


PixelMap: An Application for Flexible Electrode Selection on Neuropixels Probes

Maxime Beau ^{1,2}, Christian Tabedzki ¹, and Carlos D. Brody ^{1,2}

¹ Princeton Neuroscience Institute, Princeton University, USA ² Howard Hughes Medical Institute, USA
 Corresponding author

DOI: [10.xxxxxx/draft](https://doi.org/10.xxxxxx/draft)

Software

- [Review](#) 
- [Repository](#) 
- [Archive](#) 

Editor: [Open Journals](#) 

Reviewers:

- [@openjournals](#)

Submitted: 01 January 1970

Published: unpublished

License

Authors of papers retain copyright and release the work under a Creative Commons Attribution 4.0 International License ([CC BY 4.0](#))

Abstract

PixelMap is a browser-based application for creating custom channelmaps for Neuropixels probes that respects electrode wiring constraints. Neuropixels probes, widely used for high-density neural recordings, have more physical electrodes than can be used for simultaneous recording because they contain fewer analogue-to-digital converters (ADCs) than data lines. Each ADC is hard-wired to several electrodes, creating complex interdependencies where selecting one electrode makes others unavailable. PixelMap provides an installation-free, browser-based interface for researchers to design arbitrary recording configurations that meet their experimental requirements while satisfying these hardware constraints. The tool generates IMRO (IMec Read Out) files compatible with SpikeGLX, the most common data acquisition software for Neuropixels recordings.

Statement of need

Neuropixels probes have revolutionised systems neuroscience by enabling simultaneous recordings from hundreds of neurons at spike resolution across multiple brain regions at any depth (Beau et al., 2021, 2025; Bondy et al., 2024; Jun et al., 2017; Steinmetz et al., 2021; Ye et al., 2025). However, configuring these probes for successful recording of neural data presents challenges. Neuropixels probes fit 960 to 5120 electrodes but can only record from 384 or 1536 channels simultaneously (Table 1), limited by the number of integrated analogue-to-digital converters (ADCs). The electrode-to-ADC wiring map follows complex patterns that vary with each Neuropixels version, making manual channel selection error-prone and time-consuming.

While existing tools like SpikeGLX and Open Ephys provide ways to edit channelmaps as .imro files, they require desktop apps and do not easily allow selection of fully arbitrary electrode geometries. Researchers often need custom channel configurations to target specific brain regions or optimise spatial sampling, but creating these configurations manually requires a deep understanding of the probe's wiring architecture and careful verification to avoid wiring violations.

PixelMap addresses these needs by:

- Being available on any machine installation-free:** The tool is available as a web application at <https://pixelmap.pni.princeton.edu> as a Python package.
- Visualising wiring constraints interactively:** When users select electrodes, the interface immediately shows which other electrodes become unavailable (marked in black) due to shared ADC lines, preventing invalid configurations.
- Supporting arbitrary electrode geometries:** Users can select electrodes through (i) picking from presets for common geometries, (ii) textually entering electrode ranges, enabling repeatable selection, (iii) dragging selection boxes and clicking on the probe

visualisation itself, and (iv) loading pre-existing .imro files. These four selection methods are intercompatible so can be used together. For instance, a SpikeGLX .imro file can be loaded as a starting point, and selection boxes used to further refine the channelmap geometry.

Probe Version	Physical Channels	Simultaneously Recordable Channels
Neuropixels 1.0	960	384
Neuropixels 2.0 (single shank)	1,280	384
Neuropixels 2.0 (4-shank)	5,120 (1,280 per shank)	384
Neuropixels 2.0 Quad Base	5,120 (1,280 per shank)	1,536

Table 1: Number of physical and simultaneously addressable electrodes across Neuropixels probe versions.

Implementation

PixelMap is implemented in Python using Holoviz' Panel (Yang et al., 2022) for the web interface, providing an interactive and responsive user experience. The software architecture consists of three main components.

First, the **wiring maps** at ./wiring_maps/*.csv are custom CSV files describing the electrode-to-ADC mappings for each supported probe type. They were built from files provided by IMEC (Neuropixels manufacturer).

Second, the **core logic** at ./backend.py implements the constraint-checking algorithms that validate electrode selections against probe-specific wiring maps. This handles the complex mapping between physical electrodes and ADC channels for different probe types (Neuropixels 1.0, 2.0 single-shank, and 2.0 four-shank so far). Hash tables (Python dictionaries) are used to query incompatible electrode pairs with $O(1)$ complexity and improve performance.

