

# Guide to estimating the distribution of selection coefficients using the TdG12 site-wise mutation-selection model

December 22, 2011

## 1 Introduction

This document describes how to use the TdG12 site-wise mutation-selection model ('swMutSel0') to estimate the distribution of selection coefficients (or 'fitness effects') from an alignment of protein coding sequences.

### 1.1 Requirements

The steps in this tutorial have been tested on Linux and Mac OS X 10.6+. Our program does not estimate tree topology or perform branch length optimisation, therefore we recommend the use of two popular tools for this purpose: RAxML and PAML<sup>1</sup>. Both have Windows versions available for download, but they have not been tested by us. We assume that you already have them installed and are able to run them on your computer. Our software is written in Java and should run on all platforms that have the Java Runtime Environment (JRE) (6 or later) installed. The JRE is available for Windows, Linux and Mac OS X 10.6+.

### 1.2 Installation

You can download the executable files for the program from <http://mathbio.nimr.mrc.ac.uk/>. Download the `tdg12.zip` file and extract the contents. The download contains the files required for this tutorial and `tdg12.jar`, which is the Java binary file for the program.

### 1.3 Citation

Tamuri AU, dos Reis, M and Goldstein RA (2012) Using Site-wise Mutation-Selection Models to Estimate the Distribution of Selection Coefficients from Phylogenetic Data. **XX:YY** *GENETICS*.

---

<sup>1</sup>However, you can use other programs if you prefer.

## 2 Tutorial

### 2.1 Preparing the alignment and tree

*[[Basic background? Types of sequences, not too many gaps, no recombination etc. You need good alignments and enough divergence??]]*

For the purposes of this tutorial, we will be estimating the distribution of selection coefficients of a set of mammalian mitochondrial ATP8 protein coding genes. The download provides an alignment, atp8.phyl (which was built using PRANK) and a tree, atp8.tree (estimated by RAxML with branch lengths optimised using PAML's codeml). Full details are available from the program's websites. The options used for running RAxML were:

```
raxmlHPC -f a -x 12345 -p 12345 -N 100 -m GTRGAMMA -s atp8.phy -n  
atp8
```

and the PAML codeml control file (codeml.ct1) is available in the download. We optimised the branch lengths using the FMutSel0 model.

### 2.2 Getting estimates of global parameters from codeml

There are a number of global, site-invariant, parameters that are required for the TdG12 model. They are:

1.  $\tau$  (tau) - rate of multiple substitutions.
2.  $\kappa$  (kappa) - transition/transversion bias.
3.  $\pi$  (pi) - base nucleotide composition.
4.  $\mu$  (mu) - branch scaling.

Each of these parameters can be estimated under the TdG12 model but this requires significant computational resources and the use of a distributed version of the software (not covered here). However, you can get obtain good estimates for  $\kappa$ ,  $\pi$  and  $\mu$  from the faster PAML codeml analysis. The results file (named 'mlc') contains estimates of  $\kappa$  and  $\pi$ . An estimate of  $\mu$  can be calculated from  $\mu = 3 \times T_{dS}/T$ , where  $T_{dS}$  is the tree length in  $dS$  (synonymous changes) units and  $T$  is the total tree length, also found in the codeml 'mlc' results file.

Therefore, the estimates for the global parameters for the ATP8 gene alignment are:  $\kappa = 4.93022$ ;  $\pi = \{0.25299, 0.22268, 0.43860, 0.08572\}$ ;  $\mu = 3 \times T_{dS}/T = 3 \times 70.7207/113.24814 = 1.8734$ . We choose a small number for  $\tau$ ,  $1.0 \times 10^{-2}$ .

