

Computer-Assisted Image Analysis I

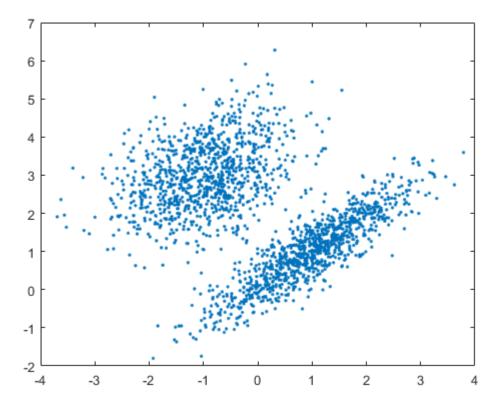
Lab4_Anam_Kumar

Md Tahseen Anam

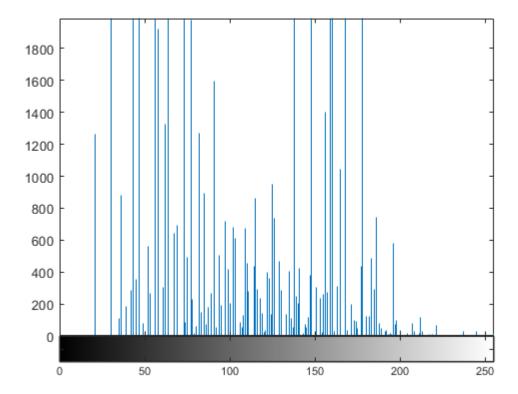
Lokesh Kumar

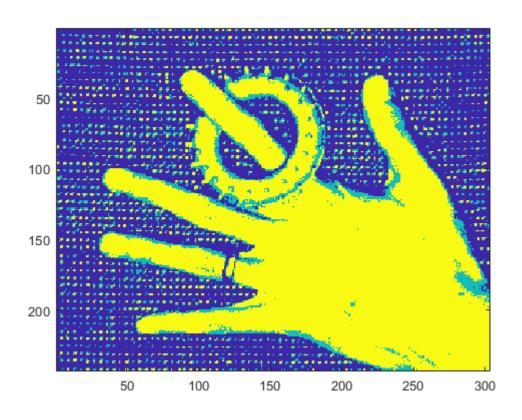
January 3, 2020

No, the two classes are not possible to separate by using either the x or y co-ordinate as a single feature. Because if we use only x co-ordinate then we will get vertical lines, using only y co-ordinates as a single feature will give us horizontal lines. This is not possible to separate because we need at least two co-ordinates to separate the classes.

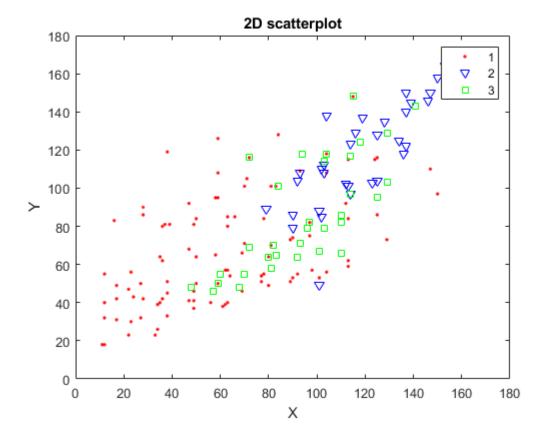


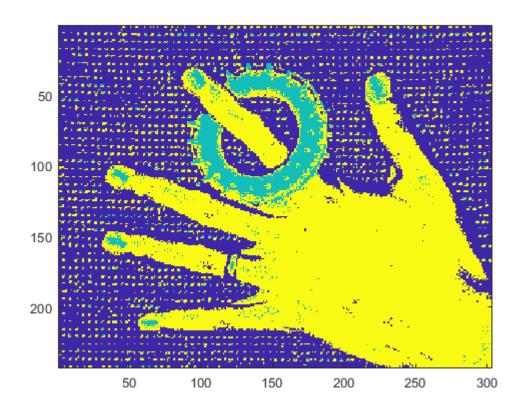
No, because for multiple objects the gray scale values overlap. That is why, in this case multiple thresholding can not successfully classify three classes. We can observe this behavior from the histogram given below.



