

¹ Supporting cells in the cochlea ² reduce hair cell excitability by ³ increasing the extracellular space

⁴ **Travis A. Babola¹, Calvin J. Kersbergen¹, Han Chin Wang^{1†}, Dwight E. Bergles^{1,2,3*}**

*For correspondence:
dbergles@jhmi.edu (DEB)

Present address: [†]Helen Wills Neuroscience Institute, University of California, Berkeley, United States

⁵ ¹The Solomon Snyder Department of Neuroscience, Johns Hopkins University,
⁶ Baltimore, Maryland 21205, USA; ²Department of Otolaryngology Head and Neck
⁷ Surgery, Johns Hopkins University, Baltimore, Maryland 21287, USA; ³Johns Hopkins
⁸ University Kavli Neuroscience Discovery Institute, Baltimore, Maryland, 21205

⁹

¹⁰ **Abstract** Neurons in developing sensory pathways exhibit spontaneous bursts of electrical
¹¹ activity that are critical for survival, maturation and circuit refinement. In the auditory system,
¹² intrinsically generated activity arises within the cochlea, but the molecular mechanisms that
¹³ initiate this activity remain poorly understood. We show that burst firing of mouse inner hair cells
¹⁴ prior to hearing onset requires P2RY1 autoreceptors expressed by inner supporting cells. P2RY1
¹⁵ activation triggers K⁺ efflux and depolarization of hair cells, as well as osmotic shrinkage of
¹⁶ supporting cells that dramatically increased the extracellular space and speed of K⁺
¹⁷ redistribution. Pharmacological inhibition or genetic disruption of P2RY1 suppressed neuronal
¹⁸ burst firing by reducing K⁺ release, but unexpectedly enhanced their tonic firing, as water
¹⁹ resorption by supporting cells reduced the extracellular space, slowing K⁺ clearance. These
²⁰ studies indicate that purinergic signaling in supporting cells regulates hair cell excitability by
²¹ controlling the volume of the extracellular space.

²²

²³ **Introduction**

²⁴ The developing nervous system must generate, organize, and refine billions of neurons and their
²⁵ connections. While molecular guidance cues forge globally precise neuronal connections between
²⁶ distant brain areas (*Stoeckli, 2018; Dickson, 2002*), the organization of local connections is initially
²⁷ coarse and imprecise (*Dhande et al., 2011; Kirkby et al., 2013; Sretavan and Shatz, 1986*). Coinci-
²⁸ dent with the refinement of topographic maps, nascent circuits experience bursts of intrinsically
²⁹ generated activity that emerge before sensory systems are fully functional (*Kirkby et al., 2013*).
³⁰ This intrinsically generated activity consists of periodic bursts of high frequency firing that pro-
³¹ motes the survival and maturation of neurons in sensory pathways (*Blankenship and Feller, 2010;*
³² *Moody and Bosma, 2005*). The precise patterning of this electrical activity appears crucial for re-
³³ finement of local connections, as its disruption results in improper formation of topographic maps
³⁴ (*Antón-Bolaños et al., 2019; Burbridge et al., 2014; Xu et al., 2011*) and impaired maturation and
³⁵ specification of sensory neurons (*Shrestha et al., 2018; Sun et al., 2018*). In all sensory systems
³⁶ that have been examined, spontaneous burst firing arises within their respective developing sen-
³⁷ sory organs, e.g. retina, olfactory bulb, spindle organ, and cochlea (*Blankenship and Feller, 2010*).
³⁸ Although the mechanisms that induce spontaneous activity in the developing retina have been ex-
³⁹ tensively explored, much less is known about the key steps involved in triggering auditory neuron
⁴⁰ burst firing in the developing cochlea. Understanding these processes may provide novel insights

41 into the causes of developmental auditory disorders, such as hypersensitivity to sounds and audi-
42 tory processing disorders that prevent children from communicating and learning effectively.

43 The mechanisms responsible for initiating spontaneous activity appear to be unique to each
44 sensory system, reflecting adaptations to the structure and cellular composition of the sensory or-
45 gans. In the cochlea, two distinct models have been proposed to initiate burst firing of inner hair
46 cells. One model proposes that burst firing results from intermittent hyperpolarization of tonically
47 active IHCs by cholinergic efferents (*Johnson et al., 2011; Wang and Bergles, 2014*), which provide
48 prominent inhibitory input to IHCs prior to hearing onset (*Glowatzki and Fuchs, 2000*). Consistent
49 with this model, activation of acetylcholine receptors in acutely isolated cochleae caused IHCs to
50 switch from sustained to burst firing (*Johnson et al., 2011*). However, *in vivo* recordings from audi-
51 tory brainstem revealed that neuronal burst firing remains, with altered features, in a9 knockout
52 mice (*Clause et al., 2014*), which lack functional efferent signaling in IHCs (*Johnson et al., 2013*), and
53 persists in IHCs and auditory neurons in cochleae maintained *in vitro* without functional efferents
54 (*Johnson et al., 2013*). Thus, cholinergic efferents appear to modulate the temporal characteristics
55 of bursts, but are not essential to initiate each event.

56 An alternative model proposes that IHCs are induced to fire bursts of action potentials by the
57 release of K⁺ from nearby inner supporting cells (ISCs), which together form a transient structure
58 known as Kölliker's organ (Greater Epithelial Ridge) that is prominent prior to hearing onset (*Tritsch
59 et al., 2010b*). K⁺ release from ISCs occurs following a cascade of events that begins with the spon-
60 taneous release of ATP and activation of purinergic autoreceptors. Purinergic receptor activation
61 induces an increase in intracellular Ca²⁺, opening of Ca²⁺-activated Cl⁻ channels (TMEM16A), and
62 efflux of Cl⁻ and subsequently K⁺ to balance charge (*Tritsch et al., 2007; Wang et al., 2015*). The loss
63 of ions during each event draws water out of ISCs through osmosis, leading to pronounced shrink-
64 age (crenation) of ISCs. While these pathways have been extensively studied *in vitro*, the molecular
65 identity of the purinergic receptors has remained elusive and few manipulations of key steps in
66 this pathway have been performed *in vivo*, limiting our understanding of how spontaneous activity
67 is generated at this critical stage of development.

68 Here, we show that the key initial step in generation of spontaneous activity in the auditory
69 system involves activation of P2RY1 autoreceptors in ISCs. These metabotropic receptors induce
70 Ca²⁺ release from intracellular stores that allow TMEM16A channels to open. Pharmacological in-
71 hibition of P2RY1 or genetic deletion of *P2ry1* dramatically reduced burst firing in spiral ganglion
72 neurons (SGNs) and blocked the coordinated, spatially restricted activation of ISCs, IHCs, and SGNs
73 in the cochlea. Unexpectedly, P2RY1 activation also promoted the clearance of K⁺ by increasing
74 the volume of extracellular space, enhancing the diffusion of K⁺ ions away from IHCs. Conversely,
75 inhibition of P2RY1 reduced the extracellular space and restricted the redistribution of K⁺ within
76 the cochlear epithelium, causing IHCs to depolarize and fire tonically, demonstrating an important
77 role for purinergic receptor-mediated extracellular space changes in controlling IHC excitability.
78 Using *in vivo* widefield epifluorescence imaging of the auditory midbrain in unanesthetized mice,
79 we show that acute inhibition of P2Y1 receptors dramatically reduced burst firing of auditory neu-
80 rons in isofrequency domains. Together, these data indicate P2RY1 autoreceptors in non-sensory,
81 cochlear supporting cells play a crucial role in generating bursts of activity among neurons that will
82 ultimately process similar frequencies of sound, providing the means to initiate the maturation of
83 auditory pathways before hearing onset.

84 Results

85 Supporting cell spontaneous currents require calcium release from intracellular 86 stores

87 Periodic release of ATP from ISCs in the developing cochlea initiates a signaling cascade in these
88 cells that increases intracellular calcium (Ca²⁺), opens Ca²⁺-activated Cl⁻ channels (TMEM16A), and
89 ultimately results in efflux of chloride and K⁺ into the extracellular space. Although the increase

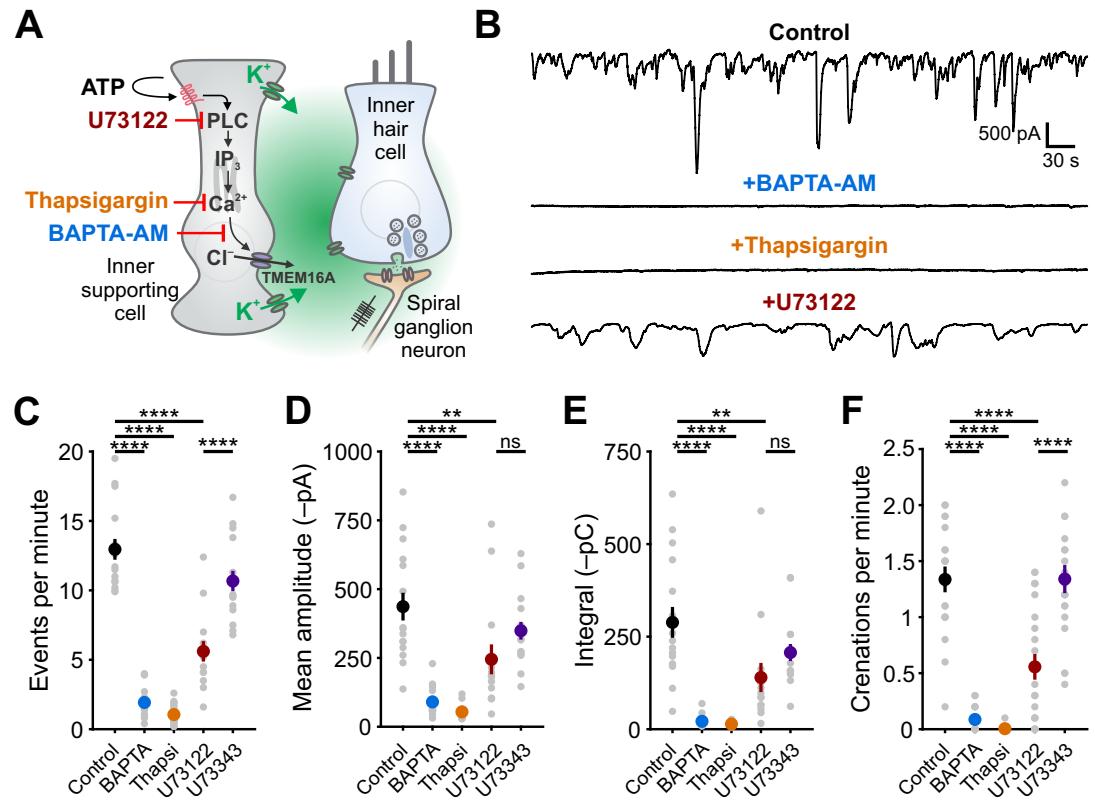


Figure 1. Ca²⁺ release from intracellular stores is required for spontaneous currents and crenation in inner supporting cells. **(A)** Model of ATP-mediated depolarization of inner hair cells. ATP: adenosine triphosphate, PLC: phospholipase C, IP₃: inositol triphosphate, TMEM16A: transmembrane member 16A (Ca²⁺-activated Cl⁻ channel). Inhibitors of key steps in this pathway are indicated. **(B)** Whole-cell voltage-clamp recordings from inner supporting cells after pre-incubating with indicated inhibitors. **(C)** Quantification of ISC spontaneous current frequency in the presence of inhibitors of the G_q pathway. Data shown as mean ± SEM. n = 16 cells (control), 16 cells (BAPTA-AM; 100µM), 20 cells (Thapsigargin; 2µM), 14 cells (U73211; 10µM), and 16 cells (U73343; 10µM). ****p<5e-5, one-way ANOVA. **(D)** Quantification of ISC spontaneous current amplitude in the presence of inhibitors of the G_q pathway. Data shown as mean ± SEM. n values are reported in (C) (one-way ANOVA; ****p<5e-5, **p<0.005, ns: not significant). **(E)** Quantification of ISC spontaneous current charge transfer (integral) in the presence of inhibitors of the G_q pathway. Data shown as mean ± SEM. n values are reported in (C) (one-way ANOVA; ****p<5e-5, **p<0.005, ns: not significant). **(F)** Quantification of ISC crenation (cell shrinkage) frequency in the presence of inhibitors of the G_q pathway. Data shown as mean ± SEM. n values are reported in (C) (one-way ANOVA; ****p<5e-5, **p<0.005, ns: not significant).

in intracellular Ca²⁺ following activation of purinergic autoreceptors is sufficient to induce both depolarization and osmotic shrinkage (crenation; *Wang et al. (2015)*), the relative contributions of Ca²⁺ influx (e.g. through Ca²⁺-permeable, ionotropic P2X receptors) and release from intracellular stores (e.g. following metabotropic P2Y receptor activation) to these cytosolic Ca²⁺ transients is unclear. To define the signaling pathways engaged by purinergic receptor activation, we examined the sensitivity of spontaneous ISC whole-cell currents and crenations to inhibitors of intracellular Ca²⁺ pathways (*Figure 1A*). Spontaneous inward currents and crenations were abolished following a 15 minute incubation of excised cochlea in BAPTA-AM (100µM), a cell permeant Ca²⁺ chelator (*Figure 1B-F*), and after depleting intracellular Ca²⁺ stores with thapsigargin (2µM), an inhibitor of endoplasmic reticulum Ca²⁺-ATPase (*Figure 1B-F*). These data suggest that Ca²⁺ release from intracellular stores is necessary for spontaneous electrical activity in ISCs.

Metabotropic G_q-coupled receptors typically induce PLC-mediated cleavage of phosphatidylinositol 4,5-bisphosphate (PIP₂) and subsequent binding of inositol trisphosphate (IP₃) to IP₃ receptor channels on the endoplasmic reticulum to release Ca²⁺ into the cytoplasm. To investigate if PLC

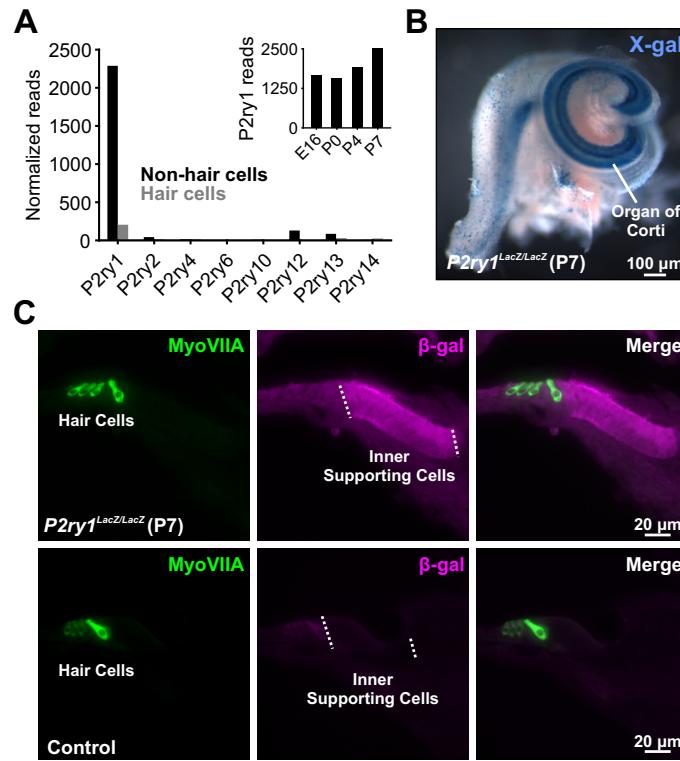


Figure 2. The metabotropic P2Y1 receptor is highly expressed by ISCs. **(A)** Expression levels of metabotropic purinergic receptors in hair cells (grey) and non-sensory cells (black) of the developing cochlea (postnatal day 7, P7). (inset) *P2y1* expression in non-sensory cells over development. Data adapted from Scheffer et al. (2015). **(B)** Image of a cochlea following X-gal reaction in *P2ry1-LacZ* reporter mice. **(C)** Immunostaining for β-galactosidase in cochleae from P7 *P2ry1-LacZ* (top) and control (bottom) cochlea.

signaling is required for generation of spontaneous activity in ISCs, we recorded spontaneous currents and crenations from ISCs in the presence of U73122 (10µM), a PLC inhibitor, and U73343 (10µM), an inactive succinimide analog. The frequency of spontaneous currents and crenations were dramatically reduced following U73122 incubation, but not U73343 (**Figure 1B-F**); the amplitudes and charge transfer of residual activity also trended lower during PLC inhibition, but this did not reach significance due to high variance in the sizes of these responses (**Figure 1B-F**). Together, these results suggest that engagement of a G_q-coupled purinergic autoreceptor is a critical first step in initiating PLC-mediated Ca²⁺ release from intracellular stores and subsequent activation of TMEM16A channels.

The metabotropic purinergic receptor P2Y1 is highly expressed by supporting cells
 There are eight members of the metabotropic purinergic receptor family in mouse, four of which are G_q-coupled (P2RY1, P2RY2, P2RY4, and P2RY6). Gene expression studies in the developing mouse cochlea revealed that non-sensory cells express *P2ry1* mRNA at high levels (Scheffer et al., 2015), >100 fold higher than any other *P2ry* (**Figure 2A**) and that expression of this receptor progressively increases during early postnatal development (**Figure 2A**, inset) concurrent with increases in spontaneous activity (Tritsch and Bergles, 2010). To determine which cells in the sensory epithelium express P2RY1, we isolated cochleae from *P2ry1-LacZ* reporter mice and performed X-gal staining. Intense blue labeling was present along the entire length of the cochlea within Källiker's organ (Greater Epithelial Ridge; **Figure 2B**). Cross-sections of cochlea revealed that staining was present within ISCs, but not IHCs (Myosin VIIA, **Figure 2C**), indicating that P2RY1 is properly localized to sense ATP release from ISCs prior to hearing onset.

