#### TANIA KURBESSOIAN

Taniakurbessoian@gmail.com | 818.497.1580 | https://www.linkedin.com/pub/Tania-kurbessoian/71/648/87a/ A motivated Ph.D. student in Microbiology, seeking for a career in Microbiology, Mycology and Genetics

#### **EDUCATION**

University of California, Riverside

PhD. In Microbiology

GPA:3.89

California State University
Master of Science in Microbiology

GPA:3.77

California State University

Bachelor of Science in Microbiology

• GPA:3.51

Riverside

2017-

**Northridge** 2014 – 2016

**Northridge** 2010 – 2013

### RELEVANT SKILLS

- Microbiology: Aseptic technique, Bright field and contrast microscopy, Staining bacteria, Enumeration and identification, Media, buffer and chemical preparation, Utilizing food microbiology, Using selective media, Plaque assay, Enzyme assay, MALDI-TOF MS as an identification tool down to the strain level
- Mycology: Aseptic technique, Bright field and contrast microscopy, staining fungi, media preparation, culturing
- **Biochemistry and Cell & Molecular Biology:** DNA extraction, Assay Preparation, Protein Extraction, Plasmid DNA preparation, Restriction enzyme digests, PCR, Agarose gel electrophoresis, using centrifugation.
- Chemistry: Preparing solutions, Titrations, Extractions, Filtration, Solubility tests

### INTERNSHIP EXPERIENCE

## Jet Propulsion Laboratory, JPL - NASA

Pasadena, CA

Summer Intern Program (SIP) Intern

July - Sept. 2016

- Collaborated with P.I. Wayne Schubert and Planetary Protection Officers to apply biological aspects to astrobiological situations.
- Created, followed up and finished Embedded Bioburden experiments on extreme heat and desiccation resistant strains of Bacillus sp. (ATCC 29669), utilized a cryogen grinder and mastered serial dilutions and plating techniques.
- Calculated varying D-values for ATCC 29669 in varying temperature and time lengths.
- Created and maintained MALDI-TOF MS protein profiles of the Bacillus sp. (ATCC 29669).
- Prepared embedded spore masses using a variety of epoxies.

### WORK EXPERIENCE

# University of California, Riverside

Riverside, CA

Graduate Researcher

July 2017-

- Working under P.I. Dr. Jason E. Stajich on observing evolutionary trends in Fungi, while also focusing on melanized yeast fungi *Hortaea werneckii*, *Knufia petricola* and others.
- Applied learned microbiology techniques to successfully isolate mycological organisms from arid regions and biological crusts.

# California State University

Northridge, CA

Graduate Research Assistant

Jan. 2014 – Dec. 2016

- Employed under P.I. Larry Baresi using numerous molecular biology techniques such as electrophoresis, plasmid preparation, and
  restriction enzyme digests to extract, isolate and digest DNA from and PCR techniques to extract 16S sequences from 57 strains of
  Sporosarcina ureae.
- Process alignments utilizing Bioinformatics tools and create trees which accurately depict the non-clonal relationship between 57 strains.
- Observed and gained extensive knowledge on varying types of spores generated from the *Sporosarcina* strains.
- Attempted different techniques to isolate spores, found it to be most successful through sonication methods.
- Appropriated the BIOLOG tool in order to observe physiologies which then were used to create another dendrogram expressing the non-clonal relationship between the 57 strains.
- Efficiently utilized MALDI-TOF MS to create protein profiles for all 57 strains of *Sporosarcina* and created dendograms based on MSP's and 97% similarities of protein profiles to group them into OTUs, comparing OTU's with each other.

## California State University

Northridge, CA

Jan 2015 - Dec 2015 Teaching Associate

- Offered constructive feedback based on student performance on laboratory classes such as: Principles of Microbiology.
- Facilitated teachings through preparing lectures, graded exams, and provided final grades to the students.
- Improved aseptic techniques for students by demonstrating how to accomplish proper aseptic technique, as well as a variety of microbial testing maneuvers.
- Promoted a dynamic learning environment, while simultaneously enhancing communication skills through student interaction.

### California State University

Northridge, CA

Graduate Assistant

Jan 2014 – Dec 2016

- Assisted undergraduate students in laboratory classes including Principles of Microbiology, Medical Microbiology, Microbial Physiology, Biology of the Fungi and Food Microbiology.
- Fostered CSU's success through preparing media and cultures that were utilized in the microbiology teaching classrooms.
- Taught students how to maneuver and accomplish proper aseptic technique.

#### **California State University**

Northridge, CA

Undergraduate Research Student

Jan - Dec 2013

- Prepared different types of media specific to certain types of Escherichia coli as well as to an archaea Methanobrevibacter smithii strain G.
- Isolated and transferred Methanobrevibacter smithii strain G through anaerobic techniques.
- Worked within an anaerobic hood infecting Methanobrevibacter smithii strain G with Phage G.
- Employed numerous molecular biology techniques such as electrophoresis, plasmid preparation, and restriction enzyme digests to extract, isolate and digest DNA from Escherichia coli, Methanobrevibacter strain G as well as its isolated phage, Phage G.

# PROFESSIONAL ORGANIZATIONS

Graduate Student Association- Microbiology Chapter (Micro-GSA)	Riverside, CA
President, Vice President, Outreach	June 2017-
Association for Women in Science- Riverside Chapter(AWIS)	Riverside, CA
President, Publicity Chair	June 2018-
Microbiology Students Association at California State University (MSA)	Northridge, CA
Secretary, Treasurer and President	Jan 2013-Aug 2016
Women in Science at California State University (WiS)	Northridge, CA
Member	Jan 2015-Dec 2016
Graduate Leadership Association at California State University (GLA)	Northridge, CA
Social Media Coordinator	Jan 2015-Dec 2016
Los Angeles Mycological Society (LAMS)	
Member	Nov 2014 - Present
American Society for Microbiology (ASM)	
Member	Aug 2013 – Present
Southern California Chapter of the American Society for Microbiology (SCASM)	
Member	Dec 2014 – Present
Mycological Society of America (MSA)	
Student Member	Jan 2016 Present

# AWARDS & ADDITIONAL INFORMATION

- Research and Graduate Department-Research Scholarship 2014
- Presented Research at American Society of Microbiology General Meeting 2015
- Presented Research at Research Symposium at California State University, Northridge 2014, 2015, 2016
- Presented Research at Southern California American Society of Microbiology General Meeting 2016
- Presented Research at JPL-NASA September 2016
- Eugene Robles Fellowship winner September 2017
- Oral Presentation at Black Yeast Workshop part of ISHAM in Amsterdam, Netherlands 2018
- Emory Simmons Fellowship winner April 2019
- Presented Research at MSA- Mycological Society of America
  - Language skills: Proficient in reading, writing and communicating in English, Armenian, Russian
  - Computer skills: Programming: Bash, Python