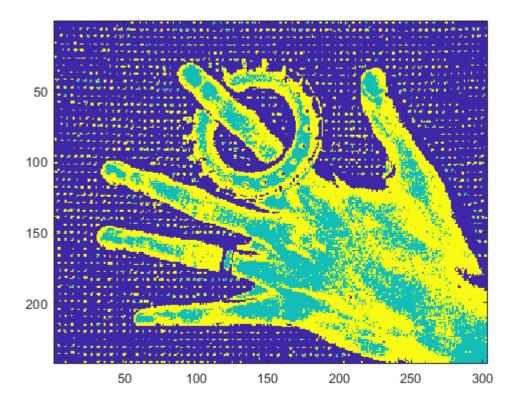


We are using linear classification model. The linear classification allows us to draw lines between classes. The assumption is that we can draw lines between different RGB bands to differentiate between classes.

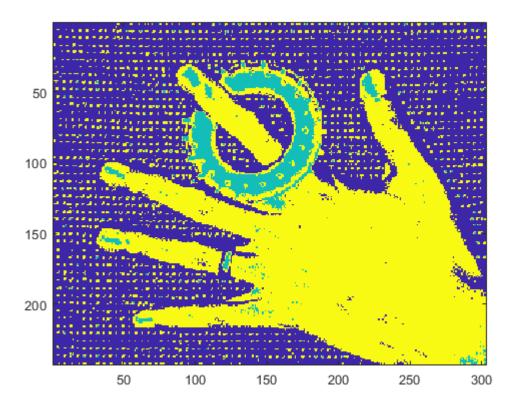




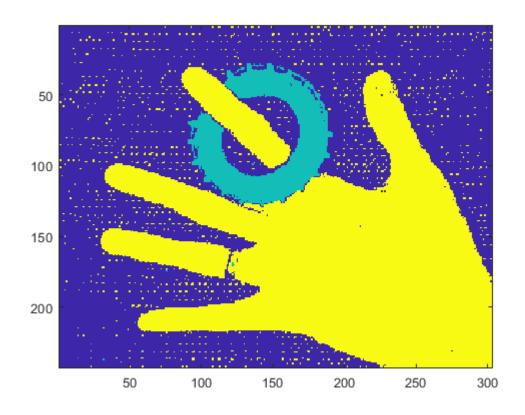
No, the result did not improve when we used classification on our grayscale image. We got the image below using grayscale image.

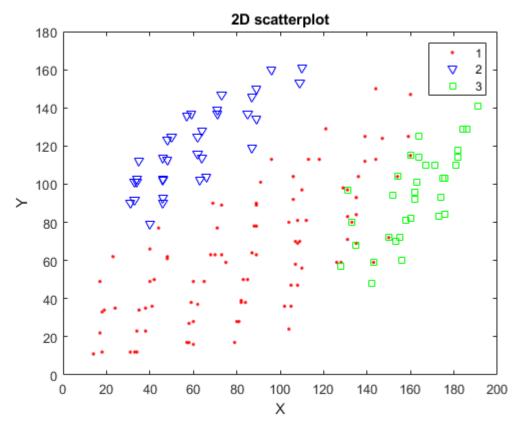


The result got improved when we used single band image instead of grayscale image. We have used green channel to create our single band image. Our classification has separated the object from the hand. This is possible as the object and the hand have different RGB values whereas they have same grayscale values in the grayscale image.



Incorporating pairs of bands will improve the classification even more. If we use full RGB information then the classification gets even better as all three channels can be used as discriminative features.