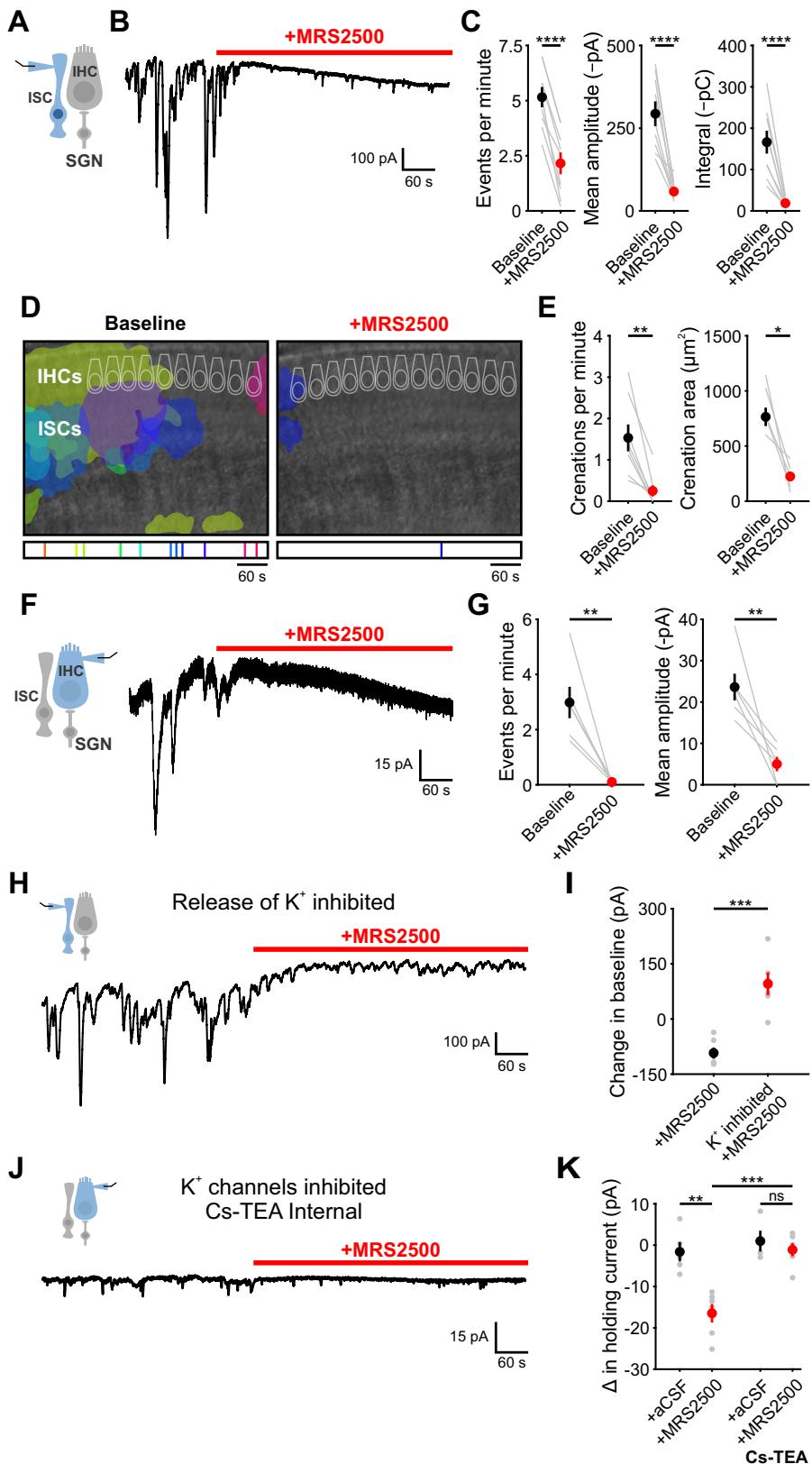


Figure 3. P2Y1 inhibition abolishes spontaneous currents in inner supporting cells and inner hair cells. **(A)** Schematic of ISC whole-cell recording configuration. **(B)** ISC spontaneous inward currents before and after application of MRS2500 (1 μ M). Recordings performed at \sim 25°C. **(C)** Plot of event frequency, amplitude, and integral (charge transfer) before and after MRS2500 application. $n = 8$ ISCs (two-tailed paired Student's t test; *** p <5e-5) **(D)** Intrinsic optical imaging performed before and after application of the P2RY1 antagonist, MRS2500 (1 μ M). Detected crenations are outlined in colors based on time of occurrence as indicated below image. Imaging performed at \sim 25°C. Caption continued on next page.

Figure 3. (E) Plot of crenation frequency and area before and after MRS2500 application. n = 7 cochleae (two-tailed paired Student's t test; **p<0.005) for frequency calculation and n = 5 cochleae (two-tailed paired Student's t test; *p<0.05) for area calculation. Cochleae that did not crenate in MRS2500 were excluded from area calculations. **(F)** Schematic of whole-cell recording configuration from IHCs. (right) Whole-cell voltage clamp recording from an IHC. **(G)** Plots of event frequency and amplitude before and after MRS2500 application. n = 6 IHCs (two-tailed paired Student's t test with Bonferroni correction; **p<0.005, *p<0.05). **(H)** Whole-cell voltage clamp recording of an ISC with MRS2500 application following pre-incubation with CdCl₂ (100μM), TTX (1μM), ouabain (10μM), and bumetanide (50μM) to limit K⁺ release. **(I)** Plot of change in holding current, defined as the 95% percentile current for each period. n = 6 ISCs for each condition (two-tailed Student's t test; ***p<0.0005). **(J)** IHC whole-cell voltage clamp recording with Cs-TEA internal solution (to inhibit K⁺ channels) before and after MRS2500 application. **(K)** Plot of the change in IHC holding current following control (aCSF) and MRS2500 with K-MeS and Cs-TEA internal. n = 5 IHCs for aCSF, n = 6 IHCs for MRS2500, n = 4 IHCs for aCSF with Cs-TEA internal, and n = 5 IHCs for MRS2500 with Cs-TEA internal (one-way ANOVA; ***p<0.005, **p<0.005, ns, not significant).

Figure 3–Figure supplement 1. P2RY1 inhibition abolishes spontaneous currents in inner supporting cells and inner hair cells near physiological temperature.

Figure 3–Figure supplement 2. Spontaneous inward currents and crenations are dramatically reduced in *P2ry1* KO mice.

125 **P2RY1 signaling is required for spontaneous activity in ISCs and IHCs**

126 To determine if P2RY1 is responsible for spontaneous ATP-mediated currents in ISCs, we examined
 127 the sensitivity of these responses and associated crenations to the P2RY1 antagonist MRS2500 (*Fig-*
 128 *ure 3A,B*). Acute inhibition of P2RY1 with MRS2500 (1 μM) markedly reduced both spontaneous ISC
 129 currents (*Figure 3B,C*) and crenations (*Figure 3D,E*); near complete inhibition occurred within min-
 130 utes at both room temperature (*Figure 3B,C*) and near physiological temperature (*Figure 3–Figure*
 131 *Supplement 1A-G*), with only sporadic, small amplitude events remaining that were not mediated
 132 by purinergic receptors (*Figure 3–Figure Supplement 1B-E*). Consistent with the involvement of
 133 P2RY1, the amplitude and total charge transfer of ISC events (*Figure 3–Figure Supplement 2A,B*)
 134 and size of spontaneous crenations (*Figure 3–Figure Supplement 2C,D*) were smaller in cochleae
 135 in *P2ry1* KO mice relative to controls. However, supporting cells *P2ry1* KO mice exhibited aberrant,
 136 gain-of-function activity consisting of frequent, small amplitude currents (*Figure 3–Figure Supple-*
 137 *ment 2A,B*), that were not blocked by MRS2500 or broad-spectrum P2 receptor antagonists (*Fig-*
 138 *ure 3–Figure Supplement 2E,F*).

139 ATP-mediated signaling in ISCs activates TMEM16A, triggering K⁺ efflux that depolarizes nearby
 140 IHCs. To assess whether P2RY1 signaling is also required for periodic excitation of IHCs prior to
 141 hearing onset, we assessed the sensitivity of spontaneous IHC inward currents to MRS2500 (*Fig-*
 142 *ure 3F*). Consistent with the supporting cell origin of IHC activity, application of MRS2500 (1 μM) also
 143 abolished spontaneous currents in IHCs (*Figure 3F,G*). Together, these data suggest that P2RY1 is
 144 the primary purinergic autoreceptor on ISCs responsible for inducing periodic excitation of hair
 145 cells prior to hearing onset.

146 **P2RY1 inhibition leads to extracellular K⁺ accumulation**

147 Although P2RY1 inhibition abolished most transient inward currents in both ISCs and IHCs, a pro-
 148 gressively increasing inward current (downward shift in baseline) appeared in both cell types with
 149 prolonged application of MRS2500 (*Figure 3B,F*). Prior studies in CNS brain slices indicated that G_q-
 150 coupled purinergic receptors in astrocytes regulate extracellular K⁺ concentration and neuronal
 151 excitability (*Wang et al., 2012*). The slowly progressing nature of the response in IHCs and ISCs
 152 suggest that it may similarly arise from accumulation of K⁺ released from cells in the organ of Corti.
 153 If this hypothesis is correct, then inhibiting the main sources of K⁺ should diminish this inward
 154 current. Indeed, when IHC and SGN excitation was inhibited with tetrodotoxin (TTX, 1 μM) and cad-
 155 dium (CdCl₂, 100μM), and the K⁺ transporters, Na,K-ATPase and NKCC, were inhibited with ouabain
 156 (10μM) and bumetanide (50μM), no inward current was induced in ISCs upon blocking P2RY1 (*Fig-*
 157 *ure 3H,I*). Similarly, if K⁺ accumulation is responsible for the current in IHCs, it should be abolished
 158 when the ability of IHCs to detect changes in K⁺ is reduced. When whole cell recordings were per-
 159 formed from IHCs using an internal solution containing Cs⁺ and TEA, which blocks most IHC K⁺
 160 channels (*Kros et al., 1998; Marcotti et al., 2003*), MRS2500 also did not induce an inward current

161 (**Figure 3J,K**). Together, these results suggest that P2RY1 has two distinct effects in the cochlea; it
 162 induces the transient inward currents that triggers IHC burst firing and it accelerates the clearance
 163 of K⁺ within the organ of Corti.

164 To directly assess the relationship between P2RY1 activity and extracellular K⁺ accumulation
 165 near IHCs, we monitored K⁺ levels in the extracellular space using IHC K⁺ channels. Focal P2RY1
 166 stimulation with a selective agonist (MRS2365, 10μM), which mimics the effect of endogenous ATP
 167 by eliciting an inward current and crenations in ISCs in control but not *P2ry1* KO mice (**Figure 4A-C**),
 168 was combined with assessments of the reversal potential of K⁺ currents in IHCs using a voltage
 169 protocol similar to one used to assess extracellular K⁺ buildup at vestibular calyceal synapses (*Lim*
 170 *et al.*, 2011). This protocol consisted of: (1) a hyperpolarizing step to -110mV to relieve K⁺ chan-
 171 nel inactivation, (2) a depolarizing step to +30mV to activate outward K⁺ currents, and (3) a step
 172 to -70mV to obtain a "tail" current (**Figure 4D-F**). Because the conductance during this last step
 173 is largely mediated by K⁺ channels, it is highly sensitive to shifts in K⁺ driving force induced by
 174 changes in extracellular K⁺ (*Contini et al.*, 2017; *Lim et al.*, 2011). Following transient stimulation
 175 of P2RY1, K⁺ tail currents immediately shifted inward, as would be expected if extracellular K⁺ in-
 176 creases (**Figure 4G,I**), and is similar to the effects of a metabotropic purinergic receptor agonist
 177 (UTP) on synaptically-evoked K⁺ currents in IHCs (*Wang et al.*, 2015). However, after a few seconds
 178 these K⁺ currents shifted outward relative to baseline, indicative of a gradual decrease in extracellu-
 179 lar K⁺ below that present prior to P2RY1 stimulation, then gradually returned to the pre-stimulation
 180 level after several minutes (**Figure 4G,I**).

181 The outward shift in K⁺ tail current followed the time course of the crenation ($\tau_{\text{decay}} = 100 \pm 14\text{s}$
 182 for tail currents and $\tau_{\text{decay}} = 38 \pm 4\text{s}$ for crenations), suggesting that the shrinkage of cells induced
 183 by P2RY1 activation results in a prolonged increase in extracellular space that may allow greater
 184 dilution and more rapid redistribution of K⁺ in the organ of Corti. Alternatively, buildup of extra-
 185 cellular K⁺ alone may stimulate greater uptake. To determine if rapid increases in extracellular
 186 K⁺ or Cl⁻ were sufficient to stimulate K⁺ redistribution in the absence of crenation, we puffed KCl
 187 (130mM) into the supporting cell syncytium in the presence of P2RY1 antagonists (**Figure 4H**). As
 188 expected, this transient increase in extracellular K⁺ induced an inward shift in K⁺ tail currents and
 189 a brief optical change induced by fluid delivery; however, K⁺ tail currents rapidly returned to base-
 190 line and did not shift outward, suggesting that K⁺ (and Cl⁻) efflux are not sufficient to enhance K⁺
 191 redistribution rates. In addition, we transiently stimulated P2RY1 in *Tecta-Cre;TMEM16A^{fl/fl}* mice, in
 192 which purinergic receptor activation is preserved, but crenations are abolished (*Wang et al.*, 2015).
 193 In these mice, ISCs failed to crenate, IHCs did not depolarize, and K⁺ tail currents remained stable
 194 throughout the duration of the recording (**Figure 4I**). These results suggest that purinergic auto-
 195 receptors on ISCs influence extracellular K⁺ levels by both triggering K⁺ release and by altering K⁺
 196 redistribution by controlling the size of the extracellular space.

197 **P2ry1 mediates coordinated neuronal activation and precise burst firing of SGNs**
 198 To evaluate the role of P2RY1 in initiating coordinated cellular activity in the cochlea, we monitored
 199 large-scale activity patterns in excised cochleae from *Pax2-Cre;R26-Isl-GCaMP3* mice, which express
 200 GCaMP3 in nearly all cells of the inner ear. Time lapse imaging revealed that the spontaneous Ca²⁺
 201 elevations that occur simultaneously within groups of ISCs, IHCs, and SGNs (*Tritsch and Bergles*,
 202 2010; *Zhang-Hooks et al.*, 2016) were abolished following inhibition of P2RY1 with MRS2500 (**Fig-**
 203 **ure 5A-C**) and were dramatically reduced in *P2ry1* KO mice (*Pax2-Cre;R26-Isl-GCaMP3;P2ry1^{-/-}*) (**Fig-**
 204 **ure 5-Figure Supplement 1A,B**). Moreover, in accordance with the progressive increase in extra-
 205 cellular K⁺ that follows P2RY1 inhibition, there was a gradual increase in spontaneous, uncoordinated
 206 Ca²⁺ transients in IHCs in the presence of MRS2500 (**Figure 5D-F**), suggesting that this K⁺ accumula-
 207 tion increases IHC firing. Similarly, IHCs in *P2ry1* KO mice displayed a higher level of uncorrelated
 208 hair cell Ca²⁺ transients (**Figure 5-Figure Supplement 1C-E**), indicative of enhanced excitability. To-
 209 gether, these results indicate that P2RY1 is required for coordinated activation of ISCs, IHCs, and
 210 SGNs before hearing onset and that P2RY1 inhibition leads to higher rates of uncorrelated activity.

