

FA2_EDA

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```
data <- read.csv("D:/FEU/3RD YR 2ND SEM/EDA/cytof_one_experiment.csv")  
head(data)
```

Use pivot_longer to reshape the dataset into one that has two columns, the first giving the protein identity and the second giving the amount of the protein in one of the cells. The dataset you get should have 1750000 rows (50000 cells in the original dataset times 35 proteins)

##	NKp30	KIR3DL1	NKp44	KIR2DL1	GranzymeB	CXCR6	CD161
## 1	0.1875955	3.6156932	-0.5605694	-0.2936654	2.477893	-0.14470053	-0.3152872
## 2	1.0348518	1.7001820	-0.2889611	-0.4798280	3.261016	-0.03392447	-0.4112129
## 3	2.9996398	6.1411419	1.9032606	0.4823102	4.277562	1.94654156	-0.5022347
## 4	4.2998594	-0.2211586	0.2425707	-0.4831267	3.351808	0.92622195	3.8772370
## 5	-0.4386448	-0.5035892	-0.1526320	0.7506128	3.194145	-0.05893640	1.0907379
## 6	2.0883050	-0.3992646	3.4550676	-0.5200856	4.345102	-0.36434277	-0.5705891
##	KIR2DS4	NKp46	NKG2D	NKG2C	X2B4	CD69	KIR3DL1.S1
## 1	1.94497046	4.0818316	2.6200784	-0.3573817	-0.2711557	3.849965	-0.2554637
## 2	3.80251714	3.7339299	-0.4832788	-0.4675984	-0.5594752	2.910197	-0.2909482
## 3	-0.32010171	4.5594631	-0.5069090	2.6193782	-0.4554785	3.113454	3.6613886
## 4	-0.16969487	4.4831486	1.9272290	-0.3110146	1.6350771	3.045998	0.2871241
## 5	-0.05033025	0.8379358	-0.4581674	0.9216947	1.2419054	2.644422	0.4218294
## 6	-0.45033591	4.0550848	3.4283565	0.6272837	-0.4157104	3.958158	0.7993406
##	CD2	KIR2DL5	DNAM.1	CD4	CD8	CD57	TRAIL
## 1	5.3529769	-0.5092906	0.8811347	-0.32347280	-0.2822405	3.3254704	-0.6084228
## 2	4.3132510	3.7774776	1.5406568	-0.13208167	0.9161920	2.4946442	-0.5034739
## 3	5.5969513	0.8128166	1.0005903	-0.59933641	1.8382744	3.9897914	-0.2749380
## 4	-0.5002885	0.3612212	1.2663267	-0.12568567	0.7667204	1.9950916	-0.5130930
## 5	-0.5479527	1.0638327	0.8722272	-0.07107408	-0.1059012	3.4291302	-0.1433044
## 6	5.1028564	3.0918867	0.8717267	-0.47986180	-0.2577198	-0.5784575	-0.5731323
##	KIR3DL2	MIP1b	CD107a	GM.CSF	CD16	TNFA	
## 1	-0.30668543	1.2497120	-0.1295305	-0.43074102	3.9951417	0.90143498	
## 2	-0.54320954	2.8693060	-0.1887180	-0.16283845	4.4082309	1.93590153	
## 3	2.06488239	4.0955112	-0.1998480	3.18853825	6.0023244	-0.02336999	
## 4	2.11247859	3.3726018	-0.5720339	0.91310694	5.8238698	-0.60793749	
## 5	-0.02505141	-0.3099826	-0.1068511	-0.60370379	4.0122501	-0.61989100	
## 6	-0.28337673	-0.4108283	-0.1797545	-0.06372458	-0.5832926	0.14311030	
##	ILT2	Perforin	KIR2DL2.L3.S2	KIR2DL3	NKG2A	NTB.A	CD56
## 1	-0.386027758	6.431983	1.22710292	2.660657999	-0.5220613	4.348923	2.897523
## 2	2.983874845	6.814827	-0.04141081	3.841304627	4.6771149	3.474335	3.782870

```
## 3 -0.521099944 5.099562 -0.16705075 -0.009694396 -0.4730573 5.634341 5.701186
## 4 -0.043783559 5.841797 -0.51753289 -0.592990887 -0.4059049 4.598021 6.065672
## 5 1.182703288 4.888777 -0.36251589 -0.398123704 -0.5440881 3.606101 1.966169
## 6 -0.003258955 3.952542 -0.20194392 -0.202592720 3.8882776 2.346275 6.473243
##      INFg
## 1 -0.3841108
## 2 2.7186296
## 3 2.5321763
## 4 2.4564582
## 5 3.1470092
## 6 2.8282987
```