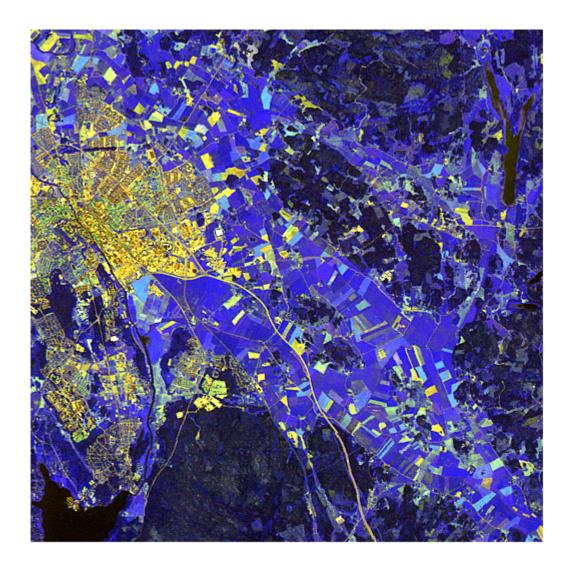


5 Source code of exercises 1,2,3,4

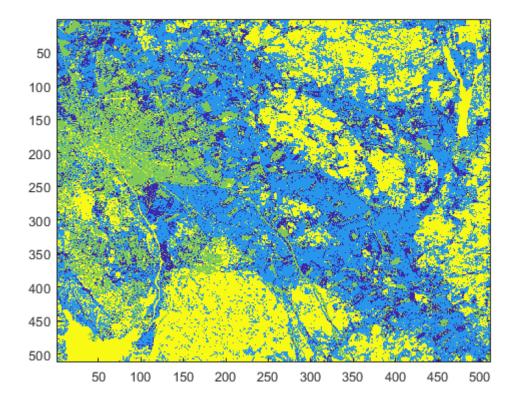
```
1 close all;
2 load cdata;
3 figure(1);
4 plot(cdata(:,1),cdata(:,2),'.');
5 %plot(cdata(:,1), '.');
6 %plot(cdata(:,2), '.');
8 I = imread ('handBW.pnm');
9 figure(2); imshow(I);
10 figure(3); imhist(I);
11 % Read the image
12 % Show the image
13 % Show the histogram
15 figure (4);
16 mtresh (I, 100, 135);
18 I2 = imread('hand.pnm');
19 % Read the image
20 figure (5);
21 imshow (I2);
22 \% Show the image
23 R = I2 (:,:,1);
24 % Separate the three layers, RGB
25 G = I2(:,:,2);
26 B = I2 (:,:,3);
27 figure (6);
28 \operatorname{plot3}(R(:),G(:),B(:),'.') % 3D scatterplot of the RGB data
30 label_im = imread('hand_training.png'); % Read image with labels
31 figure (7); imagesc(label_im);
32 \% \ View \ the \ training \ areas
33
34 I3 (:,:,1) = G;\% Create an image with two bands/features
35 I3 (:,:,2) = B;
36 [data, class] = create_training_data(I3, label_im); % Arrange the
      training data into vectors
37 figure(8);
38 %imshow(I3);
39 scatterplot2D (data, class);
40 % View the training feature vectors
42 \text{ Itest} = \text{im} 2 \text{test} \text{data} (\text{I3});
43 C = classify (double (Itest), double (data), double (class));
44 ImC = class2im (C, size (I3, 1), size (I3, 2));
45 figure (9);
46 imagesc (ImC);
```

```
47
48 I4 (:,:,1) = G;\% Create an image with single bands/features
49 [data, class] = create_training_data(I4, label_im); % Arrange the
      training data into vectors
50 figure (10);
51 %imshow(I3);
52 scatter (data, class);
53 % View the training feature vectors
55 \text{ Itest} = \text{im} 2 \text{test} \text{data} (\text{I4});
56 C = classify (double (Itest), double (data), double (class));
57 \operatorname{ImC} = \operatorname{class2im}(C, \operatorname{\mathbf{size}}(I4, 1), \operatorname{\mathbf{size}}(I4, 2));
58 figure (11);
59 imagesc(ImC);
61 \text{ I5} = \text{rgb2gray}(\text{I2});
62 [data, class] = create_training_data(I5, label_im); % Arrange the
      training data into vectors
63 figure (12);
64 %imshow(I3);
65 scatter (data, class);
66 % View the training feature vectors
67
68
69 Itest = im2testdata(I5);
70 C = classify (double (Itest), double (data), double (class));
71 ImC = class2im(C, size(I5,1), size(I5,2));
72 figure (13);
73 imagesc(ImC);
74
76 I6 (:,:,1) = R; % Create an image with two bands/features
77 I6 (:,:,2) = G;
78 I6 (:,:,3) = B;
79 [data, class] = create_training_data(I6, label_im); % Arrange the
      training data into vectors
80 figure (14);
81 \% imshow(I3);
82 scatterplot2D (data, class);
83 % View the training feature vectors
85 Itest = im2testdata(I6);
86 C = classify(double(Itest), double(data), double(class));
87 ImC = class2im (C, size (I6,1), size (I6,2));
88 figure (15);
89 imagesc(ImC);
```