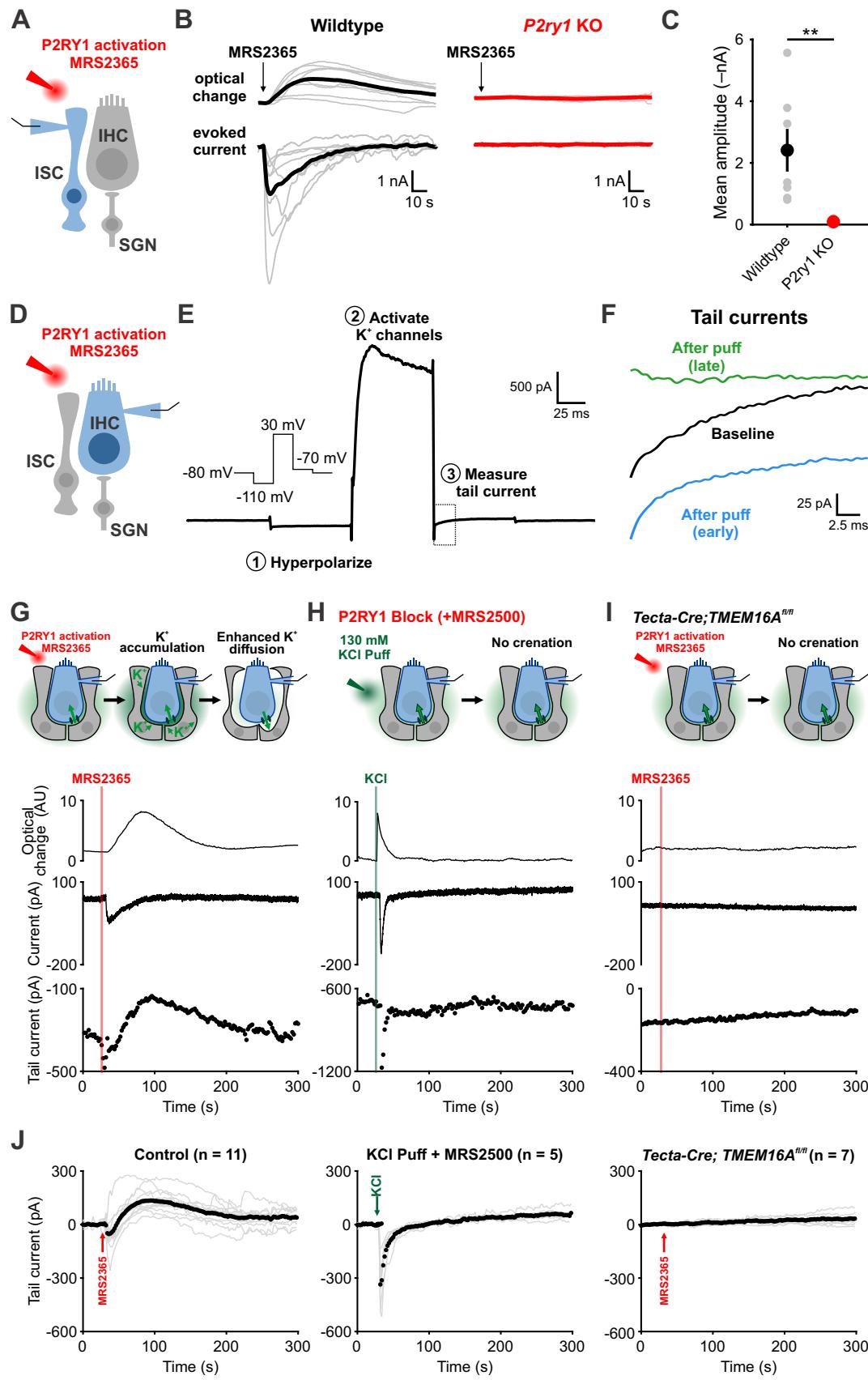


Figure 4. Caption on next page.

Figure 4. Activation of P2RY1 results in an initial accumulation of extracellular K⁺, followed by crenation and enhanced K⁺ clearance. **(A)** Schematic of whole-cell recording configuration from ISCs with puffs of MRS2365 (10μM), a P2RY1 agonist. **(B)** Optical change (crenation) and current elicited with MRS2365 puffs in wildtype and *P2ry1* KO mice. **(C)** Plot of mean current amplitude with MRS2365 puffs. n = 6 and n = 5 ISCs from wildtype and *P2ry1* KO mice, respectively (two-tailed Student's t test; **p<0.005). **(D)** Schematic of whole-cell recording configuration from IHCs with puffs of MRS2365 (10μM). **(E)** Example current trace and voltage-protocol designed to measure K⁺ accumulation. Dashed box indicated tail current measurement period indicated in (F). **(F)** Tail currents observed during baseline, immediately following the MRS2365 puff (with 2 seconds), and after the puff (30 seconds). **(G)** Model of K⁺ dynamics following MRS2365 stimulation. Initially, extracellular K⁺ rapidly increases following stimulation, but ISCs crenate, increasing the amount of extracellular space and K⁺ buffering. (bottom) Optical change (crenation), holding current, and tail current as a function of time with respect to MRS2365 puff. **(H)** Similar to G, but with KCl puffs (130μM) in cochlea treated with MRS2500. **(I)** Similar to G, but in *Tecta-Cre;TMEM16A^{fl/fl}* mice where TMEM16A has been conditionally removed from the sensory epithelium (see *Figure 4-Figure Supplement 1*). No crenations were observed with MRS2365 stimulation. **(J)** Plot of tail currents over time following MRS2365 stimulation. Grey lines indicate individual IHCs; black points indicate the mean across the population. Baseline was normalized to 0 pA for all traces.

Figure 4-Figure supplement 1. Crispr-Cas9 mediated generation of the *Tecta-Cre* mouse line.

IHCs in the developing cochlea exhibit regenerative Ca²⁺ spikes that strongly activate post-synaptic SGNs, resulting in bursts of action potentials that propagate to the CNS. To determine if P2RY1 initiates burst firing in SGNs, we recorded spontaneous activity from SGNs using juxtaglomerular recordings from their somata (*Figure 6A*). Application of MRS2500 resulted in a dramatic reduction of high frequency burst firing in SGNs, visible as a decrease in burst frequency and action potentials per burst (*Figure 6E,F*). All SGN spiking was abolished by the AMPA receptor antagonist NBQX (50μM) (*Figure 6D*), indicating that their activity requires synaptic excitation by IHCs. The precise patterning of action potentials within bursts was also disrupted by P2RY1 inhibition, as there were fewer interspike intervals in the 75–125ms range (*Figure 6C,F*), which correspond to the maximum rate of Ca²⁺ spike generation by IHCs during ATP-mediated excitation (*Tritsch et al., 2010a*). Additionally, the coefficient of variation measured for interspike intervals was significantly lower following P2RY1 inhibition, suggesting SGNs fire more randomly (*Figure 6E*). However, the average frequency of action potentials remained unchanged during P2RY1 inhibition (*Figure 6E*), due to increases in non-burst firing (*Figure 3F,Figure 4D*). SGNs in *P2ry1* KO cochlea exhibited activity similar to wildtype SGNs in the presence of MRS2500, with a lower burst firing rate, fewer interspike intervals in the 75–125ms range, and a lower coefficient of variation of interspike intervals relative to controls (*Figure 6G-I*). However, despite the profound contribution of P2RY1 to ISC and IHC activity, some burst-like behavior was still observed in SGNs (*Figure 6D,G*), suggesting that other forms of excitation emerge in the absence of P2RY1, due to an increase in overall excitability or developmental changes. Together, these data indicate that P2RY1 is required to generate discrete bursts of action potentials in SGNs and that loss of these receptors enhances uncorrelated firing.

P2RY1 promotes auditory neuron firing *in vivo*

The highly synchronized electrical activity exhibited by IHCs prior to hearing onset propagates through the entire developing auditory system to induce correlated firing of auditory neurons within isofrequency zones (*Babola et al., 2018; Tritsch et al., 2010a*). To determine if P2RY1 is required to produce this form of correlated activity, we used *in vivo* wide-field epifluorescence microscopy of the inferior colliculus (IC) in mice that express GCaMP6s in all neurons (*Snap25-T2A-GCaMP6s* and *Snap25-T2A-GCaMP6s;P2ry1^{-/-}* mice). Time lapse imaging revealed that both control and *P2ry1* KO mice exhibited correlated neuronal activity confined to stationary bands oriented along the tonotopic axis (*Figure 7A-C*). Spontaneous events were less frequent in *P2ry1* KO mice (9.7 ± 0.8 events per minute compared to 13.4 ± 0.7 events per minute in control; two-tailed Student's t test, p = 0.002), although the events were similar in amplitude and duration (half-width) (*Figure 7D*), suggesting that some compensatory amplification of events occurs in the CNS of these mice, similar to that seen in *Vglut3* KO mice (*Babola et al., 2018*). Spontaneous activity in *P2ry1* KO mice differed from controls in three other ways. First, the contralateral bias exhibited for each

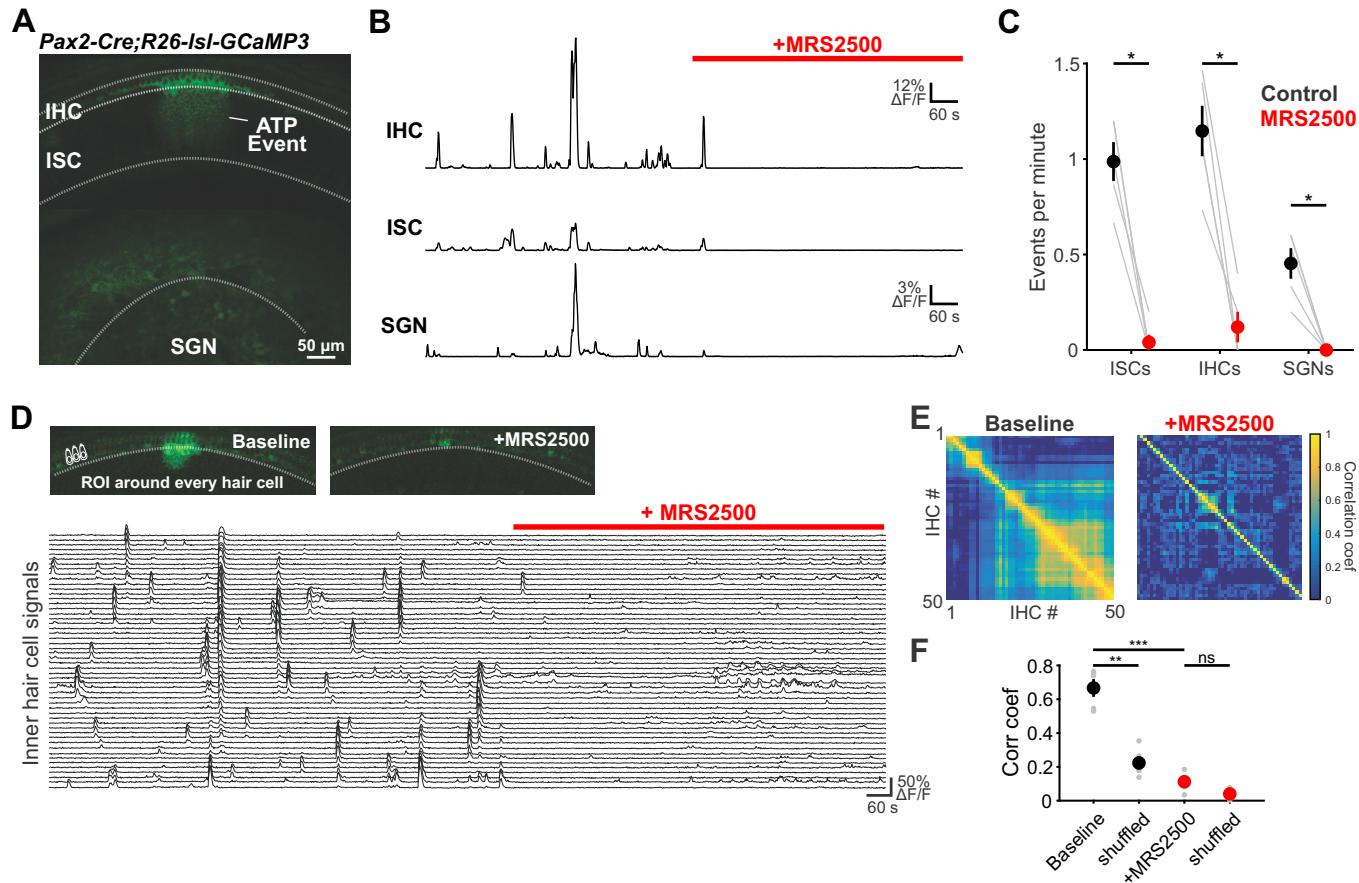


Figure 5. Large-scale coordinated activity in the cochlea requires P2RY1. **(A)** Exemplar Ca^{2+} transient in excised cochlea from *Pax2-Cre;R26-*Isl*-GCaMP3* mice. Note coordinated activation of ISCs, IHCs, and SGNs. **(B)** Traces of fluorescence intensity over time taken from ROIs that span the entire IHC, ISC, and SGN regions indicated in (A). **(C)** Plot of event frequency before and during application of MRS2500 (1 μM). $n = 5$ cochlea (two-tailed paired Student's t test with Bonferroni correction; * $p < 0.05$). **(D)** Exemplar images of IHC Ca^{2+} transients. ROIs were drawn around every IHC for subsequent analysis (bottom). **(E)** Correlation matrices generated by calculating the linear correlation coefficient for all IHC pairs before and after MRS2500 application. **(F)** Plot of average correlation coefficient calculated between the four nearest IHCs or four randomly shuffled IHCs. $n = 5$ cochleae (two-tailed paired Student's t test with Bonferroni correction; *** $p < 0.0005$, ** $p < 0.005$, ns, not significant).

Figure 5-Figure supplement 1. *P2ry1* KO mice exhibit reduced Ca^{2+} transients in ISCs.

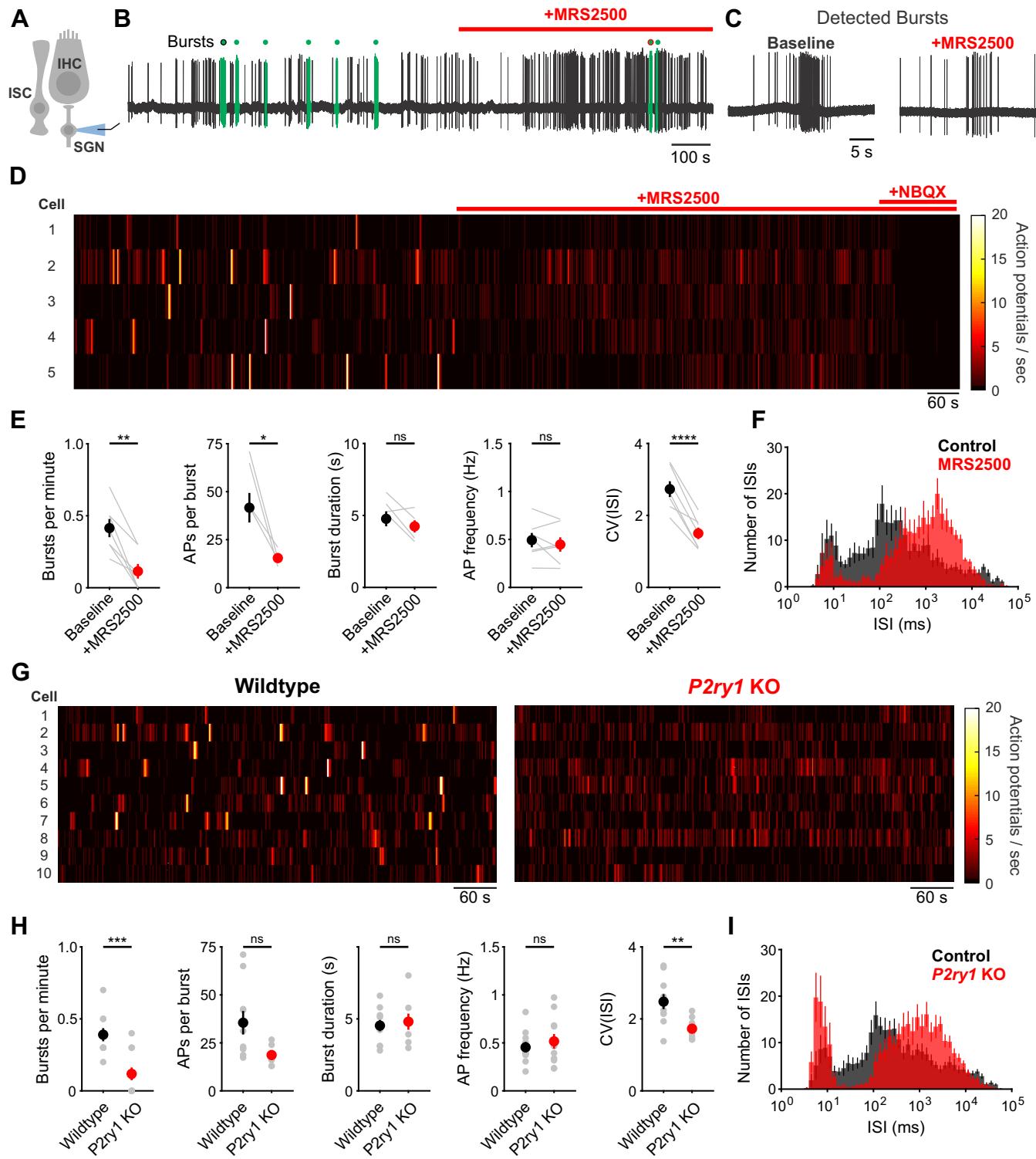


Figure 6. Inhibition of P2RY1 disrupts burst firing in spiral ganglion neurons. **(A)** Schematic of SGN juxtaglomerular recording configuration. All recordings were performed at ~25°C. **(B)** Action potentials recorded before and during MRS2500 application (1 μM). Detected bursts are indicated in green (see Methods and Materials for parameters used for burst detection). Circles with black and red outlines are expanded in **(C)**. **(C)** Action potentials within a detected burst before and after MRS2500 application. **(D)** Raster plots indicating the average firing rate of SGNs (bin: 1 s) before and after MRS2500 application (1 μM) and subsequent NBQX (50 μM). **(E)** Plots of average burst frequency, burst duration, action potentials (AP) per burst, average AP frequency, and coefficient of variation for all interspike intervals (ISIs) measured. $n = 7$ SGNs from 7 cochleae (two-tailed paired Student's t-test with Bonferroni correction; **** $p < 5e-5$, ** $p < 0.005$, * $p < 0.05$, ns, not significant). **(F)** Average log-binned interspike interval histograms before and after MRS2500 application. **(G)** Raster plots indicating the average firing rate of SGNs (bin: 1 s) in wildtype and *P2ry1* KO mice. **(H)** Plots of average burst frequency, burst duration, action potentials (AP) per burst, average AP frequency, and coefficient of variation for all ISIs measured. $n = 10$ wildtype and 11 *P2ry1* KO SGNs (two-tailed Student's t-test with Bonferroni correction; *** $p < 0.0005$, ** $p < 0.005$, ns, not significant). **(I)** Average log-binned interspike interval histograms from wildtype and *P2ry1* KO SGNs.

event was higher, with the weaker relative to stronger side amplitude decreasing from 0.61 ± 0.02 to 0.44 ± 0.02 (two-tailed Student's t test, $p = 3.0\text{e-}6$) (**Figure 7D**). Second, the coefficient of variation (ratio of standard deviation to the mean) of event amplitudes was 40% higher relative to controls (**Figure 7D**). Third, a detailed examination of the spatial location of events across the tonotopic axis (**Figure 7E**) revealed that activity in brain areas later responsible for processing higher frequency tones (~8 – 16 kHz) was reduced by 68% in *P2ry1* KO mice, while activity in low frequency areas was unaltered (**Figure 7F-H**). In *P2ry1* KO mice, bilateral removal of both cochleae abolished activity in the IC, demonstrating that activity in these mice originates in the periphery (**Figure 7–Figure Supplement 1A-C**).