Finally, the **graphical user interface** at ./gui/gui.py was built with Holoviz' Panel. The interface provides real-time visualisation of the probe layout with electrode colour-coded based on their selection state (available in grey, selected in red, or unavailable in black). The interface supports the abovementioned four selection modes, including bokeh-based interactive click-selection and box-selection to select or deselect electrodes. User interactions trigger immediate recalculation of available electrodes based on the current selection state. This design ensures users receive instant feedback about constraint violations, preventing invalid configurations before file generation.

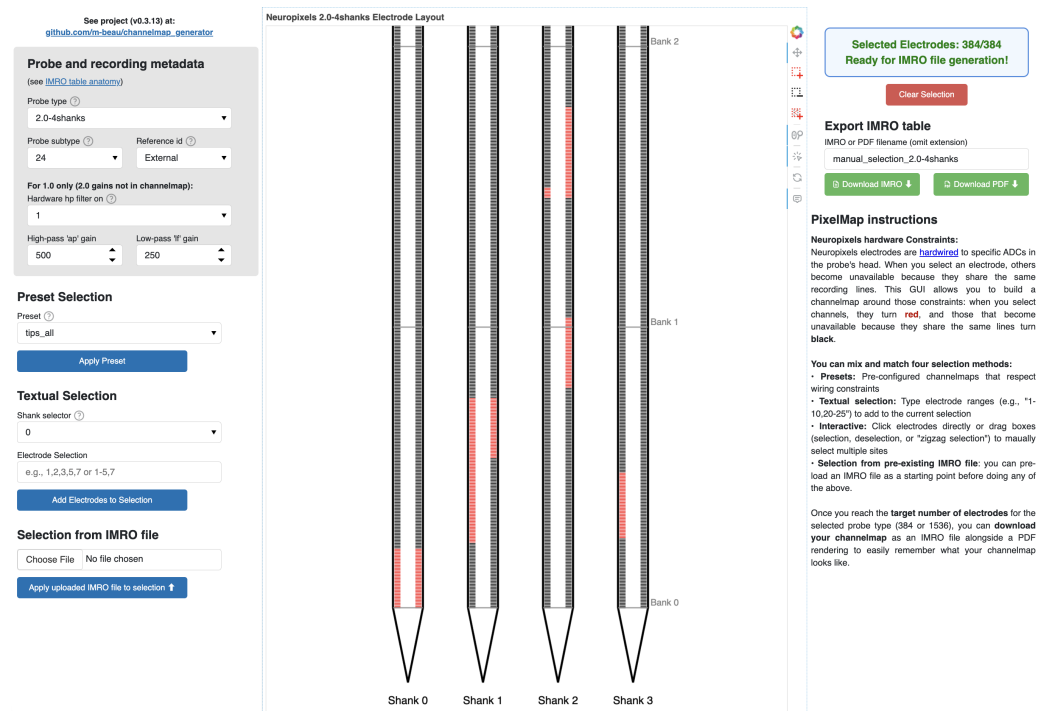


Figure 1: PixelMap's browser-based graphical user interface.

Center: Main panel featuring the probe's physical layout with one or four shanks that exhibit the 960 (Neuropixels 1.0) or 1,280 (Neuropixels 2.0) physical electrodes/shank to be selected. Electrodes available for selection are light grey, selected electrodes turn red, and electrodes that become unavailable due to hardware wiring constraints turn black. In this example, 384 electrodes have been selected (matching the maximum simultaneous recording capacity), with a distributed pattern across multiple banks, illustrating that PixelMap allows selection of arbitrary channelmap geometries.

Left: panel to input probe metadata (also part of .imro files) as well as three methods of electrode selection: preset geometries, manual textual input of electrode ranges, and pre-loading an existing .imro file. These three methods of electrode selection can be mixed together with an interactive click-and-drag box selector and deselector.

Right: electrode status indicator that turns green to confirm the selection is complete and is ready for IMRO file generation. Users can export their configuration via the "Download IMRO" button for direct use in SpikeGLX or save a PDF visualisation to easily remember the geometry of the corresponding .imro file in the future. Below the status indicator are PixelMap's instructions.

Installation and Usage

PixelMap can be used through:

1. **Web application:** Available at <https://pixelmap.pni.princeton.edu> for immediate use without installation.
2. **Local installation:** Via pip (pip install .) or uv (uv run cmap_gui) from the cloned GitHub repository.
3. **Docker container:** Users can download the image used for the website and run the container locally.
4. **Programmatic API:** Python scripts can directly call generate_imro_channelmap() for batch processing or integration into analysis pipelines.

For more details, see the project repository at https://github.com/m-beau/channelmap_generator.

