W203-2, Week 15, Lab 4

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library(car)  
library(lmtest)

## Loading required package: zoo

##   
## Attaching package: 'zoo'

## The following objects are masked from 'package:base':  
##   
## as.Date, as.Date.numeric

library(sandwich)  
library(stargazer)

##   
## Please cite as:

## Hlavac, Marek (2015). stargazer: Well-Formatted Regression and Summary Statistics Tables.

## R package version 5.2. http://CRAN.R-project.org/package=stargazer

# Introduction

The United States is known to have the highest prison population in the world. Our team has been hired by a political campaign to provide research in identifying factors that influence the probability of getting sentenced () for the offences committed. By identifying these factors, the team hopes to help the campaign formulate possible legislative actions that the government could undertake in reducing such crimes and hence the number of inmates in the prisons.

# Initial exploratory analysis

The file crime.csv contains crime statistics for a selection of counties. While there may be other factors not included in the dataset that are contributing to jail sentences and hence possibly introduce omitted variable biases in our final models, we have a pretty comprehensive set of variables given in the dataset ranging from crime, geography, economic, and demographics of the counties included in the dataset each of which we will delve into shortly.

Data <- read.csv('crime\_v2\_updated.csv')  
head(Data)

## X county year crime probarr probsen probconv avgsen police  
## 1 1 1 88 0.0356036 0.436170 0.298270 0.5275960 6.71 0.00182786  
## 2 2 3 88 0.0152532 0.450000 0.132029 1.4814800 6.35 0.00074588  
## 3 3 5 88 0.0129603 0.600000 0.444444 0.2678570 6.76 0.00123431  
## 4 4 7 88 0.0267532 0.435484 0.364760 0.5254240 7.14 0.00152994  
## 5 5 9 88 0.0106232 0.442623 0.518219 0.4765630 8.22 0.00086018  
## 6 6 11 88 0.0146067 0.500000 0.524664 0.0683761 13.00 0.00288203  
## density tax west central urban pctmin wagecon wagetuc  
## 1 2.4226327 30.99368 1 0 0 20.21870 281.4259 408.7245  
## 2 1.0463320 26.89208 1 0 0 7.91632 255.1020 376.2542  
## 3 0.4127659 34.81605 0 1 0 3.16053 226.9470 372.2084  
## 4 0.4915572 42.94759 1 0 0 47.91610 375.2345 397.6901  
## 5 0.5469484 28.05474 0 1 0 1.79619 292.3077 377.3126  
## 6 0.6113361 35.22974 0 1 0 1.54070 250.4006 401.3378  
## wagetrd wagefir wageser wagemfg wagefed wagesta wageloc mix  
## 1 221.2701 453.1722 274.1775 334.54 477.58 292.09 311.91 0.08016878  
## 2 196.0101 258.5650 192.3077 300.38 409.83 362.96 301.47 0.03022670  
## 3 229.3209 305.9441 209.6972 237.65 358.98 331.53 281.37 0.46511629  
## 4 191.1720 281.0651 256.7214 281.80 412.15 328.27 299.03 0.27362204  
## 5 206.8215 289.3125 215.1933 290.89 377.35 367.23 342.82 0.06008584  
## 6 187.8255 258.5650 237.1507 258.60 391.48 325.71 275.22 0.31952664  
## ymale  
## 1 0.07787097  
## 2 0.08260694  
## 3 0.07211538  
## 4 0.07353726  
## 5 0.07069755  
## 6 0.09891920

n <- nrow(Data)  
num\_cols <- ncol(Data)

head() confirms that the data has been succesfully loaded. The dataset contains 26 columns (variables) and 90 rows. This is sufficiently large enough to assume CLT.

# Check for NAs  
for(i in names(Data)){  
 val <- Data[[i]][is.na(Data[[i]])]  
 if(length(val)) {  
 sprintf("%s: %d NA row(s) found", i, length(val))  
 }  
}

No NAs are found in the dataset given.

## Individual variable analysis

### X

This is just an index variable and hence no analysis is required.

### Country identifier

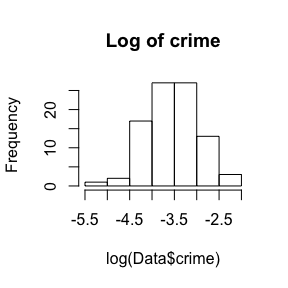
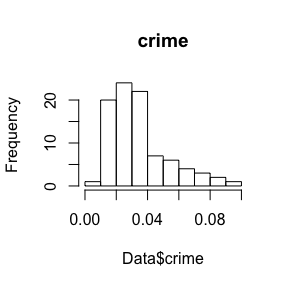
This is just an identifier and hence no analysis is required.

### Year

This is just the year when this data was collected and it is simply 88 for all rows. No analysis requried.

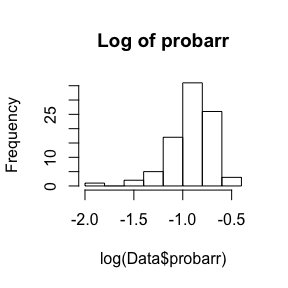
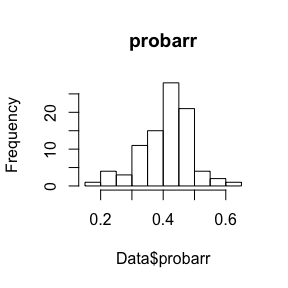
### Crime committed per person

hist(Data$crime, main = "crime")  
hist(log(Data$crime), main = "Log of crime")

 The histogram is positively skewed. No extreme outliers observed. The histogram becomes more normal when log() is applied.

