Machine learning methods for the analysis of bacterial genomes

Random Forest classification using Tidymodels in R

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- Collection of libraries for implementing predictive models
- Objective:
 - Standardize the many available procedures/libraries
 - Enable the creation of workflow
 - use the tidyverse approach (sequential operations)

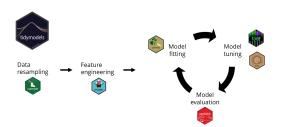


Figure 1: Tidymodels in a nutshell

	Overview of tidymodels Basics					
Package	Step	Functions				
rample	1. Split into testing and training sets	initial_split() training() testing()				
(included in the control of the cont	2. Create recipe + assign variable roles	recipe() update_role()				
panyip	3. Specify model, engine, and mode	parsnip function for specifying model (ex. decision_tree()) (https://www.tidymodels.org/find/parsnip/) set_engine() set_mode()				
	4. Create workflow, add recipe, add model	workflow() add_recipe() add_model()				
Dannig	5. Fit workflow	fit()				
Darwig	6. Get predictions	predict()				
PARTIE STATE OF THE PARTIE	7. Use predictions to get performance metrics	rmse() (continuous outcome) accuracy() (categorical outcome) metrics() (either type of outcome)				

Figure 2: Tidymodels Ecosystem

Why use Tidymodels? → Consistency, Safety, Communicability

```
# Linear Regression

model1 <- lm(outcome - ., dataset)

model2 <- linear_reg() %%
    set_engine (engine = "lm") %>%
    set_mode(mode = "regression")%>%
    fit(outcome - ., dataset)
```

```
# Regularization: Ridge-Lasso Regression
model1 <- glmnet(
    as.matrix(dataset[2:18]), # data matrix
    dataset$outcome) # vector

model2 <- linear_reg() %>%
    set_engine(engine = "glmnet") %>%
    set_mode(mode = "regression") %>%
fit(outcome - ., dataset)
```

Main steps:

- Sampling and Predictor Management: rsample, recipes
- Model Definition and Tuning: parsnip, tune, dials
- Model Evaluation: yardstick

An example of a machine learning analysis using the iris dataset

```
Load libraries
```

```
library("tidyverse")
library("tidymodels")
```

Load and check dataset

```
iris %>%
  head(n = 5) %>%
knitr::kable(caption = 'Dataset Iris')
```

Table 1: Dataset Iris

Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
5.1 4.9 4.7 4.6 5.0	3.5 3.0 3.2 3.1 3.6	1.4 1.4 1.3 1.5	0.2 0.2 0.2 0.2 0.2	setosa setosa setosa setosa setosa

Descriptive statistics

Check outcome frequency (the aim is to estimate the species variable)

```
iris %>%
  count(Species)%>%
  knitr::kable(caption = 'Species frequency')
```

Table 2: Species frequency

Species	r
setosa versicolor virginica	50 50 50

Check descriptive statistics grouped by species

Table 3: Descriptive statistics

Species	Sepal.Length_Media	Sepal.Length_Std
setosa	5.01	0.35
versicolor	5.94	0.52
virginica	6.59	0.64

Create train/test sets 1

```
set.seed(123)
iris_split <- rsample::initial_split(iris,prop = 0.7,</pre>
                                         strata = Species)
print(iris_split)
## <Training/Testing/Total>
## <105/45/150>
iris train <- training(iris split)</pre>
iris test <- testing(iris split)</pre>
```

Create train/test sets 2

Check outcome frequency in train set

```
iris_train %>%
  count(Species) %>%
  knitr::kable(caption='Species freq in train set')
```

Check outcome frequency in test set

```
iris_test %>%
  count(Species) %>%
  knitr::kable(caption='Species freq in test set')
```

Table 4: Species freq in train set

Species	r
setosa versicolor	35 35
virginica	35

Table 5: Species freq in test set

Species	r
setosa versicolor virginica	15 15 15

Rescale the predictors using the recipe() function and the step_*() command.

Examples of transformations: conversion to numeric/character/factor, normalization, rescaling, missing value imputation.

```
rec1 <-recipe(Species ~ ., data=iris_train) %>%
  step_normalize()
```

Model building 1

- Specify the type of model (e.g., linear regression, random forest...)
 - linear_reg()
 - rand_forest()
- Specify the so-called engine (i.e. package implementation of algorithm)
 - set_engine("some package's implementation")
- Declare the type of variable and therefore the type of analysis (e.g., classification vs regression)
 - When the model can be applied to both types of analysis
 - set mode("regression")
 - set_mode("classification")

Model building 2

Define the model

```
rf_mod <- rand_forest(trees = 150) %>%
set_engine('randomForest') %>%
set_mode(mode = "classification")
```

Create a so-called workflow, that is, a sequence of operations that combines the chosen model and any preprocessing steps.

```
iris_wflow <-
workflow() %>%, # create workflow
add_model(rf_mod) %>%, # model
add_recipe(rec1) # preprocessing
```

Model fit

```
iris_fit <-
  iris_wflow %>% # pipeline
fit(data = iris_train)
```

Results

```
iris_fit
```

```
## == Workflow [trained] ======
## Preprocessor: Recipe
## Model: rand forest()
##
## -- Preprocessor ------
## 1 Recipe Step
##
## * step_normalize()
##
##
## Call:
  randomForest(x = maybe_data_frame(x), y = y, ntree = ~150)
                Type of random forest: classification
##
                     Number of trees: 150
##
## No. of variables tried at each split: 2
##
         OOB estimate of error rate: 4.76%
##
## Confusion matrix:
##
            setosa versicolor virginica class.error
## setosa
                35
                                 0 0.00000000
                          0
## versicolor
               0
                          33
                                  2 0.05714286
## virginica
                                  32 0.08571429
```