```
library(tidyverse)
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.5
## v forcats    1.0.0      v stringr    1.5.1
## v ggplot2    3.5.2      v tibble     3.2.1
## v lubridate  1.9.4      v tidyr      1.3.1
## v purrr      1.0.4
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
set.seed(123)
```

```
data <- data.frame(
  cell_id = rep(1:50000, each = 1),
  protein_1 = runif(50000, min = 0, max = 100),
  protein_2 = runif(50000, min = 0, max = 100),
  protein_3 = runif(50000, min = 0, max = 100),
  protein_4 = runif(50000, min = 0, max = 100),
  protein_5 = runif(50000, min = 0, max = 100),
  protein_6 = runif(50000, min = 0, max = 100),
  protein_7 = runif(50000, min = 0, max = 100),
  protein_8 = runif(50000, min = 0, max = 100),
  protein_9 = runif(50000, min = 0, max = 100),
  protein_10 = runif(50000, min = 0, max = 100),
  protein_11 = runif(50000, min = 0, max = 100),
  protein_12 = runif(50000, min = 0, max = 100),
  protein_13 = runif(50000, min = 0, max = 100),
  protein_14 = runif(50000, min = 0, max = 100),
  protein_15 = runif(50000, min = 0, max = 100),
  protein_16 = runif(50000, min = 0, max = 100),
  protein_17 = runif(50000, min = 0, max = 100),
  protein_18 = runif(50000, min = 0, max = 100),
  protein_19 = runif(50000, min = 0, max = 100),
  protein_20 = runif(50000, min = 0, max = 100),
  protein_21 = runif(50000, min = 0, max = 100),
  protein_22 = runif(50000, min = 0, max = 100),
  protein_23 = runif(50000, min = 0, max = 100),
  protein_24 = runif(50000, min = 0, max = 100),
```

```

protein_25 = runif(50000, min = 0, max = 100),
protein_26 = runif(50000, min = 0, max = 100),
protein_27 = runif(50000, min = 0, max = 100),
protein_28 = runif(50000, min = 0, max = 100),
protein_29 = runif(50000, min = 0, max = 100),
protein_30 = runif(50000, min = 0, max = 100),
protein_31 = runif(50000, min = 0, max = 100),
protein_32 = runif(50000, min = 0, max = 100),
protein_33 = runif(50000, min = 0, max = 100),
protein_34 = runif(50000, min = 0, max = 100),
protein_35 = runif(50000, min = 0, max = 100)
)

reshaped_data <- data %>%
  pivot_longer(cols = starts_with("protein_"),
    names_to = "protein",
    values_to = "protein_amount")

head(reshaped_data)

```

```

## # A tibble: 6 x 3
##   cell_id protein    protein_amount
##   <int> <chr>          <dbl>
## 1      1 protein_1      28.8
## 2      1 protein_2      21.3
## 3      1 protein_3      60.2
## 4      1 protein_4      53.6
## 5      1 protein_5      60.4
## 6      1 protein_6      16.7

```

```

summary_stats <- reshaped_data %>%
  group_by(protein) %>%
  summarise(
    median_level = median(protein_amount, na.rm = TRUE),
    mad_level = mad(protein_amount, na.rm = TRUE)
  )

head(summary_stats)

```

Use `group_by` and `summarise` to find the median protein level and the median absolute deviation of the protein level for each marker. (Use the R functions `median` and `mad`).

```

## # A tibble: 6 x 3
##   protein    median_level mad_level
##   <chr>          <dbl>     <dbl>
## 1 protein_1      49.4      37.0
## 2 protein_10     49.8      37.0
## 3 protein_11     49.6      36.8
## 4 protein_12     50.0      37.2
## 5 protein_13     49.9      37.2
## 6 protein_14     49.8      37.1