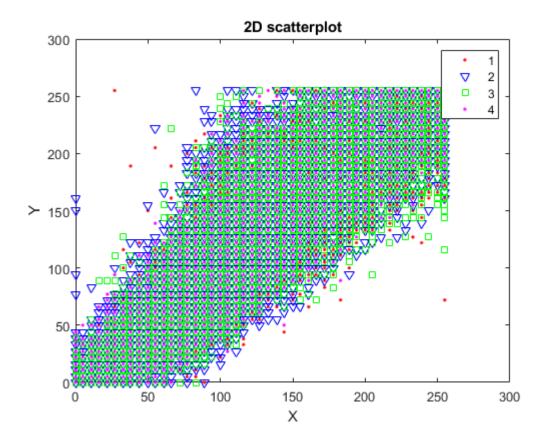
We have used bands 1, 3 and 4. Because we can discriminate buildings, forest and field using these bands.



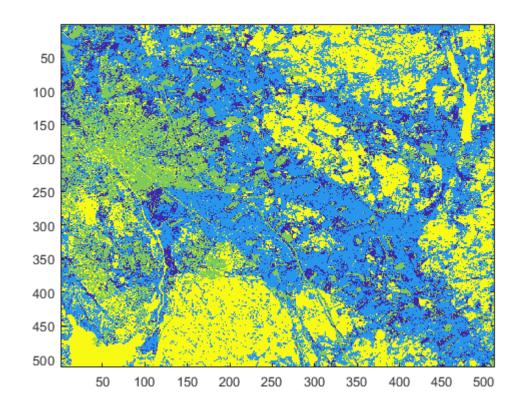
We have used quadratic classifier to do our classification and classified using the selected bands 1,3 and 4 and got the following result.

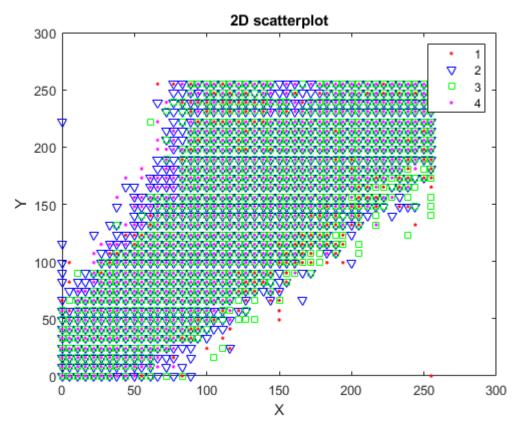


Here is our scatterplot figure.



If we use all bands for classification, we do not see much difference. Here is the result of classification using all bands.





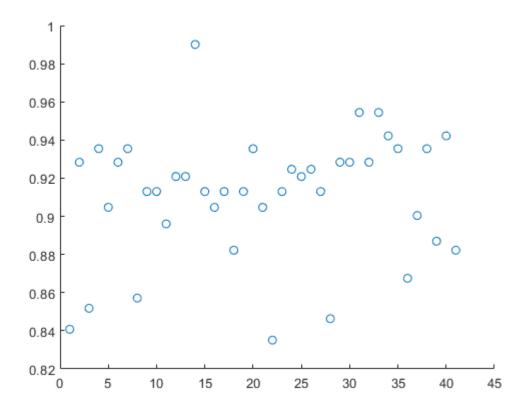
Using all bands for classification may overfit the image and the result may remove some details. It will also slow down the classification process.