### ‘Probability’ of arrest

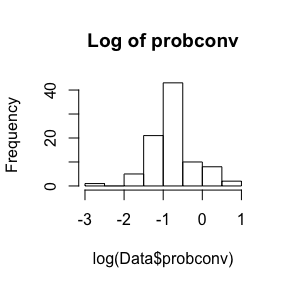
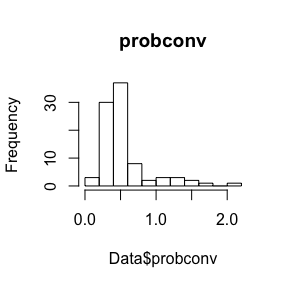
hist(Data$probarr, main = "probarr")  
hist(log(Data$probarr), main = "Log of probarr")

 The histogram is relatively normal. No extreme outliers observed. The histogram actually becomes less normal when log() is applied.

### ‘Probability’ of conviction

hist(Data$probconv, main = "probconv")  
hist(log(Data$probconv), main = "Log of probconv")  
(length(Data$probconv[Data$probconv > 1]))

## [1] 10

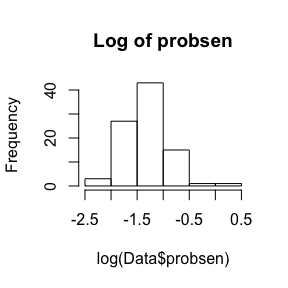
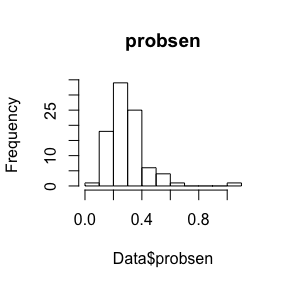


The histogram is positively skewed with extreme outliers (10 items over 1). The histogram becomes more normal when log() is applied.

### ‘Probability’ of prison sentence

hist(Data$probsen, main = "probsen")  
hist(log(Data$probsen), main = "Log of probsen")  
(length(Data$probsen[Data$probsen > 1]))

## [1] 1



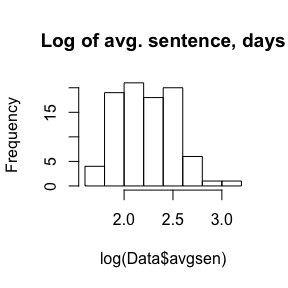
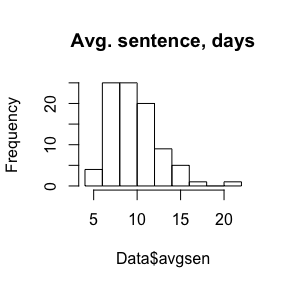
The histogram is relatively normal with an exception of one extreme outlier (10 items over 1). The histogram becomes more normal when log() is applied.

### Avg. sentence, days

hist(Data$avgsen, main = "Avg. sentence, days")  
(length(Data$probsen[Data$avgsen > 20]))

## [1] 1

hist(log(Data$avgsen), main = "Log of avg. sentence, days")



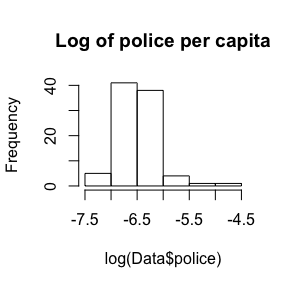
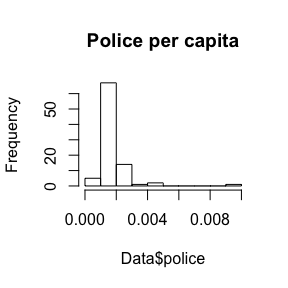
The histogram is slightly positively skewed with 1 outlier (20 >). The histogram becomes more normal when log() is applied.

### Police per capita

hist(Data$police, main = "Police per capita")  
(length(Data$probsen[Data$police > 0.009]))

## [1] 1

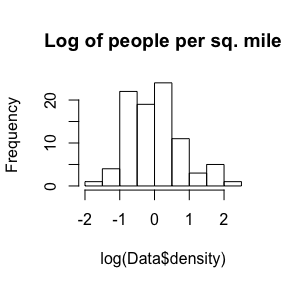
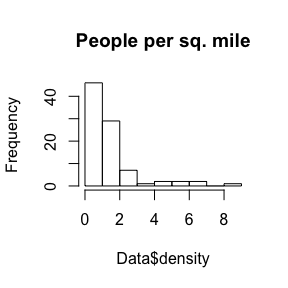
hist(log(Data$police), main = "Log of police per capita")



The histogram is positively skewed with 1 outlier. The histogram becomes slightly more normal when log() is applied.