Although *P2ry1* KO mice mimic some aspects of acute P2RY1 inhibition, the absence of P2RY1 signaling throughout life may have led to compensatory changes, such as the increase in non-purinergic ISC activity (see **Figure 3–Figure Supplement 2E**). Therefore, to better assess the role of P2RY1 in generating spontaneous activity *in vivo*, we delivered a solution containing MRS2500 into the intraperitoneal cavity of mice while imaging activity in the IC. Compared to mice injected with control solution (5% mannitol), mice injected with MRS2500 exhibited dramatic reductions in IC event frequency (from 13.3 ± 0.8 to 3.9 ± 1.1 events per minute; two-tailed Student's t test, $p = 0.0001$) and amplitude (from 9.9 ± 0.5 to $4.9 \pm 0.8\Delta F/F_0$; two-tailed Student's t test, $p = 0.0006$) ~5 minutes after administration (**Figure 8A-D**). This decrease was specific to the IC, as SC retinal wave activity (*Ackman et al., 2012*) was unaffected by acute MRS2500 administration (**Figure 8B,C,E**), indicating that the locus of action is likely within the cochlea, which has been shown to have a less intact blood-tissue barrier at this age (*Suzuki et al., 1998*). Spatial analysis revealed that unlike the selective deficit observed in higher frequency zones in *P2ry1* KO mice, the inhibition was not limited to certain tonotopic regions, but rather occurred evenly across all frequency zones (**Figure 8F,G**). Together, these data indicate that ISC P2RY1 autoreceptors within the cochlea play a critical role in initiating spontaneous bursts of neural activity in auditory centers within the brain prior to hearing onset.

Discussion

Intense periods of neuronal activity dramatically alter the ionic composition of the extracellular environment, leaving behind excess K^+ that can alter neuronal excitability, induce spontaneous activity and trigger debilitating seizures. In the CNS, homeostatic control of extracellular K^+ levels is accomplished by glial cells, which redistribute K^+ passively through ion channels and actively through facilitated transport, but much less is known about the mechanisms that control excitability in the peripheral nervous system. Sensory hair cells and primary auditory neurons in the cochlea are surrounded by supporting cells that share key features with CNS glia and are thought to redistribute K^+ that accumulates during sound detection. However, prior to hearing onset, ATP dependent K^+ release from these cells triggers periodic bursts of activity in nearby IHCs that propagate throughout the auditory system. Here, we demonstrate that this form of intrinsically generated activity is initiated through activation of P2RY1, a G_q -coupled metabotropic purinergic receptor. Acute inhibition or genetic removal of this receptor dramatically reduced spontaneous activity and disrupted burst firing in IHCs, SGNs and central auditory neurons. In addition to triggering episodic K^+ dependent depolarization of hair cells, activation of P2RY1 also enhanced K^+ clearance by increasing the volume of extracellular space, allowing more rapid dissipation of extracellular K^+ transients. This duality of purpose, to induce K^+ efflux and enhance K^+ clearance, promotes discrete bursts of activity throughout the developing auditory system.

Purinergic signaling in the developing cochlea

Before the onset of hearing, neurons in the auditory system that will process similar sound frequencies exhibit periodic bursts of highly correlated activity, an entrainment that is initiated within the cochlea (*Babola et al., 2018; Clause et al., 2014; Sonntag et al., 2009; Tritsch et al., 2010a*). Within the developing cochlear epithelium, spontaneous release of ATP from ISCs activates purinergic

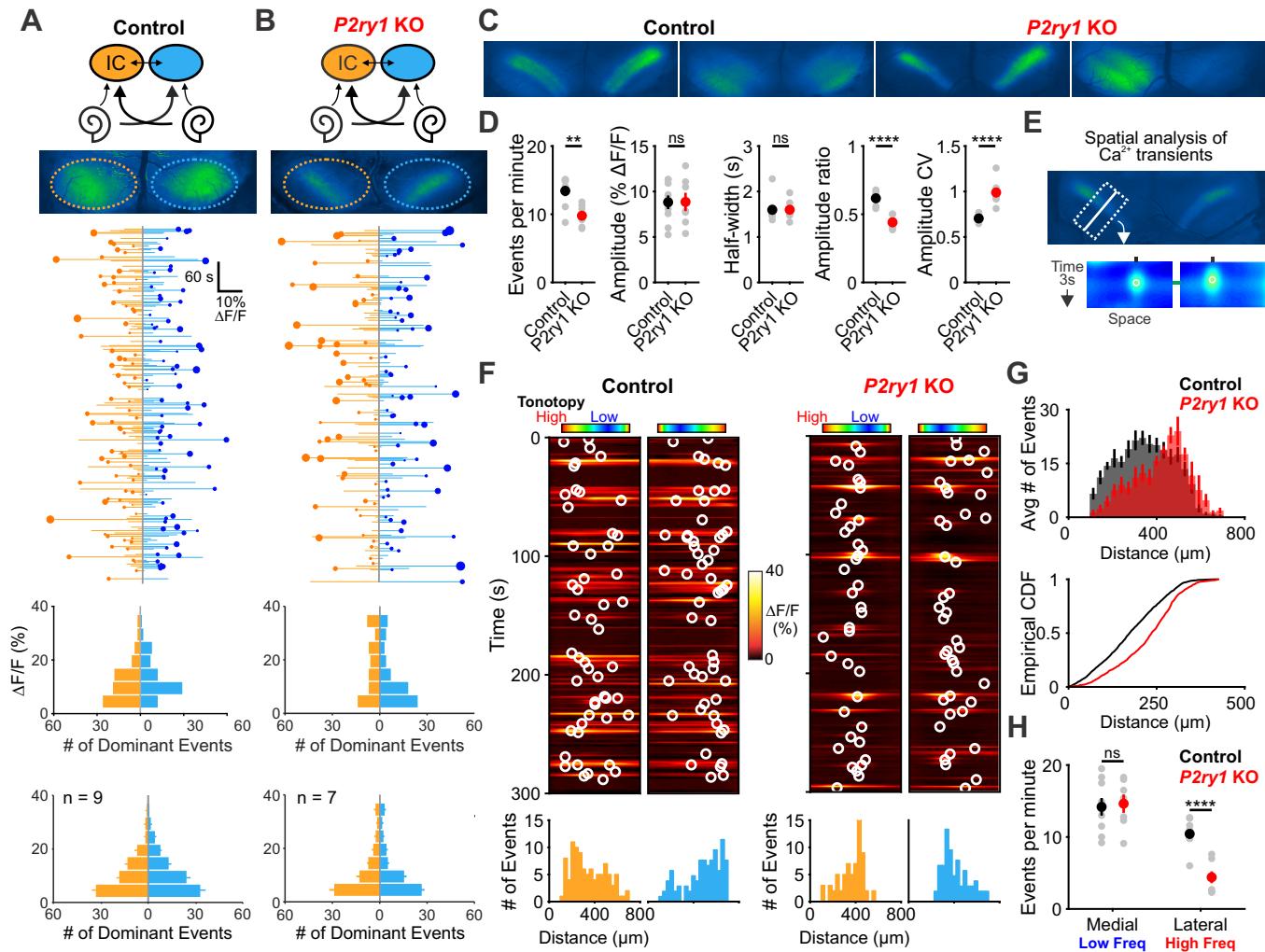


Figure 7. *P2ry1* KO mice exhibit reduced and spatially restricted spontaneous activity in the inferior colliculus. **(A)** Diagram illustrating flow of information through the auditory system and average intensity image over the 10 min imaging session. (middle) Activity over time in left and right IC in an individual where each line indicates the fluorescence intensity of each detected event; the circle indicates the dominant lobe, and the size of the circle indicated the difference in fluorescence. (bottom) Histograms showing the frequency of dominant events of a given amplitude for this experiment and for all experiments. Imaging was performed in *Snap25-T2A-GCaMP6s* mice ($n = 9$ mice). **(B)** Similar to (A), but in *Snap25-T2A-GCaMP6s;P2ry1^{-/-}* (*P2ry1* KO) mice ($n = 7$ mice). **(C)** Images of spontaneous events in the IC of in control (*Snap25-T2A-GCaMP6s*) and *P2ry1* KO mice (*Snap25-T2A-GCaMP6s;P2ry1^{-/-}*). **(D)** Comparisons of average frequency, amplitude, half-width, and event ratio from control and *P2ry1* KO mice. Bilateral amplitude ratio was calculated for events simultaneous across both lobes of the IC and defined as the ratio of the weak to the strong side amplitude. A ratio of 1 would indicate complete synchrony between lobes; a ratio of 0 would indicate complete asymmetry. $n = 9$ control and $n = 7$ *P2ry1* KO mice (two-tailed Student's t test with Bonferroni correction; **** $p < 5e-4$, ** $p < 0.005$, ns: not significant). **(E)** Exemplars of a single-banded event. Rectangular ROIs were placed as shown and averaged to create a 'line-scan' across the tonotopic axis. (bottom) Heat maps of activity as a function of time and distance; circles indicate detected peaks. **(F)** Activity over a five-minute time frame in the left and right IC of control and *P2ry1* KO mice. Circles indicate detected peaks. (bottom) Histograms of peak locations. **(G)** Histogram of average number of events across all control (black) and *P2ry1* KO (red) mice. (bottom) Cumulative distribution function of event locations across the tonotopic axis pooled from all animals. Events from left and right IC were combined for each experiment. **(H)** Quantification of event frequency in the medial (low frequency) and lateral (high frequency) regions of the IC. $n = 9$ control and 7 *P2ry1* KO mice (two-tailed Student's t test with Bonferroni correction; **** $p < 5e-5$, ns, not significant).

Figure 7-Figure supplement 1. Spontaneous activity in *P2ry1* KO mice originates in the cochlea.

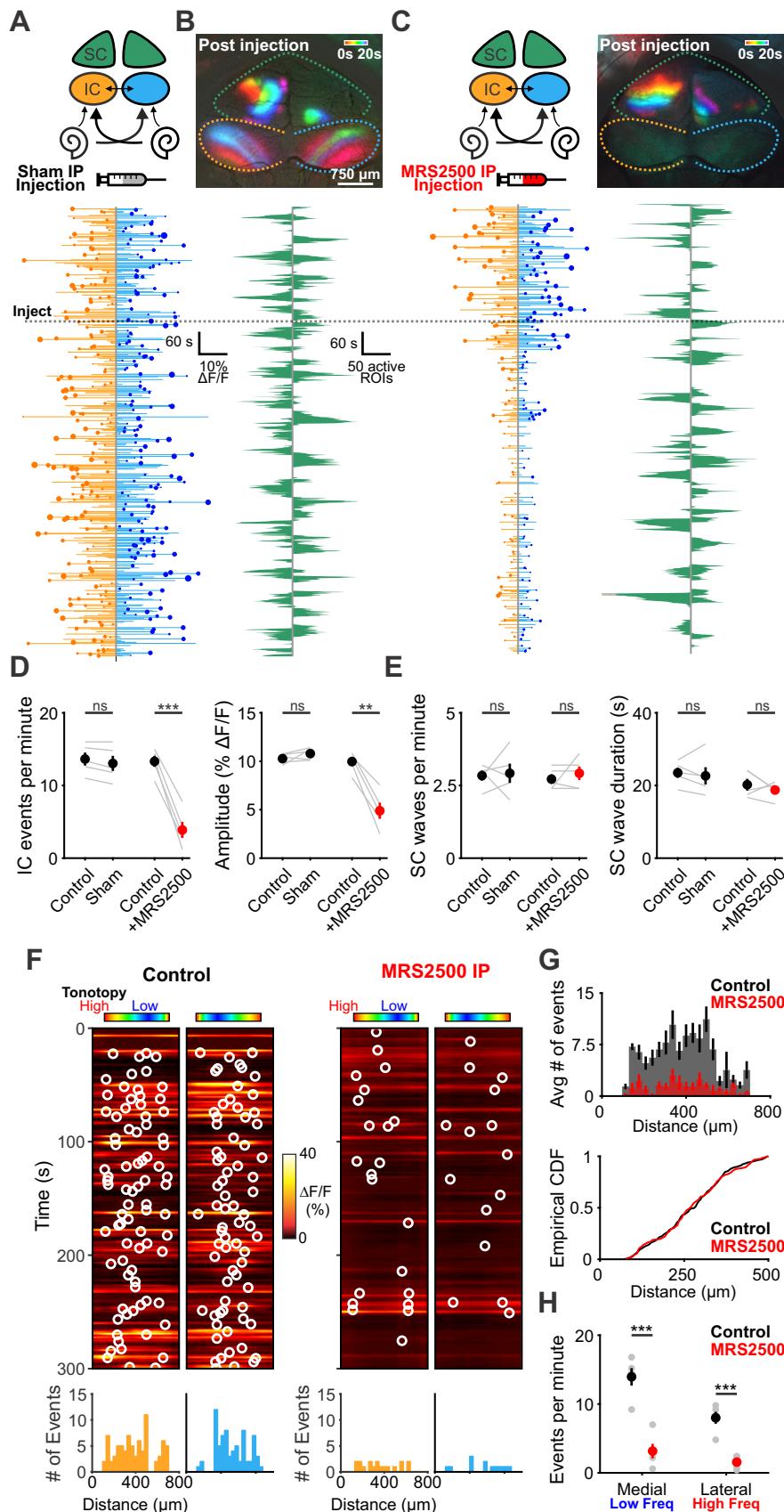


Figure 8. Delivery of MRS2500 *in vivo* dramatically reduces spontaneous activity in the developing auditory system. **(A)** Diagram illustrating flow of information to the midbrain and the visual superior colliculus. Sham solution (5% mannitol) was injected via IP catheter during imaging. (bottom) Activity over time in left and right IC in an individual. Each line indicates a detected event, circle indicates the dominant lobe, and the size of the circle indicates the difference in fluorescence. Dashed line is time of injection. Caption continued on next page.

Figure 8. **(B)** Calcium transients in the midbrain, color-coded based on time of occurrence following sham injection. (bottom) Calcium transients observed in the left and right SC. **(C)** Similar to (A) and (B), but with injection of MRS2500 (50 μ L of 500 μ M MRS2500 in 5% mannitol solution). **(D)** Plot of IC event frequency and amplitude in sham and MRS2500 injected animals. n = 5 mice for each condition (two-tailed paired Student's t test with Bonferroni correction; ***p<0.005, **p<0.005, ns: not significant). **(E)** Plot of SC wave frequency and duration in sham and MRS2500 injected animals. n = 5 mice for each condition (two-tailed paired Student's t test with Bonferroni correction; ns: not significant). **(F)** Activity along the tonotopic axis over a five-minute time frame in the left and right IC before (left) and after (right) MRS2500 injection. Circles indicate detected peaks. (bottom) Histograms of peak locations. **(G)** Histogram of average number of events before (black) and after (red) MRS2500 injection. (bottom) Cumulative distribution function of event locations across the tonotopic axis pooled from all animals. Events from left and right IC were combined for each experiment. **(H)** Quantification of event frequency in the medial (low frequency) and lateral (high frequency) regions of the IC. n = 5 mice (two-tailed Student's t test with Bonferroni correction; ***p<0.005).