8 Source code of exercises 5,6

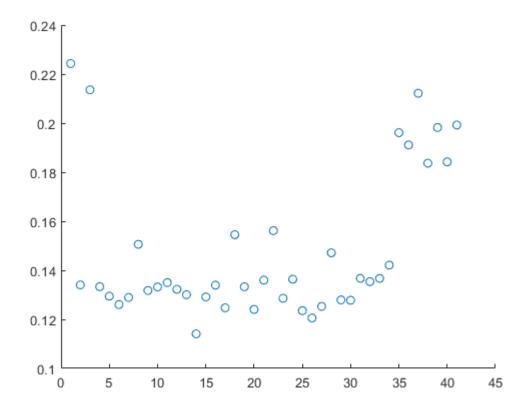
```
1 close all;
2 clear all;
3 load cdata;
4 load landsat_data;
5 I2 = landsat_data;
6 figure;
7 imshow(landsat_data(:,:,[1,3,4])./255);
9 % Show the image
10 R = I2 (:,:,1);
11 % Separate the three layers, RGB
12 G = I2 (:,:,3);
13 B = I2 (:,:,4);
14
15
16 label_im = imread('hand_training.png'); % Read image with labels
17 figure (7); imagesc(label_im);
18 \text{ label_im} = \text{imresize}(\text{label_im}, [512,512]);
19 label_im (1:100, 1:100) = 1;
20 \text{ label_im} (101:200, 101:200) = 3;
21 \text{ label_im} (101:200,201:300) = 4;
22 \text{ label_im} (201:512,301:512) = 2;
23
25 I6 (:,:,1) = R; % Create an image with three bands/features
26 \text{ I6} (:,:,2) = G;
27 I6 (:,:,3) = B;
28 [data, class] = create_training_data(I6, label_im); % Arrange the
      training data into vectors
29 figure (14);
30 \% imshow(I3);
31 scatterplot2D (data, class);
32 % View the training feature vectors
34 \text{ Itest} = \text{im} 2 \text{test} \text{data} (\text{I6});
35 C = classify (double (Itest), double (data), double (class), 'quadratic');
36 \text{ ImC} = \text{class2im} (C, \text{size} (16, 1), \text{size} (16, 2));
37 figure (15);
38 imagesc(ImC);
39
```

Using all bands for classification may overfit the image and the result may remove some details. It will also slow down the classification process.

This is the resulted Entropy.



Below is the resulted Uniformity.



From the above images of entropy and uniformity, we can say that outliers are the ones with low entropy and high uniformity. So, those objects should be removed. Real virus should have higher entropy because of the randomness of nearly same grayscale value. Images with fewer different grayscale values have higher uniformity. So, real viruses should have lower uniformity.