### People per sq. mile

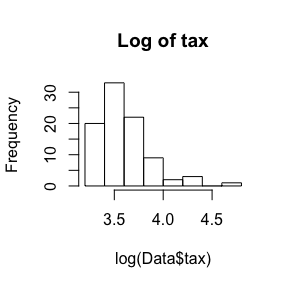
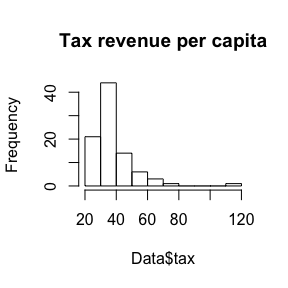
hist(Data$density, main = "People per sq. mile")  
hist(log(Data$density), main = "Log of people per sq. mile")



The histogram is positively skewed. The histogram becomes more normal when log() is applied.

### Tax revenue per capita

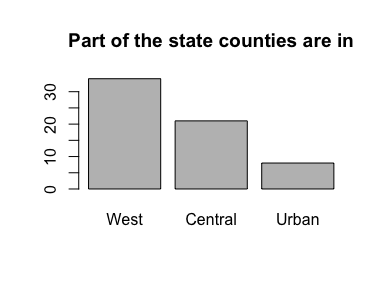
hist(Data$tax, main = "Tax revenue per capita")  
hist(log(Data$tax), main = "Log of tax")



The histogram is positively skewed. The histogram becomes slightly more normal when log() is applied however is still positively skewed.

### West/Central/Urban

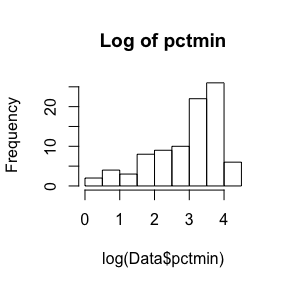
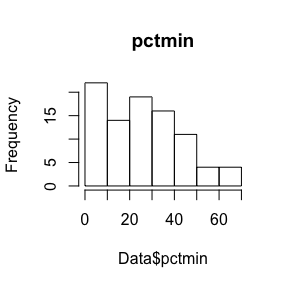
barplot(c(sum(Data$west), sum(Data$central), sum(Data$urban)),   
 names.arg = c("West", "Central", "Urban"), main = "Part of the state counties are in")  
sum\_geo <- sum(Data$west) + sum(Data$central) + sum(Data$urban)



Dummy variables indicating whether or not a given county is in the western/central/urban part of the state. Interestingly, the sum of the 3 regions only add up to 63 which is considerably less than our n of 90. There are many 27 counties in the dataset do not fall under any of these regions.

### Proportion that is minority or nonwhite

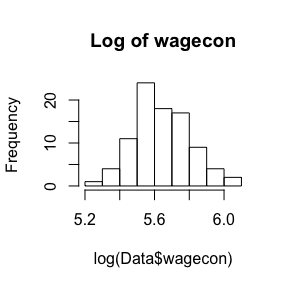
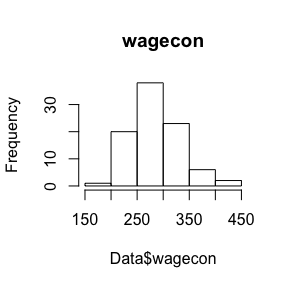
hist(Data$pctmin, main = "pctmin")  
hist(log(Data$pctmin), main = "Log of pctmin")



The histogram is positively skewed. The histogram becomes more normal when log() is applied althogh it is still negatively skewed.

### Weekly wage, construction

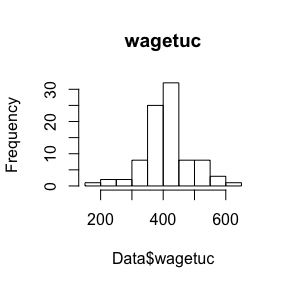
hist(Data$wagecon, main = "wagecon")  
hist(log(Data$wagecon), main = "Log of wagecon")



The histogram is pretty normal. The histogram becomes more normal when log() is applied.

### Weekly wage, transportation, utilities, communications

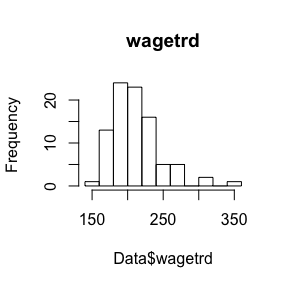
hist(Data$wagetuc, main = "wagetuc")



The histogram is relatively normal.

### Weekly wage, wholesale, retail trade

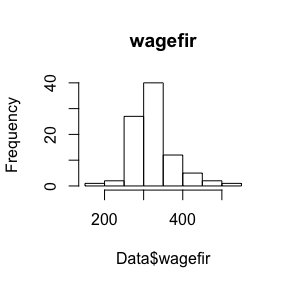
hist(Data$wagetrd, main = "wagetrd")



The histogram is relatively normal with some outliers.

### Weekly wage, finance, insurance and real estate

hist(Data$wagefir, main = "wagefir")

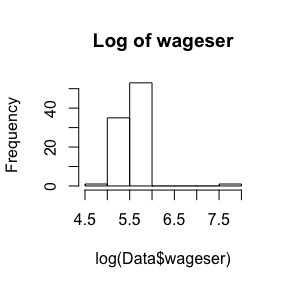
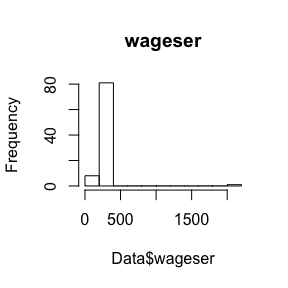


The histogram is relatively normal.