296 receptors, triggering a rapid increase of intracellular Ca²⁺, gating of TMEM16A Ca²⁺-activated Cl⁻
 297 channels, and subsequent Cl⁻ and K⁺ efflux into the extracellular space (*Tritsch et al., 2007; Wang*
 298 *et al., 2015*). This transient K⁺ efflux is sufficient to depolarize nearby IHCs, resulting in a burst of
 299 Ca²⁺ action potentials, release of glutamate, and suprathreshold activation of postsynaptic SGNs
 300 via AMPA and NMDA receptors (*Tritsch et al., 2010a; Zhang-Hooks et al., 2016*). Our results show
 301 that activation of metabotropic P2RY1 autoreceptors is a key first step in this transduction pathway.
 302 P2RY1 is highly expressed by ISCs at a time when spontaneous activity is prominent in the cochlea
 303 (*Scheffer et al., 2015; Tritsch and Bergles, 2010*) (*Figure 2A*), and spontaneous activity was reduced
 304 when intracellular Ca²⁺ stores were depleted or PLC was inhibited (*Figure 1B-F*), manipulations that
 305 disrupt canonical G_q-coupled GPCR signaling pathways (*Erb and Weisman, 2012; Fabre et al., 1999*).
 306 Moreover, our pharmacological studies indicate that P2RY1 is both necessary and sufficient for
 307 spontaneous current generation in supporting cells (*Figure 3B, Figure 4B*), and inhibition of P2RY1
 308 *in vivo* profoundly decreased cochlea-generated activity in the auditory midbrain (*Figure 8C*). This
 309 reliance on P2RY1 is unexpected, as ionotropic P2X receptors are also widely expressed in the de-
 310 veloping cochlea (*Brändle et al., 1999; Lahne and Gale, 2008; Liu et al., 2015; Nikolic et al., 2003;*
 311 *Scheffer et al., 2015; Tritsch et al., 2007*). The lack of P2X engagement may reflect the partic-
 312 ular spatial-temporal characteristics of ATP release by ISCs, which may occur in locations devoid of
 313 P2X receptors or yield ATP concentration transients that favor P2RY1 activation. Exogenous ATP
 314 can induce all of the phenomenon associated with spontaneous events (ISC currents, crenation,
 315 IHC depolarization, SGN burst firing); however, it is possible that other nucleotides are released
 316 that have greater affinity for P2RY1 (e.g. ADP), or that extracellular nucleotidases rapidly convert
 317 ATP to ADP/AMP that favor activation of these native metabotropic receptors (*von Kügelgen, 2006;*
 318 *Vlajkovic et al., 1998, 2002*).

319 **Control of extracellular K⁺ dynamics by supporting cells**

320 Pharmacological inhibition of P2RY1 unexpectedly induced IHCs to gradually depolarize and begin
 321 tonic, uncorrelated firing, a phenotype also observed in *P2ry1* KO mice (*Figure 5-Figure Supple-*
 322 *ment 1C*). Our studies indicate that this phenomenon occurs because P2RY1 controls the volume
 323 of the extracellular space in the organ of Corti. Activation of P2RY1 induces ISCs to shrink osmot-
 324 ically (crenate), a consequence of ion and water efflux that is triggered by opening of TMEM16A
 325 channels (*Figure 4G*). The resulting increase in extracellular space lasts for many seconds and en-
 326 hances dissipation of extracellular K⁺, visible through the time-dependent shift in the reversal po-
 327 tential of K⁺ mediated tail currents (*Figure 4G,I*). Conversely, inhibition of P2RY1 increased the size
 328 of ISCs, a swelling-induced “relaxation” that concomitantly decreased extracellular space around
 329 IHCs. K⁺ accumulation and depolarization of IHCs followed, an effect absent when IHC K⁺ channels
 330 were inhibited (*Figure 3J,K*). This phenomenon is consistent with the depolarizing shift in the resting
 331 membrane potential of IHCs observed in *Tmem16A* cKO mice (*Wang et al., 2015*), which similarly
 332 blocks ISC crenation, and with studies in the brain where inducing cell swelling with hypoosmotic
 333 solutions or impairing K⁺ buffering results in neuronal epileptiform activity (*Larson et al., 2018;*
 334 *Murphy et al., 2017; Thrane et al., 2013*). Basal P2RY1 activation in supporting cells therefore hy-

335 perpolarizes nearby IHCs in the developing cochlea by expanding the extracellular space and low-
 336 ering local K⁺ concentrations. These changes increase the dynamic range of IHCs and allow finer
 337 control of excitability through transient ATP mediated signaling events.

338 The tonic inward current that develops in ISCs in response to P2RY1 block was abolished when
 339 homeostatic K⁺ release pathways (Na⁺ channels, Ca²⁺ channels, Na⁺-K⁺-Cl⁻cotransporters, and Na,K-
 340 ATPase) were inhibited (**Figure 3H,I**), suggesting that K⁺ redistribution mechanisms, in the absence
 341 of ISC crenation, are weak at this stage of development. Indeed, although the membrane poten-
 342 tial of ISCs is close to E_K, their membrane conductance is dominated by intercellular gap junction
 343 channels; when uncoupled from their neighbors, they exhibit very high (1–2GΩ) input resistance
 344 (**Jagger and Forge, 2014; Wang et al., 2015**), suggesting that few K⁺ leak channels are expressed.
 345 The presence of tight junctions at the apical surface of the cochlear epithelium and the limited K⁺
 346 conductance of ISCs may restrict passive diffusion and dilution of K⁺, similar to what has been de-
 347 scribed in the vestibular epithelium (**Contini et al., 2017**), thus necessitating uptake via alternative
 348 mechanisms. Both inner phalangeal and Dieters' cells (which envelop the inner and outer hair cells,
 349 respectively) express K⁺-Cl⁻ symporters, Na,K-ATPase pumps, and inwardly-rectifying K⁺ channels
 350 that may siphon K⁺ into the supporting cell syncytium after extrusion from hair cells. However,
 351 the apparently low capacity of these systems places a greater dependence on diffusion within the
 352 extracellular volume fraction controlled by the supporting cells.

353 In the CNS, astrocytes facilitate rapid dissipation of accumulated K⁺ through the glial syncytium
 354 via gap junctions (**Kofuji and Newman, 2004**), a mechanism termed spatial buffering. Astrocytes
 355 are efficient K⁺ sinks due to their highly negative resting potential (~−85 mV) and large resting K⁺
 356 conductance dominated by inward rectifying K⁺ channels and two-pore K⁺ channels (**Ryoo and
 357 Park, 2016; Olsen, 2012**). While uptake of K⁺ through these conductances is passive, recent studies
 358 suggest that K⁺ buffering in astrocytes is also actively regulated by purinergic receptors. Following
 359 stimulation of native astrocyte purinergic receptors or foreign G_q-coupled receptors (MrgA1) and
 360 release of Ca²⁺ from intracellular stores, Na,K-ATPase activity increased, resulting in a transient
 361 decrease in extracellular K⁺, hyperpolarization of nearby neurons, and reduction in their sponta-
 362 neous activity (**Wang et al., 2012**). Although P2RY1 is expressed by some astrocytes and can trigger
 363 Ca²⁺ waves (**Gallagher and Salter, 2003**), this mechanism does not appear to regulate IHC excitabil-
 364 ity in the cochlea, as stimulation of P2RY1 in *Tmem16a* cKO mice, which have intact metabotropic
 365 receptor signaling but no crenations (**Wang et al., 2015**), did not hyperpolarize IHCs (**Figure 4I,J**).
 366 Thus, astrocytes and cochlear ISCs use purinergic signaling in different ways to maintain the ionic
 367 stability of the extracellular environment and control the excitability of nearby cells.

368 **Role of supporting cells in the generation of spontaneous activity**

369 Our understanding of how non-sensory cells contribute to spontaneous activity has been limited by
 370 a lack of *in vivo* mechanistic studies. Recent advances in visualizing cochlea-induced spontaneous
 371 activity in central auditory centers *in vivo* using genetically-encoded calcium indicators (**Babola
 372 et al., 2018**) allowed us to assess whether supporting cell purinergic receptors are involved in gen-
 373 erating this activity. Prior to hearing onset, the blood-labyrinth barrier within the inner ear is not
 374 fully formed (**Suzuki et al., 1998**), permitting pharmacological access to the cochlea at this age. In-
 375 fusion of a P2RY1 inhibitor into the intraperitoneal space dramatically decreased activity within
 376 the inferior colliculus within minutes, while retina-induced activity in the superior colliculus (**Ack-
 377 man et al., 2012**) was unaffected (**Figure 8**), suggesting that inhibition is not due to activation of
 378 astrocyte P2RY1 receptors; as noted above, inhibition of P2RY1 in astrocytes would be expected to
 379 enhance, rather than inhibit neuronal activity (**Wang et al., 2012**).

380 *In vivo* imaging in *P2ry1* KO mice recapitulated many aspects of changes seen when P2RY1 was
 381 acutely inhibited, with significantly reduced neuronal activity observed in lateral regions of the IC
 382 (later active to 8–16kHz tones; **Figure 7**). However, neuronal burst firing persisted within central
 383 regions of the IC, regions that will ultimately process lower frequency sounds (3–8kHz). Developing
 384 sensory systems exhibit a remarkable ability to preserve spontaneous activity. In the visual system,

385 cholinergic antagonists injected directly into the eye blocks retinal waves *in vivo* (*Ackman et al., 2012*),
 386 but genetic removal of the $\beta 2$ acetylcholine receptor subunit alters, but does not abolish,
 387 peripherally-generated activity (*Zhang et al., 2012*). In the auditory system, *in vivo* spontaneous
 388 activity can be blocked by acute inhibition of cochlear AMPARs, but deaf mice that lack the ability
 389 to excite SGNs (*Vglut3 KO* mice) exhibit activity patterns remarkably similar to control mice (*Babola*
 390 *et al., 2018*). These robust homeostatic mechanisms allow spontaneous activity to persist despite
 391 disruption of key transduction components. Local purinergic signaling within the cochlea may still
 392 initiate tonotopic activity in central auditory circuits of *P2ry1 KO* mice, as events in the IC exhibited
 393 spatial and temporal characteristics similar to controls. IHCs and SGNs are more depolarized in
 394 these mice, reducing the threshold for activation by other purinergic receptors. Although such
 395 gain-of-function changes in the developing nervous system present challenges for interpretation
 396 of genetic manipulations, preservation of early, patterned activity in children that carry deafness
 397 mutations may improve the outcome of later therapeutic interventions to restore hearing.

398 Purinergic receptors in the adult cochlea

399 In the adult inner ear, members of all purinergic receptors subtypes (ionotropic P2X receptors,
 400 metabotropic P2Y, and adenosine P1 receptors) are expressed by cells throughout the sensory ep-
 401 ithelium, Reissner's membrane, stria vascularis, and SGNs (*Housley et al., 2009; Huang et al., 2010*).
 402 The widespread expression of these receptors coupled with observations of increased endolym-
 403 phatic ATP concentrations following trauma (*Muñoz et al., 1995a*) have led to the hypothesis that
 404 these receptors serve a neuroprotective role. Indeed, infusion of ATP into the inner ear profoundly
 405 reduces sound-evoked compound action potentials in the auditory nerve (*Bobbin and Thompson,*
 406 *1978; Muñoz et al., 1995b*), due to decreased endolymphatic potential following shunting inhibi-
 407 tion through P2XR2 (*Housley et al., 2013*). Consistent with these observations, *P2rx2 KO* mice and
 408 humans with a P2RX2 variant (c.178G > T) experience progressive sensorineural hearing loss (*Yan*
 409 *et al., 2013*). Ca²⁺ imaging and recordings from adult cochleae have also revealed robust responses
 410 to UTP in the inner sulcus, pillar cells, and Dieters' cells (*Sirko et al., 2019; Zhu and Zhao, 2010*),
 411 suggesting that metabotropic purinergic receptors continue to be expressed. Following traumatic
 412 noise damage, ATP release could activate K⁺ buffering mechanisms in supporting cells, enhance
 413 K⁺ redistribution, reduce IHC depolarization and prevent excitotoxic damage. Purinergic receptors
 414 may also contribute to IHC gain control by influencing their membrane potential, as ATP circulates
 415 in the endolymph at low nanomolar concentrations (*Muñoz et al., 1995a*). Further studies involving
 416 conditional deletion of *P2ry1* from ISCs in the adult cochlea may help to define the role of this
 417 receptor in both normal hearing and injury contexts.

418 Methods and Materials

419 Both male and female mice and rats of postnatal days P6–P15 were used for all experiments and
 420 randomly allocated to experimental groups. Transgenic breeders were crossed to female FVB mice
 421 to improve litter survival. Mice were housed on a 12 hour light/dark cycle and were provided food
 422 ad libitum. This study was performed in accordance with the recommendations provided in the
 423 Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All experi-
 424 ments and procedures were approved by the Johns Hopkins Institutional Care and Use Committee
 425 (protocol M018M350). All surgery was performed under isoflurane anesthesia and every effort was
 426 made to minimize suffering.

427 Electrophysiology

428 For inner supporting cell recordings, apical segments of the cochlea were acutely isolated from
 429 P6-P8 rat (*Figure 1*) and mouse pups (all other figures) and used within 2 hours of the dissection.
 430 Cochleae were moved into a recording chamber and continuously superfused with bicarbonate-
 431 buffered artificial cerebrospinal fluid (1.5–2mL/min) consisting of the following (in mM): 119 NaCl,
 432 2.5 KCl, 1.3 MgCl₂, 1.3 CaCl₂, 1 NaH₂PO₄, 26.2 NaHCO₃, 11 D-glucose and saturated with 95% O₂ / 5%

433 CO₂ to maintain a pH of 7.4. Solutions were superfused at either room temperature or near phys-
 434 iological temperature (32–34°C) using a feedback-controlled in-line heater (Warner Instruments),
 435 as indicated in figure legends. Whole-cell recordings of inner supporting cells (ISCs) were made
 436 under visual control using differential interference contrast microscopy (DIC). Electrodes had tip
 437 resistances between 3.5–4.5MΩ when filled with internal consisting of (in mM): 134 KCH₃SO₃, 20
 438 HEPES, 10 EGTA, 1 MgCl₂, 0.2 Na-GTP, pH 7.3. Spontaneous currents were recorded with ISCs held
 439 at -80mV.

440 For inner hair cell recordings, apical segments of the cochlea were acutely isolated from P6-
 441 P8 mouse pups and used within 2 hours of the dissection. Cochleae were moved into a recording
 442 chamber and continuously superfused with bicarbonate-buffered artificial cerebrospinal fluid (1.5–
 443 2mL/min) consisting of the following (in mM): 115 NaCl, 6 KCl, 1.3 MgCl₂, 1.3 CaCl₂, 1 NaH₂PO₄,
 444 26.2 NaHCO₃, 11 D-glucose. Solutions were saturated with 95% O₂ / 5% CO₂ to maintain a pH of
 445 7.4. Solutions were superfused at room temperature. Electrodes had tip resistances between 4.5–
 446 6.0MΩ when filled with internal consisting of (in mM): 134 KCH₃SO₃, 20 HEPES, 10 EGTA, 1 MgCl₂,
 447 0.2 Na-GTP, pH 7.3. For hair cell recordings with K⁺ channels inhibited with cesium and TEA, the
 448 internal solution consisted of (in mM): 100 cesium methanesulfonate, 20 TEA-Cl, 10 EGTA in CsOH,
 449 20 HEPES, 1 MgCl₂, 0.2 Na-GTP, pH 7.3 with CsOH. Spontaneous currents were recorded with IHCs
 450 held at near their resting membrane potential (-75 to -80mV).

451 Errors due to the voltage drop across the series resistance and the liquid junction potential
 452 were left uncompensated for recordings of spontaneous activity. For IHC recordings with K⁺ accu-
 453 mulation voltage protocols (Figure 4), the amplifier compensation circuit was used to compensate
 454 70% of the access resistance. Recordings that displayed more than a 10% increase in access re-
 455 sistance or access resistances > 30 MΩ were discarded. ISC and IHC spontaneous currents were
 456 recorded with pClamp 10 software using a Multiclamp 700B amplifier, low pass filtered at 2kHz,
 457 and digitized at 5kHz with a Digidata 1322A analog-to-digital converter (Axon Instruments).

458 Action potentials were analyzed offline using custom routines written in Matlab 2017b (Math-
 459 works). Briefly, raw traces were high-pass filtered to remove baseline drift and spikes were identi-
 460 fied using an amplitude threshold criterion. As described previously (*Tritsch et al., 2010a*), bursts
 461 were identified by classifying interspike intervals into non-bursting intervals (> 1s), burst intervals
 462 (30ms–1s), and mini-burst intervals (< 30ms). Bursts were defined as clusters of at least 10 con-
 463 secutive burst intervals (with mini-burst intervals being ignored in the context of burst detection).
 464 Spikes within mini-bursts were included when calculating the number of spikes within a burst. Col-
 465 ored raster plots were generated by grouping spikes into one-second bins and applying a color
 466 map to the resulting data (modified 'hot' colormap; Matlab).

467 **Cochlear explant culture**

468 Cochleae were dissected from postnatal day 5-6 control (*P2ry1*^{+/+} or *Pax2-Cre;R26-*lsl*-GCaMP3*) and
 469 *P2ry1* KO (*P2ry1*^{-/-} or *Pax2-Cre;R26-*lsl*-GCaMP3;P2ry1*^{-/-}) mice in ice-cold, sterile HEPES-buffered arti-
 470 ficial cerebrospinal fluid (aCSF) consisting of the following (in mM): 130 NaCl, 2.5 KCl, 10 HEPES,
 471 1 NaH₂PO₄, 1.3 MgCl₂, 2.5 CaCl₂, and 11 D-Glucose. Explants were mounted onto Cell-Tak (Corn-
 472 ing) treated coverslips and incubated at 37°C for 24 hours in Dulbecco's modified Eagle's medium
 473 (F-12/DMEM; Invitrogen) supplemented with 1% fetal bovine serum (FBS) and 10U/mL penicillin
 474 (Sigma) prior to recording or imaging.

475 **Transmitted light imaging**

476 Cochlear segments were imaged with a Olympus 40x water immersion objective (LUMPlanFl/IR)
 477 and recorded using MATLAB and a USB capture card (EZ Cap). Difference movies were generated
 478 by subtracting frames at time t_n and t_{n+5} seconds using ImageJ software to generate an index of
 479 transmittance change over time. To quantify transmittance changes, a threshold of three standard
 480 deviations above the mean was applied to the values. To calculate the frequency of these events,
 481 the whole field was taken as an ROI and peaks were detected using MATLAB (findpeaks function).