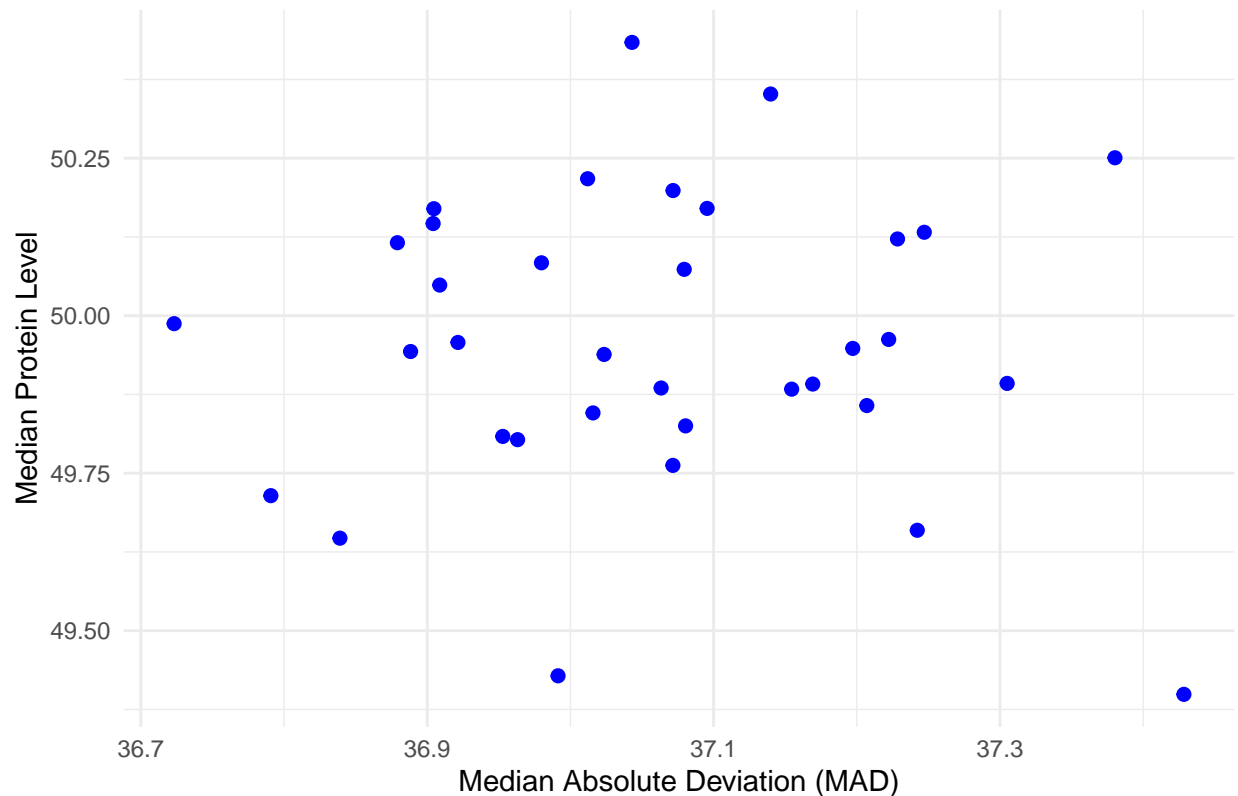
```

```
library(ggplot2)

ggplot(summary_stats, aes(x = mad_level, y = median_level)) +
  geom_point(color = "blue", size = 2) +
  labs(
    x = "Median Absolute Deviation (MAD)",
    y = "Median Protein Level",
    title = "Spread-Location Plot: MAD vs Median Protein Level"
  ) +
  theme_minimal()
```

Make a plot with mad on the x-axis and median on the y-axis. This is known as a spreadlocation (s-l) plot. What does it tell you about the relationship between the median and the mad?

Spread-Location Plot: MAD vs Median Protein Level



```
library(dcldata)
library(tidyr)
library(dplyr)
reshaped_data <- example_gymnastics_2 %>%
  pivot_longer(cols = starts_with("vault") | starts_with("floor"),
    names_to = "event_year",
    values_to = "score") %>%
```

```
separate(col = event_year,
         into = c("event", "year"),
         sep = "_")

head(reshaped_data)
```

The MAD values range between roughly 36.7 and 37.3, while the median protein level is around 49.5 to 50.3 suggesting that the variation of median protein levels are higher when compared to the variation in MAD.

```
## # A tibble: 6 x 4
##   country      event year  score
##   <chr>      <chr> <chr> <dbl>
## 1 United States vault 2012   48.1
## 2 United States vault 2016   46.9
## 3 United States floor 2012   45.4
## 4 United States floor 2016   46.0
## 5 Russia      vault 2012   46.4
## 6 Russia      vault 2016   45.7
```

```
str(reshaped_data)
```

```
## tibble [12 x 4] (S3: tbl_df/tbl/data.frame)
## $ country: chr [1:12] "United States" "United States" "United States" "United States" ...
## $ event : chr [1:12] "vault" "vault" "floor" "floor" ...
## $ year : chr [1:12] "2012" "2016" "2012" "2016" ...
## $ score : num [1:12] 48.1 46.9 45.4 46 46.4 ...
```