### Weekly wage, service industry

hist(Data$wageser, main = "wageser")  
hist(log(Data$wageser), main = "Log of wageser")  
max(Data$wageser)

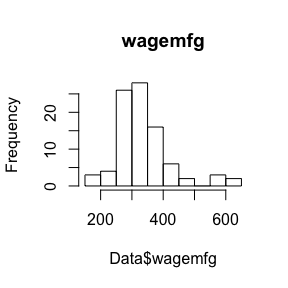
## [1] 2177.068



The histogram is positively skewed with one extreme outlier. The histogram becomes slightly more normal when log() is applied.

### Weekly wage, manufacturing

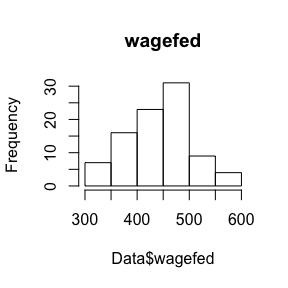
hist(Data$wagemfg, main = "wagemfg")



The histogram is relatively normal but with some outiers.

### Weekly wage, federal employees

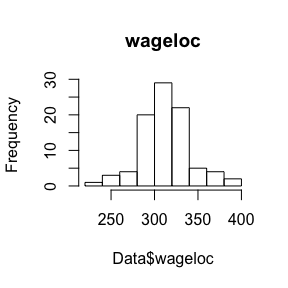
hist(Data$wagefed, main = "wagefed")



The histogram is relatively normal.

### Weekly wage, local government employees

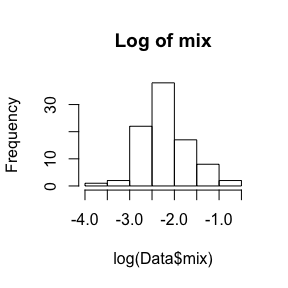
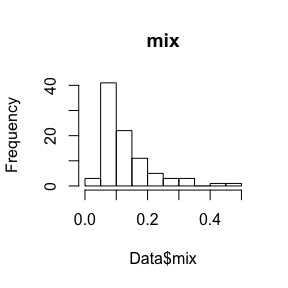
hist(Data$wageloc, main = "wageloc")



The histogram is pretty normal.

### Ratio of face to face/all other crimes

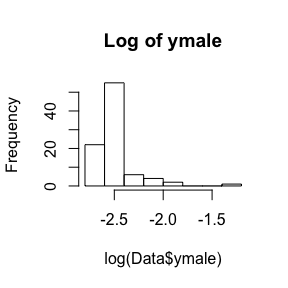
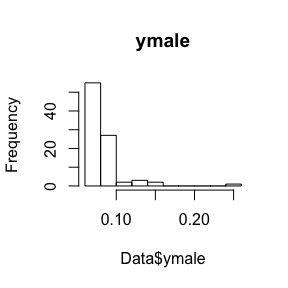
hist(Data$mix, main = "mix")  
hist(log(Data$mix), main = "Log of mix")



The histogram is positively skewed. The histogram becomes more normal when log() is applied.

### Proportion of county males between the ages of 15 and 24

hist(Data$ymale, main = "ymale")  
hist(log(Data$ymale), main = "Log of ymale")

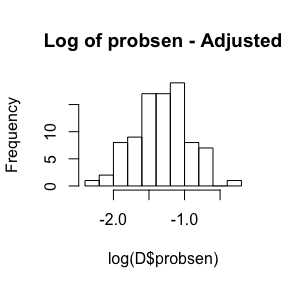
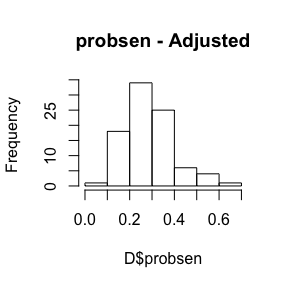
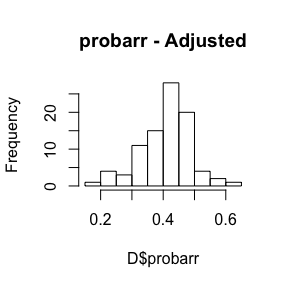


The histogram is positively skewed. The histogram becomes more normal when log() is applied however it is still positively skewed.

## Variable transformations

and contain values > 1 which are difficult to intrepret. For the purpose of the study, I am omitting them.

D <- Data[Data$probarr < 1 & Data$probsen < 1,]  
hist(D$probarr, main = "probarr - Adjusted")  
hist(D$probsen, main = "probsen - Adjusted")  
hist(log(D$probsen), main = "Log of probsen - Adjusted")



Without the outliers, the 2 histograms are looking relatively normal. is still looking positively skewed so I am taking the log of it which makes the distribution more normal.

Now I will apply log() to the below variables as they are not very normally distributed and store them in the newly created dataframe D so they will be available for my models.