482 To calculate area of these events, a Gaussian filter ($\sigma= 2.0$) was applied to the image after thresh-
 483 olding and the borders detected using MATLAB (bwlabel function). The area was then calculated as
 484 the number of pixels within the border multiplied by the area scaling factor ($\mu\text{m}/\text{pixel}$)² measured
 485 with a stage micrometer.

486 **Immunohistochemistry and X-gal Reaction**

487 Mice were deeply anesthetized with isoflurane and perfused with freshly prepared paraformalde-
 488 hyde (4%) in 0.1 M phosphate buffer. Cochleae were post-fixed for 45 minutes at room tempera-
 489 ture and stored at 4°C until processing. For X-gal reactions, P6-P8 cochleae were removed from
 490 the temporal bone and washed 3 x 5 minutes with PBS. Tissue was then incubated for 24 hours
 491 in the dark at 37°C in X-gal working solution consisting of (in mM): 5 K⁺ ferricyanide crystalline, 5
 492 K⁺ ferricyanide trihydrate, 2 magnesium chloride, and 0.1% X-gal (GoldBio) dissolved in DMSO. Af-
 493 ter washing 3 x 5 minutes with PBS, images of cochleae were acquired on a dissecting microscope
 494 (Zeiss Stemi 305). For immunohistochemistry, fixed tissue was washed 3 x 5 minutes in PBS, placed
 495 in 30% sucrose solution overnight, and incubated in OCT mounting medium overnight at 4°C. Ten
 496 micron thick cross-sections of the cochlea were made on a cryostat and mounted on Superfrost
 497 Plus slides (Fisher), which were then allowed to dry for 1 hour before processing. Cross-sections
 498 were incubated overnight with primary antibodies against β-gal (anti-Chicken; 1:4000, Aves) and
 499 Myosin-VIIa (anti-Rabbit; 1:500, Proteus BioSciences) for detection of β-gal and Myosin-VIIa only for
 500 qualitative analysis of the *Tecta-Cre;TdT* reporter mouseline (**Figure 4–Figure Supplement 1**). Sec-
 501 tions were then rinsed three times with PBS and incubated for two hours at room temperature with
 502 secondary antibodies raised in donkey (Alexa-488 and Alexa-546; 1:2000, Life Technologies). Slides
 503 were washed three times in PBS (second wash with PBS + 1:10,000 DAPI), allowed to dry, and sealed
 504 using Aqua Polymount (Polysciences, Inc.). Images were captured using a laser scanning confocal
 505 microscope (LSM 510 or 880, Zeiss).

506 **Confocal imaging of explants**

507 After one day *in vitro*, cochleae were moved into a recording chamber and continuously superfused
 508 with bicarbonate-buffered artificial cerebrospinal fluid (1.5 - 2mL/min) consisting of the following
 509 (in mM): 119 NaCl, 2.5 KCl, 1.3 MgCl₂, 1.3 CaCl₂, 1 NaH₂PO₄, 26.2 NaHCO₃, 11 D-glucose, and sat-
 510 urated with 95% O₂ / 5% CO₂ to maintain a pH of 7.4. A piezo-mounted objective was used to
 511 rapidly alternate between SGN cell bodies and ISCs/IHCs. Images were captured at 1 frame per
 512 second using a Zeiss laser scanning confocal microscope (LSM 710, Zeiss) through a 20X objective
 513 (Plan APOCHROMAT 20x/1.0 NA) at 512 x 512 pixel (354 x 354μm; 16-bit depth) resolution. Sec-
 514 tions were illuminated with a 488nm laser (maximum 25mW power). MRS2500 (1μM, Tocris) was
 515 applied by addition to the superfusing ACSF.

516 **Analysis of *in vitro* Ca²⁺ transients**

517 Images were imported into ImageJ and image registration (MultiStackReg) was used to correct for
 518 drifts in the imaging field. Since images were obtained at two different z-planes, images were
 519 combined into one stack for analysis. This was done by eliminating the empty bottom half of the
 520 imaging field containing ISCs and IHCs and the empty top half of the field containing SGN cell bodies
 521 and merging the two images. For analysis of coordinated activity throughout the cochlea, regions
 522 of interest were drawn around the entirety of ISCs, IHCs, and SGNs. Fluorescence changes were
 523 normalized as $\Delta F/F_0$ values, where $\Delta F = F - F_0$ and F_0 was defined as the fifth percentile value for
 524 each pixel. Peaks in the signals were detected in MATLAB using the built-in peak detection function
 525 (findpeaks) with a fixed value threshold criterion (mean + 3 standard deviations for each cell).

526 To quantify frequency and areas of Ca²⁺ transients, a threshold of three standard deviations
 527 above the mean was applied to each pixel within the ROI. To calculate the frequency of these events,
 528 the whole field was taken as an ROI and peaks were detected using MATLAB (findpeaks function) on
 529 the number of thresholded pixels per frame. To calculate area of these events, a Gaussian filter ($\sigma=$

530 2.0) was applied to the image after thresholding and the borders detected using MATLAB (bwlabel
 531 function). The area was then calculated as the number of pixels within the border multiplied by an
 532 area scaling factor ($1\mu\text{m}/\text{pixel}$)² measured with a stage micrometer.

533 For correlation analysis, ROIs were drawn around every IHCs in the field of view. Pairwise cor-
 534 relation coefficients were performed between every hair cell pair and represented as correlation
 535 matrices.

536 **Installation of cranial windows**

537 Inhalation anesthesia was induced with vaporized isoflurane (4% for 5 minutes, or until mice are
 538 non-responsive to toe-pinch) and surgical plane maintained during the procedure (with 1-2% isoflu-
 539 rane) with a stable respiration rate of 80 breaths per minute. A midline incision beginning posterior
 540 to the ears and ending just anterior to the eyes was made. Two subsequent cuts were made to
 541 remove the dorsal surface of the scalp. A headbar was secured to the head using super glue (Krazy
 542 Glue). Fascia and neck muscles overlying the interparietal bone were resected and the area bathed
 543 in sterile, HEPES-buffered artificial cerebrospinal fluid that was replaced as necessary throughout
 544 the surgery. Using a 28G needle and microblade, the sutures circumscribing the interparietal bone
 545 were cut and removed to expose the midbrain. The dura mater was removed using fine scissors
 546 and forceps, exposing the colliculi and extensive vasculature. A 5 mm coverslip (CS-5R; Warner In-
 547 struments) was then placed over the craniotomy, the surrounding bone was dried using a Kimwipe,
 548 and super glue was placed along the outer edges of the coverslip for adhesion to the skull. Replace-
 549 ment 0.9% NaCl solution was injected IP and a local injection of lidocaine was given to the back of
 550 the neck. Animals were weaned off isoflurane, placed under a warming lamp, and allowed to re-
 551 cover for a minimum of 1 hour prior to imaging. Spontaneous activity was not seen in deeply anes-
 552 thetized animals and emerged ~30 minutes after recovery from isoflurane exposure, as reported
 553 previously (Ackman et al., 2012).

554 ***In vivo* calcium imaging**

555 After 1 hour of post-surgical recovery from anesthesia, pups were moved into a swaddling 15 mL
 556 conical centrifuge tube. The top half of this tube was removed to allow access to the headbar and
 557 visualization of the midbrain. Pups were head-fixed and maintained at 37°C. using a heating pad
 558 and temperature controller (TC-1000; CWE). During the experiments, pups were generally immo-
 559 bile; however, occasional limb and tail twitching did occur.

560 For wide field epifluorescence imaging, images were captured at 10 Hz using a Hamamatsu
 561 ORCA-Flash4.0 LT digital CMOS camera attached to a Zeiss Axio Zoom.V16 stereo zoom microscope.
 562 For midbrain imaging, a 4 x 4mm field of view was illuminated continuously with a mercury lamp
 563 (Zeiss Illuminator HXP 200C) and visualized through a 1X PlanNeoFluar Z 1.0x objective at 17x zoom.
 564 Images were captured at a resolution of 512 x 512 pixels (16-bit pixel depth) after 2 x 2 binning to
 565 increase sensitivity. Each recording consisted of uninterrupted acquisition over 10 minutes or 20
 566 minutes if injected with pharmacological agents.

567 **Catheterization of animals for *in vivo* imaging**

568 After induction of anesthesia and before installing the cranial window, a catheter was placed in
 569 the intraperitoneal (IP) space of neonatal mouse pups. A 24G needle was used to puncture the
 570 peritoneum and a small-diameter catheter (SAI Infusion Technologies, MIT-01) was placed. A drop
 571 of Vetbond secured the catheter to the pup's belly. Installation of cranial window proceeded as
 572 described above. Imaging sessions consisted of 5 minutes of baseline activity measurements, fol-
 573 lowed by a slow push of either 50µL of sham (5% mannitol solution) or MRS2500 solution (500µM
 574 in 5% mannitol solution). Imaging was continuous throughout and 20 minutes of activity total were
 575 collected. No discernable diminishment of activity was observed in sham animals.

576 **Image processing**

577 For wide field imaging, raw images were imported into the ImageJ environment and corrected for
 578 photobleaching by fitting a single exponential to the fluorescence decay and subtracting this com-
 579 ponent from the signal (Bleach Correct function, exponential fit). Images were then imported into
 580 MATLAB (Mathworks) and intensities were normalized as $\Delta F/F_0$ values, where $\Delta F = F - F_0$ and F_0 was
 581 defined as the fifth percentile value for each pixel. Ovoid regions of interest (ROIs) encompassing
 582 the entire left and right inferior colliculi were drawn. Across all conditions, the size of the ROIs was
 583 invariant, however, due to small differences in the imaging field between animals, the ROIs were
 584 placed manually for each imaging session. Peaks in the signals were detected in MATLAB using
 585 the built-in peak detection function (findpeaks) using a fixed value threshold criterion; because
 586 fluorescence values were normalized, this threshold was fixed across conditions (2% $\Delta F/F_0$). Occa-
 587 sionally, large events in the cortex or superior colliculus would result in detectable fluorescence
 588 increases in the IC. These events broadly activated the entire surface of the IC and did not exhibit
 589 the same spatially-confined characteristics as events driven by the periphery. These events were
 590 not included in the analysis.

591 **Analysis of spatial distribution of activity in the IC**

592 As shown in *Figure 7D*, a rectangle of size 125 x 50 pixels was placed perpendicular to the tonotopic
 593 axis of the IC ($\pm 55^\circ$ rotation, respectively). The columns of the resulting matrix were averaged
 594 together to create a line scan (125 pixels x 1 pixel) for the entire time series. Peaks were detected
 595 using MATLAB's imregionalmax function with a constant threshold of 3% $\Delta F/F_0$ across all animals.
 596 Histograms of events along the tonotopic axis were generated by summing the number of events in
 597 25 μm bins. Lateral and medial designations were assigned by splitting the area evenly between the
 598 lateral edge and the location of defined single-band events in the medial portion of the IC. Events
 599 detected on the medial edge of single-band events, reflective of the bifurcation of this information,
 600 was not included in the medial/lateral analysis.

601 **Analysis of retinal wave activity in the superior colliculus**

602 ROIs (200 x 150 pixels) were placed over each lobe of the superior colliculus and downsampled by
 603 a factor of five. Signals were normalized as $\Delta F/F_0$ values, where $\Delta F = F - F_0$ and F_0 was defined as the
 604 fifth percentile value for each pixel. In order to eliminate periodic whole-sample increases in fluo-
 605 rescence, the mean intensity of all pixels was subtracted from each individual pixel. Following this,
 606 pixels were considered active if they exceeded the mean + 3 standard deviations. For each point in
 607 time, the number of active pixels was summed. Retinal waves were defined as prolonged periods
 608 (> 1 second), where more than 5 pixels were active simultaneously. Retinal wave durations were
 609 defined as the total continuous amount of time that more than 5 pixels were active. Frequencies
 610 and durations are similar to earlier reports (*Ackman et al., 2012*).

611 **Generation of the Tecta-Cre mouseline**

612 A crRNA (TAATGATGAATAATTCCATCC) targeted near exon 2 of the Tecta gene, tracrRNA, Cas9 re-
 613 combinase, and a donor plasmid containing an iCre-WPRE-polyA sequence (500 base pair homol-
 614 ogy arms) were injected into single-cell embryos that were then transferred to pseudopregnant
 615 recipient mothers. After birth, mouse pups were screened for insertion of the gene at the correct
 616 locus with two pairs of primers: one pair amplified DNA beginning 5' of the 5' homology arm and
 617 ending within the Cre sequence and the other amplified DNA within the polyA sequence and end-
 618 ing 3' of the 3' homology arm. These primers were then used to sequence the junctions. Of these,
 619 all mice used for experiments were derived from a single founder that was positive for both sets
 620 of primers and had 100% sequence validation. Mice were crossed to a TdTomato reporter line to
 621 examine cell-specific recombination (*Figure 4–Figure Supplement 1*).

622 Quantification and statistical analysis

623 All statistics were performed in the MATLAB (Mathworks) programming environment. All statistical
 624 details, including the exact value of n, what n represents, and which statistical test was performed,
 625 can be found in the figure legends. To achieve statistical power of 0.8 with of a 30% effect size with
 626 means and standard deviations similar to those observed in previous studies, power calculations
 627 indicated that 7 animals in each condition were necessary ($\mu_1 = 10$, $\mu_2 = 7$, $\sigma = 2$, sampling ratio = 1).
 628 While this number was used as a guide, power calculations were not explicitly performed before
 629 each experiment; many experiments had much larger effect sizes and sample sizes were adjusted
 630 accordingly. For transparency, all individual measurements are included in the figures. Unless
 631 otherwise noted, data are presented as mean \pm standard error of the mean. All datasets were
 632 tested for Gaussian normality using the D'Agostino's K² test. For single comparisons, significance
 633 was defined as p <= 0.05. When multiple comparisons were made, the Bonferroni correction was
 634 used to adjust p-values accordingly to lower the probability of type I errors. For multiple condition
 635 datasets, one-way ANOVAs were used, followed by Tukey's multiple comparison tests.

636 Acknowledgments

637 We thank Dr. M. Pucak and N. Ye for technical assistance, T. Shelly for machining expertise, and
 638 members of the Bergles laboratory for discussions and comments on the manuscript. TAB was
 639 supported by grants from the NIH (DC016497) and by departmental training grants (NS091018,
 640 DC000023). HCW and CJK were supported by a departmental training grant (DC000023). Funding
 641 was also provided the NIH (DC008860, NS050274), Otonomy Inc., the Brain Science Institute at
 642 Johns Hopkins University, and the Rubenstein Fund for Hearing Research to DB.

643 Author contributions

644 Travis A. Babola, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation,
 645 Methodology, Software, Validation, Visualization, Writing-original draft, Writing-review and
 646 editing; Calvin J. Kersbergen, Data curation, Formal analysis, Investigation, Methodology, Validation,
 647 Writing-review and editing; Han Chin Wang, Methodology, Data curation, Formal analysis, In-
 648 vestigation, Methodology, Writing-review and editing; Dwight E. Bergles, Conceptualization, Data
 649 curation, Funding acquisition, Methodology, Project administration, Supervision, Writing-review
 650 and editing

651 Declaration of Interests

652 The authors declare no competing financial interests.

653 References

- 654 **Ackman JB**, Burbridge TJ, Crair MC. Retinal waves coordinate patterned activity throughout the developing visual system. *Nature*. 2012; 490(7419):219–25. doi: 10.1038/nature11529.
- 655 **Antón-Bolaños N**, Sempere-Ferrández A, Guillamón-Vivancos T, Martíni FJ, Pérez-Saiz L, Gezelius H, Filipchuk A, Valdeolmillos M, López-Bendito G. Prenatal activity from thalamic neurons governs the emergence of functional cortical maps in mice. *Science*. 2019; 364(6444):987–990. doi: 10.1126/science.aav7617.
- 656 **Babola TA**, Li S, Gribizis A, Lee BJ, Issa JB, Wang HC, Crair MC, Bergles DE. Homeostatic control of spontaneous activity in the developing auditory system. *Neuron*. 2018; 99(3):511–524. doi: 10.1016/j.neuron.2018.07.004.
- 657 **Blankenship AG**, Feller MB. Mechanisms underlying spontaneous patterned activity in developing neural circuits. *Nature Reviews Neuroscience*. 2010; 11(1):18–29. doi: 10.1038/nrn2759.
- 658 **Bobbin RP**, Thompson MH. Effects of putative transmitters on afferent cochlear transmission. *Annals of Otolaryngology, Rhinology & Laryngology*. 1978; 87(2):185–190. doi: 10.1177/000348947808700207.
- 659 **Brändle U**, Zenner HP, Ruppertsberg JP. Gene expression of P2X-receptors in the developing inner ear of the rat. *Neuroscience Letters*. 1999; 273(2):105–108. doi: 10.1016/S0304-3940(99)00648-5.