The software includes an automated test suite with 41 tests covering hardware constraint validation, all preset configurations, IMRO file generation for all supported probe types, and

81 end-to-end workflows. Tests run automatically via GitHub Actions continuous integration
82 on every code change, ensuring software reliability. See the repository's tests/ directory for
83 details.

84 Author Contributions

	Maxime Beau	Christian Tabedzki	Carlos D. Brody
Conceptualisation	X		
Backend and GUI	X		
App hosting		X	
Supervision and funding			X

85 Acknowledgements

86 We thank Julie Fabre for discussions and designing PixelMap's logo. We also thank Jesse
87 C. Kaminsky, Jorge Yanar, and members of the Brody laboratory for testing and feedback
88 during development, and PNI IT members Garrett McGrath and Gary Lyons for their advice
89 concerning hosting. Finally, we thank the Princeton Neuroscience Institute for hosting the web
90 application. This work was supported by Howard Hughes Medical Institute and the National
91 Institutes of Health.

92 References

- 93 Beau, M., D'Agostino, F., Lajko, A., Martínez, G., Häusser, M., & Kostadinov, D. (2021).
94 *NeuroPyxels: Loading, processing and plotting Neuropixels data in Python*. Zenodo.
95 <https://doi.org/10.5281/zenodo.5509733>
- 96 Beau, M., Herzfeld, D. J., Naveros, F., Hemelt, M. E., D'Agostino, F., Oostland, M.,
97 Sánchez-López, A., Chung, Y. Y., Maibach, M., Kyranakis, S., Stabb, H. N., Martínez
98 Lopera, M. G., Lajko, A., Zedler, M., Ohmae, S., Hall, N. J., Clark, B. A., Cohen,
99 D., Lisberger, S. G., & others. (2025). A deep learning strategy to identify cell types
100 across species from high-density extracellular recordings. *Cell*, 188(8), 2218–2234.e22.
101 <https://doi.org/10.1016/j.cell.2025.01.041>
- 102 Bondy, A. G., Charlton, J. A., Luo, T. Z., Kopec, C. D., Stagnaro, W. M., Venditto, S. J. C.,
103 Lynch, L., Janarthanan, S., Oline, S. N., Harris, T. D., & Brody, C. D. (2024). Coordinated
104 cross-brain activity during accumulation of sensory evidence and decision commitment.
105 *bioRxiv*. <https://doi.org/10.1101/2024.08.21.609044>
- 106 Jun, J. J., Steinmetz, N. A., Siegle, J. H., Denman, D. J., Bauza, M., Barbarits, B., Lee, A. K.,
107 Anastassiou, C. A., Andrei, A., Aydin, Ç., Barbic, M., Blanche, T. J., Bonin, V., Couto, J.,
108 Dutta, B., Gratiy, S. L., Gutnisky, D. A., Häusser, M., Karsh, B., & others. (2017). Fully
109 integrated silicon probes for high-density recording of neural activity. *Nature*, 551(7679),
110 232–236. <https://doi.org/10.1038/nature24636>
- 111 Steinmetz, N. A., Aydin, C., Lebedeva, A., Okun, M., Pachitariu, M., Bauza, M., Beau,
112 M., Bhagat, J., Böhm, C., Broux, M., Chen, S., Colonell, J., Gardner, R. J., Karsh, B.,
113 Kloosterman, F., Kostadinov, D., Mora-Lopez, C., O'Callaghan, J., Park, J., & others.
114 (2021). Neuropixels 2.0: A miniaturized high-density probe for stable, long-term brain
115 recordings. *Science*, 372(6539), eabf4588. <https://doi.org/10.1126/science.abf4588>
- 116 Yang, S., Madsen, M. S., & Bednar, J. A. (2022). HoloViz: Visualization and Interactive
117 Dashboards in Python. *Proceedings of the 28th ACM SIGKDD Conference on Knowledge
118 Discovery and Data Mining*, 4846–4847. <https://doi.org/10.1145/3534678.3542621>

119 Ye, Z., Shelton, A. M., Shaker, J. R., Boussard, J., Colonell, J., Birman, D., Manavi, S., Chen,
120 S., Windolf, C., Hurwitz, C., Yu, H., Namima, T., Pedraja, F., Weiss, S., Raducanu, B. C.,
121 Ness, T. V., Jia, X., Mastroberardino, G., Rossi, L. F., & others. (2025). Ultra-high-density
122 Neuropixels probes improve detection and identification in neuronal recordings. *Neuron*.
123 <https://doi.org/10.1016/j.neuron.2025.08.030>

DRAFT