D$logcrime <- log(D$crime)  
D$logavgsen <- log(D$avgsen)  
D$logpolice <- log(D$police)  
D$logprobconv <- log(D$probconv)  
D$logprobsen <- log(D$probsen)  
D$logdensity <- log(D$density)  
D$logtax <- log(D$tax)  
D$logpctmin <- log(D$pctmin)  
D$logwagecon <- log(D$wagecon)  
D$logwageser <- log(D$wageser)  
D$logmix <- log(D$mix)  
D$logymale <- log(D$ymale)

# Models

The team wants to explore how much is accounted for by the crime and police-related variables such as number of crimes committed (), police per capita () and ratio of face-to-face/all other crimes () and how much of it can be attributed to other demographic variables such as race, gender, age, economic standings (wages).

## Proposed Model 1 - Minimum specification

: We can intuitively anticipate probsen to go up as , and increase. My intuition would be and to have a positive correlation as increased would suggest there would be more severe crimes happening in a given county.

: I expect the other 2 probability variables and to have strong correlations with probsen and hence they will also be included in the model so we can measure how much influence the other variables have on holding and fixed.

For this initial model, I will exclude other demographic variables.

model1 <- lm(logprobsen ~ logcrime + probarr + logprobconv + logavgsen + logpolice + logmix, data = D)

### CLM Assessment

#### CLM 1 - A linear model

The model is specified such that the dependent variable is a linear function of the explanatory variables.

Is the assumption valid?

#### CLM 2 - Random samling

As the dataset has been provided for a selection of counties, the data is not truly randomly sampled. We are not given much information about how the data in the CSV file has been collected. We will assume here that the data has been collected from the relevant random samples in these counties.

Is the assumption valid?

#### CLM 3 - Multicollinearity

X <- data.matrix(subset(  
 D, select = c("logprobsen", "logcrime", "probarr", "logprobconv", "logavgsen", "logpolice", "logmix")))  
(Cor = cor(X))

## logprobsen logcrime probarr logprobconv logavgsen  
## logprobsen 1.00000000 -0.360492812 -0.04064202 -0.31311633 -0.12188311  
## logcrime -0.36049281 1.000000000 0.06321588 -0.32628681 0.13418145  
## probarr -0.04064202 0.063215878 1.00000000 -0.02533560 -0.17225398  
## logprobconv -0.31311633 -0.326286811 -0.02533560 1.00000000 -0.05986352  
## logavgsen -0.12188311 0.134181452 -0.17225398 -0.05986352 1.00000000  
## logpolice -0.16102212 0.542713183 -0.05647614 -0.29508551 0.29487049  
## logmix 0.56189540 -0.006115974 0.09256607 -0.38424287 -0.13105421  
## logpolice logmix  
## logprobsen -0.16102212 0.561895402  
## logcrime 0.54271318 -0.006115974  
## probarr -0.05647614 0.092566073  
## logprobconv -0.29508551 -0.384242869  
## logavgsen 0.29487049 -0.131054206  
## logpolice 1.00000000 0.062667253  
## logmix 0.06266725 1.000000000

We are not seeing any obvious signs of multicollinearity. We will now compute VIF.

vif(model1)

## logcrime probarr logprobconv logavgsen logpolice logmix   
## 1.534288 1.049640 1.368897 1.151492 1.574976 1.241359

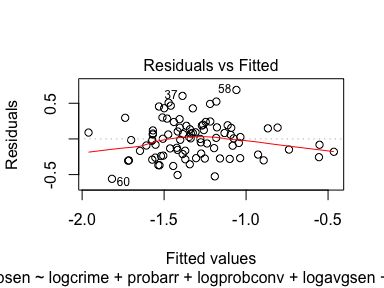
The VIF is < 4 and R is not flagging perfect multicollinearity.

Is the assumption valid?

#### CLM 4 - Zero conditional mean

We’ll now plot our model in order to assess if the model has zero conditional mean.

plot(model1, which=1)



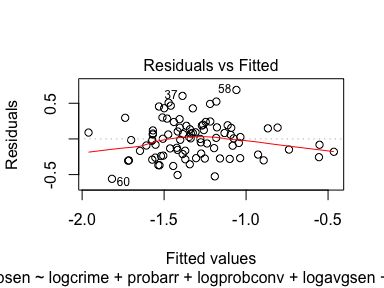
The red line is staying relatively close to the X-axis for the most part although it is influenced by the outliers on the ends.

Is the assumption valid?

#### CLM 5 - Homoscedasticity

We will use the same plot to assess the model’s homoscedasticity.

plot(model1, which=1)



The plot is relatively scattered about the fitted values with some extreme outliers. It is a little bit difficult to determine if we have achieved homoscedasticity from this plot alone. We will run a couple of additional tests to determine the homoscedasticity of the model.

bptest(model1)

##   
## studentized Breusch-Pagan test  
##   
## data: model1  
## BP = 4.0657, df = 6, p-value = 0.6678

ncvTest(model1)

## Non-constant Variance Score Test   
## Variance formula: ~ fitted.values   
## Chisquare = 0.6267145 Df = 1 p = 0.428563

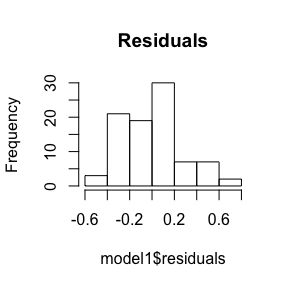
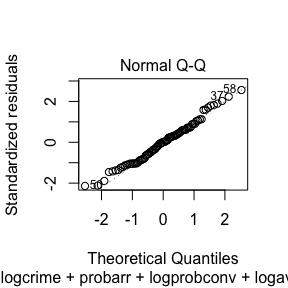
Neither test is showing a small enough P-value suggesting we fail to reject the null hypothesis of homoscedasticity. Therefore we most likely have homoscedasticity however looking at the plot, it is a little bit questionable.