- 667 **Burbridge TJ**, Xu HP, Ackman JB, Ge X, Zhang Y, Ye MJ, Zhou ZJ, Xu J, Contractor A, Crair MC. Visual circuit development requires patterned activity mediated by retinal acetylcholine receptors. *Neuron*. 2014; 84(5):1049–1064. doi: [10.1016/j.neuron.2014.10.051](https://doi.org/10.1016/j.neuron.2014.10.051).
- 670 **Clause A**, Kim G, Sonntag M, Weisz CJC, Vetter DE, Rübsamen R, Kandler K. The precise temporal pattern of prehearing spontaneous activity is necessary for tonotopic map refinement. *Neuron*. 2014; 82(4):822–35. doi: [10.1016/j.neuron.2014.04.001](https://doi.org/10.1016/j.neuron.2014.04.001).
- 673 **Contini D**, Price SD, Art JJ. Accumulation of K⁺ in the synaptic cleft modulates activity by influencing both vestibular hair cell and calyx afferent in the turtle. *The Journal of Physiology*. 2017; 595(3):777–803. doi: [10.1113/JP273060](https://doi.org/10.1113/JP273060).
- 676 **Dhande OS**, Hua EW, Guh E, Yeh J, Bhatt S, Zhang Y, Ruthazer ES, Feller MB, Crair MC. Development of single retinofugal axon arbors in normal and β 2 knock-out mice. *Journal of Neuroscience*. 2011; 31(9):3384–3399. doi: [10.1523/JNEUROSCI.4899-10.2011](https://doi.org/10.1523/JNEUROSCI.4899-10.2011).
- 679 **Dickson BJ**. Molecular mechanisms of sxon guidance. *Science*. 2002; 298(5600):1959–1964. doi: [10.1126/science.1072165](https://doi.org/10.1126/science.1072165).
- 681 **Erb L**, Weisman GA. Coupling of P2Y receptors to G proteins and other signaling pathways. *Wiley interdisciplinary reviews Membrane transport and signaling*. 2012; 1(6):789–803. doi: [10.1002/wmts.62](https://doi.org/10.1002/wmts.62).
- 683 **Fabre JE**, Nguyen M, Latour A, Keifer JA, Audoly LP, Coffman TM, Koller BH. Decreased platelet aggregation, increased bleeding time and resistance to thromboembolism in P2Y1-deficient mice. *Nature Medicine*. 1999; 5(10):1199–1202. doi: [10.1038/13522](https://doi.org/10.1038/13522).
- 686 **Gallagher CJ**, Salter MW. Differential properties of astrocyte calcium waves mediated by P2Y1 and P2Y2 receptors. *Journal of Neuroscience*. 2003; 23(17):6728–6739.
- 688 **Glowatzki E**, Fuchs PA. Cholinergic synaptic inhibition of inner hair cells in the neonatal mammalian cochlea. *Science*. 2000; 288(5475):2366–2368. doi: [10.1126/science.288.5475.2366](https://doi.org/10.1126/science.288.5475.2366).
- 690 **Housley GD**, Bringmann A, Reichenbach A. Purinergic signaling in special senses. *Trends in Neurosciences*. 2009; 32(3):128–141.
- 692 **Housley GD**, Morton-Jones R, Vlajkovic SM, Telang RS, Paramanathanasivam V, Tadros SF, Wong ACY, Froud KE, Cederholm JME, Sivakumaran Y, Snguanwongchai P, Khakh BS, Cockayne DA, Thorne PR, Ryan AF. ATP-gated ion channels mediate adaptation to elevated sound levels. *Proceedings of the National Academy of Sciences*. 2013; 110(18):7494–9. doi: [10.1073/pnas.1222295110](https://doi.org/10.1073/pnas.1222295110).
- 696 **Huang LC**, Thorne PR, Vlajkovic SM, Housley GD. Differential expression of P2Y receptors in the rat cochlea during development. *Purinergic Signalling*. 2010; 6(2):231–48. doi: [10.1007/s11302-010-9191-x](https://doi.org/10.1007/s11302-010-9191-x).
- 698 **Jagger DJ**, Forge A. Connexins and gap junctions in the inner ear – it's not just about K⁺ recycling. *Cell and Tissue Research*. 2014; p. 633–644. doi: [10.1007/s00441-014-2029-z](https://doi.org/10.1007/s00441-014-2029-z).
- 700 **Johnson SL**, Eckrich T, Kuhn S, Zampini V, Franz C, Ranatunga KM, Roberts TP, Masetto S, Knipper M, Kros CJ, Marcotti W. Position-dependent patterning of spontaneous action potentials in immature cochlear inner hair cells. *Nature Neuroscience*. 2011; 14(6):711–717. doi: [10.1038/nn.2803](https://doi.org/10.1038/nn.2803).
- 703 **Johnson SL**, Wedemeyer C, Vetter DE, Adachi R, Holley MC, Elgoyhen AB, Marcotti W. Cholinergic efferent synaptic transmission regulates the maturation of auditory hair cell ribbon synapses. *Open Biology*. 2013; 3(NOV):130163. doi: [10.1098/rsob.130163](https://doi.org/10.1098/rsob.130163).
- 706 **Kirkby LA**, Sack GS, Firl A, Feller MB. A role for correlated spontaneous activity in the assembly of neural circuits. *Neuron*. 2013; 80(5):1129–44. doi: [10.1016/j.neuron.2013.10.030](https://doi.org/10.1016/j.neuron.2013.10.030).
- 708 **Kofuji P**, Newman EA. Potassium buffering in the central nervous system; 2004. doi: [10.1016/j.neuroscience.2004.06.008](https://doi.org/10.1016/j.neuroscience.2004.06.008).
- 710 **Kros CJ**, Ruppertsberg JP, Rüscher A. Expression of a potassium current in inner hair cells during development of hearing in mice. *Nature*. 1998; 394(6690):281–284.
- 712 **von Kugelgen I**. Pharmacological profiles of cloned mammalian P2Y-receptor subtypes. *Pharmacology & Therapeutics*. 2006; 110(3):415–432. doi: <https://doi.org/10.1016/j.pharmthera.2005.08.014>.

- 714 **Lahne M**, Gale JE. Damage-induced activation of ERK1/2 in cochlear supporting cells is a hair cell death-
715 promoting signal that depends on extracellular ATP and calcium. *Journal of Neuroscience*. 2008; 28(19):4918–
716 28. doi: [10.1523/JNEUROSCI.4914-07.2008](https://doi.org/10.1523/JNEUROSCI.4914-07.2008).
- 717 **Larson VA**, Mironova Y, Vanderpool KG, Waisman A, Rash JE, Agarwal A, Bergles DE. Oligodendrocytes control
718 potassium accumulation in white matter and seizure susceptibility. *Elife*. 2018; 7:e34829.
- 719 **Lim R**, Kindig AE, Donne SW, Callister RJ, Brichta AM. Potassium accumulation between type I hair cells and calyx
720 terminals in mouse crista. *Experimental Brain Research*. 2011 may; 210(3):607–621. doi: [10.1007/s00221-011-2592-4](https://doi.org/10.1007/s00221-011-2592-4).
- 722 **Liu C**, Glowatzki E, Fuchs PA. Unmyelinated type II afferent neurons report cochlear damage. *Proceedings of
723 the National Academy of Sciences*. 2015; 112(47):14723–14727. doi: [10.1073/pnas.1515228112](https://doi.org/10.1073/pnas.1515228112).
- 724 **Marcotti W**, Johnson SL, Holley MC, Kros CJ. Developmental changes in the expression of potassium currents
725 of embryonic, neonatal and mature mouse inner hair cells. *The Journal of Physiology*. 2003; 548(2):383–400.
- 726 **Moody WJ**, Bosma MM. Ion channel development, spontaneous activity, and activity-dependent development
727 in nerve and muscle cells. *Physiological Reviews*. 2005; 85(3):883–941. <https://doi.org/10.1152/physrev.00017.2004>, doi: [10.1152/physrev.00017.2004](https://doi.org/10.1152/physrev.00017.2004).
- 729 **Muñoz DJ**, Thorne PR, Housley GD, Billett TE. Adenosine 5'-triphosphate (ATP) concentrations in the endolymph
730 and perilymph of the guinea-pig cochlea. *Hearing Research*. 1995; 90(1-2):119–125.
- 731 **Muñoz DJB**, Thorne PR, Housley GD, Billett TE, Battersby JM. Extracellular adenosine 5'-triphosphate (ATP) in
732 the endolymphatic compartment influences cochlear function. *Hearing Research*. 1995; 90(1-2):106–118.
733 doi: [10.1016/0378-5955\(95\)00152-3](https://doi.org/10.1016/0378-5955(95)00152-3).
- 734 **Murphy TR**, Binder DK, Fiacco TA. Turning down the volume: astrocyte volume change in the generation and
735 termination of epileptic seizures. *Neurobiology of Disease*. 2017; 104:24–32.
- 736 **Nikolic P**, Housley GD, Thorne PR. Expression of the P2X7 receptor subunit of the adenosine 5'-triphosphate-
737 gated ion channel in the developing and adult rat cochlea. *Audiology and Neurotology*. 2003; 8(1):28–37. doi:
738 [10.1159/000067891](https://doi.org/10.1159/000067891).
- 739 **Olsen M**. Examining potassium channel function in astrocytes. *Methods in Molecular Biology*. 2012; 814:265–
740 281.
- 741 **Ryoo K**, Park JY. Two-pore domain potassium channels in astrocytes. *Experimental Neurobiology*. 2016;
742 25(5):222–232. doi: [10.5607/en.2016.25.5.222](https://doi.org/10.5607/en.2016.25.5.222).
- 743 **Scheffer DI**, Shen J, Corey DP, Chen ZY. Gene expression by mouse inner ear hair cells during development.
744 *Journal of Neuroscience*. 2015; 35(16):6366–6380. doi: [10.1523/JNEUROSCI.5126-14.2015](https://doi.org/10.1523/JNEUROSCI.5126-14.2015).
- 745 **Shrestha BR**, Chia C, Wu L, Kujawa SG, Liberman MC, Goodrich LV. Sensory neuron diversity in the inner ear
746 is shaped by activity. *Cell*. 2018; 174(5):1229–1246.e17. doi: [10.1016/j.cell.2018.07.007](https://doi.org/10.1016/j.cell.2018.07.007).
- 747 **Sirko P**, Gale JE, Ashmore JF. Intercellular Ca²⁺ signalling in the adult mouse cochlea. *Journal of Physiology*.
748 2019; 597(1):303–317. doi: [10.1113/JP276400](https://doi.org/10.1113/JP276400).
- 749 **Sonntag M**, Englitz B, Kopp-Scheinpflug C, Rübsamen R. Early postnatal development of spontaneous and
750 acoustically evoked discharge activity of principal cells of the medial nucleus of the trapezoid body: an in vivo
751 study in mice. *Journal of Neuroscience*. 2009; 29(30):9510–9520. doi: [10.1523/JNEUROSCI.1377-09.2009](https://doi.org/10.1523/JNEUROSCI.1377-09.2009).
- 752 **Sretavan DW**, Shatz CJ. Prenatal development of cat retinogeniculate axon arbors in the absence of binocular
753 interactions. *Journal of Neuroscience*. 1986; 6(4):990–1003. doi: [10.1523/JNEUROSCI.06-04-00990.1986](https://doi.org/10.1523/JNEUROSCI.06-04-00990.1986).
- 754 **Stoeckli ET**. Understanding axon guidance: are we nearly there yet? *Development*. 2018; 145(10):dev151415.
755 doi: [10.1242/dev.151415](https://doi.org/10.1242/dev.151415).
- 756 **Sun S**, Babola T, Pregering G, So KS, Nguyen M, Su SM, Palermo AT, Bergles DE, Burns JC, Müller U. Hair cell
757 mechanotransduction regulates spontaneous activity and spiral ganglion subtype specification in the auditory system.
758 *Cell*. 2018; 174(5):1–17. doi: [10.1016/j.cell.2018.07.008](https://doi.org/10.1016/j.cell.2018.07.008).
- 759 **Suzuki M**, Yamasoba T, Kaga K. Development of the blood-labyrinth barrier in the rat. *Hearing Research*. 1998;
760 116(1):107–112. doi: [https://doi.org/10.1016/S0378-5955\(97\)00208-6](https://doi.org/10.1016/S0378-5955(97)00208-6).

- 761 **Thrane VR**, Thrane AS, Wang F, Cotrina ML, Smith NA, Chen M, Xu Q, Kang N, Fujita T, Nagelhus EA. Ammonia
762 triggers neuronal disinhibition and seizures by impairing astrocyte potassium buffering. *Nature Medicine*.
763 2013; 19(12):1643.
- 764 **Tritsch NX**, Bergles DE. Developmental regulation of spontaneous activity in the Mammalian cochlea. *Journal
765 of Neuroscience*. 2010; 30(4):1539–50. doi: [10.1523/JNEUROSCI.3875-09.2010](https://doi.org/10.1523/JNEUROSCI.3875-09.2010).
- 766 **Tritsch NX**, Rodríguez-Contreras A, Crins TH, Wang HC, Borst JGG, Bergles DE. Calcium action potentials in hair
767 cells pattern auditory neuron activity before hearing onset. *Nature Neuroscience*. 2010; 13(9):1050–2. doi:
768 [10.1038/nn.2604](https://doi.org/10.1038/nn.2604).
- 769 **Tritsch NX**, Yi E, Gale JEJ, Glowatzki E, Bergles DE. The origin of spontaneous activity in the developing auditory
770 system. *Nature*. 2007; 450(7166):50–5. doi: [10.1038/nature06233](https://doi.org/10.1038/nature06233).
- 771 **Tritsch NX**, Zhang YX, Ellis-Davies G, Bergles DE. ATP-induced morphological changes in supporting cells of the
772 developing cochlea. Purinergic signalling. 2010; 6(2):155–66. doi: [10.1007/s11302-010-9189-4](https://doi.org/10.1007/s11302-010-9189-4).
- 773 **Vlajkovic SM**, Thorne PR, Housley GD, Muñoz DJB, Kendrick IS. The pharmacology and kinetics of ecto-
774 nucleotidases in the perilymphatic compartment of the guinea-pig cochlea. *Hearing Research*. 1998;
775 117(1):71–80. doi: [https://doi.org/10.1016/S0378-5955\(98\)00004-5](https://doi.org/10.1016/S0378-5955(98)00004-5).
- 776 **Vlajkovic SM**, Thorne PR, Sévigny J, Robson SC, Housley GD. NTPDase1 and NTPDase2 Immunolocalization in
777 Mouse Cochlea: Implications for Regulation of P2 Receptor Signaling. *Journal of Histochemistry & Cytochemistry*.
778 2002; 50(11):1435–1441. doi: [10.1177/002215540205001102](https://doi.org/10.1177/002215540205001102).
- 779 **Wang F**, Smith NA, Xu Q, Fujita T, Baba A, Matsuda T, Takano T, Bekar L, Nedergaard M. Astrocytes modulate
780 neural network activity by Ca²⁺-dependent uptake of extracellular K⁺. *Science Signaling*. 2012; 5(218):ra26.
781 doi: [10.1126/scisignal.2002334](https://doi.org/10.1126/scisignal.2002334).
- 782 **Wang HC**, Bergles DE. Spontaneous activity in the developing auditory system. *Cell and Tissue Research*. 2014;
783 361(1):66–75. doi: [10.1007/s00441-014-2007-5](https://doi.org/10.1007/s00441-014-2007-5).
- 784 **Wang HC**, Lin CC, Cheung R, Zhang-Hooks Y, Agarwal A, Ellis-Davies G, Rock J, Bergles DE. Spontaneous activity of
785 cochlear hair cells triggered by fluid secretion mechanism in adjacent support cells. *Cell*. 2015; 163(6):1348–
786 1359. doi: [10.1016/j.cell.2015.10.070](https://doi.org/10.1016/j.cell.2015.10.070).
- 787 **Xu HP**, Furman M, Mineur YS, Chen H, King SL, Zenisek D, Zhou ZJ, Butts DA, Tian N, Picciotto MR, Crair MC. An
788 instructive role for patterned spontaneous retinal activity in mouse visual map development. *Neuron*. 2011;
789 70(6):1115–1127. doi: [10.1016/j.neuron.2011.04.028](https://doi.org/10.1016/j.neuron.2011.04.028).
- 790 **Yan D**, Zhu Y, Walsh T, Xie D, Yuan H, Sirmaci A, Fujikawa T, Wong ACY, Loh TL, Du L, Grati M, Vlajkovic SM,
791 Blanton S, Ryan AF, Chen ZY, Thorne PR, Kachar B, Tekin M, Zhao HB, Housley GD, et al. Mutation of the ATP-
792 gated P2X2 receptor leads to progressive hearing loss and increased susceptibility to noise. *Proceedings of
793 the National Academy of Sciences*. 2013; 110(6):2228–2233. doi: [10.1073/pnas.1222285110](https://doi.org/10.1073/pnas.1222285110).
- 794 **Zhang J**, Ackman JB, Xu HP, Crair MC. Visual map development depends on the temporal pattern of binocular
795 activity in mice. *Nature neuroscience*. 2012; 15(2):298.
- 796 **Zhang-Hooks YX**, Agarwal A, Mishina M, Bergles DE. NMDA receptors enhance spontaneous activity
797 and promote neuronal survival in the developing cochlea. *Neuron*. 2016; 89(2):337–350. doi:
798 [10.1016/j.neuron.2015.12.016](https://doi.org/10.1016/j.neuron.2015.12.016).
- 799 **Zhu Y**, Zhao HB. ATP-mediated potassium recycling in the cochlear supporting cells. Purinergic Signalling. 2010;
800 6(2):221–229. doi: [10.1007/s11302-010-9184-9](https://doi.org/10.1007/s11302-010-9184-9).

