \frac{\phi_{\nu} \Delta P x_{bp}}{P_{o} t_{bp}} \tag{23}$$

where ΔP is the pressure fluctuation range [F L⁻²], P_o is mean atmospheric pressure [F L⁻²], t_{bp} is the period of pressure fluctuations [T], and x_{bp} is the depth [L] of pressure fluctuation propagation estimated as

$$x_{bp} = \min\left(L_u, \frac{k_a \Delta P t_{bp}}{2\phi_v \mu_a}\right) \tag{24}$$

in which k_a is the unsaturated zone vertical air permeability [L²], and μ_a is the dynamic air viscosity [ML⁻¹T⁻¹]. For cases in which water table fluctuations have a large magnitude and/or frequency (e.g., tidal zones), vapor dispersion may be described alternatively by

$$a = \frac{2\phi_{v}\Delta x_{wt}}{t_{wt}} \tag{25}$$

in which Δx_{wt} is the magnitude of water table fluctuations [L], and t_{wt} is the period [T]. The mass transfer efficiency in (19) is defined by

$$K_{sat} = \frac{1}{1 + I_{sat} / I_{soil}}$$

$$I_{sat} = \frac{HL_a / 2}{D_{wo} \phi_a^{4/3} + A_v q_a}$$

$$I_{soil} = \frac{H}{q_u} \left(\exp\left(\frac{q_u L_u}{D_{eff} H}\right) - 1 \right) \quad \text{for } q_u \neq 0$$

$$I_{soil} = \frac{L_u}{D_{eff}} \quad \text{for } q_u = 0$$

$$(26)$$

where I_{sat} and I_{soil} are impedances to transport in the saturated and unsaturated zones, respectively, D_{wo} is the free water molecular diffusion coefficient [L T⁻²], A_v is aquifer vertical dispersivity [L], and it has been assumed that the vertically-averaged dissolved concentration in (18) corresponds to the concentration at a depth $L_a/2$ below the water table.

2.3 Dissolved Phase Transport

We consider advective-dispersive dissolved phase transport in a steady-state planar velocity field downgradient of the source zone. If locations of interest are sufficiently far downgradient of the source that vertical mixing may be assumed, a 2-D transport equation may be employed as

$$R\frac{\partial C}{\partial t} = v\frac{\partial C}{\partial x} - A_L v\frac{\partial^2 C}{\partial x^2} - A_T v\frac{\partial^2 C}{\partial y^2} - \lambda_T(x)C + \frac{s(t)}{\phi_a}$$
(27)

where C is dissolved phase concentration [ML⁻³], R is a retardation factor [-], v is aquifer pore velocity [LT⁻¹] (darcy velocity divided by aquifer porosity), A_L is longitudinal dispersivity [L], A_T is transverse dispersivity [L], s(t) is the time-dependent source net flux per unit aquifer volume [MT⁻¹L⁻³], x is distance from the downgradient plane of the source [L], y is distance from the center of the source in the transverse direction [L], t is time [T], and $\lambda_T(x) = \lambda_{vol} + \lambda_{bio}$ where λ_{vol} is the groundwater volatilization coefficient [T⁻¹] and λ_{bio} is a first-order biodecay coefficient for the dissolved plume [T⁻¹], both treated as functions of longitudinal position. Specifically, we consider multiple "zones" along the axis of the plume reflecting variations in geochemical conditions (e.g., redox, DOC) that affect microbial activity or reflecting differences in conditions that affect volatilization. We also consider changes in coefficients that may occur following remedial action.

Falta et al. (2005b) presented a solution for dissolved transport with a depleting DNAPL source with constant coefficients based on Dominico (1987). Here we employ a time-convolution method for a time-dependent flux applied uniformly over the thickness of the aquifer at x=0 between $-L_y/2 < y < L_y/2$ for a contaminant-free initial condition to obtain a 2-D solution for the first aquifer "zone" located within the region $0 < x \le L_I$ as

$$C_{1}(x, y, t) = \int_{0}^{t} \frac{J_{net1}(t - \tau)}{4L_{z}L_{y}\phi_{a}(\pi RA_{L}\nu)^{1/2}} \exp\left(-\frac{\lambda_{TI}(\tau)\tau}{R} - \frac{(Rx - \nu\tau)^{2}}{4RA_{L}\nu\tau}\right) \times \left[erfc\left(-\frac{y + L_{y}/2}{2(A_{T}\nu\tau/R)^{1/2}}\right) - erfc\left(-\frac{y - L_{y}/2}{2(A_{T}\nu\tau/R)^{1/2}}\right)\right] \frac{d\tau}{\tau^{1/2}}$$
(28)