Is the assumption valid?

#### CLM 6 - Normality of residuals

We will now lok at the QQ-plot to assess the normality of residuals.

plot(model1, which=2)  
hist(model1$residuals, main="Residuals")

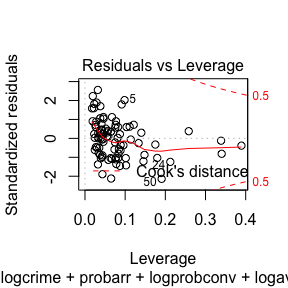
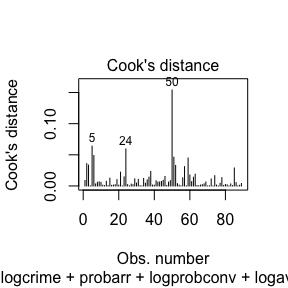


The values are staying close to the slope for the most part however are deviating on both ends. However the distribution of the residuals is relatively normal and our sample size n is 90 and hence CLM 6 is achieved.

Is the assumption valid?

### Cook’s distance

plot(model1, which = 4)  
plot(model1, which = 5)

 There is a influencial value at 50 however it is still well within the bounds of Cook’s distance.

### AIC

(model1$AIC <- AIC(model1))

## [1] 30.08571

The AIC for this model is 30.0857125.

## Propposed Model #2 - Optimal specification

In addition to the set of explanatory variables introduced in Proposed Model #1, I have decided to include the following variables in this model:

: I am interested to see if demographics information such as race, gender and age would influence the probability of prison sentence and therefore including , in this model.

: I suspect population density would have a negative influence on by introducing more complexity in crimes.

: I anticipate would have a negative coefficient as people with more money would be able to afford better lawyers and hence would have lower chances of ending up with prison sentences.

model2 <- lm(logprobsen ~ logcrime + probarr + logprobconv + logavgsen + logpolice  
 + logdensity + logtax + logpctmin + logmix + logymale, data = D)

Holding the other variables such as other probabilities such as probarr and probconv, crime-related variables such as log(crime) and log(police), we can see pctmin and ymale as well as pctmin:ymale are actually statistically significant.

It is interesting that including probarr and probconv reduces the stastical significance of the covariate mix drastically.

model2\_1 <- lm(mix ~ probarr + probconv, data = D)  
summary(model2\_1)

##   
## Call:  
## lm(formula = mix ~ probarr + probconv, data = D)  
##   
## Residuals:  
## Min 1Q Median 3Q Max   
## -0.10755 -0.05036 -0.02638 0.03199 0.30525   
##   
## Coefficients:  
## Estimate Std. Error t value Pr(>|t|)   
## (Intercept) 0.12092 0.04566 2.648 0.00963 \*\*  
## probarr 0.11704 0.10396 1.126 0.26336   
## probconv -0.07310 0.02453 -2.979 0.00375 \*\*  
## ---  
## Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1  
##   
## Residual standard error: 0.07854 on 86 degrees of freedom  
## Multiple R-squared: 0.107, Adjusted R-squared: 0.08628   
## F-statistic: 5.155 on 2 and 86 DF, p-value: 0.007684

Regressing on probarr and probconv, R-squared shows 0.107, indicating that probarr and probconv are accountable for 10.7% of the variance in the variable mix.

### CLM

No change in CLM1-2.

#### CLM 3 - Multicollinearity

We’ll compute VIF

vif(model2)

## logcrime probarr logprobconv logavgsen logpolice logdensity   
## 3.724762 1.070785 1.796106 1.159418 2.126302 2.456174   
## logtax logpctmin logmix logymale   
## 1.540660 1.809699 1.712168 1.318453

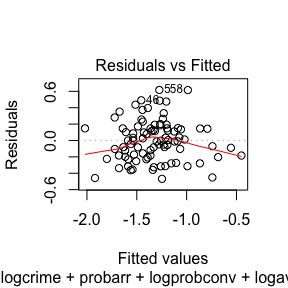
VIF is flagging pctmin, ymale and pctmin:ymale which is expected as pctmin:ymale is an interaction term made up of pctmin and ymale.

Is the assumption valid?

#### CLM 4 - Zero conditional mean

We’ll now plot our model in order to assess if the model has zero conditional mean.

plot(model2, which=1)



The fitted line is staying relatively close to the X-axis for the most part however is influenced by the outliers on the both sides.

Is the assumption valid?