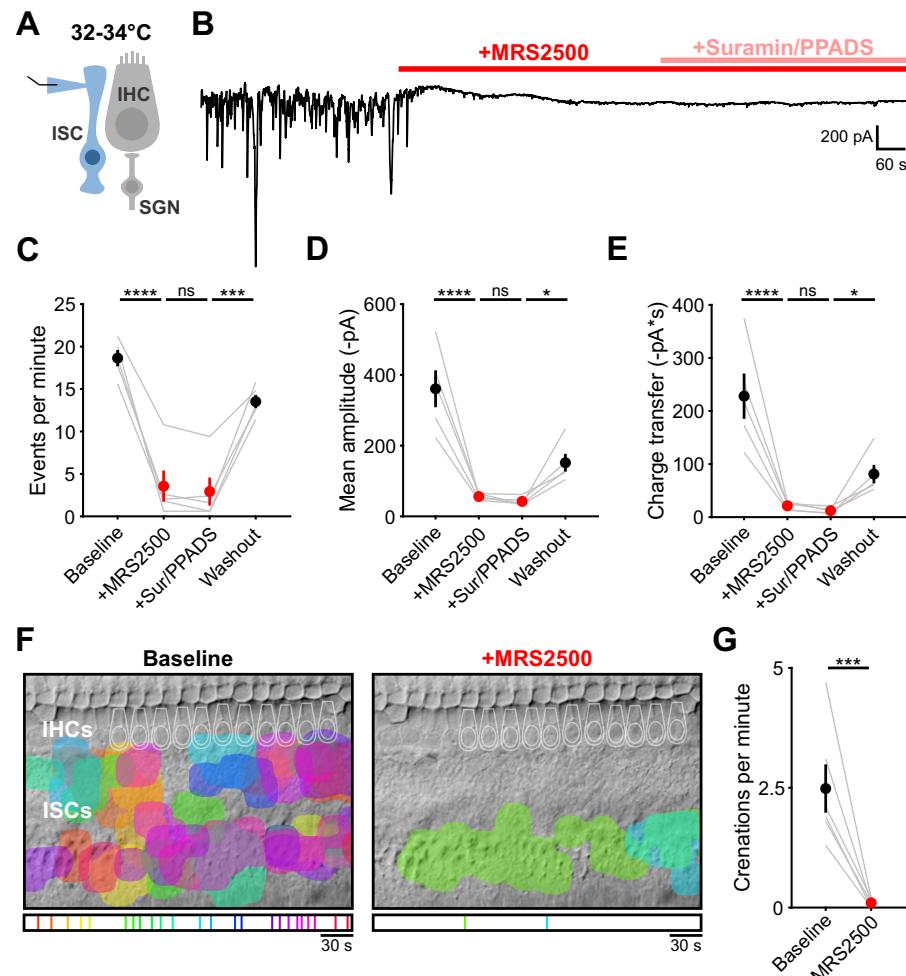


Figure 3-Figure supplement 1. P2RY1 inhibition abolishes spontaneous inward currents near physiological temperature. **(A)** Schematic of whole-cell recording configuration from ISCs. **(B)** Spontaneous inward currents recorded from an ISC before and after application of MRS2500 (1 μ M) and subsequent broad spectrum purinergic antagonists suramin (10 μ M) and PPADS (50 μ M). Recordings were performed near physiological temperature (32–34°C). **(C)** Plot of event frequency. Each window measured was 5 minutes in length, washout was taken 20 minutes after superfusion of aCSF. n = 5 ISCs from 5 cochleae (one-way ANOVA; ****p<5e-5, ***p<0.0005, ns, not significant). **(D)** Plot of event amplitude. n = 5 ISCs from 5 cochleae (one-way ANOVA; ****p<5e-5, *p<0.05, ns, not significant). **(E)** Plot of average integral (charge transfer). n = 5 ISCs from 5 cochleae (one-way ANOVA; ****p<5e-5, *p<0.05, ns, not significant). **(F)** Intrinsic optical imaging performed before and after application of the P2Y1 antagonist, MRS2500 (1 μ M). Detected crenations are outlined in colors based on time of occurrence as indicated by timeline below image. Imaging was performed near physiological temperature (32–34°C). **(G)** Plot of crenation frequency before and after MRS2500 application. n = 6 cochleae (two-tailed paired Student's t test; **p<0.005).

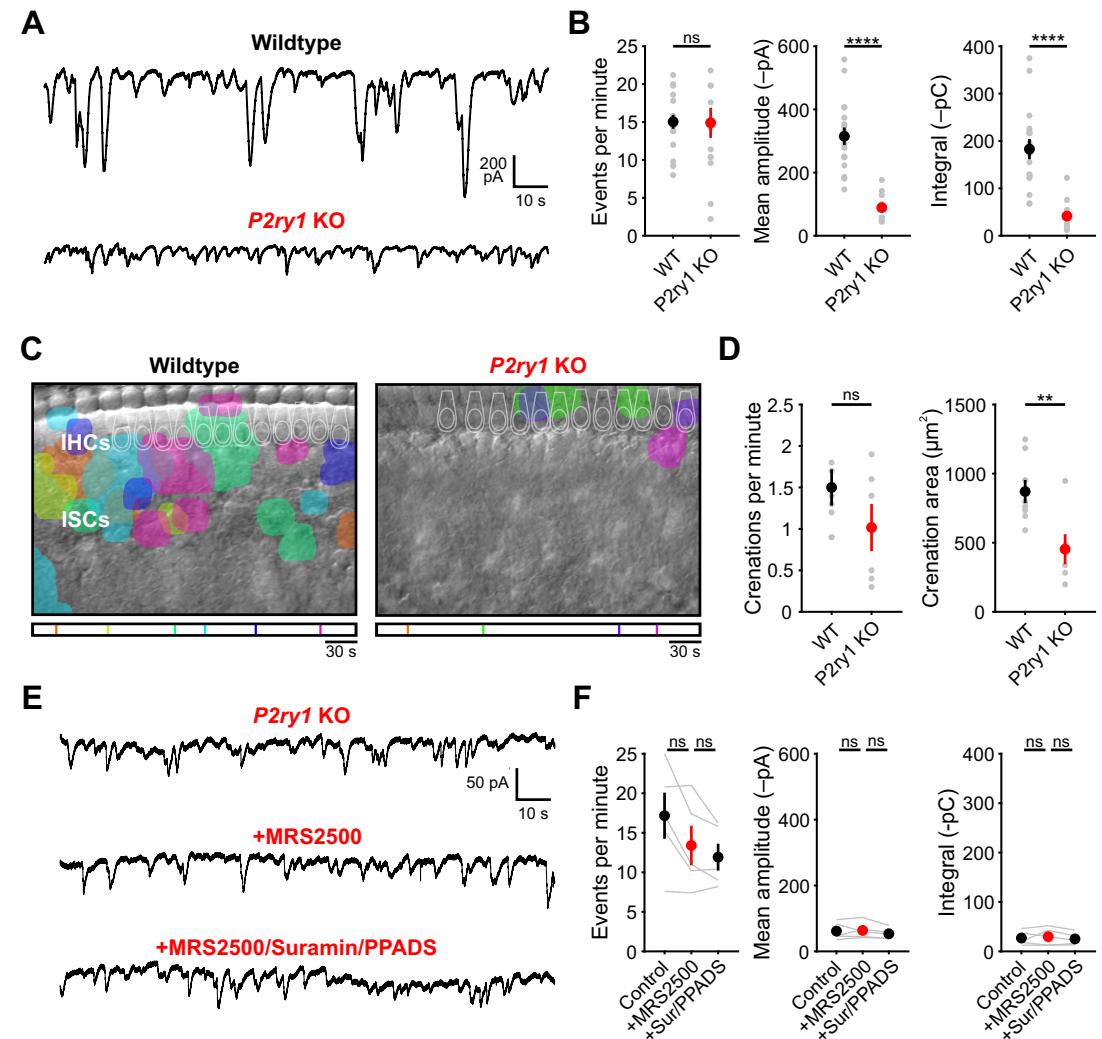


Figure 3-Figure supplement 2. Spontaneous inward currents and crenations are dramatically reduced in *P2ry1* KO mice. **(A)** Spontaneous inward currents recorded from ISCs in wildtype and *P2ry1* KO mice. Recordings were performed near physiological temperature (32–34°C). **(B)** Plots of event frequency, amplitude, and integral (charge transfer). n = 17 wildtype and 14 *P2ry1* KO ISCs (two-tailed Student's t-test with Bonferroni correction; ****p<0.0005, ns, not significant). **(C)** Intrinsic optical imaging performed in wildtype and *P2ry1* KO mice. Detected crenations are outlined in colors based on time of occurrence as indicated by the timeline below image. Imaging was performed at room temperature. **(D)** Plots of crenation frequency and area in wildtype and *P2ry1* KO mice. n = 8 wildtype cochleae and 6 *P2ry1* KO cochleae (two-tailed paired Student's t test with Bonferroni correction; **p<0.005, ns, not significant). **(E)** Spontaneous inward currents recorded from an inner supporting cell in *P2ry1* KO mice before and during application of MRS2500 (1μM) and subsequent broad spectrum purinergic antagonists suramin (100μM) and PPADS (50μM). **(F)** Plots of event frequency, amplitude, and charge transfer. n = 5 *P2ry1* KO ISCs (one-way ANOVA; ns, not significant).

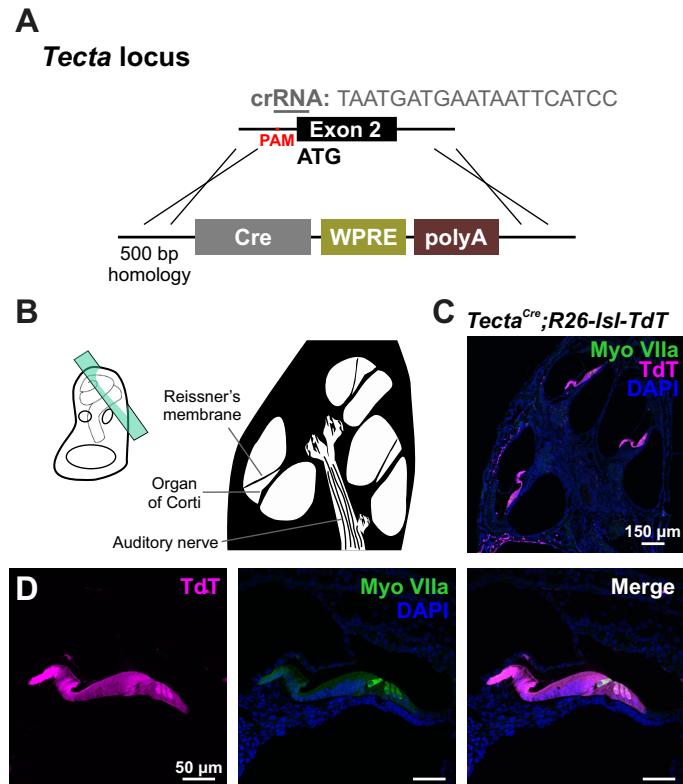


Figure 4–Figure supplement 1. Crispr-Cas9 mediated generation of the *Tecta*-Cre mouse line. **(A)** Targeting strategy for introducing an *iCre* coding sequence into the endogenous *Tecta* locus. Note: start ATG is located in exon 2. **(B)** Schematic of temporal bone with sectioning orientation indicated with green plane. **(C)** TdT reporter expression observed along the entire length of a P7 cochlea. Expression was absent in stria vascularis and very sparse in apical SGNs. **(D)** TdT reporter expression observed in nearly all cells within the sensory epithelium, including hair cells (MyoVIIa, green).

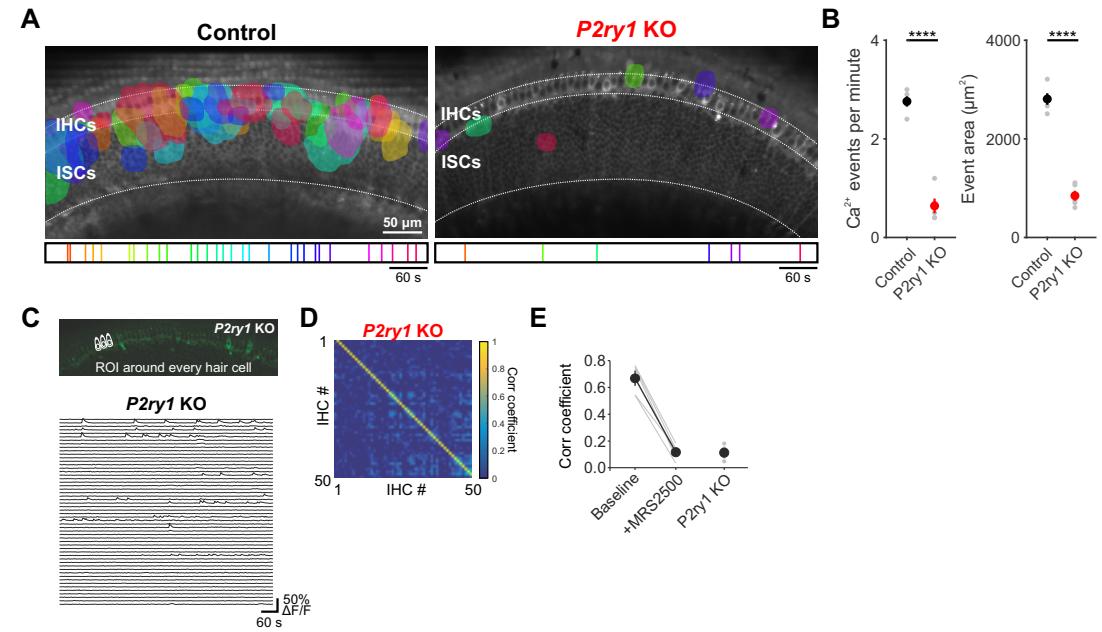


Figure 5-Figure supplement 1. *P2ry1* KO mice exhibit reduced Ca^{2+} transients in ISCs. **(A)** Maps showing maximum area of spontaneous Ca^{2+} transients in control (*Pax2-Cre;R26-*Isl-GCaMP3**) and *P2ry1* KO (*Pax2-Cre;R26-*Isl-GCaMP3*; P2ry1^{-/-}*) mice. Ca^{2+} transients in the ISC and IHC regions are color-coded based on time of occurrence as indicated in timeline below image. Imaging was performed at room temperature. **(B)** Plots of Ca^{2+} event frequency and area in control and *P2ry1* KO mice. $n = 5$ control and 5 *P2ry1* KO mice (two-tailed Student's t test with Bonferroni correction, **** $p < 0.05$). **(C)** Exemplar images of IHC Ca^{2+} transients. ROIs were drawn around every IHC for subsequent analysis (bottom). **(D)** Correlation matrices generated by calculating the linear correlation coefficient for all IHC pairs in *P2ry1* KO mice. **(E)** Plot of average correlation coefficient among the four nearest neighboring hair cells. Data from MRS2500 experiment (Figure 4) is reproduced here for comparison. $n = 5$ cochleae (two-tailed paired Student's t test with Bonferroni correction; *** $p < 0.0005$).

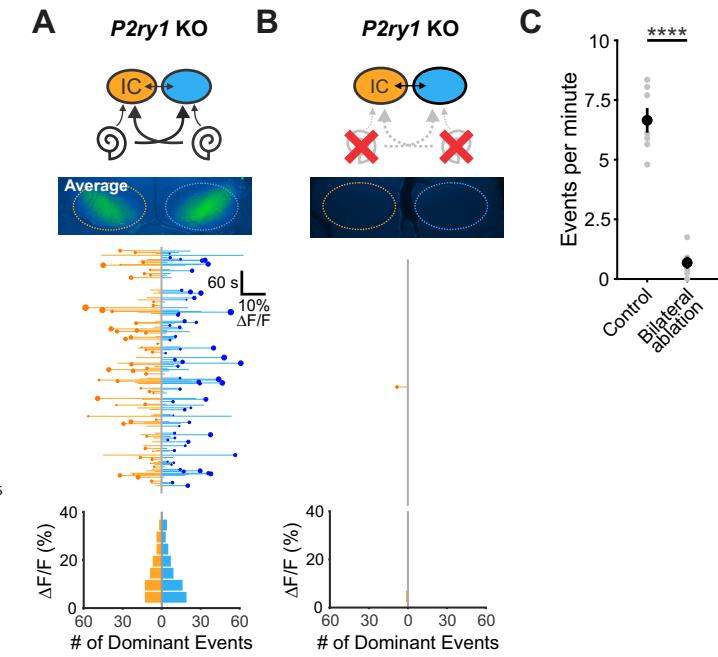


Figure 7-Figure supplement 1. Spontaneous activity in *P2ry1* KO mice originates in the cochlea. **(A)** Top: Diagram illustrating flow of information through the auditory system and average intensity image over the 10 min imaging session. Middle: Activity over time in left and right IC in an individual where each line indicates the fluorescence intensity of each detected event; the circle indicates the dominant lobe, and the size of the circle indicated the difference in fluorescence. Bottom: Histograms showing the frequency of dominant events of a given amplitude. **(B)** Similar to A, but with bilateral ablation of the cochleae. **(C)** Plot of IC event frequency in control (*P2ry1* KO) and bilaterally ablated (*P2ry1* KO) mice. $n = 7$ control and $n = 5$ bilaterally ablated *P2ry1* KO mice (two-tailed Student's t test; **** $p < 5e-5$).