Generate predictions on test set

```
iris_pred <- iris_fit %>%
  predict(new_data = iris_test)
```

Use the 'augment' function from the 'broom' package to create predictions and automatically generate a data frame.

```
augment(iris_fit,iris_test) -> estimates
head(estimates)%>%
knitr::kable(caption = 'Dataset with predictions', digits =1)
```

Table 6: Dataset with predictions

.pred_class	.pred_setosa .pr	ed_versicolor.pre	d_virginica S	epal.Length	Sepal.Width I	Petal.Length	Petal.Widt	h Specie
setosa	1	0	0	5.1	3.5	1.4	0.2	setosa
setosa	1	0	0	4.9	3.0	1.4	0.2	setosa
setosa	1	0	0	5.4	3.9	1.7	0.4	setosa
setosa	1	0	0	5.7	4.4	1.5	0.4	setosa
setosa	1	0	0	5.1	3.5	1.4	0.3	setosa
setosa	1	0	0	5.1	3.8	1.5	0.3	setosa

Model evaluation 1

Calculate the confusion matrix

```
## Truth

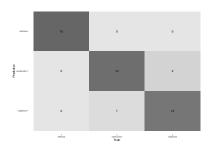
## Prediction setosa versicolor virginica

## setosa 15 0 0

## versicolor 0 14 2

## virginica 0 1 13
```

Visualize a heatmap



Model evaluation 2

Visualize the metrics

```
cm %>%
summary()%>%
knitr::kable(caption = 'Metrics for evaluation')
```

Table 7: Metrics for evaluation

.metric	.estimator	.estimate
accuracy	multiclass	0.9333333
kap	multiclass	0.9000000
sens	macro	0.9333333
spec	macro	0.9666667
ppv	macro	0.9345238
npv	macro	0.9670004
mcc	multiclass	0.9006674
j_index	macro	0.9000000
bal_accuracy	macro	0.9500000
detection_prevalence	macro	0.3333333
precision	macro	0.9345238
recall	macro	0.9333333
f_meas	macro	0.9332592

Apply a 5-fold cross validation (CV) to the train set

```
cv_folds <-
vfold_cv(iris_train,
    v = 5,
    strata = Species)</pre>
```

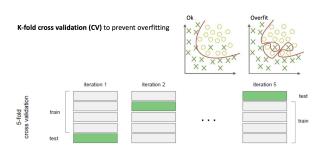


Figure 3: k-fold cross validation

Run the previously created model on all resamples using the fit_resamples function.

```
rf res <-
  iris fit %>%
  fit_resamples(
    resamples = cv_folds,
    metrics = metric_set(
      recall, precision, f meas,
      accuracy, kap,
      roc auc, sens, yardstick::spec),
    control = control resamples(save pred = TRUE)
```

Sum up the metrics of the CV with the collect_metrics function.

```
rf_res %>%
collect_metrics(summarize = TRUE)%>%
knitr::kable(caption = 'CV - Results')
```

Table 8: CV - Results

.metric	.estimator	mean	n	std_err	.config
accuracy f_meas kap precision recall roc_auc sens spec	multiclass macro multiclass macro macro hand_till macro macro	0.9523810 0.9519353 0.9285714 0.9578042 0.9523810 0.9897959 0.9523810 0.9761905	5 5 5 5 5 5	0.0212959 0.0215201 0.0319438 0.0192236 0.0212959 0.0069707 0.0212959 0.0106479	Preprocessor1_Model1 Preprocessor1_Model1 Preprocessor1_Model1 Preprocessor1_Model1 Preprocessor1_Model1 Preprocessor1_Model1 Preprocessor1_Model1

Complete fit

```
last_fit_rf %>%
  collect_metrics()%>%
  knitr::kable(caption = 'CV - Final Fit Results')
```

Table 9: CV - Final Fit Results

.metric	.estimator	.estimate	.config
recall	macro	0.9333333	Preprocessor1_Model1
precision	macro	0.9345238	Preprocessor1_Model1
f_meas	macro	0.9332592	Preprocessor1_Model1
accuracy	multiclass	0.9333333	Preprocessor1_Model1
кар	multiclass	0.9000000	Preprocessor1_Model1
sens	macro	0.9333333	Preprocessor1_Model1
spec	macro	0.9666667	Preprocessor1_Model1
roc_auc	hand_till	0.9962963	Preprocessor1_Model1

Generate predictions on test set using the final model

```
final_model <- last_fit_rf %>%
  extract_workflow()

rf_pred <- final_model %>%
  predict(new_data = iris_test)
```

Take-home message: Tidymodels pros & cons

Advantages:

- Possibility to carry out all steps of a predictive analysis using a single framework
- Output is in data.frame format rather than lists or other structures
- Easy to switch between different algorithms by changing a single parameter
- Built-in functions to collect and summarize results
- Use of the workflow concept (sequential analysis)

Take-home message: Tidymodels pros & cons

Disadvantages:

- The core of the libraries is "hidden", which can lead to the risky approach of "using a function without knowing what it does"
- The tidyverse approach may not be intuitive, especially for those familiar only with base R
- Working with data.frames may impact performance (very large datasets can slow down computation)
- Other tools are available (e.g., Scikit-Learn in Python) is it worth learning another language?

Thank you for the attention!