#### CLM 5 - Homoscedasticity

The plot is relatively distributed evenly about the fitted values.

bptest(model2)

##   
## studentized Breusch-Pagan test  
##   
## data: model2  
## BP = 12.476, df = 10, p-value = 0.2544

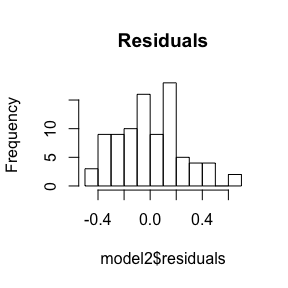
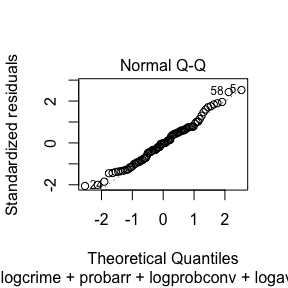
Checking the BP test result, the P-value is not small enough to reject the null hypothesis of homoscedasticity.

Is the assumption valid?

#### CLM 6 - Normality of residuals

We will now lok at the QQ-plot to assess the normality of residuals.

plot(model2, which=2)  
hist(model2$residuals, main = "Residuals")

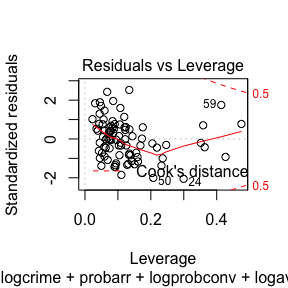
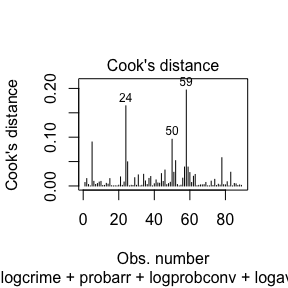


The both plots are showing we have normality of residuals.

Is the assumption valid?

### Cook’s distance

plot(model2, which = 4)  
plot(model2, which = 5)

 There is a influencial value at 59 however it is still well within the bounds of Cook’s distance.

### AIC

(model2$AIC <- AIC(model2))

## [1] 26.70092

The AIC for this model is 26.7009226 which is lower compared to model 1 indicating this is an improved model.

## Proposed Model 3 - Comprehensive specification

This model includes all variables present in the dataset to show the robustness of my modeling process and the underlying assumptions to model specification.

model3 <- lm(logprobsen ~ logcrime + probarr + logprobconv + logavgsen + logpolice  
 + logdensity + logtax + logpctmin + logmix + logymale + west + central + urban  
 + logwagecon + wagetuc + wagetrd + wagefir + logwageser + wagemfg + wagefed  
 + wagesta + wageloc, data = D)

### CLM

No change in CLM1-2.

#### CLM 3 - Multicollinearity

We’ll compute VIF

vif(model3)

## logcrime probarr logprobconv logavgsen logpolice logdensity   
## 4.863753 1.174475 2.054750 1.557634 2.889421 6.081971   
## logtax logpctmin logmix logymale west central   
## 2.455541 4.448054 2.071487 1.698017 2.267788 4.878311   
## urban logwagecon wagetuc wagetrd wagefir logwageser   
## 2.928001 2.186521 1.745930 3.228493 2.931556 1.663908   
## wagemfg wagefed wagesta wageloc   
## 2.008753 3.522783 1.699442 2.354271

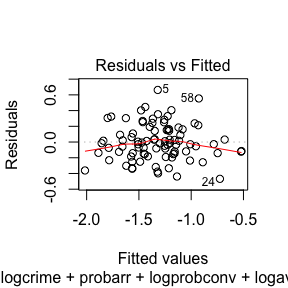
All the values are < 10.

Is the assumption valid?

#### CLM 4 - Zero conditional mean

We’ll now plot our model in order to assess if the model has zero conditional mean.

plot(model3, which=1)



The fitted line is staying relatively close to the X-axis for the most part.

Is the assumption valid?

#### CLM 5 - Homoscedasticity

The plot is relatively distributed evenly about the fitted values.

bptest(model3)

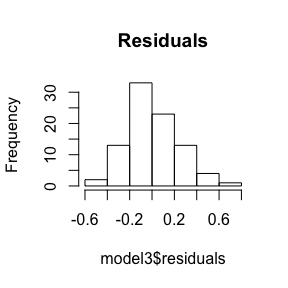
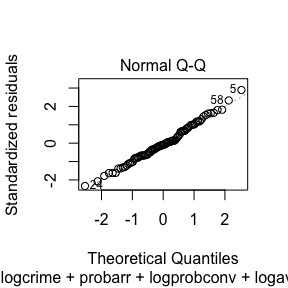
##   
## studentized Breusch-Pagan test  
##   
## data: model3  
## BP = 25.311, df = 22, p-value = 0.2825

Checking the BP test result, the P-value is not small enough to reject the null hypothesis of homoscedasticity.

Is the assumption valid?

#### CLM 6 - Normality of residuals

plot(model3, which=2)  
hist(model3$residuals, main = "Residuals")

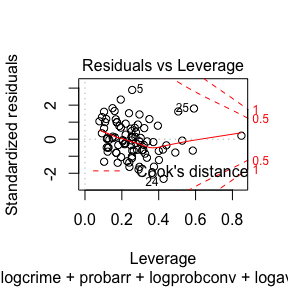
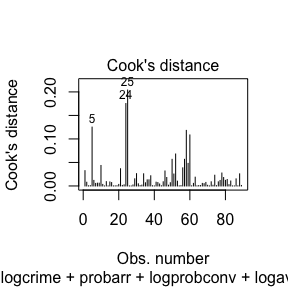


The both plots are showing we have normality of residuals.