10 Source code of exercises 7

```
1 close all;
3 %% Read image and template from disk
5 % Read image
6 I = imread('viruses.tif');
8 % Read template
9 template = imread('virusTemplate.tif');
11 % Show image and template
12 figure ('name', 'Original image'); imshow(I);
13 figure ('name', 'Template'); imshow (template);
15 % Template matching
17 % Select step length (How many pixels the template is moved in each
18\% iteration)
19 \text{ stepSize} = 10;
20
21 % Do the template matching
22 ccimg = templatematching(I, template, stepSize);
23
24 % Show the resulting correlation image
25 \ \% figure \ ('name', 'correlation coefficients'); imshow \ (ccimg, []); colormap
      (copper); colorbar;
26
27
28 % Find localmaxima in the correlation image
30 maxima = imextendedmax (ccimg, 0, 4);
32 % Shrink objects to points
33 maxima = bwmorph(maxima, 'shrink', inf);
35~\%~Calculate~the~correlation~coefficients~for~the~maxima
36 maxvals = maxima .* ccimg;
37
38~\%~Pick~the~corresponding~correlation~values~and~threshold~them~sung
      Otsu's
39 \% method.
40 h = \mathbf{hist} (maxvals (maxvals > 0),128);
41 \text{ h2} = \text{imfilter}(h, [1 \ 1 \ 1 \ 1 \ 1] \ ./ \ 5);
42 \text{ thresh} = \text{graythresh}(h2);
44 % Create a binary image keeping only those with a correlation value
      that
```

```
45\%\ lies\ above\ the\ threshold.
46 \text{ maxvals}(\text{maxvals} < \text{thresh}) = 0;
47 \text{ maxvals} (\text{maxvals} = 0) = 1;
48
49 \% Label the binary image and obtain their positions
50 \text{ maxlbl} = \log \operatorname{ical}(\text{maxvals});
51 maxcentroids = regionprops(maxlbl, 'centroid');
52
53 % Create a circular mask which will be used to cutout individual
      objects
54 % from the original image.
55 [h,w] = size(template);
56 d = \max([h w]);
57 \text{ mask} = \text{imcircle}(d);
59 % Cut out all objects using the mask and store it in a cell array
60 % named "cutouts".
61 nrOfObjects = length(maxcentroids);
62 cutouts { nrOfObjects } = [];
63 figure ('name', 'Segmentation result'); imshow(I); hold on;
64 \text{ for } i = 1 : \text{nrOfObjects}
       % The position corresponding to the centroid has to be
65
           calculated since
       % the centroid position depends on the step size (stepSize) used
66
       realpos = [((maxcentroids(i).Centroid(1)-1)*stepSize+1)]
67
           \maxcentroids (i). Centroid (2) -1)*stepSize+1)];
68
       cutouts\{i\} = I(realpos(2): realpos(2)+d-1, realpos(1): realpos(1)+d
69
           -1);
       cutouts{i} = cutouts{i} .* uint8(mask);
70
71
72
       rectangle ('Position', [realpos(1), realpos(2), d, d],...
73
            'Curvature', [1,1],...
74
            "edgecolor", "w", \dots
75
            'linewidth', 1);
76
77 end
78 hold off;
79
80 %% Adding one noisy disk and one gray disk
81
82 %refimg = noise(100 .* ones(size(mask)), 'mu', 1);
83 \% cutouts \{1, end+1\} = uint8 (refimg.* mask);
84 \% cutouts \{1, end+1\} = uint8 (mask) .* 128;
86 % Collect all cutouts in an image
87
88 imagesPerRow = 10;
```

```
89 \text{ cols} = \text{imagesPerRow};
90 rows = \mathbf{ceil}(\mathbf{size}(\text{cutouts}, 2) / \text{cols});
92 objectMap = uint8(zeros(size(mask).*[rows cols]));
93 \text{ counter} = 1;
94
95 for i = 1 : rows
        for j = 1 : cols
96
            if counter <= size(cutouts,2)
97
                 objectMap((i-1)*d+1:(i-1)*d+d,(j-1)*d+1:(j-1)*d+d) =
98
                    cutouts { counter } (:,:);
                 counter = counter + 1;
99
100
            end
       end
101
102 end
103
104 figure ('name', 'Cutouts of segmented objects'); imshow (object Map);
105
106 %% cleaning the workspace from surplus variables
107 keep('I', 'template', 'mask', 'cutouts', 'nrOfObjects'); %'objectMap');
109 % Texture analysis
110 %
111 \% I
                     - original image
112 \% template
                     - the template used in the template matching
      procedure
113 \% cutouts
                     - a cell array with all the segmented objects
114 % nrOfObjects
                     - the number of objects in 'cutouts'
115 % mask
                     - the binary mask used to cutout the objects
116 % objectMap
                     - a compiled image of all the objects in 'cutouts'
117 %
118
120 % Place for your own texture analysis.
121
122 figure, imshow(cutouts {1});
123
   for n = 1:nrOfObjects
124
       gcm = graycomatrix(cutouts\{n\}, 'Offset', [0\ 1;\ -1\ 1;\ 1\ -1];
125
           -1]);
        en(n) = entropy(gcm);
126
        energy = graycoprops(gcm, 'Energy Correlation');
127
       un(n) = mean(energy.Energy);
128
129 end
130
131
132 figure;
133 title ('Uniformity');
134 scatter ([1:41], un);
```

```
135
136 figure;
137 title('Entropy');
138 scatter([1:41],en);
```