where τ is a dummy variable, $J_{netI}(t-\tau)$ is the net source mass flux (M T⁻¹) at time t- τ computed from (14) through (16), and $\lambda_{TI}(\tau)$ is the zone 1 decay coefficient ($\lambda_{volI} + \lambda_{bioI}$) that is taken as the pre-remediation value ($\lambda_{TI \ 0}$) at $\tau \leq t_{rem}$ and as the post-remediation ($\lambda_{TI \ f}$) value thereafter. To obtain the solution in the second zone at $x > L_I$, (28) is employed using the zone 2 decay coefficient ($\lambda_{T2 \ 0}$ and $\lambda_{T2 \ f}$ before and after remediation, respectively) in lieu of λ_{TI} and with a modified source function J_{net2} adjusted to correct for the difference in mass flux at $x = L_I$ using

 λ_{T1} versus λ_{T2} as follows

$$C_{2}(x, y, t) = C_{1}(x, y, t; \lambda_{T2}, J_{net2}(\tau))$$

$$J_{net2}(\tau) = J(\tau; F_{2}J_{cal1}, F_{2}M_{cal1})$$

$$F_{2} = \frac{C_{1}(L_{1}, 0, t; \lambda_{T1}, J_{net1}(\tau))}{C_{1}(L_{1}, 0, t; \lambda_{T2}, J_{net1}(\tau))}$$
(29)

where $C_1(x, y, t; \lambda, J(\tau))$ denotes the solution to (28) for specified values for x, y and t with the specified decay coefficient λ and net source flux function versus time $J(\tau)$ with other parameters identical for both zones; $J_{net2}(\tau)$ is a fictitious source function that adjusts for mass

lost in zone 1; $J(\tau; J_{cal}, M_{cal})$ represents the solution to (14) - (16) with specified values of J_{cal}

and M_{cal} ; J_{call} and M_{call} represent values of J_{cal} and M_{cal} that describe the actual source behavior; $J_{net1}(\tau) = J(\tau; J_{cal1}, M_{cal1})$ is the actual net source function; and F_2 is a flux adjustment factor. Note that (29) constrains the solution to conserve mass and maintain continuity of concentrations in time and space. Additional zones may be considered using the preceding zone solution by incrementing subscripts in (29). Spatial variations in other parameters can be treated in a similar manner subject to the constraint that the ratio of darcy velocity to aquifer thickness must remain constant to maintain a mass balance with the planar flow field.

In general, the average groundwater flow direction over the history of the plume is not precisely known, resulting in uncertainty in the coordinate system orientation. Accordingly, we consider the translation from a working coordinate system (x'-y') to a plume-oriented system (x-y) as

$$x = x'\cos\alpha - y'\sin\alpha$$

$$y = x'\sin\alpha + y'\cos\alpha$$
(30)

where the coordinate origins are assumed to be identical (i.e., source location is known) and α is a clockwise rotation angle.

3. Model Implementation

The DNAPL source and dissolved transport model described above was implemented in Excel/VBA as a function call. The format of the function is:

= CNAPL(
$$J_{cal}$$
, M_{cal} , β , L_y , L_z , t_0 , t_{cal} , q_w , ϕ , R , A_L , A_T , $F_{bio\ 0}$, λ_l , L_{12} , λ_2 , t_{rem} , ΔM_{rem} , F_{mt} , $F_{bio\ f}$, x , y , t , $mode$)

where x is distance (m) from the source downstream boundary in the direction of groundwater flow, y is lateral distance form the plume centerline, t is the date in decimal years, mode is a switch controlling function output, and all other parameters are described in Table 1.

The output modes are as follows:

If mode = 0 then CNAPL = Dissolved concentration at location (x,y) and time $t (\mu g/L)$

If mode = 1 then $CNAPL = M_{rem0}$ (kg)

If mode = 2 then $CNAPL = M_{remf}(kg)$

If mode = 3 then $CNAPL = R_{jrem}$ (-)

If mode = 4 then CNAPL = Source mass remaining at time t (kg)

If mode = 5 then CNAPL = Source flux from the source at time t (kg/d)

If mode = 6 then CNAPL = Flow-average exit concentration from source

Table 1. Summary of model parameters.