Is the assumption valid?

### Cook’s distance

plot(model3, which = 4)  
plot(model3, which = 5)



There are some spikes however they are still well within the bounds of Cook’s distance.

### AIC

(model3$AIC <- AIC(model3))

## [1] 37.99468

The AIC for this model is 37.9946771 which is the highest of the 3 models.

# Model Adjustments

We will now be adjusting the models in order as there were some CLM assumptions that were violated or not entirely met.

In order to address the possible violations of CLM 5 Homoscedasticity assumption, we will be converting our coefficients using heteroscedasticity robust standard errors.

## Model 1

model1$coefficients

## (Intercept) logcrime probarr logprobconv logavgsen logpolice   
## -1.99145420 -0.30408208 -0.32580565 -0.20125641 -0.03140236 -0.05106572   
## logmix   
## 0.30975736

(model1$adjusted\_coefficients <- sqrt(diag(vcovHC(model1))))

## (Intercept) logcrime probarr logprobconv logavgsen logpolice   
## 0.67896991 0.08267105 0.34124047 0.06952464 0.10253441 0.09683524   
## logmix   
## 0.06779809

## Model 2

model2$coefficients

## (Intercept) logcrime probarr logprobconv logavgsen logpolice   
## -2.81273396 -0.41212132 -0.35930841 -0.30460079 -0.02495748 0.06231808   
## logdensity logtax logpctmin logmix logymale   
## 0.02258665 -0.07331550 0.10476483 0.21423888 -0.33614134

(model2$adjusted\_coefficients <- sqrt(diag(vcovHC(model2))))

## (Intercept) logcrime probarr logprobconv logavgsen logpolice   
## 1.43429852 0.12265221 0.38445249 0.08437148 0.10627715 0.13583936   
## logdensity logtax logpctmin logmix logymale   
## 0.05989955 0.14400446 0.04552969 0.07354445 0.24169374

## Model 3

model3$coefficients

## (Intercept) logcrime probarr logprobconv logavgsen   
## -4.491754e+00 -4.978365e-01 -3.138610e-01 -3.269633e-01 -2.102672e-02   
## logpolice logdensity logtax logpctmin logmix   
## 1.349940e-02 8.814882e-02 7.119438e-02 1.768710e-01 2.166907e-01   
## logymale west central urban logwagecon   
## -1.927436e-01 -4.400895e-02 1.566188e-01 -2.046996e-01 2.107010e-01   
## wagetuc wagetrd wagefir logwageser wagemfg   
## -5.890375e-05 -8.433569e-07 -5.703936e-04 -9.128650e-02 -3.135048e-05   
## wagefed wagesta wageloc   
## 1.209783e-03 -9.266370e-04 -7.510987e-05

(model3$adjusted\_coefficients <- sqrt(diag(vcovHC(model3))))

## (Intercept) logcrime probarr logprobconv logavgsen   
## 2.3887671032 0.1904656805 0.4114521222 0.1131892621 0.1429948150   
## logpolice logdensity logtax logpctmin logmix   
## 0.1650621562 0.1043631263 0.2305953809 0.0901974581 0.0924087356   
## logymale west central urban logwagecon   
## 0.2796174017 0.1064263357 0.1649583437 0.1931151236 0.2744396082   
## wagetuc wagetrd wagefir logwageser wagemfg   
## 0.0006598016 0.0017896417 0.0011538875 0.1111826014 0.0004584581   
## wagefed wagesta wageloc   
## 0.0010681425 0.0010781599 0.0025036122

# Model Analysis

stargazer(model1, model2, model3, omit.stat = "f", header=FALSE,   
 title = "Models for predicting probability of prison sentences",  
 se =   
 list(model1$adjusted\_coefficients, model2$adjusted\_coefficients,  
 model3$adjusted\_coefficients),  
 star.cutoffs = c(0.05, 0.01, 0.001), no.space = TRUE)

## Model 1

, and have very small P-values suggesting strong stastical significance. Interestingly, all the original co-efficients except for logmix are negative contrary to my initial hypothesis. 1% increase in and results in -30.4% and -20.1% impact on the dependent variable which are both practically significant. Once adjusted using robust standard errors, all the co-efficients became positive.

Adjusted is 0.475 which is the lowest of the 3 models, explanining 47.5% of the variation in .

## Model 2

In addition to , , and has a P-value < 0.05 in this model. It has a positive coefficient indicating in 1% increase in will translate into 10.5% increase in which is a practically significant result.

Adjusted is 0.515 which is the highest of the 3 models, explanining 51.5% of the variation in . The model also has the lowest AIC of the 3 models at 26.701 indicating this is the best model of the 3 according to Akaike’s Information Criterion.

## Model 3

The same set of variables as Model 2, , , and are showing statistical significance although not as strongly. One thing to note is that the co-efficient values for the statistically significant covariates in this model appear to be larger in the magnitude and hence practical significance than those of Model 2. For example, is showing -0.498 which is greater vs -0.412 for Model 2.

Adjusted is 0.503 which is the second highest of the 3 models, explanining 50.3% of the variation in . The model also has the highest AIC of the 3 models at 37.995 indicating this is the worst model of the 3 according to Akaike’s Information Criterion.

# Causality

In my analysis, I have tried to show

# Conclusion