### 2.3 Analysing the data using the site-wise mutation-selection model

You can now run the TdG12 program to estimate the site-wise fitness of each amino acid in our alignment. The available options for running the program are:

- t** The tree file in NEWICK format (required).
- s** The protein coding alignment file in PHYLIP sequential format (required).
- gc** The genetic code, 'standard' or 'vertebrate\_mit' (required).
- tau** Rate of multiple substitutions (required).
- kappa** Transition/transversion bias (required).
- pi** Comma-separated (with no spaces) base nucleotide frequencies (T,C,A,G) (required).
- mu** Branch scaling factor (required).
- useApprox** Use the approximation for unobserved residues which allows for faster computation (optional).
- site** Location of the site to analyse (optional, default = all sites in the alignment).
- optimruns** The number of restarts for the optimisation routine (optional, default = 1).
- threads** The number of threads to use for processing in a multicore environment (optional, default = 1).

You start the analysis of the set of ATP8 genes by running:

```
java -cp tdg12.jar tdg.Analyse -s atp8.phy -t atp8.tree -gc vertebrate_mit
-tau 1e-2 -kappa 4.93 -pi 0.2530,0.2227,0.4386,0.0857 -mu 1.8734 >
tdg.out
```

You must add '> tdg.out' to redirect the analysis output to a file named tdg.out.

### Using multicore/multiple CPUs

If you are running the program on a computer with multicore or multiple CPUs, you can specify the **-threads** option. Usually, this would be the number of available cores - 1. For example, to utilise 3 cores you would run:

```
java -cp tdg12.jar tdg.Analyse -s atp8.phy -t atp8.tree -gc vertebrate_mit
-tau 1e-2 -kappa 7.8 -pi 0.25,0.25,0.25,0.25 -mu 2.3 -threads 3 >
tdg.out
```

## 2.4 Parsing the results and calculating the distribution

Once the program completes, the results saved in tdg.out need to be processed to calculate the distribution of selection coefficients. The output looks something like (truncated):

```
Site 1 - Residues: [1/20] { 1:(12, M), 2:(0, A), 3:(1, R), ... }
Site 1 - Optimisation run (267 evaluations). lnL = -4.939901E-5, Params = {0.0, ...}
Site 1 - Homogeneous model lnL: -4.939901011180276E-5
Site 1 - Fitness: { -13.06494, -18.20550, -16.32137, ... }
Site 1 - Pi: { 4.79931E-6, 5.79345E-8, 2.70189E-7, ... }
...
```

The output is quite verbose. Run the following command (specifying the name of the result file with '-o tdg.out'):

```
java -cp tdg12.jar tdg.results.All -o tdg.out -gc vertebrate_mit -tau
1e-6 -kappa 7.8 -pi 0.25,0.25,0.25,0.25 -mu 2.3
```

The global parameters should be the same as were specified when you run the analysis. This command reads the results file and the global parameters from the command-line, and writes the files:

**F.txt** Fitness values for each amino acid at each site

**Q0.txt** Neutral mutation matrix for entire alignment

**S.txt** Selection coefficients matrix for each site

**QS.txt** Mutation with selection matrix for each site

**PiS.txt** Codon frequencies at each site

**PiAA.txt** Amino acid frequencies at each site

**distribution.mutations.csv** the distribution of selection coefficients for mutations

**distribution.substitutions.csv** the distribution for substitutions.

The last two are comma-separated value files that can be opened in a program like Excel to plot the distribution. The three columns in the distribution files are (i) the histogram bins for  $S$  (ii) all mutations or substitutions and (iii) non-synonymous mutations or substitutions.

### 3 Colophon

TdG12 uses the following libraries:

1. Colt Project (<http://acs.lbl.gov/software/colt/>). For linear algebra.
2. PAL: Phylogenetic Analysis Library (<http://www.cebl.auckland.ac.nz/pal-project/>). reading/traversing/writing NEWICK trees and PHYLIP alignments.
3. Apache Commons (<http://commons.apache.org/>). Specifically: collections, configuration, lang, logging and math.
4. Guava: Google Core Libraries (<http://code.google.com/p/guava-libraries/>).
5. JCommander (<http://jcommander.org/>). Parsing and managing command-line options.
6. Simple Java HTTP server (<http://www.simpleframework.org/>). Used by `tdg.distributed.Slave`.
7. Asynchronous Http Client library for Java (<https://github.com/sonatype/async-http-client>). Used by `tdg.distributed.Master` to make asynchronous HTTP requests to the slaves.