Description	Symbol	Units
Plume darcy velocity	$q_{\scriptscriptstyle W}$	m/d
Retardation factor	R	-
Longitudinal dispersivity	A_L	m
Transverse dispersivity	A_T	m
Zone 1 total decay coefficient	λ_{TI}	1/d
Zone 2 total decay coefficient	λ_{T2}	1/d
Ratio of source dissolution to source biodecay rates prior to remediation	$F_{bio\ 0}$	-
Source mass removed by source remediation	ΔM_{rem}	kg
Ratio of mass transfer coefficient post- vs pre-remediation	f_{mt}	-
Date of DNAPL release	t_o	yr
Calibration date	t_{cal}	yr
Date of source remediation	t_{rem}	yr
Dissolution flux on calibration date	$oldsymbol{J}_{cal}$	kg/d
Source mass on calibration date	M_{cal}	kg
Source depletion exponent	β	-
Coordinate rotation	α	degrees
Source width	L_{y}	m
Distance from downstream source boundary to zone 2	L_{12}	m
Aquifer thickness	$L_{a/}$	m
Aquifer porosity	ϕ	-
Ratio of source dissolution to source biodecay rates after remediation	F_{biof}	-

Decay coefficients are lumped values representing biodecay within the aquifer and volatilization losses from the water table and ground surface, which may be computed from an associated worksheet external to the VBA code. Multiple DNAPL sources (which may be co-located in time and space or not) may be modeled by adding solutions for two or more function calls. Electron donor (ED) limited biodecay may similarly be modeling by superposition of solutions for contaminant transport and ED after adjusting the ED solution for the reaction stoichiometry (i.e., mass of TCE degraded per mass of ED).

4. Model Calibration

Model calibration may be performed using a nonlinear regression method to minimize an objective function formulated as

$$O(\overline{p}) = \sum_{i=1}^{N_o} \frac{(Y_{obs\ i} - Y_i(\overline{p}))^2}{s_{oi}^2} + \sum_{j=1}^{N_p} \frac{(p_{prior\ j} - p_j)^2}{s_{pj}^2}$$
(31)

where \bar{p} is a vector of model parameters, $Y_{obs\,i}$ is the i^{th} of N_o field observations with variance s_{oi}^2 and corresponding model predictions $Y_i(\bar{p})$, and $p_{prior\,j}$ is the prior estimate of the j^{th} of N_p model parameters with estimation variance s_{pj}^2 and current regression parameter value p_j . The first term in (31) reflects errors in model predictions and the second term is a "penalty function" that constrains parameters from moving too far from prior estimates considering the uncertainty in prior information (Sadeghipour and Yeh, 1984). We assume uncertainty in the depletion exponent, release date and coordinate rotation angle to be normally distributed. Predicted concentrations and all other model parameters are assumed to be log-normally distributed. Variables in (31) corresponding to the latter therefore represent natural log-transformed populations. For log-transformed variables, the inverse weighting factors therefore become the respective ln-variances, i.e., $s_{\ln x}^2$ rather than s_x^2 . Minimization of (31) is performed using the Levenberg-Marquardt algorithm (Marquardt, 1963).

Logarithms of predicted and observed concentrations are included in the first term in the objective function. The $s_{\ln o}^2$ for concentration data may be initially assumed to be 1 and subsequently refined using the root mean square $\ln C$ regression error. The logarithm of remediation flux ratio (R_{Jrem}) can be computed by eq (17) and incorporated in the first term of

the objective function. A prior estimate may be computed as the geometric mean ratio of dissolved concentrations after remediation versus those before remediation from near-source wells.

An estimate of the post-remediation source biodecay factor (F_{biof}) may be obtained by measuring the ratio of TCE daughter products to TCE plus decay products in near-source wells. A prior estimate of the aquifer darcy velocity (q_w) may be obtained from aquifer pump tests and natural gradient measurements. A prior estimate of the retardation factor for TCE (R) may be determined from *in situ* push-pull tests, lab batch data or calculations from organic carbon data. Longitudinal and transverse dispersivities (A_L and A_L) may be initially estimated from correlations with plume size (e.g., ~ 5 and 1 %, respectively). Estimates of aquifer biodecay coefficients may be obtained from push-pull or other field tests, and volatilization coefficients may be estimated from the volatilization model in conjunction with unsaturated zone data.

A prior estimate of the pre-remediation source decay ratio ($F_{bio\,0}$ – line 8) may be computed from measurements of TCE and daughter products in wells near the source and an estimate of source mass removal during source remedial action (ΔM_{rem}) may be obtained by analysis of source treatment process data, pre-/post-remediation soil boring data, or experience from similar sites and source treatment methods (e.g., thermal, chemical oxidation, etc.). The post- versus pre-remediation mass transfer ratio (f_{ml}) is expected to be greater than 1 with various field and lab studies indicating values between 2-4 most likely.

The release date (t_o) can often be bracketed based on site operational history and refined by calibration. The dissolution flux on the calibration date (J_{cal}) may be estimated from measurements of primary contaminant and decay product concentrations near the source in conjunction with estimates of groundwater velocity and aquifer thickness, or via direct flux measurements (Annable et al., 2005). If the calibration date is selected to immediately precede remediation, the source mass on the calibration date (M_{cal}) will correspond to the pre-remediation mass (M_{cal}) . Source mass is difficult to characterize accurately. Data from soil boring may be helpful, but even so, prior estimates will be uncertain and will need to be refined by calibration.

Site characterization data will often indicate whether DNAPL pools and/or lenses are likely at the site. The source depletion exponent (β) will generally be between 0.5 and 1.0 for pools and between 1.0 and 1.5 for residual DNAPL. If both occur, then two source functions may be necessary for accurate calibration. Multi-level concentration or flux data will facilitate dual source calibration.

5. Uncertainty Analysis

We turn now to the quantification of model prediction uncertainty to assess the practical utility of the calibrated model. Following Hill (1998) and assuming log-normally distributed model predictions, we estimate total uncertainty for prediction k as

$$s_{T \ln C_{k}} = s_{R \ln C_{k}} + s_{P \ln C_{k}} \tag{32}$$

where $s_{T \ln Ci}$ is the total $\ln C$ uncertainty, $s_{R \ln Ci}$ is the regression error, and $s_{P \ln Ci}$ is the error due to parameter uncertainty. Note that the k^{th} prediction may correspond to a member of the calibration data set as well as to out-of-sample predictions.

The regression variance for prediction k with independent variables (x_k, y_k, t_k) is computed as

$$s_{R \ln C_k}^2 = MSE \left(1 + \frac{1}{N_o} + \frac{(x_k - \overline{x})^2}{\sum_{i=1}^{N_o} (x_i - \overline{x})^2} + \frac{(y_k - \overline{y})^2}{\sum_{i=1}^{N_o} (y_i - \overline{y})^2} + \frac{(t_k - \overline{t})^2}{\sum_{i=1}^{N_o} (t_i - \overline{t})^2} \right)$$
(33)

where i = 1 to N_o refers to observations used for calibration, overbars signify averages over the calibration data set, and MSE is the mean square error in concentrations used for calibration, computed as

$$MSE = \frac{\sum_{i=1}^{N_o} (\ln C_{obs i} - \ln C_{pred i})^2}{N_o - N_p}.$$
 (34)

where N_p is the number of calibrated parameters. Error due to parameter uncertainty is estimated using a first-order error propagation method (e.g., Unlu et al., 1995), which may be given in matrix notation as

$$\mathbf{s}_{\mathbf{P}\,\mathbf{ln}\,\mathbf{C}}^{2} = \mathbf{J}^{\mathbf{T}}\mathbf{Cov}\,\mathbf{J} \tag{35}$$

where the left-hand side is the vector of variances due to parameter uncertainty for all predictions of interest, J is a Jacobian matrix, and Cov is the parameter covariance matrix. Terms in the Jacobian are defined by

$$J_{ij} = \frac{\partial \ln C_i}{\partial P_j} \tag{36}$$

where P_j denotes uncertain parameter P, which as previously noted is either the actual parameter

value or its natural logarithm depending on the variable. The covariance matrix is estimated as

$$\mathbf{Cov} = MSE\left(\mathbf{J}^{\mathsf{T}}\mathbf{J}\right)^{-1}.\tag{37}$$

Confidence limits for predicted concentrations are estimated assuming log-normally distributed error as

$$CL_{\alpha} = \exp(\ln C_i \pm t_{\alpha} s_{T \ln C_i}) \tag{38}$$

where C_i is prediction i using best estimates of all model parameters, and t_{α} is the t-statistic for probability 1- α .

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